Supporting Information

for

Selective methylation of kaempferol via benzylation and deacetylation of kaempferol acetates

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Experimental section; NMR and ESI-HRMS spectra of 1–23; NOESY spectra of 4 and 13; HMBC spectra of 4, 5, 7, 9, 12, 13, 16, 18, 22 and 23

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Experimental section

Melting points were determined with an X-6 apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer Spectrum One spectrometer in KBr disks. NMR spectra, including NOESY and $^{1}\text{H-}^{13}\text{C}$ HMBC experiments, were carried out at Bruker AC-400 (400 MHz for ^{1}H NMR and 100 MHz for ^{13}C NMR) or Bruker AM-600 (600 MHz for ^{1}H NMR and 150 MHz for ^{13}C NMR) spectrometers in CDCl₃ or in DMSO- d_{6} using TMS as internal reference at ambient temperature; chemical shifts (δ) and coupling constants (J) are given in ppm and Hz, respectively. ESI-HRMS spectra were obtained on a Bruker Bio TOF IIIQ (quadrupole time of flight) mass spectrometer. Analytical TLC was performed on plates precoated with 0.15–0.2 mm of silica gel GF₂₅₄ from QOCEC (Qingdao Ocean Chemical Engineering Company, P. R. China). HPLC was equipped with Perkin-Elmer series 200 pump, Perkin-Elmer series UV/Vis detector, 200 µL manual injector, and Phenomenex C₁₈ column (10 mm × 250 mm, 10 µm). Reagents were analytical reagent grade and were used without further purification unless otherwise noted. CH₂Cl₂ and NMP were freshly distilled over CaH₂.

3,4',5,7-Tetra-*O*-acetylkaempferol (1) [1]

Acetic anhydride (60 mL) was added to the solution of dry kaempferol (4 g, 13.99 mmol) in pyridine (30 mL), and the mixture was allowed to stand overnight at ambient temperature. After complete conversion monitored by TLC, the solvent was removed and the residue was poured into crushed ice with vigorous stirring. The abundant resulting off-white precipitate was recovered by filtration and washed with cold water and then methanol, and dried in air. The crude product was crystallized from acetone/95% ethanol (1:3) to afford **1** (5.95 g, 94%) as colorless needles, mp: 183-184 °C. IR (cm⁻¹): 1775, 1661, 1631, 1503, 1476, 1435, 1370, 1186, 1158, 1122,

1081, 1013.¹H NMR (600 MHz, CDCl₃): δ = 7.84 (d, J = 8.7 Hz, 2H), 7.32 (d, J = 2.1 Hz, 1H), 7.26 (d, J = 8.7 Hz, 2H), 6.87 (d, J = 2.1 Hz, 1H), 2.43 (s, 3H), 2.34 (s, 6H), 2.32 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): δ = 170.24, 169.39, 168.99, 167.98, 167.91, 157.10, 154.91, 154.38, 153.08, 150.61, 134.08, 129.76, 127.16, 122.21, 114.97, 113.97, 109.10, 21.30, 21.18, 20.71. ESI-HRMS: 477.0807 [M+Na]⁺ (calcd. for C₂₃H₁₈O₁₀Na: 477.0792).

3,4',5-Tri-O-acetylkaempferol (2)

This was carried out according to the reported procedure [2]. To a solution of kaempferol tetraacetate (1) (1g, 2.2 mmol) in NMP (5 mL) was added imidazole (60 mg, 0.88 mmol) followed by thiophenol (0.31 mL, 3.08 mmol) at 0 °C. The mixture was stirred at 0 °C for 11 h and then diluted with EtOAc (100 mL), and washed successively with 1 M HCl (aq) (50 mL), water (40 mL) and brine (40 mL). The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The residue was purified over silica gel column using acetone/petroleum ether (1:2) as solvents to give **2** (825 mg, 91%) as white powder, mp: 136-138 °C. IR (cm⁻¹): 3348, 2935, 1765, 1728, 1636, 1501, 1453, 1371, 1249, 1201, 1179, 1077. ¹H NMR (400 MHz, DMSO- d_6): δ = 11.32 (s, 1H), 7.91 (d, J = 8.8 Hz, 2H), 7.37 (d, J = 8.8 Hz, 2H), 6.93 (d, J = 2.3 Hz, 1H), 6.64 (d, J = 2.3 Hz, 1H), 2.32 (s, 3H), 2.30 (s, 6H). ¹³C NMR (100 MHz, DMSO- d_6): δ = 169.00, 168.99, 168.79, 167.99, 162.78, 157.61, 153.30, 152.61, 150.22, 132.61, 129.48, 126.64, 122.63, 109.26, 109.04, 100.96, 20.91, 20.90, 20.32. ESI-HRMS: 435.0695 [M+Na]⁺ (calcd. for C₂₁H₁₆O₉Na: 435.0687).

7-O-Methyl-3,4',5-tri-O-acetylkaempferol (3) [3]

A suspension of compound **2** (200 mg, 0.49 mmol), anhydrous K₂CO₃ (108 mg, 0.78 mmol) and Me₂SO₄ (0.061 mL, 0.64 mmol) in dry acetone (6 mL) was stirred at room

temperature for 12 h. The reaction mixture was diluted with water (10 mL) and extracted with EtOAc (20 mL). The organic layer was washed with brine, dried over MgSO₄ and concentrated under reduced pressure. The dried residue was purified over silica gel column using acetone/petroleum ether (1:4) as solvents to provide **3** (180 mg, 87%) as white powder, mp: 186-187 °C. IR (cm⁻¹): 2960, 2902, 1763, 1658, 1600, 1494, 1368, 1290, 1202, 1155, 1014. ¹H NMR (400 MHz, CDCl₃): δ = 7.84 (d, J = 8.8 Hz, 2H), 7.25 (d, J = 8.8 Hz, 2H), 6.84 (d, J = 2.4 Hz, 1H), 6.64 (d, J = 2.4 Hz, 1H), 3.91 (s, 3H), 2.43 (s, 3H), 2.34 (s, 3H), 2.32 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): δ = 170.27, 169.72, 169.06, 168.15, 163.98, 158.36, 154.30, 152.88, 150.92, 133.87, 129.68, 127.48, 122.15, 111.27, 108.85, 99.03, 56.22, 21.31, 21.27, 20.77. ESI-HRMS: 449.0850 [M+Na]⁺ (calcd. for C₂₂H₁₈O₉Na: 449.0843).

7-O-Methylkaempferol (rhamnocitrin) (4) [4]

The solution of compound **3** (150 mg, 0.35 mmol) in 7.0 M methaniloc ammonia (2 mL) was stirred at room temperature for 3 h. The solution was diluted with MeOH (4 mL) and neutralized cautiously with 0.5 M HCl (aq) and then left at room temperature for 12 h. The abundant resulting precipitate was recovered by filtration, washed with water and dried over P_2O_5 under vacuum to give **4** (102 mg, 97%) as yellow powder, mp: 172-173 °C. IR (cm⁻¹): 3481, 3274, 2926, 2854, 1609, 1587, 1503, 1415, 1355, 1231, 1164. H NMR (400 MHz, DMSO- d_6): δ = 12.47 (s, 1H, OH-5), 10.14 (s, 1H, OH-4'), 9.51 (s, 1H, OH-3), 8.08 (d, J = 8.8 Hz, 2H, H-2',6'), 6.93 (d, J = 8.8 Hz, 2H, H-3',5'), 6.75 (d, J = 2.0 Hz, 1H, H-8), 6.35 (d, J = 2.0 Hz, 1H, H-6), 3.86 (s, 3H, OCH₃-7). ¹³C NMR (150 MHz, DMSO- d_6): δ = 176.04 (C-4), 164.91 (C-7), 160.38 (C-5), 159.32 (C-4'), 156.11 (C-9), 147.28 (C-2), 135.96 (C-3), 129.58 (C-2',6'), 121.58 (C-1'), 115.46 (C-3',5'), 104.04 (C-10), 97.46 (C-6), 92.03 (C-8), 56.02 (OCH₃-7). ESI-HRMS: 299.0568 [M-H]⁻ (calcd. for C₁₆H₁₁O₆: 299.0561).

4',7,-Di-*O*-methylkaempferol (5) [5]

A mixture of compound 2 (166 mg, 0.4 mmol), dimethyl sulfate (0.12 mL, 1.25 mmol), anhydrous K₂CO₃ (199 mg, 1.44 mmol) and acetone (6 mL) was refluxed for 2 h. Then methanol (2 mL) was added to the mixture and the reaction continued for 24 h. Excess of solvent was removed under reduced pressure and ice water (5 mL) was poured onto the residue with stirring. The mixture was acidified cautiously with 0.5 M HCl (aq) until pH = 5 and extracted with CHCl₃ (8 mL). The organic extract was washed with saturated NaHCO₃ solution, brine and dried over Mg₂SO₄, filtered and evaporated under reduced pressure. Crystallization of the dried residue from acetone/petroleum (1:4) ether afforded 5 (113 mg, 90%) as yellow needles, mp: 157-158 °C. IR (cm⁻¹): 3317, 2926, 2851, 1660, 1596, 1506, 1311, 1260, 1167, 1031. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.44 (s, 1H, OH-5), 9.67 (s, 1H, OH-3), 8.18 (d, J_1 $= 9.1 \text{ Hz}, 2H, H-2',6'), 7.13 \text{ (d, } J = 9.1 \text{ Hz}, 2H, H-3',5'), 6.78 \text{ (d, } J = 2.2 \text{ Hz}, 1H, H-8),}$ 6.37 (d, J = 2.2 Hz, 1H, H-6), 3.87 (s, 3H, OCH₃-7), 3.85 (s, 3H, OCH₃-4'). ¹³C NMR (150 MHz, DMSO- d_6): $\delta = 176.13$ (C-4), 164.98 (C-7), 160.59 (C-4'), 160.37 (C-5), 156.15 (C-9), 146.69 (C-2), 136.35 (C-3), 129.38 (C-2',6'), 123.15 (C-1'), 114.04 (C-3',5'), 104.08 (C-10), 97.49 (C-6), 92.06 (C-8), 56.02 (OCH_3-7) , 55.37 (OCH_3-4') . ESI-HRMS: $315.0873 \, [M+H]^{+}$ (calcd. for $C_{17}H_{15}O_6$: 315.0863).

4'-O-Acetylkaempferol (6)

To a solution of compound $\mathbf{2}$ (251 mg, 0.61 mmol) in CH_2Cl_2 (1 mL) and CH_3CN (3 mL) was added anhydrous $AlCl_3$ (201 mg, 1.52 mmol). The mixture was refluxed for 3 h. After cooling down to room temperature, 1 M HCl (aq) (3 mL) was added and the mixture was stirred further for 1 h. The reaction mixture was diluted with $CHCl_3$ and the pH was adjusted to 4–5 with saturated $NaHCO_3$ solution. After evaporation of the

organic layer, the amorphous residue was purified over silica gel column using EtOAc/CH₂Cl₂/petroleum ether (1:3:6) as solvents to yield **5** (178 mg, 89%) as yellow needles, mp: 164-165 °C. IR (cm⁻¹): 3403, 3298, 2928, 1722, 1653, 1623, 1603, 1504, 1370, 1313, 1245, 1168. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.36 (s, 1H), 10.87 (s, 1H), 9.76 (s, 1H), 8.20 (d, J = 8.9 Hz, 2H), 7.33 (d, J = 2.0 Hz, 2H), 6.47 (d, J = 2.0 Hz, 1H), 6.22 (d, J = 2.0 Hz, 1H), 2.31 (s, 3H). ¹³C NMR (100 MHz, DMSO- d_6): δ = 176.30, 169.09, 164.28, 160.80, 156.41, 151.43, 145.21, 137.07, 128.97, 128.53, 122.11, 103.27, 98.38, 93.64, 20.92. ESI-HRMS: 327.0510 [M-H]⁻ (calcd. for C₁₇H₁₁O₇: 327.0510).

3,7-Di-O-methylkaempferol (kumatakenin) (7) [6]

Methylation of compound **6** (151 mg, 0.46 mmol) with dimethyl sulfate (0.115 mL, 1.2 mmol) and anhydrous K_2CO_3 (190 mg, 1.38 mmol) in dry acetone (6 mL) was carried out at room temperature for 12 h. The mixture was diluted with water (15 mL) and extracted with EtOAc (20 mL). The organic layer was washed with brine, dried over MgSO₄ and concentrated under reduced pressure. Without purification, the dried residue was stirred with 7.0 M methanolic ammonia (3 mL) for 3 h. The precipitate formed was filtered off and crystallized from acetone/petroleum ether (1:2) to provide **7** (121 mg, 84%) as yellow crystals, mp: 228-230 °C. IR (cm⁻¹): 3410, 3261, 2928, 2854, 1658, 1602, 1498, 1346, 1287, 1170. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.67 (s, 1H, OH-5), 10.32 (s, 1H, OH-4'), 7.98 (d, J = 8.9 Hz, 2H, H-2',6'), 6.95 (d, J = 8.9 Hz, 2H, H-3',5'), 6.75 (d, J = 2.0 Hz, 1H, H-8), 6.37 (d, J = 2.0 Hz, 1H, H-6), 3.86 (s, 3H, OCH₃-7), 3.80 (s, 3H, OCH₃-3). ¹³C NMR (100 MHz, DMSO- d_6): δ = 178.11 (C-4), 165.16 (C-7), 160.96 (C-5), 160.32 (C-4'), 156.34 (C-9), 155.99 (C-2), 137.86 (C-3), 130.25 (C-2',6'), 120.50 (C-1'), 115.70 (C-3',5'), 105.22 (C-10), 97.81 (C-6), 92.39 (C-8), 59.74 (OCH₃-3), 56.12 (OCH₃-7). ESI-HRMS: 315.0862 [M+H]* (calcd. for

C₁₇H₁₅O₆: 315.0863).

7-O-Benzyl-3,4',5-tri-O-acetylkaempferol (8)

A mixture of kaempferol tetraacetate (1) (2.5 g, 5.51 mmol), potassium iodide (450 mg, 2.71 mmol), benzyl bromide (1.25 mL, 10.59 mmol), anhydrous K_2CO_3 (1925 mg, 13.95 mmol) and dry acetone (80 mL) was refluxed for 24 h. The filtered solution was concentrated to give an amorphous solid, which was crystallized from absolute ethanol to give **8** as white crystals (2.35 g, 85%), mp: 163-164 °C. IR (cm⁻¹): 1771, 1636, 1621, 1505, 1442, 1369, 1290, 1193, 1170, 1077, 1017. ¹H NMR (600 MHz, CDCl₃): δ = 7.83 (d, J = 8.8 Hz, 2H), 7.43–7.35 (m, 5H), 7.24 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 2.3 Hz, 1H), 6.71 (d, J = 2.3 Hz, 1H), 5.16 (s, 2H), 2.43 (s, 3H), 2.34 (s, 3H), 2.31 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): δ = 170.23, 169.70, 169.05, 168.13, 163.02, 158.30, 154.33, 152.88, 150.96, 135.37, 133.88, 129.69, 128.99, 128.74, 127.70, 127.46, 122.15, 111.45, 109.41, 100.00, 71.01, 21.31, 21.28, 20.76. ESI-HRMS: 525.1152 [M+Na]* (calcd. for $C_{28}H_{22}O_9Na$: 525.1156).

7-O-Benzyl-4'-O-acetylkaempferol (9)

To a solution of compound **8** (300 mg, 0.6 mmol) in CH₂Cl₂ (1 mL) and CH₃CN (3 mL) was added anhydrous AlCl₃ (201 mg, 1.52 mmol). The mixture was refluxed for 3 h. After cooling down to room temperature, 1 M HCl (aq) (3 mL) was added and the mixture was stirred further for 1 h. The reaction mixture was diluted with CHCl₃ and the pH was adjusted to 4–5 with saturated NaHCO₃ solution. After evaporation of the organic layer, the residue was purified over silica gel column using acetone/petroleum ether (1:4) as solvents to yield compound **9** (216 mg, 86%) as pale yellow powder, mp: 172-173 °C. IR (cm⁻¹): 3324, 2925, 2856, 1756, 1647, 1623, 1598, 1503, 1368, 1308,1212, 1171. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.35 (s, 1H, OH-5), 9.91 (s, 1H,

OH-3), 8.23 (d, J = 8.9 Hz, 2H, H-2',6'), 7.48 (d, J = 7.3 Hz, 2H, H-2",6"), 7.42 (t, J = 7.3 Hz, 2H, H-3",5"), 7.36 (overlapped, 3H, H-4",3',5'), 6.89 (d, J = 2.1 Hz, 1H, H-8), 6.47 (d, J = 2.1 Hz, 1H, H-6), 5.24 (s, 2H, H-7"), 2.31 (s, 3H, CH₃COO-4'). ¹³C NMR (100 MHz, DMSO- d_6): δ = 176.45 (C-4), 169.04 (CH₃COO-4'), 164.21 (C-7), 160.48 (C-5), 156.27 (C-9), 151.53 (C-4'), 145.65 (C-2), 137.40 (C-3), 136.13 (C-1"), 128.99 (C-2',6'), 128.56 (C-3",5"), 128.43 (C-1'), 128.16 (C-4"), 127.88 (C-2",6"), 122.13 (C-3',5'), 104.37 (C-10), 98.20 (C-6), 93.06 (C-8), 70.02 (C-7"), 20.90 (CH₃COO-4'). ESI-HRMS: 419.1115 [M+H]⁺ (calcd. for C₂₄H₁₉O₇: 419.1125).

