Supporting Information

for

Versatile synthesis of amino acid functionalized

nucleosides via a domino carboxamidation reaction

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Experimental procedures, characterization data, and ¹H and ¹³C NMR spectra of new compounds

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Experimental Section

General Remarks: All chemicals and solvents (Sigma-Aldrich, Fluka, Acros, Novabiochem, Iris Biotech GmbH) were purchased and used without any further purification, except dichloromethane, which was distilled from CaH₂ prior to use. All reactions were performed under argon or nitrogen with magnetic stirring and were monitored by thin-layer chromatography (TLC) using SIL G-25 UV254 pre-coated silica gel plates (0.25 mm thickness). Reactions under CO atmosphere were carried out in a Parr-High-Pressure reaction vessel with particular care. The ultra sonication was performed using a sonic bath (42 kHz, Bransonic, 2510E-MT, USA) at ambient temperature. TLC plates were visualized by using anisaldehyde (5% anisaldehyde in ethanol with 1% sulfuric acid) or PMA (5% phosphomolybdic acid in ethanol) solutions. Flash column chromatography was performed using BIOSOLVE silica gel (0.063-0.200 mm particle size, 20-30 g silica/1 g compound/ KieselgelMerck, 230–400 mesh, Type 9385, 60 Angström). ¹H NMR spectra were recorded at 300 MHz, ¹³C NMR spectra were recorded at 75 MHz. Chemical shifts (δ) are reported in units of parts per million (ppm), with the residual ¹H or ¹³C peaks of the solvent used as internal standards (CDCl₃: $\delta H = 7.26$ ppm and $\delta C = 77.16$ ppm, CD₃OD: $\delta H = 3.31$ ppm and $\delta C = 49.00$ ppm). The following abbreviations are used to explain the observed multiplicities: s, singlet; d, doublet; t, triplet; q, quadruplet; m, multiplet; br, broad; band, several overlapping signals; AB, AB system with strongly skewed signals; app, indicates an "apparent" multiplicity, for which only the observed average coupling constant can be quoted, in absence of information on the real J values. Where given, assignments of resonances were confirmed by standard COSY and HSQC 2D NMR experiments. High resolution mass spectra (HRMS) were recorded with an Agilent Accurate-Mass Quadrupole Time-of-Flight mass spectrometer.

Synthesis of the histidine modified nucleoside (3): The TBDMS-protected nucleoside 1 (500 mg, 0.86 mmol) was suspended in THF (20 mL) in a Parr-High-Pressure reaction vessel and the protected histidine 2 (622 mg, 2.6 mmol), Et₃N (1.2 mL, 8.6 mmol) and Pd(PPh₃)₄ (99.4 mg, 0.09 mmol) were added in the order listed. The reaction mixture was incubated at 70 °C under 50 psi (~4 bar) carbon monoxide for 48 h. The reaction mixture was cooled down to room temperature, filtered over celite and concentrated under vacuum. The residue was taken up in anhydrous DMF (20 mL) and to the stirring solution under argon, Et₃N (350