3-O-Methyl-7-O-benzyl-4'-O-acetylkaempferol (10)

A suspension of compound **9** (201 mg, 0.48 mmol), anhydrous K_2CO_3 (105 mg, 0.76 mmol) and Me_2SO_4 (0.06 mL, 0.63 mmol) in dry acetone (6 mL) was stirred at room temperature for 12 h. The mixture was diluted with water (10 mL), and extracted with EtOAc (20 mL). The organic layer was washed with brine, dried over $MgSO_4$ and then concentrated. The residue was crystallized from acetone/petroleum ether (1:3) to afford **10** (176 mg, 85%) as yellowish-brown crystals, mp: 115-117 °C. IR (cm⁻¹): 3444, 2925, 2853, 1744, 1661, 1609, 1503, 1461, 1384, 1342, 1230, 1171, 1005. ¹H NMR (400 MHz, DMSO- d_6): $\bar{\delta}$ = 12.53 (s, 1H), 8.09 (d, J = 8.7 Hz, 2H), 7.47 (d, J = 7.4 Hz, 2H), 7.41 (t, J = 7.4 Hz, 2H), 7.35 (t, J = 8.5 Hz, 3H), 6.86 (d, J = 1.9 Hz, 1H), 6.48 (d, J = 1.9 Hz, 1H), 5.23 (s, 2H), 3.84 (s, 3H), 2.32 (s, 3H). ¹³C NMR (100 MHz, DMSO- d_6): $\bar{\delta}$ = 178.31, 169.02, 164.36, 161.02, 156.46, 154.86, 152.42, 139.01, 136.09, 129.75, 128.57, 128.18, 127.87, 127.49, 122.34, 105.61, 98.60, 93.40, 70.06, 60.08, 20.92. ESI-HRMS: 433.1261 [M+H]⁺ (calcd. for $C_{25}H_{21}O_7$: 433.1282).

3-O-Methyl-7-O-benzylkaempferol (11)

A solution of compound **10** (152 mg, 0.35 mmol) in 7.0 M methanolic ammonia (3 mL)

was stirred at room temperature for 3 h. The precipitate formed was filtered off and crystallized from acetone/petroleum ether (1:3) to give **11** (132 mg, 96%) as yellow needles, mp: 155-157 °C. IR (cm⁻¹): 3421, 2930, 1603, 1494, 1371, 1339, 1287, 1220, 1172. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.67 (s, 1H), 10.30 (s, 1H), 7.97 (d, J = 8.9 Hz, 2H), 7.47 (d, J = 7.1 Hz, 2H), 7.41 (t, J = 7.1 Hz, 2H), 7.35 (t, J = 7.1 Hz, 1H), 6.96 (d, J = 8.9 Hz, 2H), 6.84 (d, J = 2.2 Hz, 1H), 6.46 (d, J = 2.2 Hz, 1H), 5.23 (s, 2H), 3.80 (s, 3H). ¹³C NMR (100 MHz, DMSO- d_6): δ = 178.05, 164.09, 160.97, 160.30, 156.24, 156.00, 137.85, 136.12, 130.20, 128.54, 128.13, 127.84, 120.46, 115.68, 105.32, 98.39, 93.23, 69.98, 59.70. ESI-HRMS: 389.1036 [M-H]⁻ (calcd. for C₂₃H₁₇O₆: 389.1031).

3-O-Methylkaempferol (12) [7]

A mixture of **11** (90 mg, 0.23 mmol), 10% Pd/C (10 mg), MeOH (9 mL) and EtOAc (3 mL) was stirred at ambient temperature under a hydrogen balloon for 5 h. The resulting mixture was filtered off through celite eluting with EtOAc (3 mL). The filtrate was concentrated to give the crude product, which was crystallized from acetone/petroleum ether (1:2) to afford **12** (63 mg, 91%) as pale yellow needles, mp: 237-239 °C. IR (cm⁻¹): 3502, 3118, 2925, 2855, 1467, 1606, 1489, 1360, 1308, 1224, 1161. 1 H NMR (400 MHz, DMSO- d_6): δ = 12.69 (s, 1H, OH-5), 10.85 (s, 1H, OH-7), 10.27 (s, 1H, OH-4'), 7.94 (d, J = 8.9 Hz, 2H, H-2',6'), 6.94 (d, J = 8.9 Hz, 2H, H-3',5'), 6.44 (d, J = 2.0 Hz, 1H, H-8), 6.20 (d, J = 2.0 Hz, 1H, H-6), 3.78 (s, 3H, OCH₃-3). 13 C NMR (150 MHz, DMSO- d_6): δ = 177.89 (C-4), 164.10 (C-7), 161.23 (C-5), 160.14 (C-4'), 156.36 (C-9), 155.60 (C-2), 137.59 (C-3), 130.11 (C-2',6'), 120.55 (C-1'), 115.63 (C-3',5'), 104.20 (C-10), 98.55 (C-6), 93.69 (C-8), 59.68 (OCH₃-3). ESI-HRMS: 299.0563 [M-H]⁻ (calcd. for C₁₆H₁₁O₆: 299.0561).

3-O-Acetyl-7-O-benzylkaempferol (13)

To the solution of compound 8 (300 mg, 0.6 mmol) in acetone (3 mL) and MeOH (1 mL) was added 1 M HCl (aq) (3 mL). The mixture was refluxed for 3.5 h, and then diluted with CHCl₃ (5 mL), neutralized with aqueous NaHCO₃, and extracted with CHCl₃. After evaporation of the organic layer, the dried residue was purified over silica gel column using acetone/petroleum ether (1:3) as solvents to provide 13 (205 mg, 82%) as yellow needles, mp: 170-171 °C. IR (cm⁻¹): 3313, 2935, 2876, 1781, 1655, 1603, 1583, 1494, 1373, 1339, 1283, 1165. ¹H NMR (600 MHz, DMSO- d_6): $\delta = 12.19$ (s, 1H, OH-5), 10.41 (s, 1H, OH-4'), 7.80 (d, J = 8.8 Hz, 2H, H-2',6'), 7.48 (d, J = 7.5Hz, 2H, H-2",6"), 7.42 (t, J = 7.5 Hz, 2H, H-3",5"), 7.36 (t, J = 7.5 Hz, 1H, H-4"), 6.97 (d, J = 8.8 Hz, 2H, H-3',5'), 6.90 (d, J = 2.0 Hz, 1H, H-8), 6.53 (d, J = 2.0 Hz, 1H, H-6),5.26 (s, 2H, H-7"), 2.34 (s, 3H, CH₃COO-3). ¹³C NMR (150 MHz, DMSO- d_6): δ = 175.07 (C-4), 167.91 (CH₃COO-3), 164.55 (C-7), 160.85 (C-4'), 160.77 (C-5), 156.49 (C-9), 156.36 (C-2), 136.00 (C-1"), 130.10 (C-2',6'), 129.82 (C-3), 128.52 (C-3",5"), 128.14 (C-4"), 127.84 (C-2",6"), 119.27 (C-1'), 115.97 (C-3',5'), 104.57 (C-10), 98.90 (C-6), 93.76 (C-8), 70.10 (C-7"), 20.21 (CH₃COO-3). ESI-HRMS: 417.0993 [M-H] (calcd. for C₂₄H₁₇O₇: 417.0980).

3-O-Acetyl-7-O-benzyl-4',5-di-O-methylkaempferol (14)

To a suspension of **13** (171 mg, 0.41 mmol), K_2CO_3 (206 mg, 1.49 mmol) and acetone (8 mL) was added Me_2SO_4 (0.11 mL, 1.15 mmol). After the mixture was stirred at 30 °C for 12 h, another batch of Me_2SO_4 (0.02 mL, 0.2 mmol) was added. After another 12 h at 30 °C, the reaction mixture was diluted with water (20 mL) and extracted with EtOAc (30 mL). After removal of the solvent, the residue obtained was purified over silica gel column using acetone/petroleum ether (1:3) as solvents to

afford compound **14** (168 mg, 92%) as white powder, mp: 94-95 °C. IR (cm⁻¹): 2955, 2919, 2851, 1776, 1607, 1467, 1349, 1298, 1258, 1170. ¹H NMR (400 MHz, DMSO- d_6): $\delta = 7.86$ (d, J = 9.0 Hz, 2H), 7.51 (d, J = 7.1 Hz, 2H), 7.43 (t, J = 7.1 Hz, 2H), 7.37 (t, J = 7.1 Hz, 1H), 7.14 (d, J = 9.0 Hz, 2H), 6.97 (d, J = 2.1 Hz, 1H), 6.64 (d, J = 2.1 Hz, 1H), 5.26 (s, 2H), 3.85 (s, 3H), 3.84 (s, 3H), 2.30 (s, 3H). ¹³C NMR (150 MHz, DMSO- d_6): $\delta = 169.08$, 168.03, 163.22, 161.46, 160.48, 158.39, 152.35, 136.07, 132.65, 129.52, 128.57, 128.23, 128.03, 121.32, 114.51, 107.76, 96.94, 94.17, 70.14, 56.28, 55.50, 20.37. ESI-HRMS: 447.1455 [M+H]⁺ (calcd. for C₂₆H₂₃O₇: 447.1438).

3-O-Acetyl-4',5-di-O-methylkaempferol (15)

A mixture of **14** (150 mg, 0.34 mmol), 10% Pd/C (15 mg), MeOH (15 mL) and EtOAc (5 mL) was stirred at ambient temperature under a hydrogen balloon for 5 h. The mixture was filtered off through celite eluting with EtOAc (6 mL). The filtrate was concentrated crude product, to give the which was crystallized from acetone/petroleum ether (1:2) to afford 15 (110 mg, 92%) as off-white crystals, mp: 145-147 °C. IR (cm⁻¹): 3263, 2926, 2855, 1730, 1603, 1509, 1459, 1298, 1257, 1175, 1102, 1030. ¹H NMR (400 MHz, DMSO- d_6): $\delta = 10.86$ (s, 1H), 7.82 (d, J = 9.0 Hz, 2H), 7.12 (d, J = 9.0 Hz, 2H), 6.53 (d, J = 2.0 Hz, 1H), 6.41 (d, J = 2.0 Hz, 1H), 3.84 (s, 3H), 3.81 (s, 3H), 2.29 (s, 3H). ¹³C NMR (150 MHz, DMSO- d_6): δ = 169.39, 168.42, 163.29, 161.58, 161.05, 158.58, 152.39, 132.53, 129.69, 121.59, 114.69, 106.68, 96.83, 95.36, 56.21, 55.67, 20.54. ESI-HRMS: $379.0770 \text{ [M+Na]}^+ \text{ (calcd. for } C_{19}H_{16}O_7\text{Na}$: 379.0788).

4',5-Di-*O*-methylkaempferol (16) [8]

Compound **15** (100 mg, 0.28 mmol) was stirred with 7.0 M methanolic ammonia (2 mL) at room temperature for 3 h. The precipitate formed was filtered off and crystallized

from acetone/petroleum ether (1:3) to give **16** (83 mg, 94%) as yellowish crystals, mp: 280-282 °C. IR (cm⁻¹): 3179, 2934, 2838, 1602, 1563, 1501, 1378, 1297, 1256, 1207, 1183, 1031. ¹H NMR (400 MHz, DMSO- d_6): δ = 10.51 (s, 1H, OH-7), 8.75 (s, 1H, OH-3), 8.09 (d, J = 8.9 Hz, 2H, H-2′,6′), 7.10 (d, J = 8.9 Hz, 2H, H-3′,5′), 6.50 (d, J = 1.8 Hz, 1H, H-8), 6.36 (d, J = 1.8 Hz, 1H, H-6), 3.83 (s, 6H, OCH₃-5,4′). ¹³C NMR (150 MHz, DMSO- d_6): δ = 171.00 (C-4), 162.55 (C-7), 160.57 (C-5), 159.96 (C-4′), 158.04 (C-9), 141.38 (C-2), 137.41 (C-3), 128.61 (C-2′,6′), 123.59 (C-1′), 114.03 (C-3′,5′), 105.20 (C-10), 95.92 (C-6), 94.76 (C-8), 55.98 (OCH₃-5 or 4′), 55.31 (OCH₃-5 or 4′). ESI-HRMS: 315.0852 [M+H]⁺ (calcd. for C₁₇H₁₅O₆: 315.0863).

7-O-Benzyl-4'-O-methylkaempferol (17)

The mixture of compound **8** (1 g, 1.99 mmol), dimethyl sulfate (0.29 mL, 3.04 mmol), anhydrous K_2CO_3 (994 mg, 7.2 mmol) and acetone (27 mL) was refluxed for 2 h. Methanol (9 mL) was added to the mixture and the reaction continued for 24 h. Excess of solvent was removed and ice water (30 mL) was poured onto the obtained residue with stirring. The mixture was neutralized with 0.5 M HCl (aq), and extracted with CHCl₃ (40 mL). After evaporation of solvents, the yellow powder was crystallized from $CH_2CI_2/MeOH$ (1:4) to provide compound **17** (637 mg, 82%) as yellow needles, mp: 168-170 °C. IR (cm⁻¹): 3293, 2927, 1652, 1617, 1589, 1501, 1351, 1310, 1260, 1225, 1162, 1091, 1036. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.44 (s, 1H), 9.64 (s, 1H), 8.17 (d, J = 9.1 Hz, 2H), 7.48 (d, J = 7.1 Hz, 2H), 7.42 (t, J = 7.1 Hz, 2H), 7.36 (t, J = 7.1 Hz, 1H), 7.13 (d, J = 9.1 Hz, 2H), 6.86 (d, J = 2.1 Hz, 1H), 6.45 (d, J = 2.1 Hz, 1H), 5.24 (s, 2H), 3.85 (s, 3H). ¹³C NMR (150 MHz, DMSO- d_6): δ = 176.13, 163.98, 160.62, 160.43, 156.08, 146.76, 136.39, 136.16, 129.39, 128.52, 128.11, 127.83, 123.15, 114.08, 104.22, 98.06, 92.98, 69.97, 55.39. ESI-HRMS: 389.1018 [M-H]⁻ (calcd. for $C_{23}H_{17}O_6$; 389.1031).

4'-O-Methylkaempferol (kaempferide) (18) [9]

A mixture of 17 (410 mg, 1.05 mmol), 10% Pd/C (45 mg), MeOH (27 mL) and EtOAc (9 mL) was stirred at room temperature under a hydrogen balloon for 5 h. The mixture was filtered off through celite eluting with EtOAc (10 mL). The filtrate was the crude product, which was crystallized concentrated to give acetone/petroleum ether (1:2) to yield 18 (293 mg, 93%) as yellow crystals, mp: 177-178 °C. IR (cm⁻¹): 3445, 3291, 2925, 2854, 1615, 1510, 1374, 1310, 1253, 1172. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.44 (s, 1H, OH-5), 10.83 (s, 1H, OH-7), 9.53 (s, 1H, OH-3), 8.14 (d, J = 9.1 Hz, 2H, H-2',6'), 7.11 (d, J = 9.1 Hz, 2H, H-3',5'), 6.46 (d, J= 2.0 Hz, 1H, H-8), 6.20 (d, J = 2.0 Hz, 1H, H-6), 3.84 (s, 3H, OCH₃-4'). ¹³C NMR (150 MHz, DMSO- d_6): $\delta = 176.01$ (C-4), 164.00 (C-7), 160.73 (C-5), 160.50 (C-4'), 156.25 (C-9), 146.28 (C-2), 136.06 (C-3), 129.34 (C-2',6'), 123.27 (C-1'), 114.06 (C-3',5'), 103.10 (C-10), 98.25 (C-6), 93.53 (C-8), 55.37 (OCH₃-4'). ESI-HRMS: 299.0560 $[M-H]^{-}$ (calcd. for $C_{16}H_{11}O_6$: 299.0561).

7-O-Benzylkaempferol (19)

The solution of **8** (400 mg, 0.8 mmol) in 7.0 M methanolic ammonia (9 mL) was stirred at room temperature for 3 h. The precipitate formed was filtered off and crystallized from acetone/MeOH (1:4) to give **19** (284 mg, 95%) as yellow crystals, mp: 183-184 °C. IR (cm⁻¹): 3436, 2925, 1655, 1605, 1586, 1498, 1351, 1309, 1227, 1172. ¹H NMR (400 MHz, DMSO- d_6): δ = 12.48 (s, 1H), 10.14 (s, 1H), 9.53 (s, 1H), 8.08 (d, J = 8.9 Hz, 2H), 7.48 (d, J = 7.1 Hz, 2H), 7.42 (t, J = 7.1 Hz, 2H), 7.36 (t, J = 7.1 Hz, 1H), 6.94 (d, J = 8.9 Hz, 2H), 6.84 (d, J = 2.1 Hz, 1H), 6.44 (d, J = 2.1 Hz, 1H), 5.24 (s, 2H). ¹³C NMR (100 MHz, DMSO- d_6): δ = 176.02, 163.89, 160.42, 159.34, 156.02, 147.31, 136.19, 136.01, 129.58, 128.54, 128.13, 127.86, 121.56, 115.47, 104.17, 98.03,

92.91, 69.95. ESI-HRMS: 375.0883 [M-H]⁻ (calcd. for C₂₂H₁₅O₆: 375.0874).

7-O-Benzyl-3,4'-di-O-methylkaempferol (20) [10]

A suspension of **19** (120 mg, 0.32 mmol), anhydrous K_2CO_3 (138 mg, 1 mmol) and Me_2SO_4 (0.08 mL, 0.84 mmol) in dry acetone (5 mL) was stirred for 12 h at room temperature. The mixture was diluted with water (8 mL), neutralized with 0.5 M HCl (aq) and extracted with EtOAc. After evaporation of solvents, the yellow powder was purified over silica gel column using acetone/petroleum ether (1:5) as solvents to provide **20** (112 mg, 87%) as pale yellow granular crystals, mp: 113-114 °C. IR (cm⁻¹): 3461, 2928, 1658, 1603, 1497, 1378, 1259, 1204, 1177. ¹H NMR (400 MHz, DMSO- d_6): $\bar{\delta}$ = 12.63 (s, 1H), 8.05 (d, J = 9.0 Hz, 2H), 7.47 (d, J = 7.1 Hz, 2H), 7.42 (t, J = 7.1 Hz, 2H), 7.36 (t, J = 7.1 Hz, 1H), 7.15 (d, J = 9.0 Hz, 2H), 6.87 (d, J = 2.1 Hz, 1H), 6.47 (d, J = 2.1 Hz, 1H), 5.24 (s, 2H), 3.87 (s, 3H), 3.81 (s, 3H). ¹³C NMR (150 MHz, DMSO- d_6): $\bar{\delta}$ = 178.10, 164.16, 161.46, 160.97, 156.29, 155.60, 138.19, 136.09, 130.04, 128.51, 128.10, 127.81, 122.07, 114.28, 105.39, 98.43, 93.30, 69.99, 59.77, 55.46. ESI-HRMS: 427.1153 [M+Na]⁺ (calcd. for $C_{24}H_{20}O_6$ Na: 427.1152).

7-O-Benzyl-3,4',5-tri-O-methylkaempferol (21) [11]

The mixture of **19** (120 mg, 0.32 mmol), Me₂SO₄ (0.11 mL, 1.15 mmol) and K₂CO₃ (177 mg, 1.28 mmol) in acetone (6 mL) was stirred for 12 h at 30 °C. Another batch of Me₂SO₄ (0.03 mL, 0.32 mmol) was then added. After another 12 h at 30 °C, the mixture was diluted with water (10 mL) and extracted with EtOAc (20 mL). After evaporation of solvents, the residue was purified over silica gel column using acetone/petroleum ether (1:3) as solvents to provide **21** (127 mg, 95%) as white powder, mp: 133-135 °C. IR (cm⁻¹): 2925, 2853, 1628, 1604, 1455, 1348, 1295, 1256, 1211, 1176, 1013. ¹H NMR (400 MHz, CDCl₃): δ = 8.06 (d, J = 9.1 Hz, 2H), 7.48–7.37

(m, 5H), 7.01 (d, J = 9.1 Hz, 2H), 6.59 (d, J = 2.0 Hz, 1H), 6.43 (d, J = 2.0 Hz, 1H), 5.14 (s, 2H), 3.96 (s, 3H), 3.89 (s, 3H), 3.88 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): $\delta = 174.20$, 163.06, 161.32, 161.24, 158.91, 152.88, 141.27, 135.90, 129.98, 128.95, 128.62, 127.77, 123.42, 114.07, 109.85, 96.45, 93.55, 70.67, 60.05, 56.57, 55.54. ESI-HRMS: 419.1482 [M+H]⁺ (calcd. for $C_{25}H_{23}O_6$: 419.1489).

3,4'-Di-O-methylkaempferol (22) [7]

A mixture of **20** (100 mg, 0.25 mmol), 10% Pd/C (10 mg), MeOH (9 mL) and EtOAc (3 mL) was stirred at room temperature under a hydrogen balloon for 5 h. The mixture was filtered off through celite eluting with EtOAc (4 mL). After concentration of the filtrate, the yellow powder was crystallized from acetone/petroleum ether (1:4) to afford **22** (71 mg, 91%) as pale yellow needles, mp: 216-218 °C. IR (cm⁻¹): 3428, 3111, 2933, 1651, 1607, 1574, 1497, 1365, 1305, 1263, 1223, 1164, 1019. ¹H NMR (400 MHz, DMSO- d_6): $\bar{\delta}$ = 12.64 (s, 1H, OH-5), 10.88 (s, 1H, OH-7), 8.02 (d, J = 9.0 Hz, 2H, H-2',6'), 7.13 (d, J = 9.0 Hz, 2H, H-3',5'), 6.46 (d, J = 2.0 Hz, 1H, H-8), 6.21 (d, J = 2.0 Hz, 1H, H-6), 3.86 (s, 3H, OCH₃-4'), 3.79 (s, 3H, OCH₃-3). ¹³C NMR (100 MHz, DMSO- d_6): $\bar{\delta}$ = 177.96 (C-4), 164.22 (C-7), 161.36 (C-4'), 161.27 (C-5), 156.44 (C-9), 155.24 (C-2), 137.94 (C-3), 130.02 (C-2',6'), 122.20 (C-1'), 114.27 (C-3',5'), 104.29 (C-10), 98.63 (C-6), 93.79 (C-8), 59.79 (OCH₃-3), 55.46 (OCH₃-4'). ESI-HRMS: 315.0858 [M+H]* (calcd. for C₁₇H₁₅O₆: 315.0863).

3,4',5-Tri-*O*-methylkaempferol (23) [5]

A mixture of **21** (100 mg, 0.24 mmol), 10% Pd/C (10 mg), MeOH (12 mL) and EtOAc (4 mL) was stirred at room temperature under a hydrogen balloon for 5 h. The mixture was filtered off through celite eluting with EtOAc (6 mL). After concentration of the filtrate, the pale green powder was crystallized from acetone/MeOH (1:3) to afford **23**

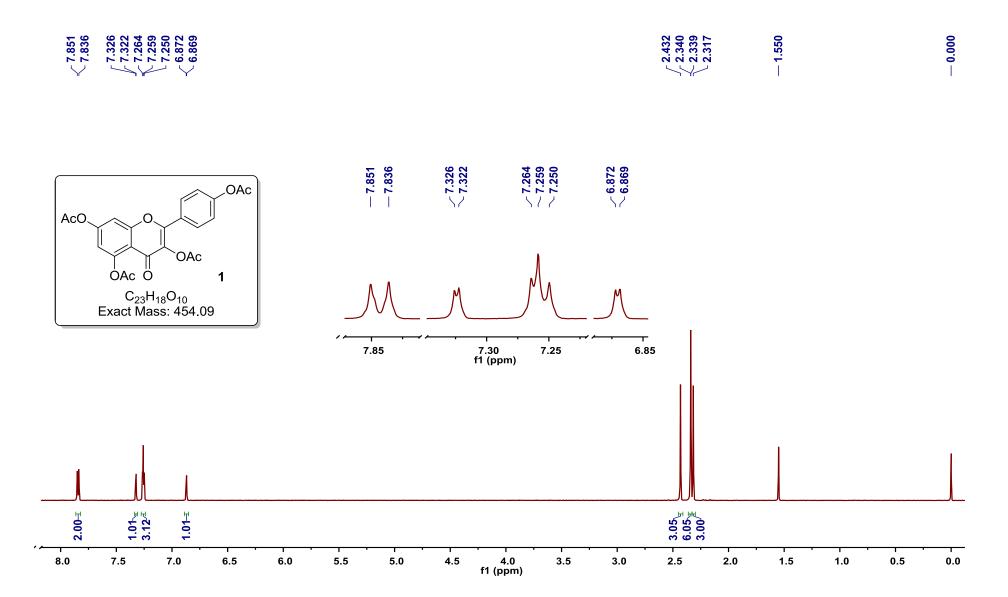
(70 mg, 89%) as off-white crystals, mp: 151-152 °C. IR (cm⁻¹): 3134, 2923, 2852, 1605, 1575, 1470, 1356, 1297, 1256, 1189. ¹H NMR (400 MHz, DMSO- d_6): δ = 10.73 (s, 1H, OH-7), 7.97 (d, J = 9.0 Hz, 2H, H-2′,6′), 7.11 (d, J = 9.0 Hz, 2H, H-3′,5′), 6.48 (d, J = 2.0 Hz, 1H, H-8), 6.36 (d, J = 2.0 Hz, 1H, H-6), 3.84 (s, 3H, OCH₃-5), 3.81 (s, 3H, OCH₃-4′), 3.71 (s, 3H, OCH₃-3). ¹³C NMR (100 MHz, DMSO- d_6): δ = 172.09 (C-4), 162.50 (C-7), 160.80 (C-5), 160.74 (C-4′), 158.11 (C-9), 151.51 (C-2), 139.99 (C-3), 129.52 (C-2′,6′), 122.63 (C-1′), 114.15 (C-3′,5′), 107.42 (C-10), 96.15 (C-6), 94.88 (C-8), 59.28 (OCH₃-3), 55.90 (OCH₃-4′), 55.39 (OCH₃-5). ESI-HRMS: 329.1016 [M+H]⁺ (calcd. for C₁₈H₁₇O₆: 329.1020).

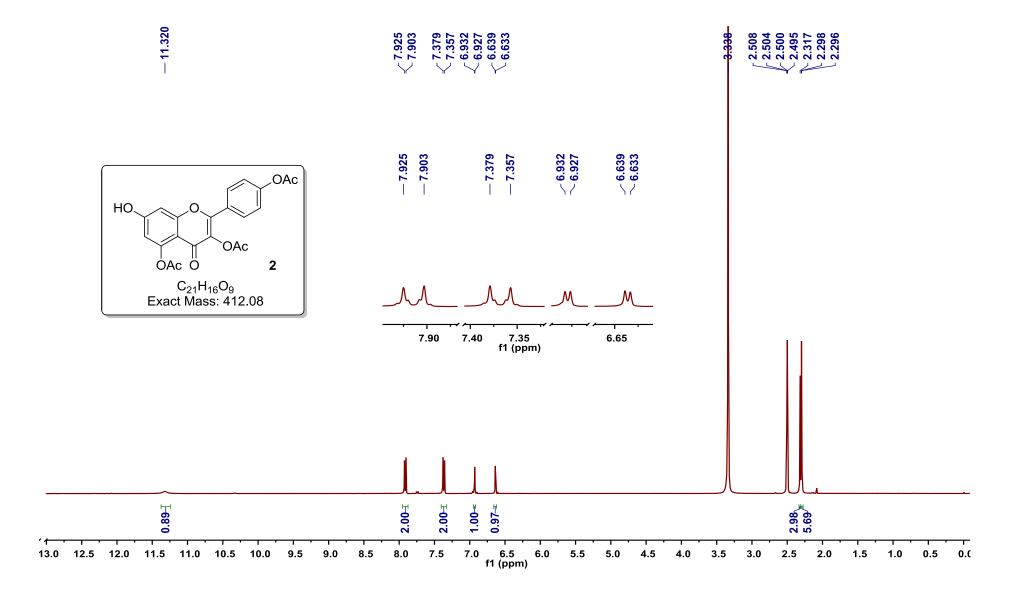
References

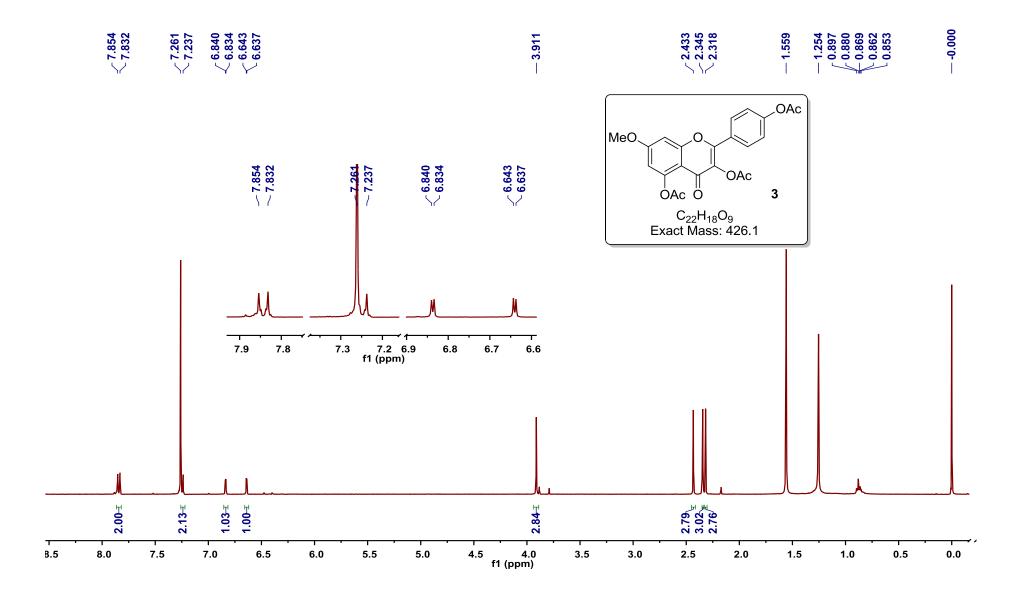
- Prawat, H.; Mahidol, C.; Ruchirawat, S.; Prawat, U.; Tuntiwachwuttikul, P.;
 Tooptakong, U.; Taylor, W. C.; Pakawatchal, C.; Skelton, B. W.; White, A. H.
 Phytochemistry 1995, 40, 1167–1173.
- 2. Kawamura, T.; Hayashi, M.; Mukai, R.; Terao, J.; Nemoto, H. *Synthesis* **2014**, *46*, 170–174.
- 3. Kimura, Y.; Takido, M.; Takahashi, S.; Kimishima, M. *Yakugaku Zasshi* **1967**, *87*, 440–443.
- Zhang, X.-F.; Hung, T. M.; Phuong, P. T.; Ngoc, T. M.; Min, B.-S.; Song, K.-S.;
 Seong, Y. H.; Bae, K.-H. Arch. Pharm. Res. 2006, 29, 1102–1108.
- 5. Rossi, M. H.; Yoshida, M.; Maia, J. G. S. *Phytochemistry* **1997**, *45*, 1263–1269.
- 6. Wang, Y.; Hamburger, M.; Gueho, J.; Hostettmann, K. *Phytochemistry* **1989**, *28*, 2323–2327.
- 7. Nakatani, N.; Jitoe, A.; Masuda, T.; Yonemori, S. Agric. Biol. Chem. 1991, 55,

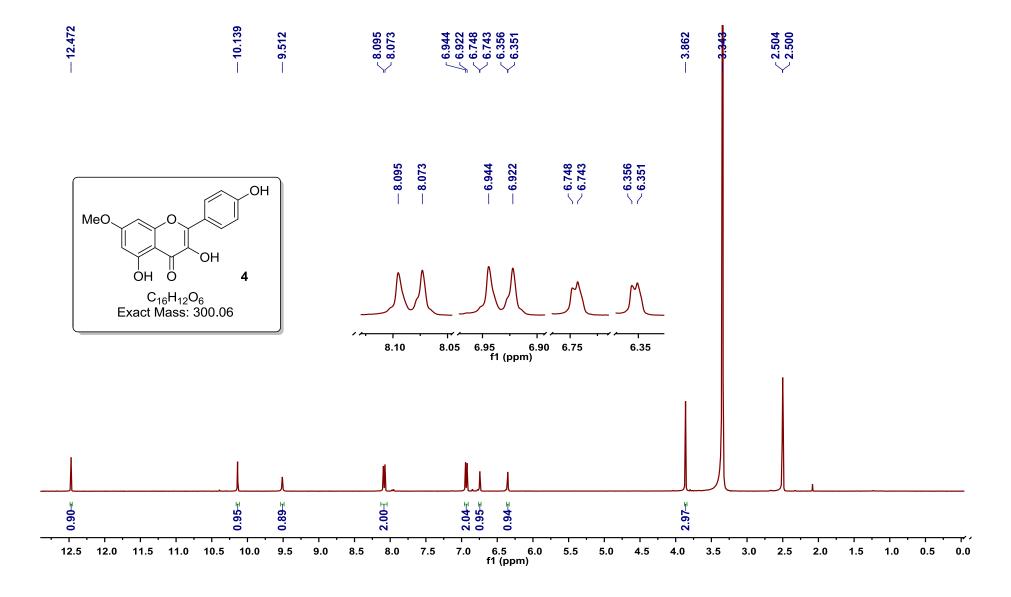
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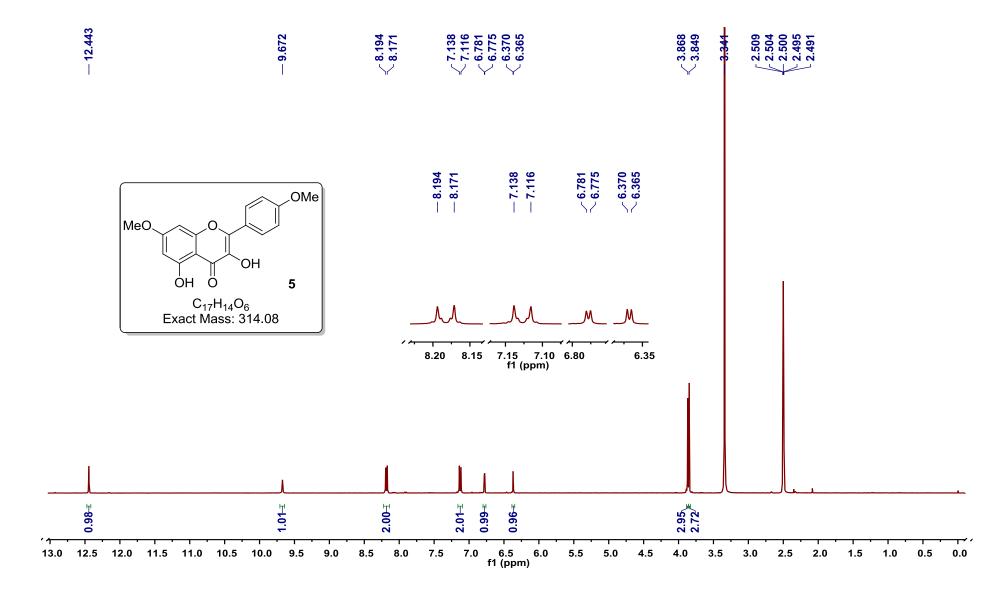
- 8. Kanao, M.; Shimokoriyama, M. Acta Phytochimica 1949, 15, 229–231.
- 9. Wadher, S. J.; Tapas, A. R.; Yeole, P. G. Int. J. Chem. Sci. 2006, 4, 761–766.
- 10. Bhrara, S. C.; Jain, A. C.; Seshadri, T. R. *Indian J. Chem.* **1965**, *3*, 68–70.
- Rajagopalan, S.; Rao, P. R.; Rao, K. V.; Seshadri, T. R. *Proc. Indian AS*, *Sect. A* 1949, 29A, 9–15.