µL, 2.6 mmol) and tert-butyl dicarbonate (282 mg, 1.3 mmol) was added. After 15 min, the reaction was quenched with anhydrous methanol (1 mL) and again concentrated to an oil. The oil was purified using flash chromatography on silica gel with 3 % MeOH in DCM to yield product **3** as a yellow oil (511 mg, 0.68 mmol, 79%). R_f (DCM: MeOH 9:1) = 0.63; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.21 (d, J = 7.5 Hz, 1H, -NH), 8.65 (s, 1H, H6), 8.33 (s br, 1H, pyrimidine-NH), 8.00 (app d, J = 1.1 Hz, 1H, H10), 7.17 (s br, 1H, H9), 6.20 (dd, J = 5.7 and 7.8 Hz, 1H, H1'), 4.98 (app dt, J = 5.8 and 7.5 Hz, 1H, H7), 4.40 (dt, J = 2.0 and 5.9 Hz, 1H, *H3*'), 4.03 (td, J = 2.0 and 3.7 Hz, 1H, *H4*'), 3.77 (m, 2H, *H5*'/*H5*''), 3.73 (s, 3H, -OCH₃), 3.16 (d, J = 5.8 Hz, 2H, H8'/H8''), 2.39 (ddd AB, J = 2.0, 5.7 and 13.3 Hz, 1H, H2'/H2''), 2.06 (ddd AB, J = 5.8, 7.8 and 13.3 Hz, 1H, H2'/H2''), 0.89 (s, 9H, -tBu), 0.87 (s, 9H, -tBu), 0.09 (s, 6H, -CH₃), 0.07 (s, 6H, -CH₃); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 171.7 (C), 162.7 (C), 161.6 (C), 149.7 (C), 147.0 (C), 146.3 (CH), 138.8 (C), 132.2 (CH), 128.5 (CH), 114.8 (CH), 106.0 (C), 88.8 (CH), 87.0 (CH), 85.5 (C), 73.1 (CH₂), 63.4 (CH₂), 52.5 (CH), 52.4 (CH₃), 41.6 (CH₂), 30.4 (CH₂), 28.0 (CH₃), 26.0 (CH₃), 25.8 (CH₃), 18.5 (C), 18.1 (C), -4.6 (CH₃), -4.7 (CH₃), -5.4 (CH₃), -5.5 (CH₃); HRMS (ESI): m/z: calcd for C₃₄H₅₆O₁₀N₅Si₂: 750.3565 [M-H]⁻; found: 750.3578.

General synthesis for the deprotection of TBDMS-protected nucleosides 3, 7 and 11 to afford the modified compounds 4, 8 and 12: The protected nucleoside (0.4–0.6 mmol) was dissolved in dry THF (3–5 mL) and $Et_3N\cdot 3HF$ with an additional amount of dry Et_3N were added. The reaction mixture was stirred overnight at room temperature under argon atmosphere. The solvent was removed under reduced pressure and the resulting crude was purified with flash chromatography on silicagel using a gradient 2–5% MeOH in DCM to yield a white foam (43–86%).

Experimental details of product (4): Reaction of product **3** (409 mg, 0.54 mmol) and Et₃N·3HF (0.53 mL, 3.3 mmol) with an additional amount of dry Et₃N (0.45 mL, 3.3 mmol) according to the general procedure described above, gave the deprotected nucleoside **4** (204 mg, 72%). R_f (DCM: MeOH 9:1) = 0.37; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.94 (d, *J* = 7.9 Hz, 1H, -N*H*), 8.83 (s, 1H, *H*6), 8.08 (app d, *J* = 1.1 Hz, 1H, H10), 7.16 (s br, 1H, H9), 6.13 (t, *J* = 5.2 Hz, 1H, *H1*'), 5.09 (app dt, *J* = 5.0 and 7.9 Hz, 1H, *H7*), 4.56 (q, *J* = 5.2 Hz, 1H, *H3*'), 4.07 (q, *J* = 5.2 Hz, 1H, *H4*'), 3.89 (m, 2H, *H5*'/*H5*''), 3.71 (s, 3H, -OCH₃), 3.29 (dd AB, *J* = 5.0 and 14.7 Hz, 1H, *H8*'/*H8*''), 3.15 (m, 1H, *H8*'/*H8*''), 2.44 (q, *J* = 5.2 Hz, 2H, *H2*'/*H2*''), 0.89 (s, 9H, *-tBu*); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 171.5 (C), 162.9 (C),

162.2 (C), 149.7 (C), 148.4 (CH), 146.9 (C), 138.2 (C), 115.0 (CH), 105.2 (C), 88.7 (CH), 88.3 (CH), 86.0 (C), 70.7 (CH), 61.9 (CH₂), 52.6 (CH/CH₃), 52.5 (CH₃), 40.4 (CH₂), 30.2 (CH₂), 28.0 (CH₃); HRMS (ESI): m/z: calcd for C₂₂H₂₈O₁₀N₅: 522.1836 [M-H]⁻; found: 522.1856.

General synthesis of the modified dimethoxytrityl analogues 5, 9 and 13 from the corresponding modified nucleosides 4, 8 and 12: The starting nucleoside (0.16–0.20 mmol) was co-evaporated three times with pyridine and dried overnight under vacuum. The modified nucleoside was dissolved in dry pyridine (1–4 mL) and dry DCM (0.5–2 mL). 4,4'- dimethoxytrityl chloride, dissolved in dry pyridine (1–2 mL) and dry DCM (0.5–1 mL), was added dropwise to this solution at 0 °C. The reaction was stirred overnight at room temperature under argon atmosphere for 7 h. The reaction was quenched with anhydrous MeOH (1–2 mL) at 0 °C and the solvent was removed under reduced pressure. The remaining residue (yellow oil) was dissolved in DCM (5 mL). The organic phase was washed with 2x sat. sol. NaHCO₃ (10 mL) and 1x with brine (10 mL). The collected organic phases were dried on Na₂SO₄ and the solvent was evaporated in vacuo. The remaining residue was purified by flash chromatography on silica gel using a gradient of 0-5 % methanol in DCM + 1% Et₃N to give the desired product (9–82%).

Experimental details of product (5): Reaction of product **4** (102 mg, 0.20 mmol) with DMTr-Cl (135 mg, 0.40 mmol) in pyridine 4 mL and DCM 2 mL according to the general procedure described above, gave the protected nucleoside **5** (84 mg, 52%). R_f (DCM: MeOH 9:1) = 0.43; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.29 (d, *J* = 7.4 Hz, 1H, -N*H*), 8.53 (s, 1H, *H*6), 8.01 (app d, *J* = 1.1 Hz, 1H, H10), 7.41-7.19 (band, 10H, -DMTr*H* + *H*9), 8.85 (d, *J* = 8.9 Hz, 4H, -DMTr*H*), 6.15 (app t, *J* = 6.4 Hz, 1H, *H1*'), 4.93 (app dt, *J* = 5.7 and 7.4 Hz, 1H, *H7*), 4.33 (dt, *J* = 5.3 and 6.4 Hz, 1H, *H3*'), 3.95 (app q, *J* = 5.3 Hz, 1H, *H4*'), 3.78 (s, 6H, -OC*H*₃), 3.69 (s, 3H, -OC*H*₃), 3.48 (dd AB, *J* = 5.3 and 10.2 Hz, 1H, *H5*'/*H5*''), 3.21 (dd AB, *J* = 5.7 and 14.9 Hz, 1H, *H8*'/*H8*''), 3.12 (dd AB, *J* = 5.7 and 14.9 Hz, 1H, *H5*'/*H5*''), 2.20 (dt AB, *J* = 6.4 and 13.7 Hz, 1H, *H2*'/*H2*''), 1.58 (s, 9H, -*tBu*); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 171.7 (C), 162.4 (C), 161.5 (C), 158.7 (C), 149.4 (C), 147.0 (C), 146.3 (CH), 144.7 (C), 138.7 (C), 135.7 (C), 130.2 (CH), 128.2 (CH), 128.1 (CH), 127.1 (CH), 114.8 (CH), 113.43 (CH), 106.1 (C), 87.0 (C), 86.2 (CH), 85.6 (CH), 72.6 (CH), 63.8 (CH₂),

55.4 (CH/CH₃), 52.5 (CH/CH₃), 46.0 (C), 40.5 (CH₂), 30.4 (CH₂), 28.0 (CH₃); HRMS (ESI): *m/z*: calcd for C₄₃H₄₈O₁₂N₅: 826.3299 [M+H]⁺; found: 826.3279.

General synthesis of 5-carboxamide modified analogues 7 and 11 from the corresponding protected TBDMS-protected compound 1: The protected nucleoside 1 (0.9–1.2 mmol) was suspended in THF (25 mL) in a Parr-High-Pressure reaction vessel and the protected amino acid 6 and 10, Et_3N and $Pd(PPh_3)_4$ were added in the order listed. The reaction mixture was incubated at 70 °C under 50 psi (~4 bar) carbon monoxide for 48 h. The reaction mixture was cooled down to room temperature, filtered over celite and concentrated under vacuum. The remaining yellow oil was then purified on a silica gel column using 0–2% methanol in DCM as eluent to give the modified analogue (68–90%).