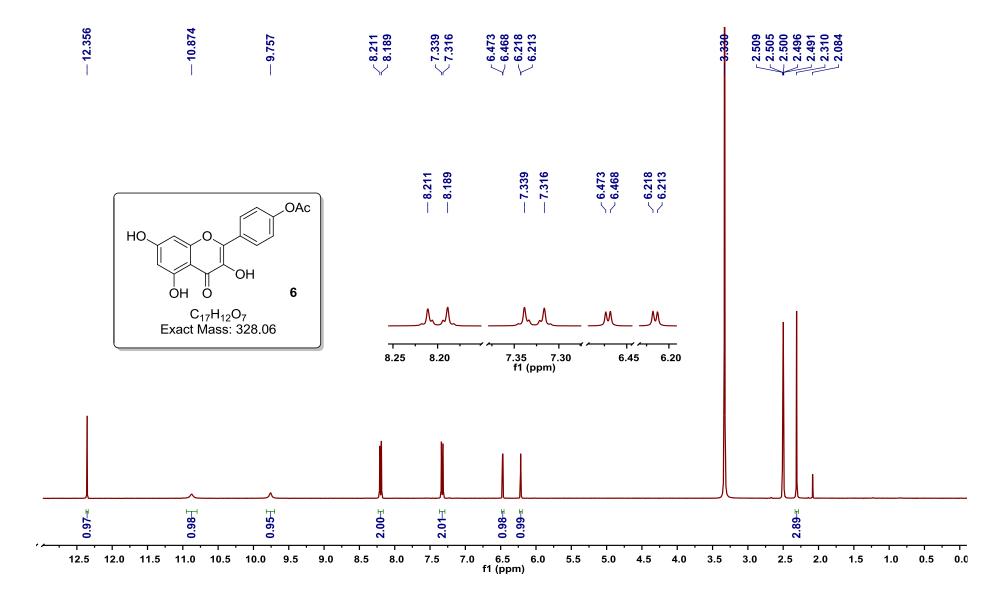


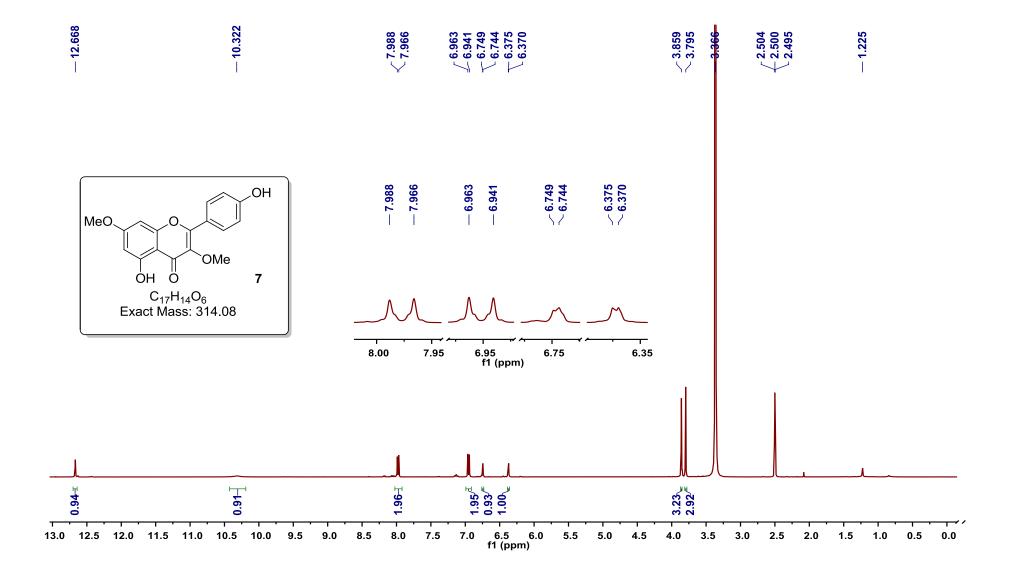


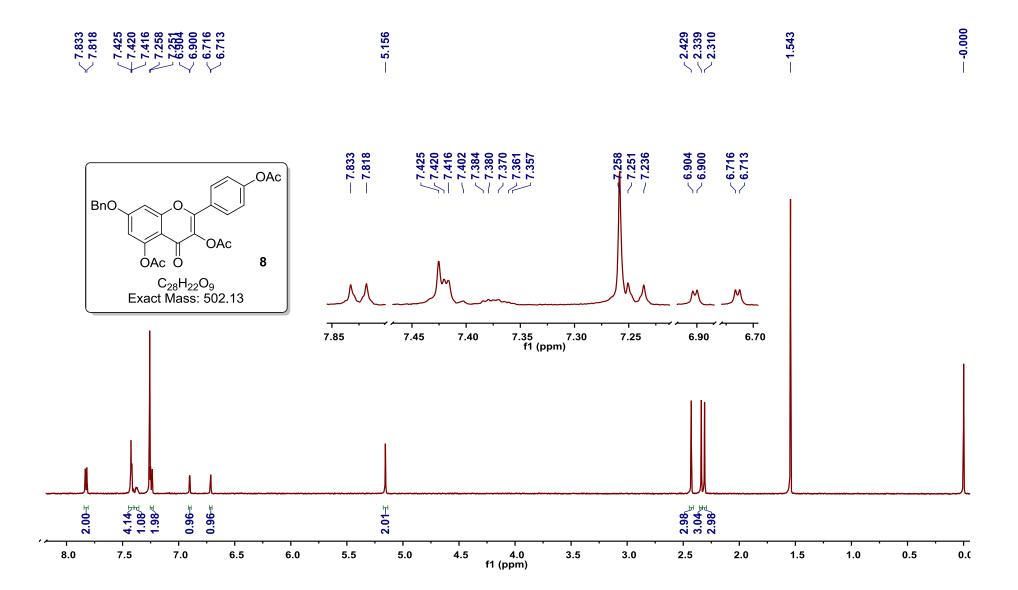


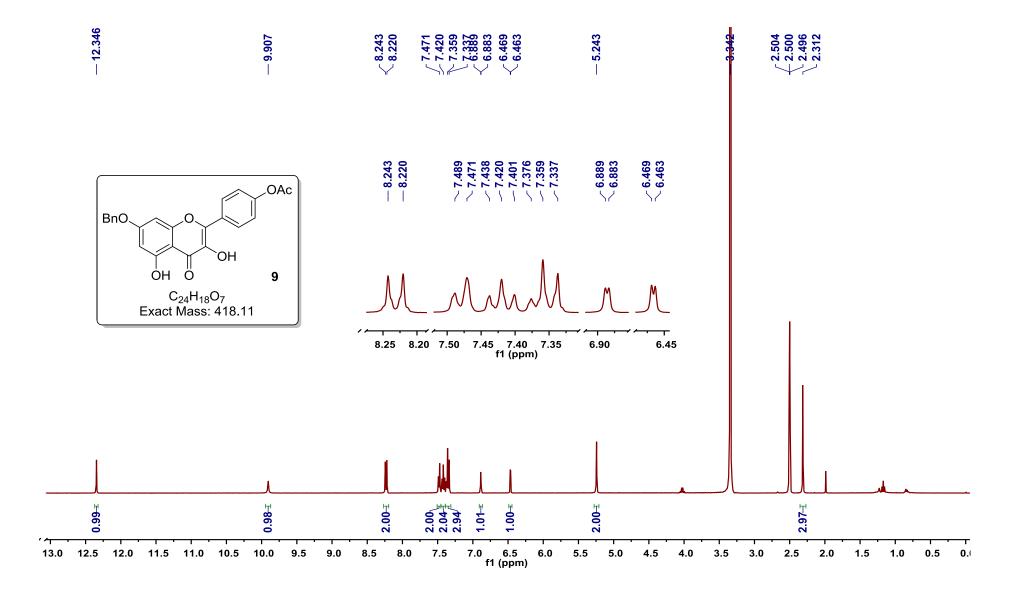


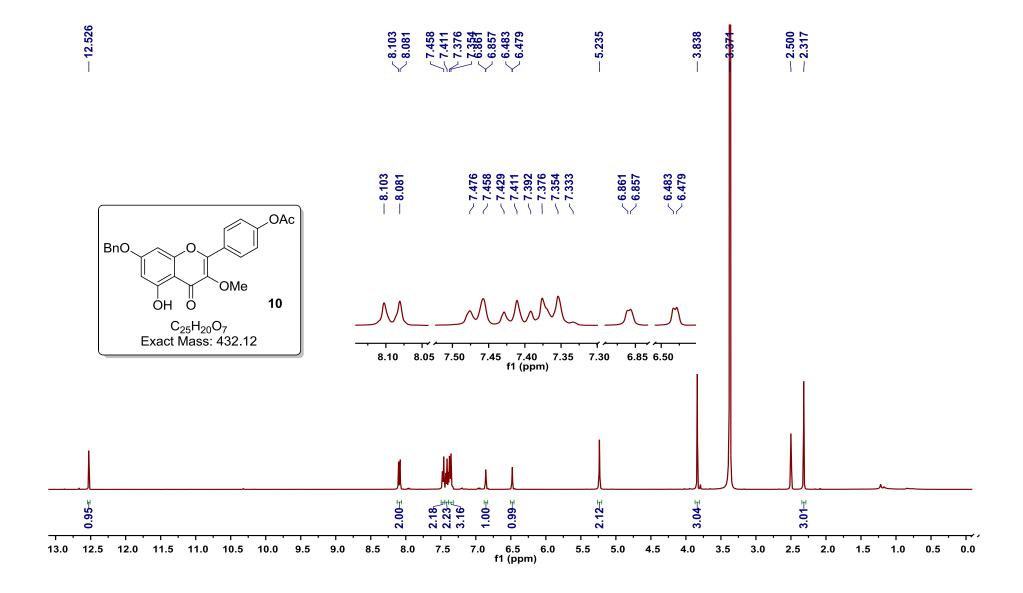


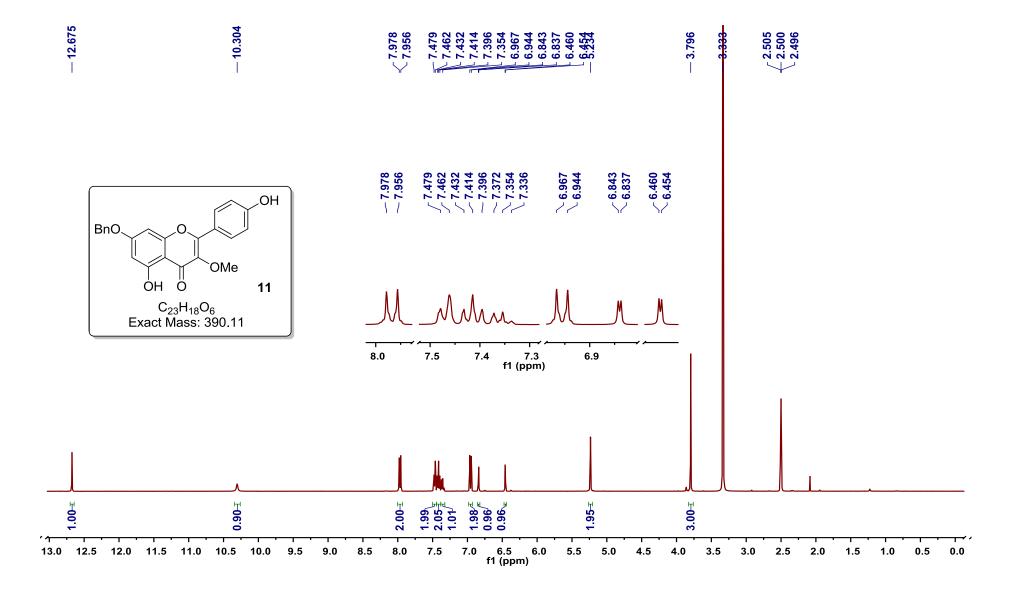


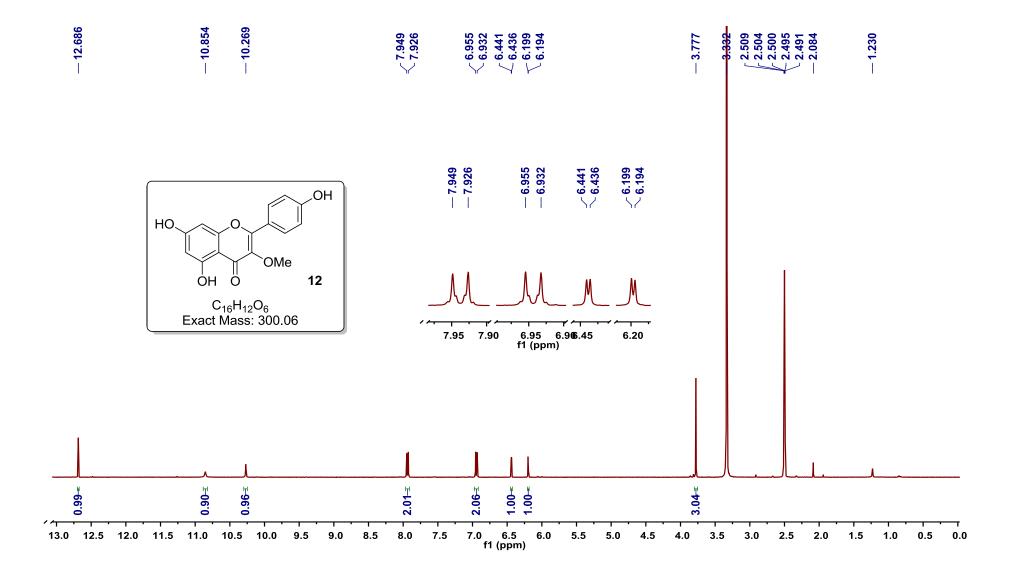


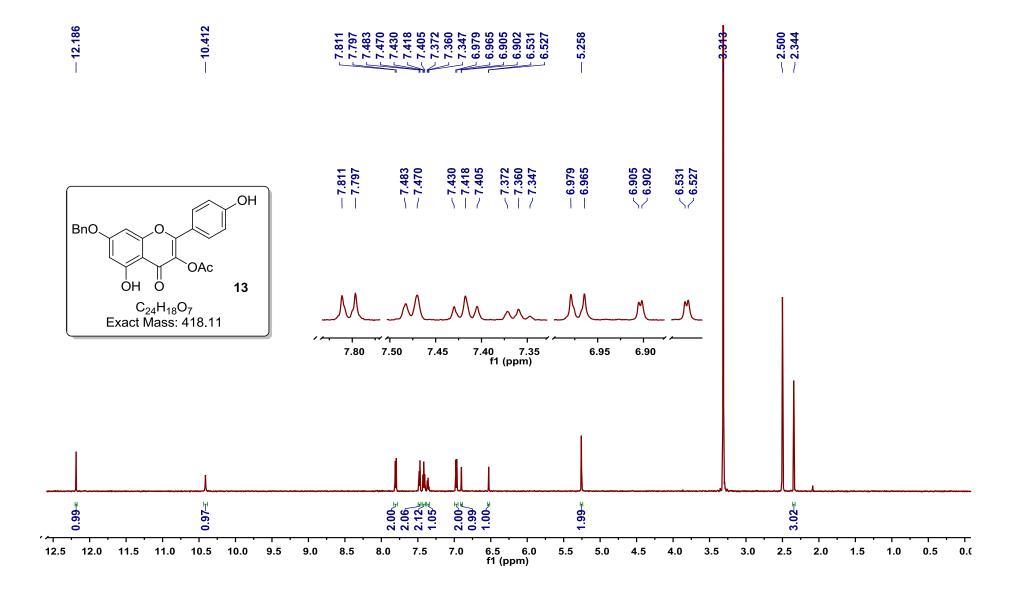


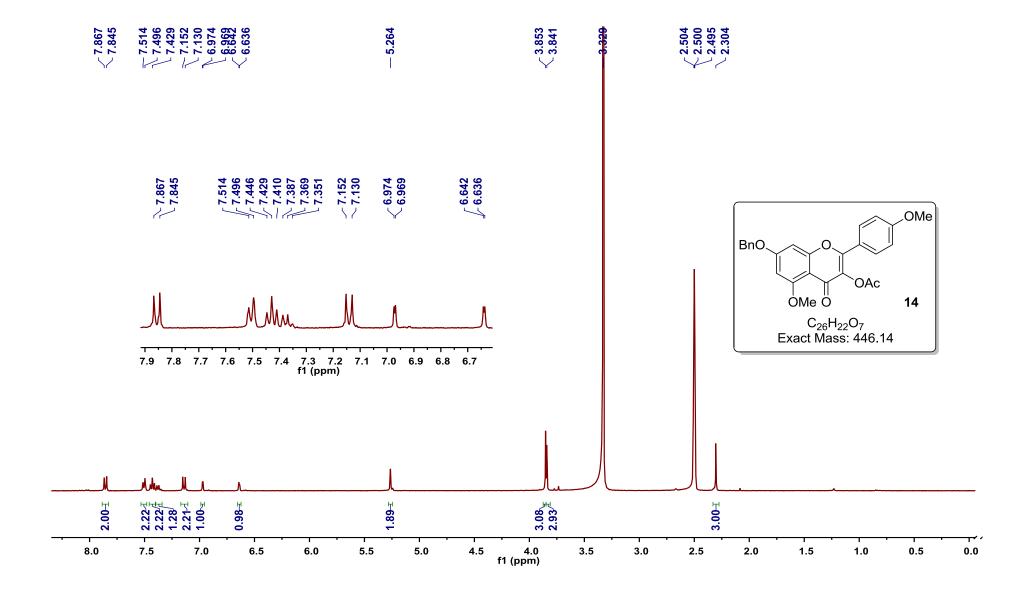


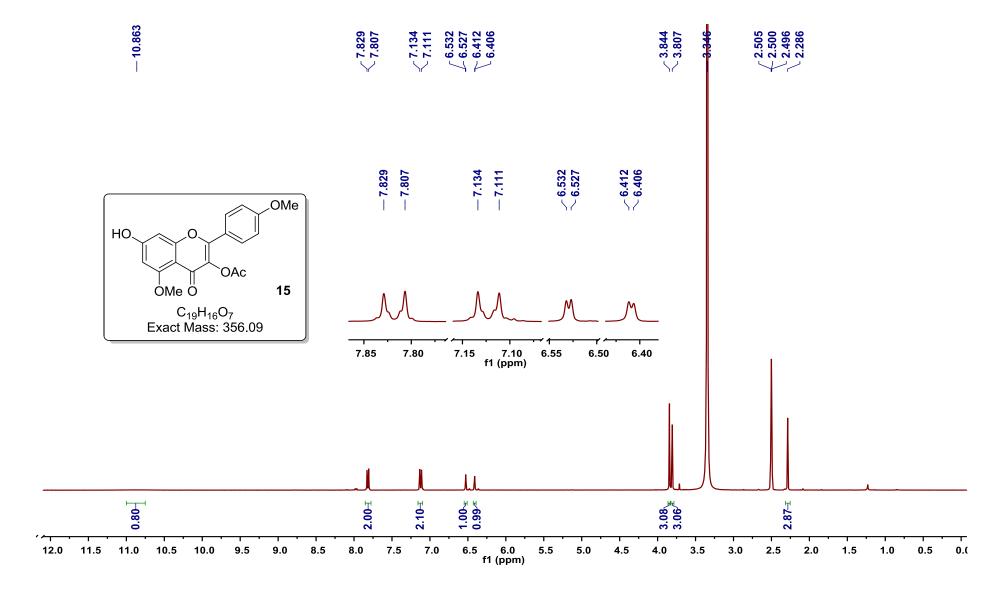


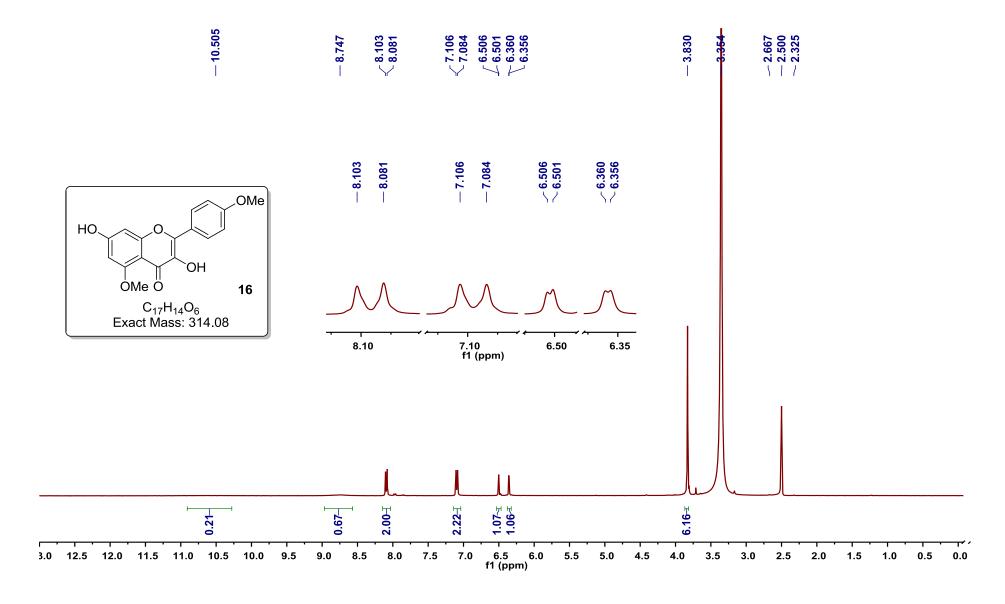


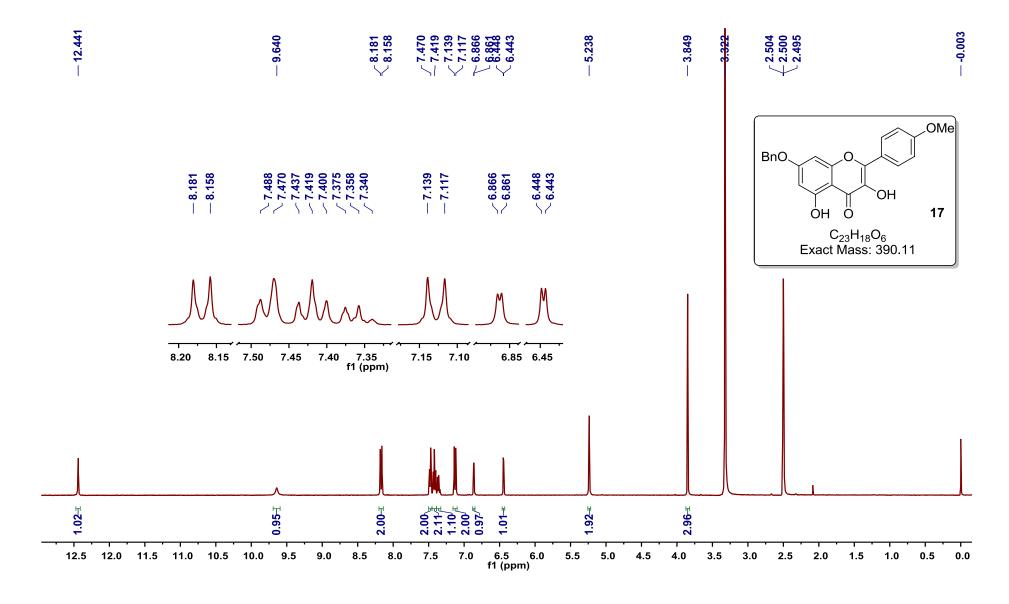


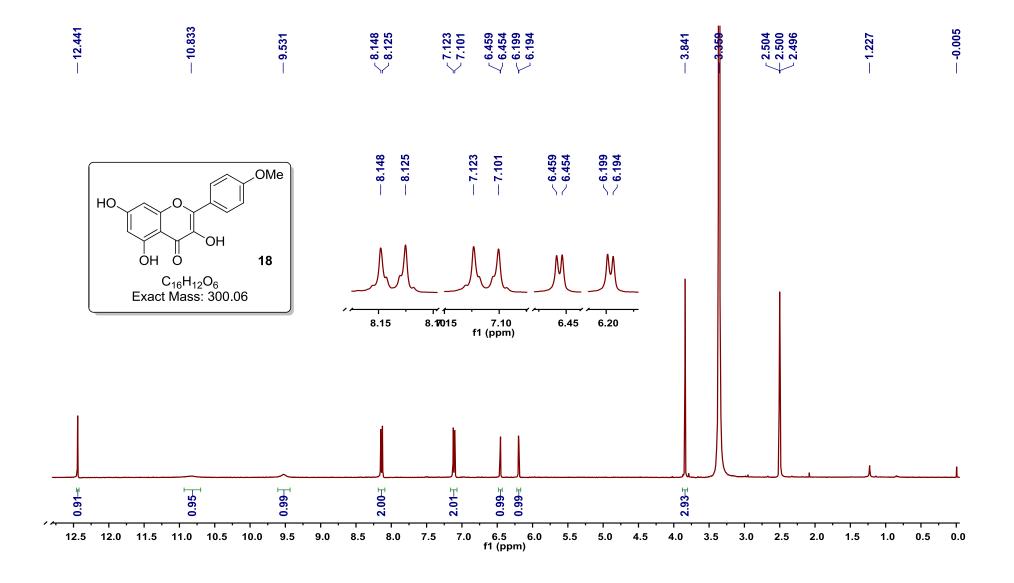


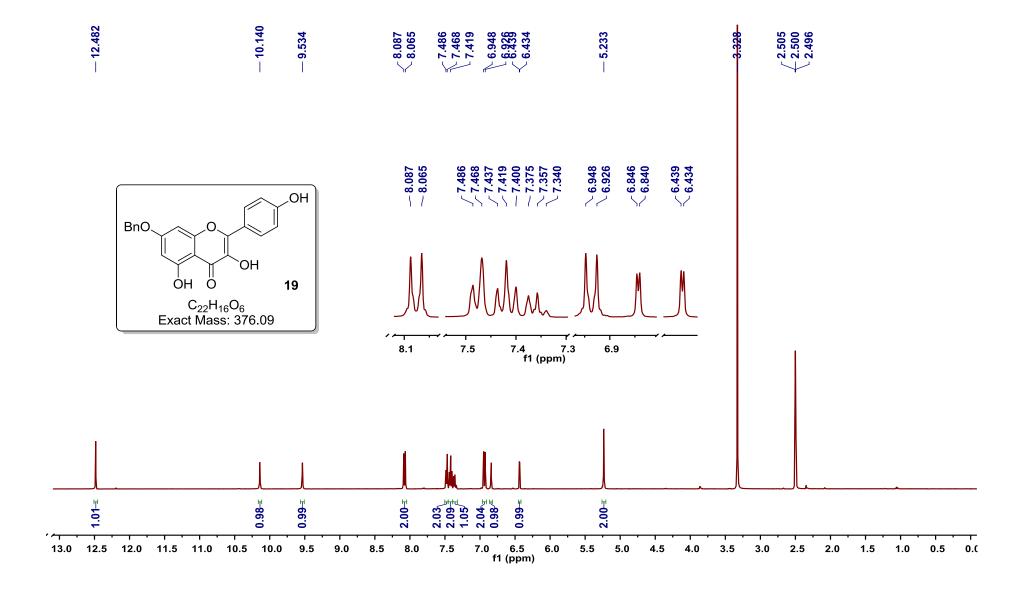


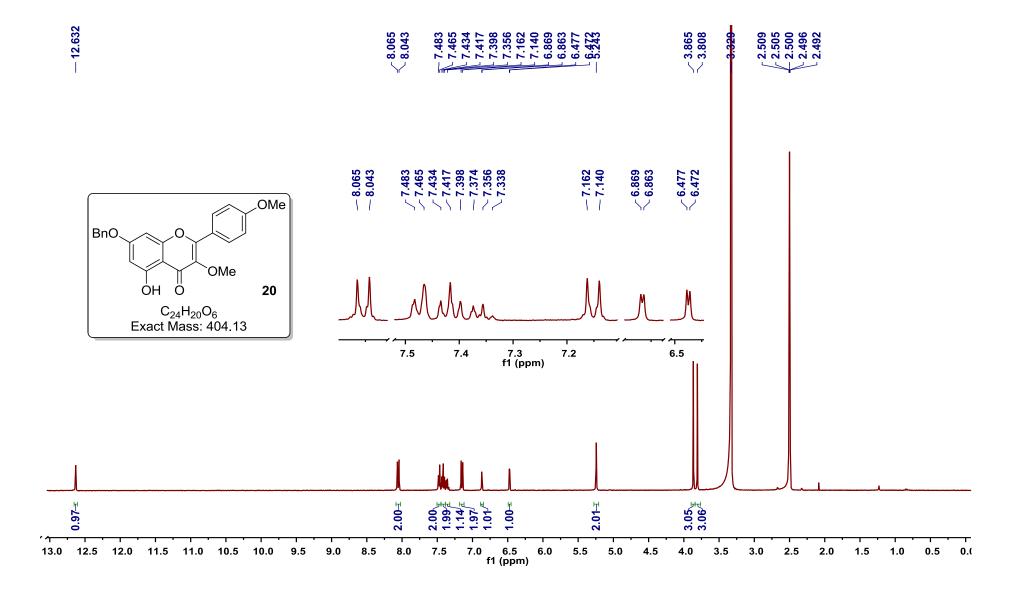


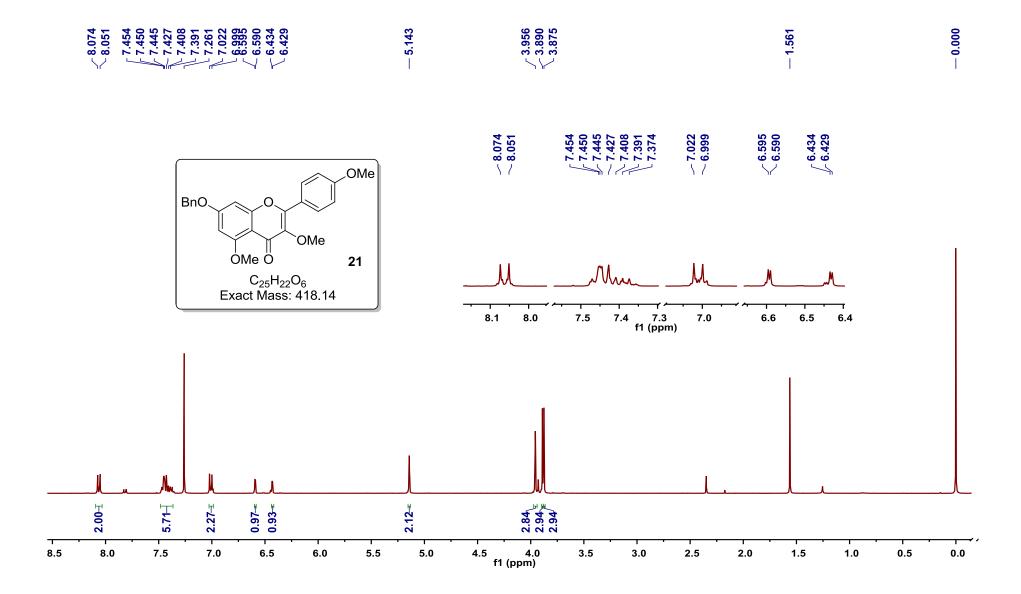


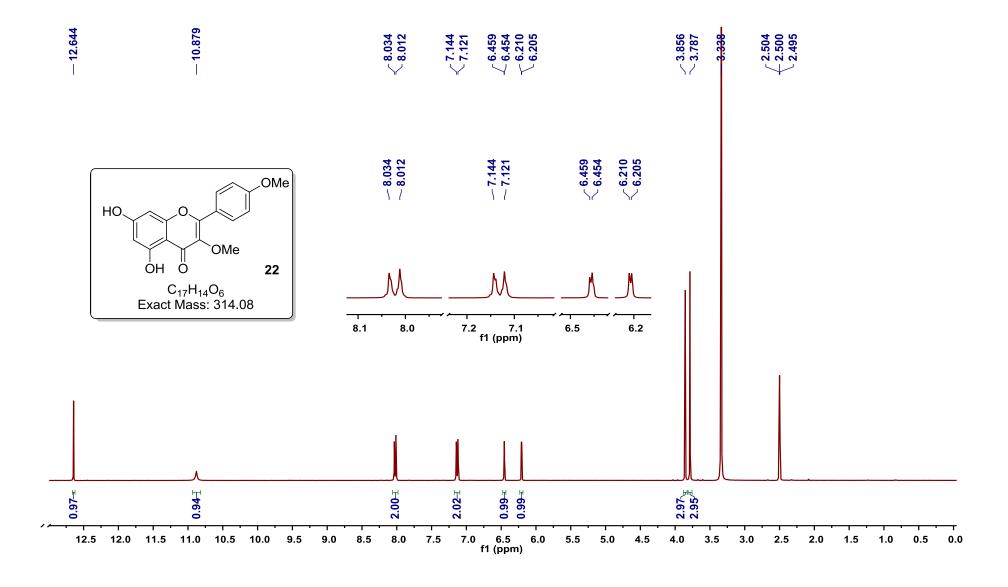


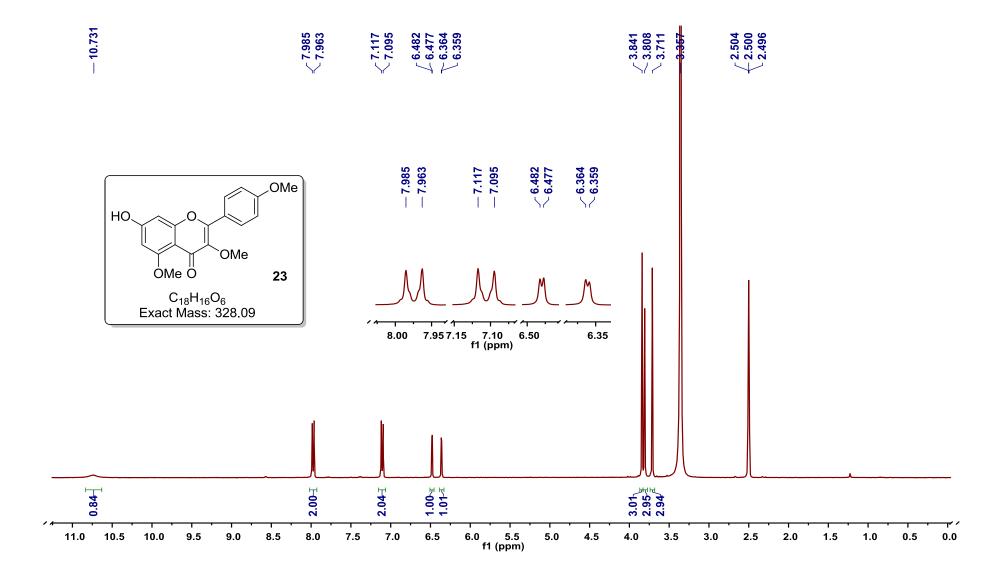


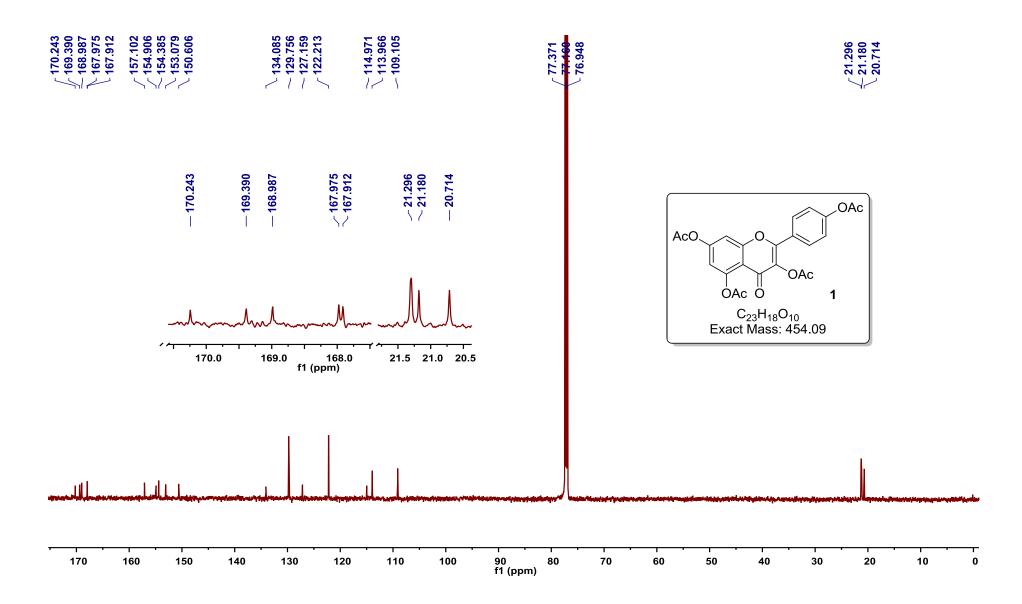


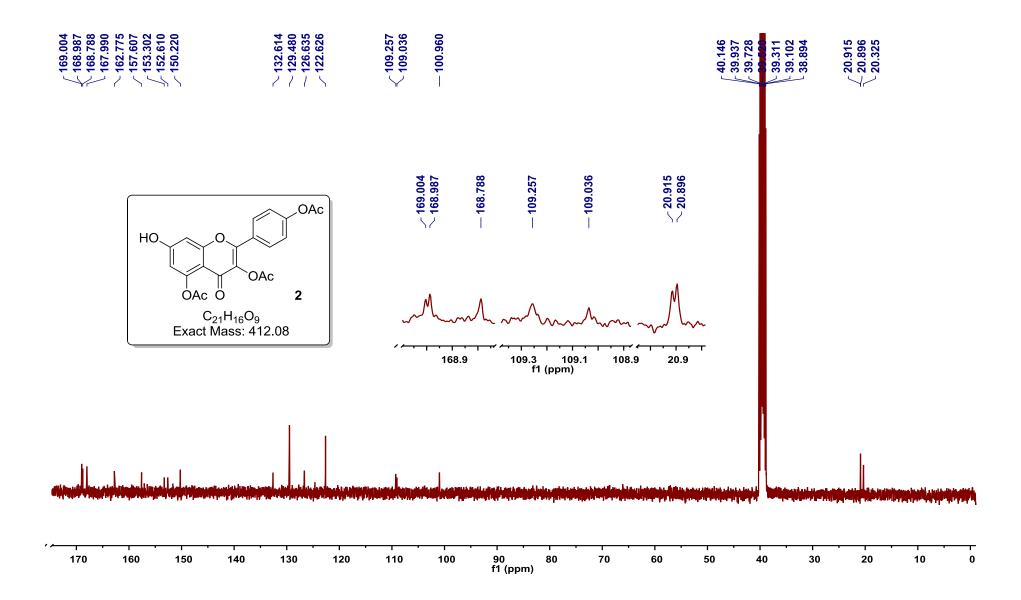


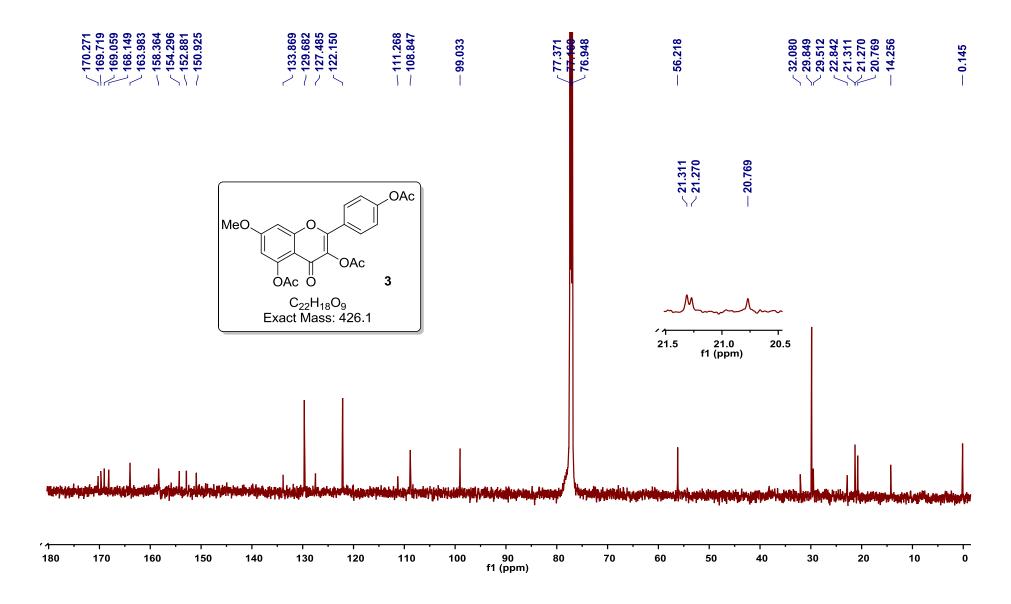


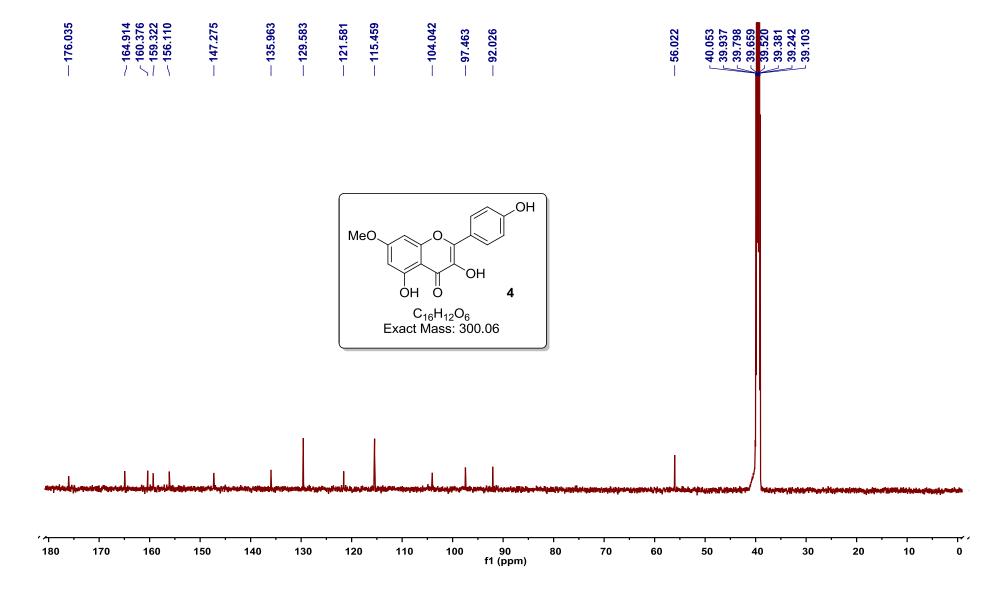


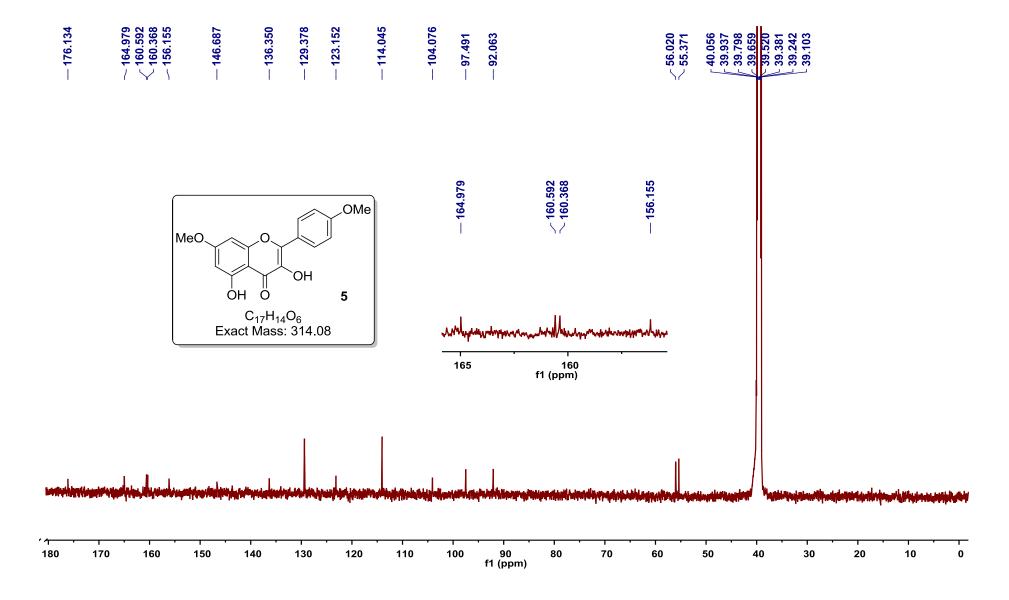


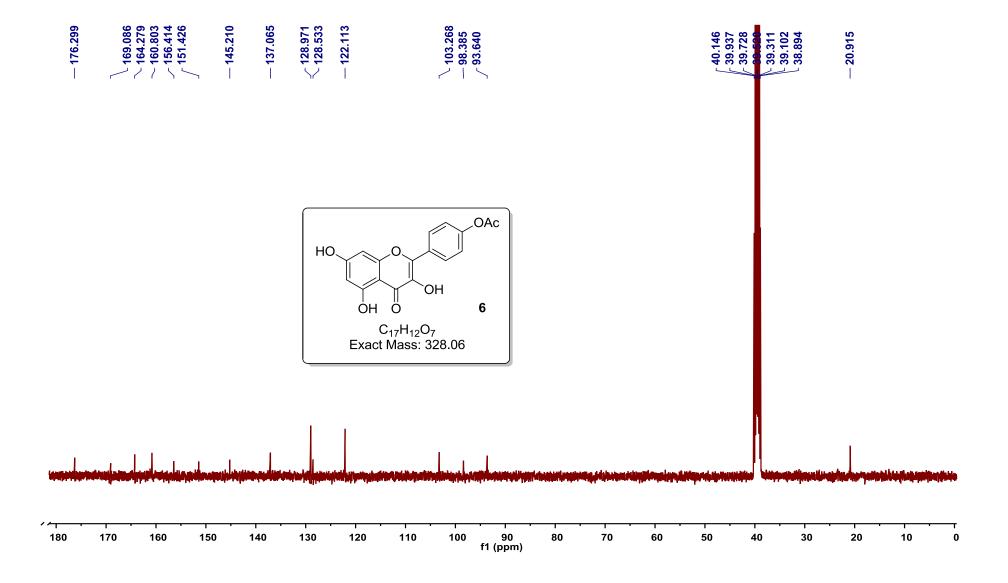