Synthesis of the serine modified nucleoside (7): Reaction of 5-iodo-3',5'-(O-di-tertbutyldimethylsilyl)-2'-deoxyuridine (1, 528 mg, 0.91 mmol) and O-benzyl-L-serine methyl ester 6 (634 mg, 2.6 mmol) according to the general procedure described above, gave the deprotected nucleoside 7 as a white solid (430 mg, 68%). R_f (DCM: MeOH 9:1) = 0.64; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.31 (d, J = 7.7 Hz, 1H, -NH), 9.22 (s br, 1H, pyrimidine-NH), 8.61 (s, 1H, H6), 7.30-7.18 (band, 5H, -ArH), 6.17 (dd, J = 5.8 and 7.5 Hz, 1H, H1'), 4.83 (app dt, J = 3.7 and 7.7 Hz, 1H, H7), 4.54 (d AB, J = 12.4 Hz, 1H, H9'/H9''), 4.48 (d AB, J = 12.4 Hz, 1H, H9'/H9''), 4.37 (dt, J = 2.4 and 6.0 Hz, 1H, H3'), 4.00 (td, J = 2.4 and 4.1 Hz, 1H, H4'), 3.89 (dd AB, J = 3.7 and 9.7 Hz, 1H, H8'/H8''), 3.74 (d, J = 4.1 Hz, 2H, H5'/H5''), 3.70 (s, 3H, -OCH₃), 3.66 (dd AB, J = 3.7 and 9.7 Hz, 1H, H8'/H8''), 2.37 (ddd AB, J = 2.4, 5.8 and 13.5 Hz, 1H, H2'/H2''), 2.03 (ddd AB, J = 5.8, 6.0 and 13.5 Hz, 1H, H2'/H2''), 0.86 (s, 9H, -tBu), 0.82 (s, 9H, -tBu), 0.05 (s, 3H, -CH₃), 0.04 (s br, 6H, -CH₃), 0.03 (s, 3H, -CH₃); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 170.6 (C), 162.6 (C), 161.6 (C), 149.5 (C), 146.5 (CH), 137.6 (C), 128.4 (CH), 127.8 (CH), 127.6 (CH), 105.9 (C), 88.9 (CH), 87.2 (CH), 73.3 (CH₂), 73.0 (CH), 69.6 (CH₂), 53.0 (CH), 52.6 (CH₃), 52.5 (CH), 41.7 (CH₂), 26.0 (CH₃), 25.9 (CH₃), 18.5 (C), 18.1 (C), -4.6 (CH₃), -4.8 (CH₃), -5.4 (CH₃), -5.5 (CH₃); HRMS (ESI): *m/z*: calcd for C₃₃H₅₄O₉N₃Si₂: 692.3398 [M+H]⁺; found: 692.3390.

Experimental details of the deprotected product (8): Reaction of the modified nucleoside **7** (353 mg, 0.51 mmol) and Et₃N·3HF (0.50 mL, 3.1 mmol) with an additional amount of dry Et₃N (0.40 mL, 3.0 mmol) according to the general procedure described above, gave the deprotected nucleoside **8** as a white foam (204 mg, 86%). R_f (DCM: MeOH 9:1) = 0.29; ¹H