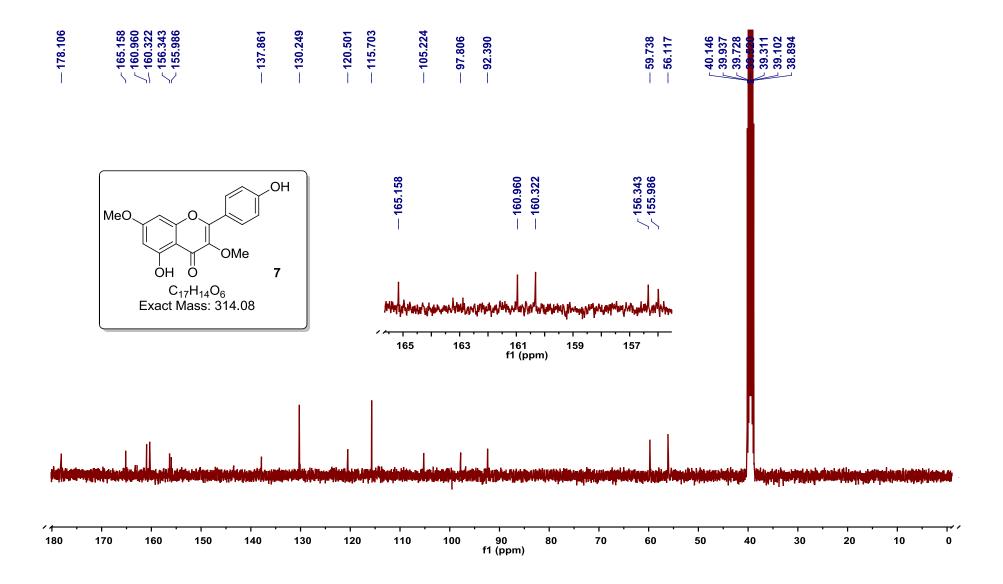


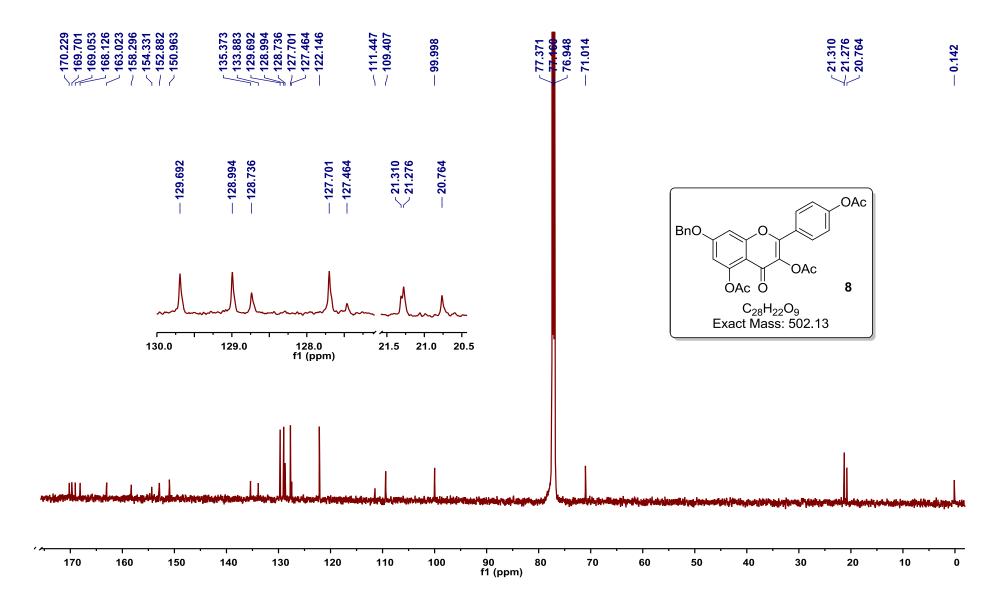


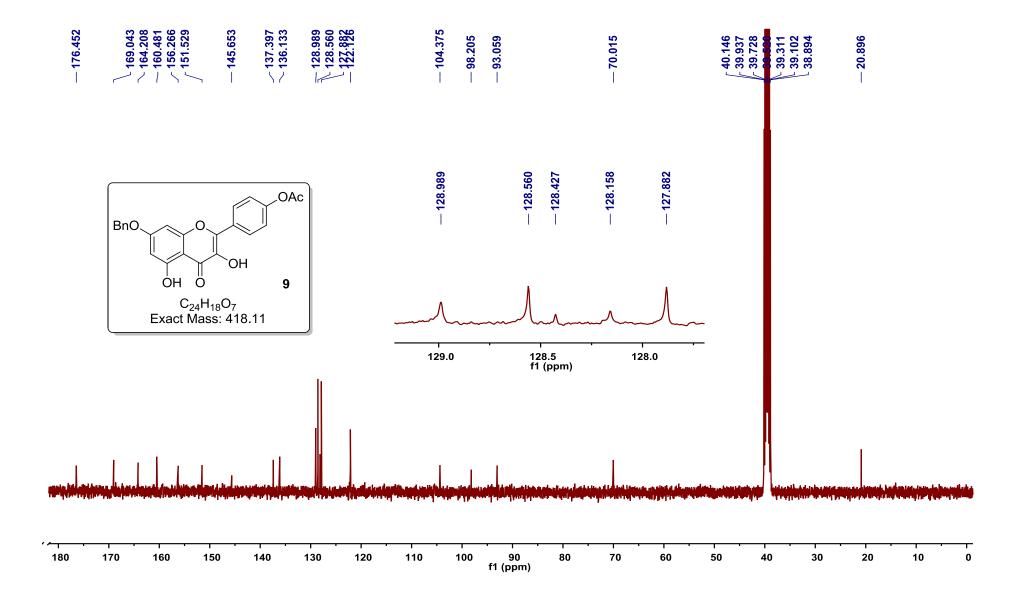


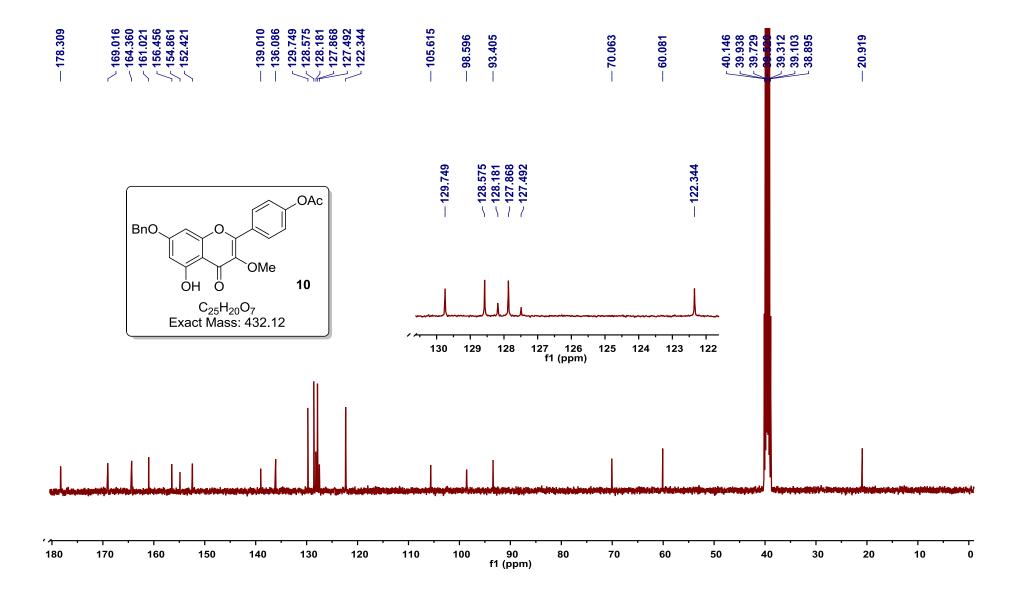


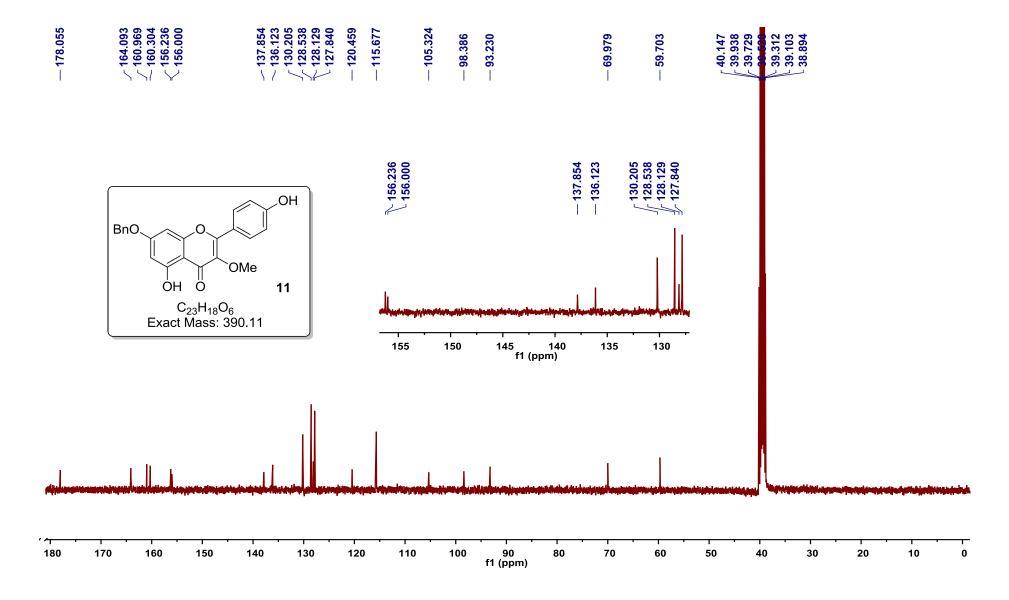


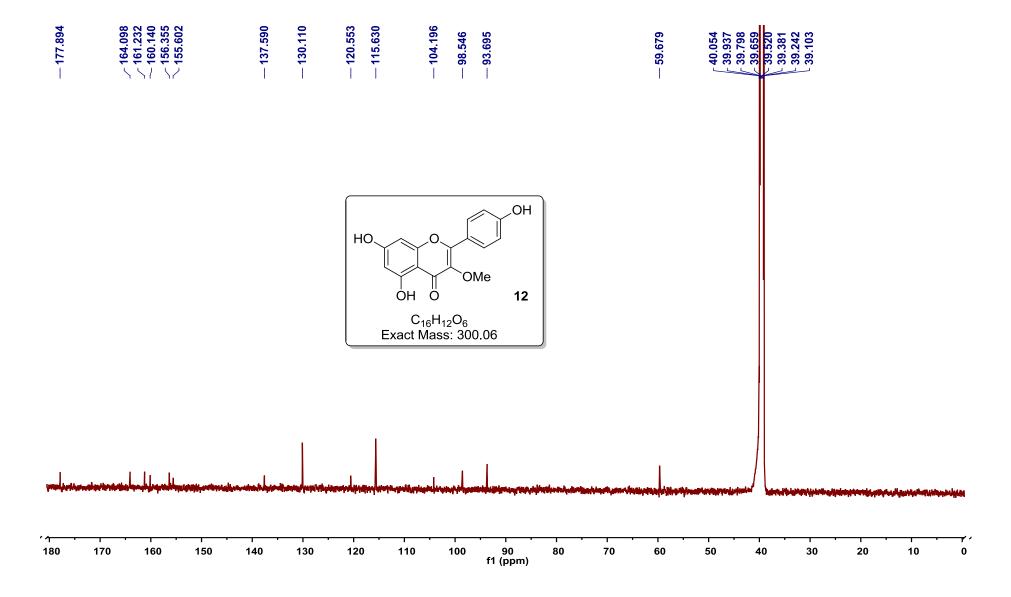


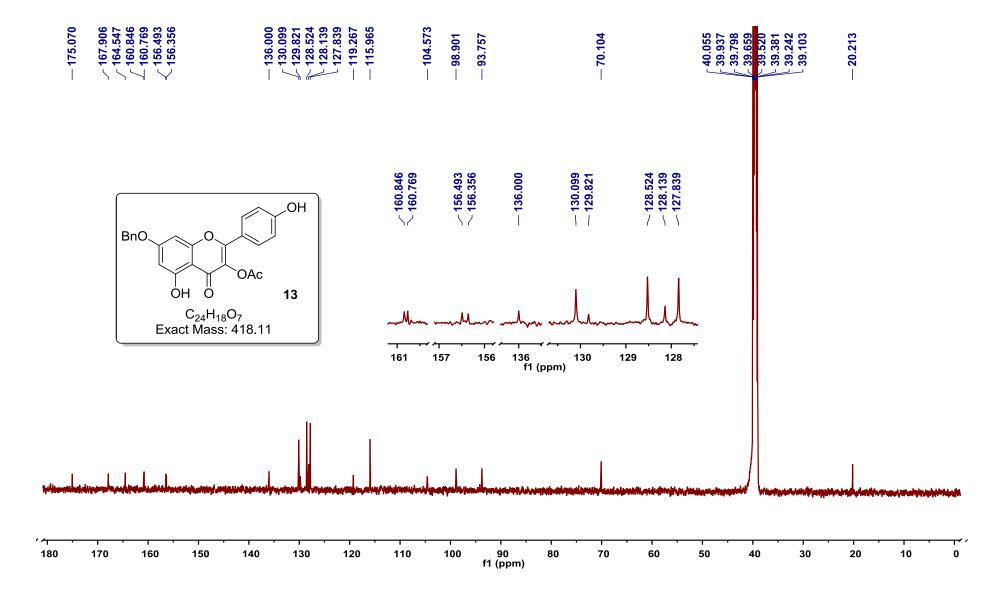


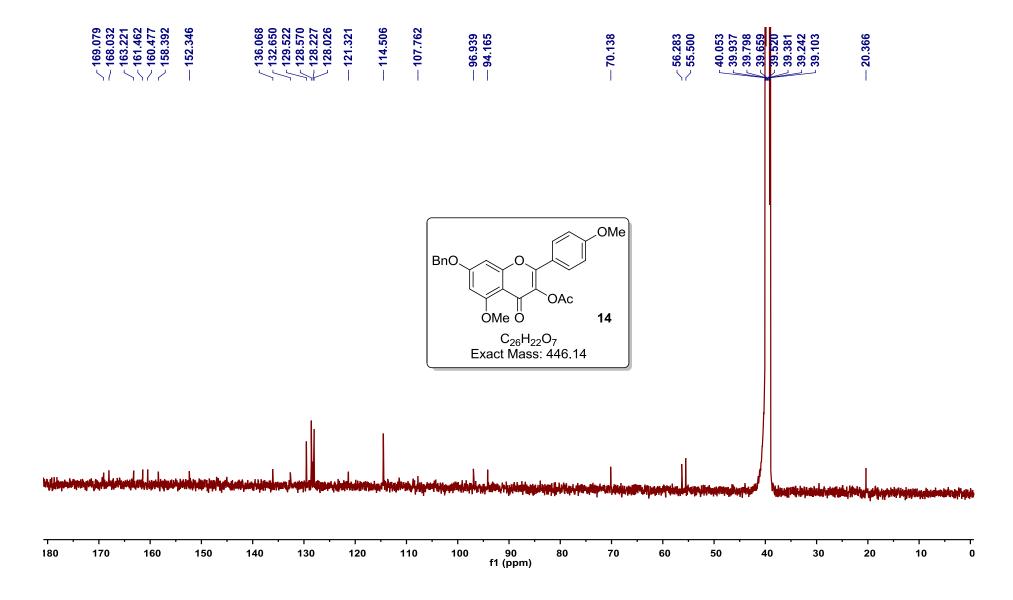


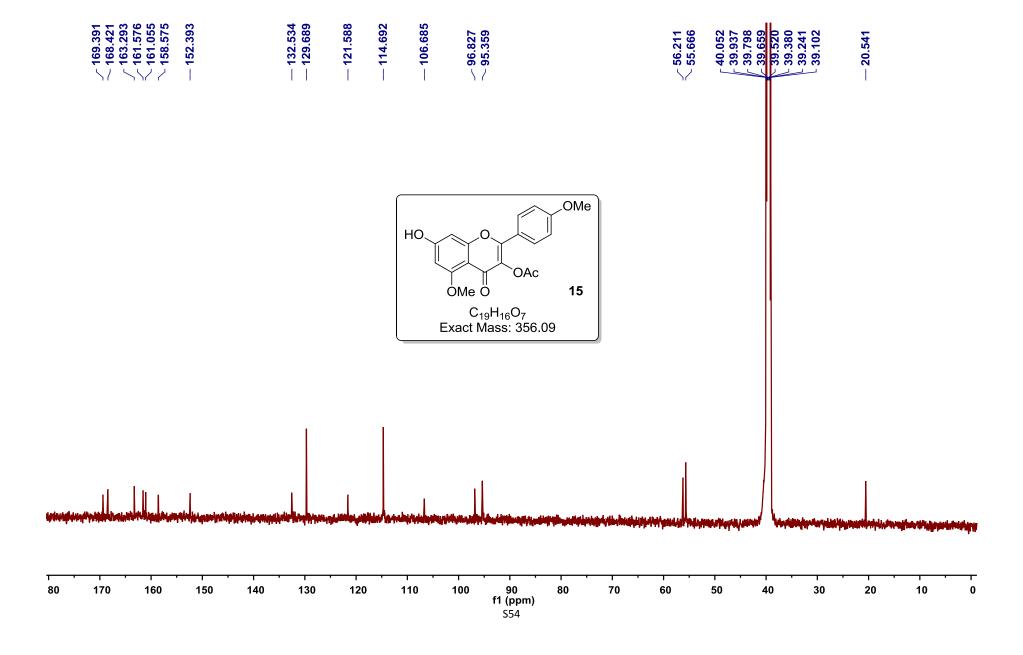


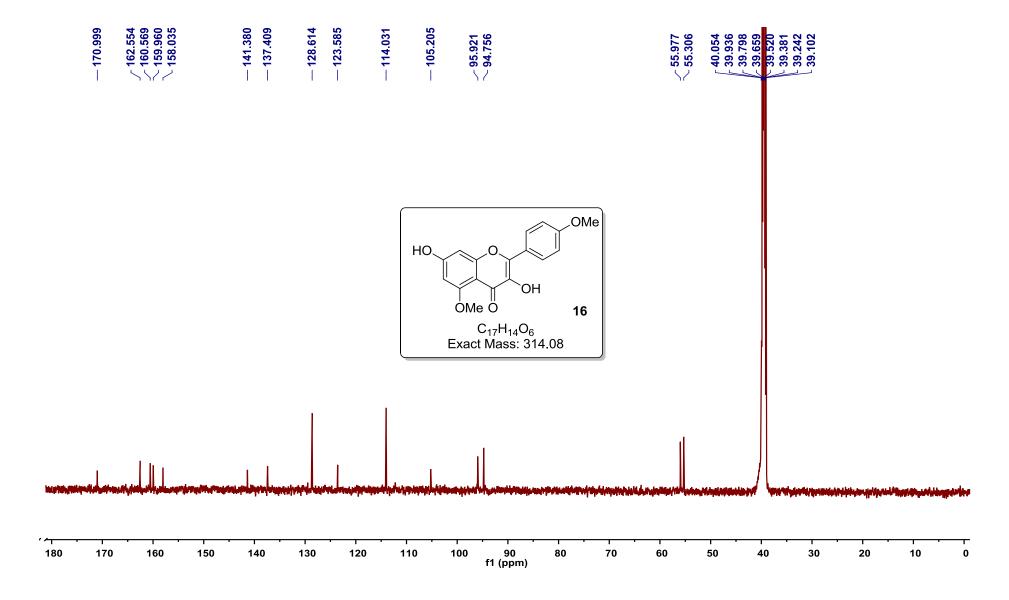


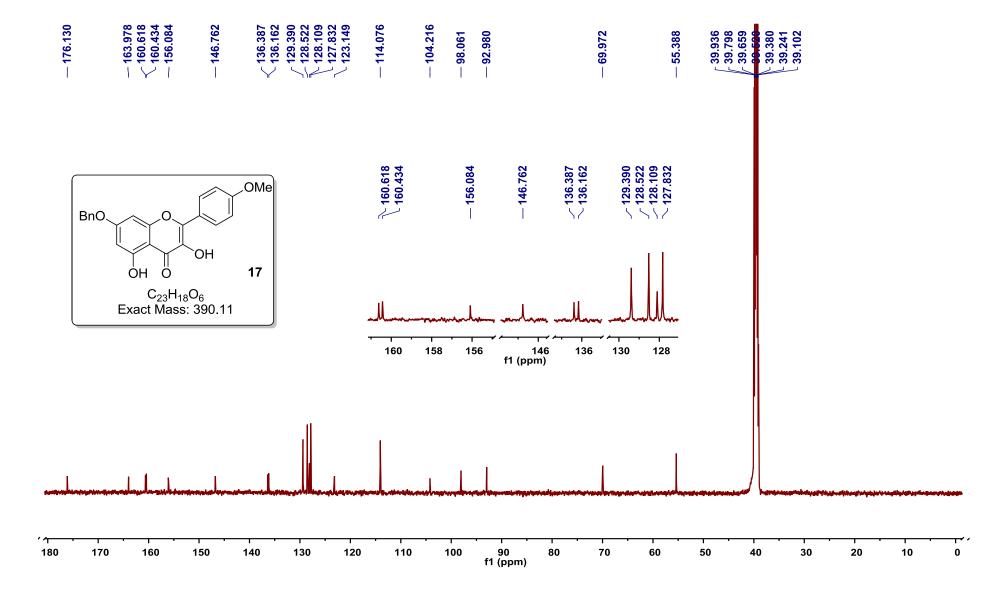


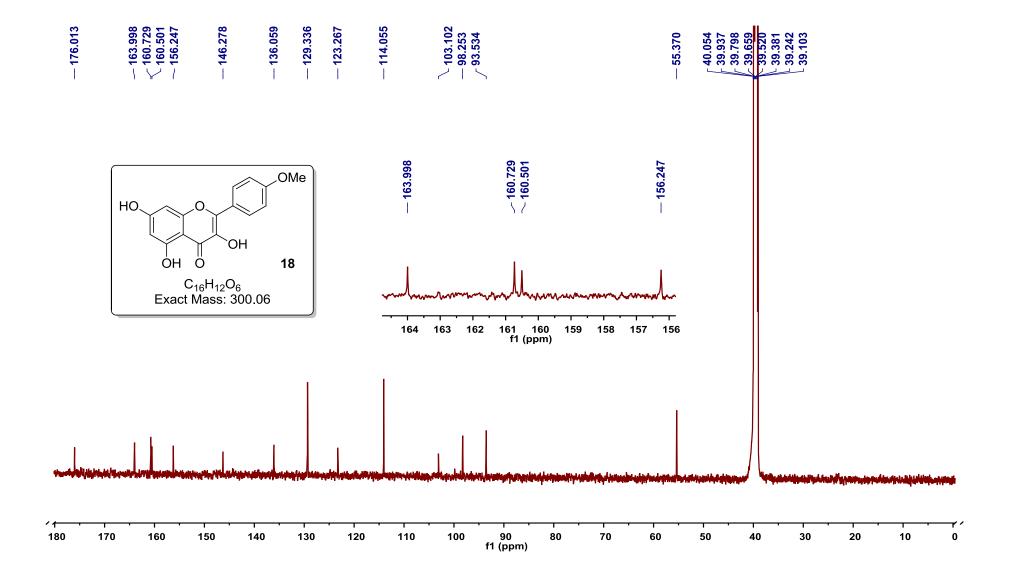


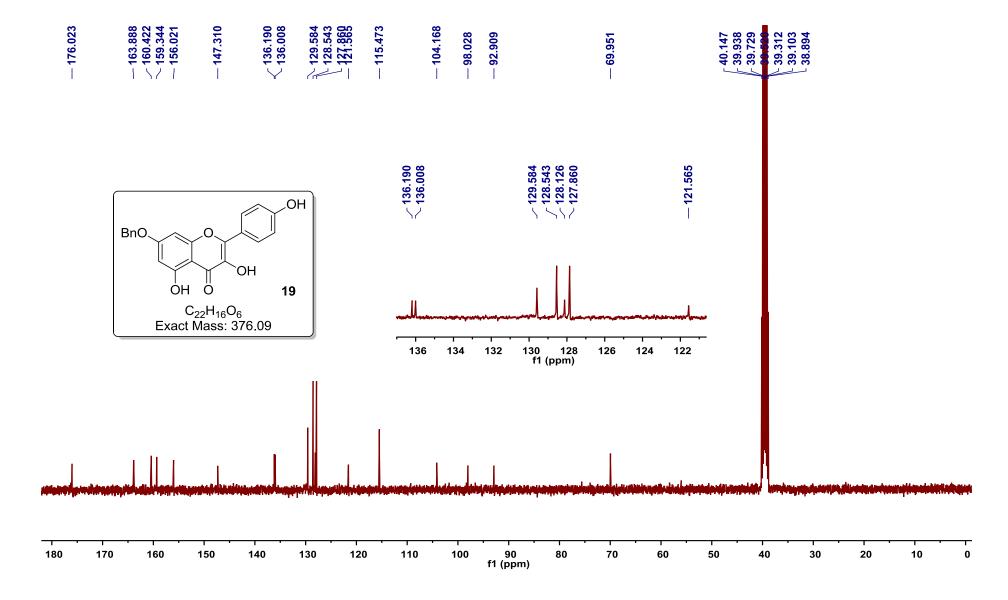


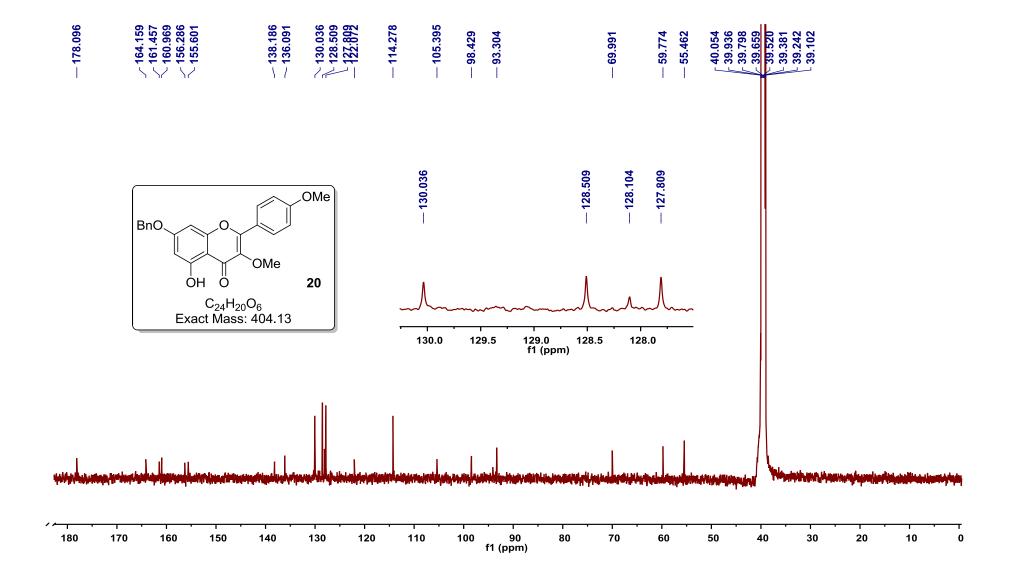


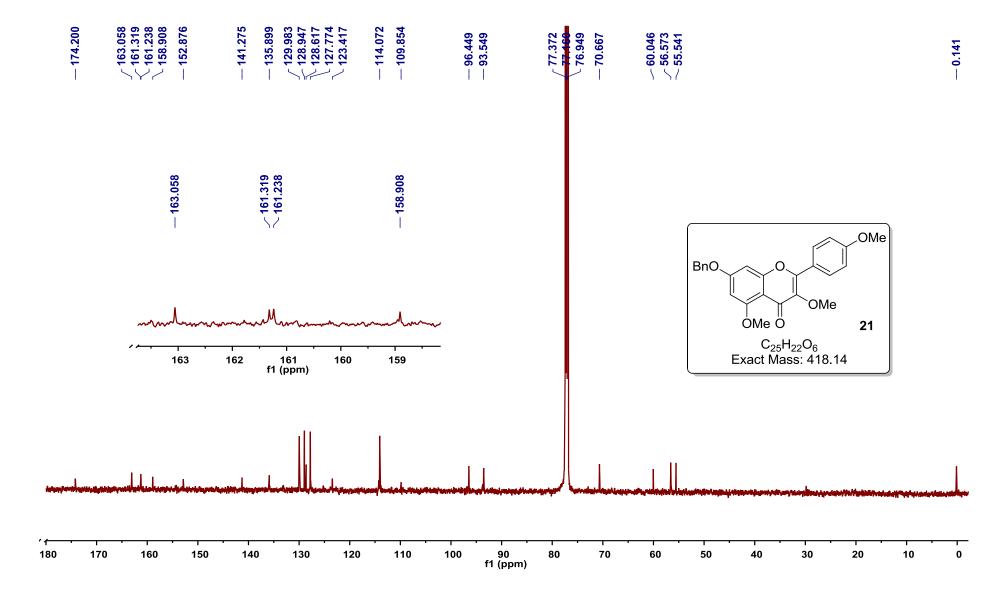


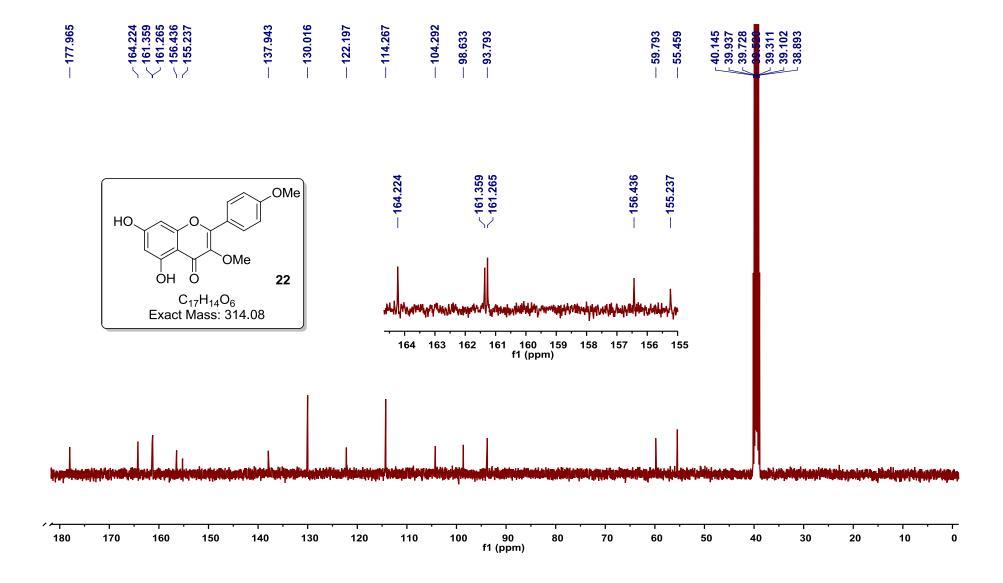


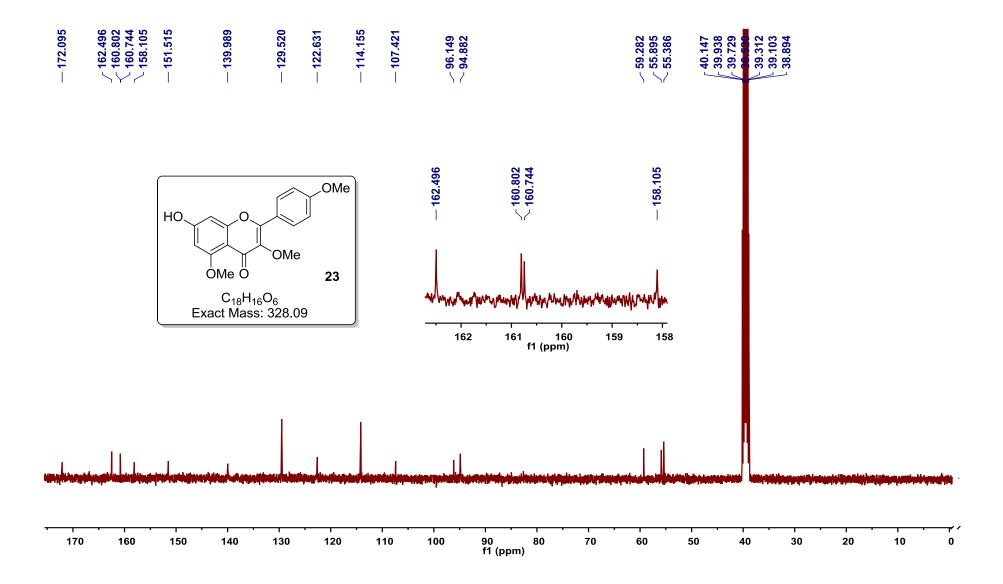


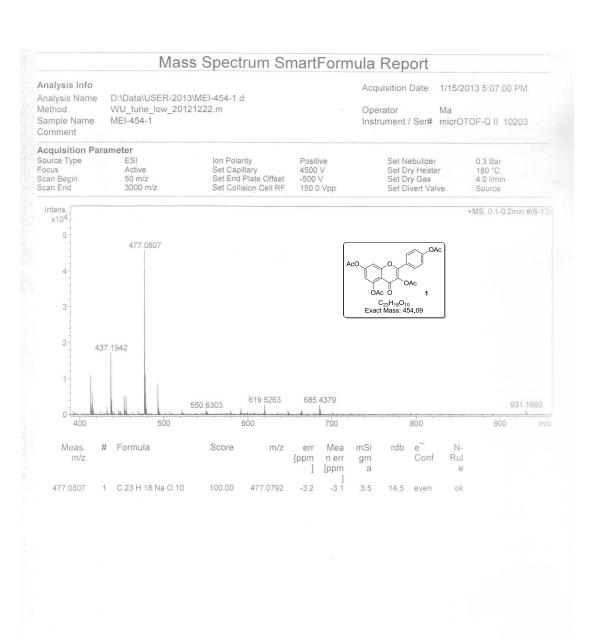












Analysis Info

Analysis Name Method

D:\Data\USER-2014\MEI412+.d

Sample Name Comment

WU_tune_low_20121222.m

MEI412+

Operator Ma

Instrument / Ser# micrOTOF-Q II 10203

Acquisition Date 3/19/2014 3:14:49 PM

Acquisition Parameter

Source Type Focus Scan Begin Scan End