NMR (300 MHz, CD₃OD, 25°C): δ 8.84 (s, 1H, *H6*), 7.28 (m, 5H, -Ar*H*), 6.26 (app t, *J* = 6.3 Hz, 1H, *H1*'), 4.78 (app t, *J* = 3.3 Hz, 1H, *H7*), 4.54 (d, *J* = 1.7 Hz, 2H, *H9*'/*H9*''), 4.40 (dt, *J* = 3.4 and 6.3 Hz, 1H, *H3*'), 3.98 (app q, *J* = 3.4 Hz, 1H, *H4*'), 3.95 (dd, *J* = 3.3 and 9.8 Hz, 1H, *H8*'/*H8*''), 3.75 (dd, *J* = 3.3 and 9.8 Hz, 1H, *H8*'/*H8*''), 3.74 (s, 3H, -OC*H*₃), 3.83-3.70 (band, 2H, *H5*'/*H5*''), 2.40 (ddd AB, *J* = 3.4, 6.3 and 13.5 Hz, 1H, *H2*'/*H2*''), 2.25 (dt AB, *J* = 6.3 and 13.5 Hz, 1H, *H2*'/*H2*''); ¹³C NMR (75 MHz, CD₃OD, 25°C): δ 171.9 (C), 164.8 (C), 164.5 (C), 151.3 (C), 148.0 (CH), 139.1 (C), 129.4 (CH), 128.9 (CH), 128.8 (CH), 106.1 (C), 89.5 (CH), 87.7 (CH), 74.2 (CH₂), 72.3 (CH), 70.4 (CH₂), 62.8 (CH₂), 54.2 (CH), 53.0 (CH₃), 41.9 (CH₂); HRMS (ESI): *m/z*: calcd for C₂₁H₂₄O₉N₃: 462.1512 [M-H]⁻; found: 462.1518.

Experimental details of the DMTr-protected modified nucleoside (9): Reaction of product 8 (90 mg, 0.20 mmol) and DMTr-Cl (152 mg, 0.45 mmol) according to the general procedure described above, gave the deprotected nucleoside 9 as an off-white pink colored foam (122 mg, 82%). R_f (DCM: MeOH 9:1) = 0.57; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.36 (d, J = 7.8 Hz, 1H, -NH), 8.56 (s, 1H, H6), 7.41-7.16 (band, 14H, -ArH/-DMTrH), 6.83 (d, J = 8.5 Hz, 4H, -DMTrH), 6.15 (app t, J = 6.5 Hz, 1H, H1'), 4.84 (app dt, J = 3.7 and 7.8 Hz, 1H, H7), 4.56 (d AB, J = 12.2 Hz, 1H, H9'/H9''), 4.49 (d AB, J = 12.2 Hz, 1H, H9'/H9''), 4.32 (dt, J = 4.2 and 6.5 Hz, 1H, H3'), 3.95 (ddd, J = 4.2, 4.7 and 5.7 Hz, 1H, *H4*'), 3.92 (dd, J = 3.7 and 9.6 Hz, 1H, *H8*'/*H8*''), 3.78 (s, 3H, -OCH₃), 3.77 (s, 3H, -OCH₃), 3.73 (s, 3H, -OCH₃), 3.71 (dd, J = 3.7 and 9.6 Hz, 1H, H8'/H8''), 3.48 (dd AB, J = 4.7 and 10.4 Hz, 1H, H5'/H5''), 3.36 (dd AB, J = 5.7 and 10.4 Hz, 1H, H5'/H5''), 2.47 (ddd AB, J =4.2, 6.5 and 14.0 Hz, 1H, H2'/H2''), 2.22 (dt AB, J = 6.5 and 14.0 Hz, 1H, H2'/H2''); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 170.7 (C), 162.6 (C), 161.7 (C), 158.8 (C), 149.6 (C), 146.2 (CH), 144.7 (C), 137.8 (C), 135.7 (C), 130.2 (CH), 130.1 (CH), 128.5 (CH), 128.2 (CH), 128.1 (CH), 127.9 (CH), 127.8 (CH), 127.1 (CH), 113.5 (CH), 106.1 (C), 87.1 (C), 86.4 (CH), 85.8 (CH), 73.4 (CH₂), 72.7 (CH), 69.6 (CH₂), 63.8 (CH₂), 55.4 (CH₃), 53.0 (CH), 52.7 (CH₃), 40.6 (CH₂); HRMS (ESI): m/z: calcd for C₄₂H₄₂O₁₁N₃: 764.2819 [M-H]⁻; found: 764.2814.

Synthesis of the lysine modified nucleoside (11): Reaction of 5-iodo-3',5'-(O-di-*tert*-butyldimethylsilyl)-2'-deoxyuridine (1, 722 mg, 1.2 mmol) and the commercialy available protected lysine 10 (1.04 g, 3.5 mmol) according to the general procedure described above, gave the deprotected nucleoside 11 as a yellow foam (864 mg, 90%). R_f (DCM: MeOH 95:5)

= 0.33; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.86 (s br, 1H, pyrimidine-N*H*), 9.01 (d, *J* = 7.6 Hz, 1H, -N*H*), 8.59 (s, 1H, *H6*), 7.31-7.22 (band, 5H, -Ar*H*), 6.15 (dd, *J* = 5.7 and 7.3 Hz, 1H, *H1*'), 5.01 (s, 2H, *H12'/H12''*), 4.64 (dt, *J* = 5.3 and 7.6 Hz, 1H, *H7*), 4.34 (dt, *J* = 2.0 and 5.6 Hz, 1H, *H3'*), 3.98 (td, *J* = 2.0 and 3.6 Hz, 1H, *H4'*), 3.72 (d, *J* = 3.6 Hz, 2H, *H5'/H5''*), 3.67 (s, 3H, -OC*H*₃), 3.14 (app q, *J* = 6.3 Hz, 2H, *H11'/H11''*), 2.34 (ddd AB, *J* = 2.0, 5.7 and 13.2 Hz, 1H, *H2'/H2''*), 1.99 (ddd AB, *J* = 5.5, 7.3 and 13.2 Hz, 1H, *H2'/H2''*), 1.94-1.70 (band, 2H, *H8'/H8''*), 1.48 (m, 2H, *H10'/H10''*), 1.36 (m, 2H, *H9'/H9''*), 0.84 (s, 9H, *-tBu*), 0.81 (s, 9H, *-tBu*), 0.03 (s br, 9H, *-CH*₃), 0.02 (s, 3H, *-CH*₃); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 172.3 (C), 163.1 (C), 161.3 (C), 156.4 (C), 149.4 (C), 146.5 (CH), 136.6 (C), 128.4 (CH), 128.0 (CH), 105.6 (C), 88.8 (CH), 86.9 (CH), 72.8 (CH), 66.5 (CH₂), 63.1 (CH₂), 52.3 (CH₃), 52.0 (CH), 41.5 (CH₂), 40.7 (CH₂), 31.7 (CH₂), 29.3 (CH₂), 25.9 (CH₃), 25.7 (CH₃), 22.4 (CH₂), 18.3 (C), 17.9 (C), *-4.8* (CH₃), *-4.9* (CH₃), *-5.5* (CH₃), *-5.7* (CH₃); HRMS (ESI): *m/z*: calcd for C₃₇H₆₁O₁₀N₄Si₂: 777.3926 [M+H]⁺; found: 777.3936.

Experimental details of the deprotected product (12): Reaction of product **11** (320 mg, 0.41 mmol) and Et₃N·3HF (0.47 mL, 2.9 mmol) with an additional amount of dry Et₃N (0.40 mL, 3.0 mmol) according to the general procedure described above, gave the deprotected nucleoside **12** as a white foam (97 mg, 43%). R_f (DCM: MeOH 9:1) = 0.37; ¹H NMR (300 MHz, CD₃OD, 25°C): δ 8.84 (s, 1H, *H*6), 7.30 (m, 5H, -Ar*H*), 6.24 (app t, *J* = 6.4 Hz, 1H, *H1*²), 5.05 (s, 2H, *H12'/H12''*), 4.59 (dd, *J* = 5.3 and 7.7 Hz, 1H, *H7*), 4.39 (dt, *J* = 3.4 and 6.4 Hz, 1H, *H3'*), 3.99 (app q, *J* = 3.4 Hz, 1H, *H4'*), 3.81 (dd AB, *J* = 3.4 and 11.9 Hz, 1H, *H5'/H5''*), 3.73 (dd AB, *J* = 3.4 and 11.3 Hz, 1H, *H5'/H5''*), 3.73 (s, 3H, -OC*H*₃), 3.11 (app t, *J* = 6.6 Hz, 2H, *H11'/H11''*), 2.38 (ddd AB, *J* = 3.4, 6.4 and 13.5 Hz, 1H, *H2'/H2''*), 2.23 (dt AB, *J* = 6.4 and 13.5 Hz, 1H, *H2'/H2''*), 1.97-1.75 (band, 2H, *H8'/H8''*), 1.52 (m, 2H, *H10'/H10''*), 1.41 (m, 2H, *H9'/H9''*); ¹³C NMR (75 MHz, CD₃OD, 25°C): δ 173.8 (C), 164.8 (C), 164.4 (C), 158.9 (C), 151.2 (C), 148.0 (CH), 138.5 (C), 129.5 (CH), 129.0 (CH), 128.7 (CH), 106.0 (C), 89.5 (CH), 87.8 (CH), 72.3 (CH), 67.3 (CH₂), 62.8 (CH₂), 53.6 (CH), 52.8 (CH₃), 41.9 (CH₂), 41.4 (CH₂), 32.7 (CH₂), 30.4 (CH₂), 23.7 (CH₂); HRMS (ESI): *m/z*: calcd for C₂₅H₃₃O₁₀N₄: 549.2196 [M+H]⁺; found: 549.2191.