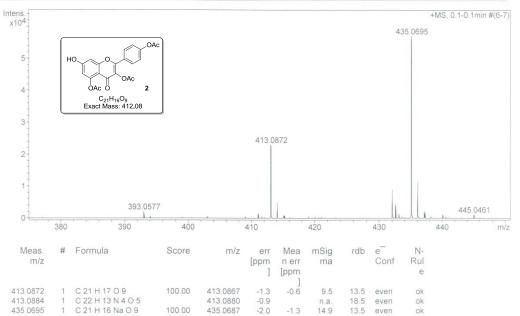
ESI Not active 50 m/z 3000 m/z

lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF

Positive 4500 V -500 V 150.0 Vpp

Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.4 Bar 180 °C 3.0 I/min Source



Analysis Info

D:\Data\USER-2014\mei83008.d WU_tune_low_20121222.m mei83008 Analysis Name Method

Sample Name

Comment

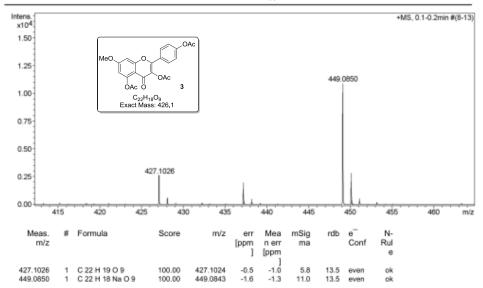
Acquisition Date 9/18/2014 10:25:37 AM

Operator Ma

Instrument / Ser# micrOTOF-Q II 10203



Acquisition Parameter
Source Type ES
Focus Not
Scan Begin 30
Scan End 300 lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve ESI Not active 30 m/z 3000 m/z Positive 4500 V -500 V 150.0 Vpp 0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

printed: 9/23/2014 9:37:33 AM

Analysis Info

D:\Data\USER-2014\0619.d

Analysis Name Method WU_tune_low_20121222.m 0619

Sample Name Comment

Acquisition Date 9/27/2014 1:02:53 PM

Operator Ma Instrument / Ser# micrOTOF-Q II 10203



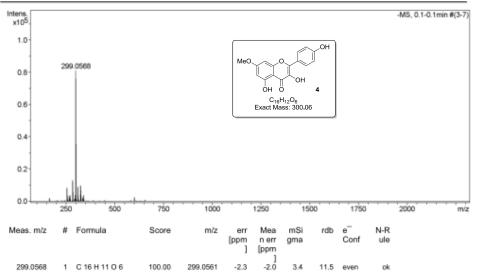
 Acquisition Parameter

 Source Type
 ESI

 Focus
 Not active

 Scan Begin
 30 m/z

 Scan End
 3000 m/z
 lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Set Nebulizer Set Dry Healer Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source Negative 4000 V -500 V 150.0 Vpp



Bruker Compass DataAnalysis 4.0

printed: 9/27/2014 1:17:49 PM

Analysis Info

Analysis Name

Method

D:\Data\USER-2014\MEI314-2.d WU_tune_low_20121222.m MEI314-2

Sample Name

Comment

Acquisition Date 3/26/2014 2:54:50 PM

Operator

Instrument / Ser# micrOTOF-Q II 10203

Acquisition Parameter

Source Type Focus Scan Begin Scan End

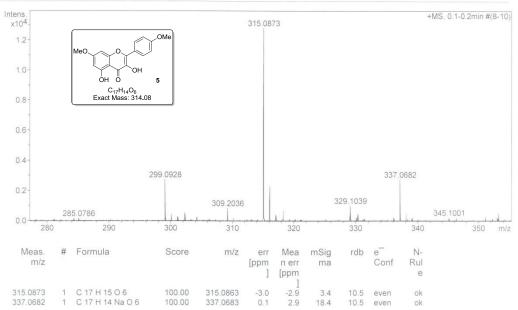
ESI Not active 50 m/z 3000 m/z

lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF

Positive 4500 V -500 V 150.0 Vpp

Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.4 Bar 180 °C 3.0 I/min



Analysis Info

D:\Data\USER-2014\mei328.d WU_tune_low_20121222.m mei328

Analysis Name Method Sample Name Comment

Acquisition Date 4/23/2014 4:51:34 PM

Operator Ma Instrument / Ser# micrOTOF-Q II 10203