Synthesis of the DMTr-protected lysine modified nucleoside (13): Reaction of the modified nucleoside 12 (88 mg, 0.16 mmol) and DMTr-Cl (143 mg, 0.42 mmol) according to the general procedure described above, gave the deprotected nucleoside 13 as a white foam (13 mg, 9%). R_f (DCM: MeOH 9:1) = 0.38; ¹H NMR (300 MHz, CDCl₃, 25°C): δ 9.09 (d, *J* =

7.4 Hz, 1H, -N*H*), 8.58 (s, 1H, *H*6), 7.40-7.15 (band, 14H, -DMTr*H*/ -Ar*H*), 6.82 (d, 4H, J = 8.9 Hz, -DMTr*H*), 6.15 (app t, J = 6.2 Hz, 1H, *H1*'), 5.06 (s br, 2H, *H12'/H12'*'), 4.85 (app t, J = 5.3 Hz, 2H, *H11'/H11'*'), 4.67 (td, J = 5.0 and 7.4 Hz, 1H, *H7*), 4.29 (dt, J = 4.5 and 5.5 Hz, 1H, *H3*'), 4.01 (q, J = 4.5 Hz, 1H, *H4*'), 3.77 (s, 6H, -OCH₃), 3.71 (s, 3H, -OCH₃), 3.43 (dd AB, J = 4.5 and 10.2 Hz, 1H, *H5'/H5'*'), 3.36 (dd AB, J = 4.5 and 10.2 Hz, 1H, *H5'/H5''*), 3.08 (m, 2H, *H10'/H10''*), 2.50 (ddd AB, J = 4.5, 6.2 and 13.7 Hz, 1H, *H2'/H2''*), 2.19 (m, 1H, *H2'/H2''*), 1.87 (m, 1H, *H8'/H8''*), 1.76 (m, 1H, *H8'/H8''*), 1.35 (m, 2H, *H9'/H9''*); ¹³C NMR (75 MHz, CDCl₃, 25°C): δ 172.6 (C), 163.2 (C), 158.7 (C), 149.7 (C), 144.8 (C), 135.8 (C), 130.2 (CH), 128.6 (CH), 128.2 (CH), 113.4 (CH), 105.9 (C), 87.0 (C), 86.5 (CH), 86.0 (CH), 72.6 (CH), 66.7 (CH₂), 63.9 (CH₂), 55.4 (CH₃), 53.2 (CH₂), 52.5 (CH/CH₃), 40.9 (CH₂), 32.1 (CH₂), 29.5 (C), 22.7 (CH₂); HRMS (ESI): *m/z*: calcd for C₄₆H₄₉O₁₂N₄: 849.3347 [M-H]⁻; found: 849.3365.





Product 3: ¹³C NMR, 300 MHz, CDCl₃, 25 °C





























Product 9: ¹³C NMR, 300 MHz, CDCl₃, 25 °C





Product 11: ¹³C NMR, 300 MHz, CDCl₃, 25 °C



Product 12: ¹H NMR, 300 MHz, CD₃OD, 25 °C