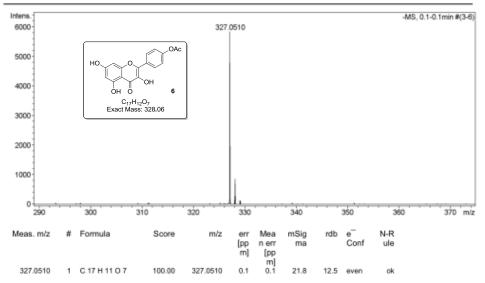
 Acquisition Parameter

 Source Type
 ESI

 Focus
 Not active

 Scan Begin
 50 m/z

 Scan End
 3000 m/z
 Negative 4000 V -500 V 150.0 Vpp 0.4 Bar 180 °C 3.0 l/min Source Ion Polarity Set Nebulizer Set Capillary Set End Plate Offset Set Collision Cell RF Set Dry Heater Set Dry Gas Set Divert Valve



Bruker Compass DataAnalysis 4.0

printed: 4/23/2014 5:24:54 PM

Analysis Info

D:\Data\USER-2014\mei0606.d WU_tune_low_20121222.m

Analysis Name Method

Sample Name

Comment

Acquisition Date 9/17/2014 2:53:23 PM

Operator Ma

Instrument / Ser# micrOTOF-Q II 10203

 Acquisition Parameter

 Source Type
 ESI

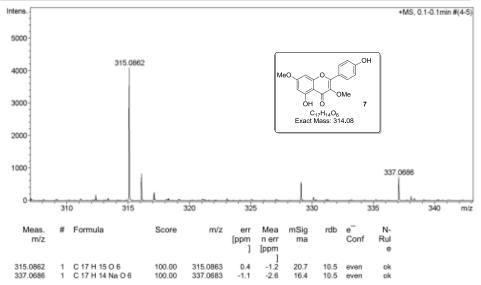
 Focus
 Not active

 Scan Begin
 30 m/z

 Scan End
 3000 m/z

lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

printed: 9/17/2014 3:39:36 PM

Analysis Info

D:\Data\USER-2013\M502.d Analysis Name Method

WU_tune_low_20121222.m

Sample Name Comment

Acquisition Date 3/11/2013 4:35:28 PM

Operator

Ма

even

Instrument / Ser# micrOTOF-Q II 10203

Acquisition Parameter

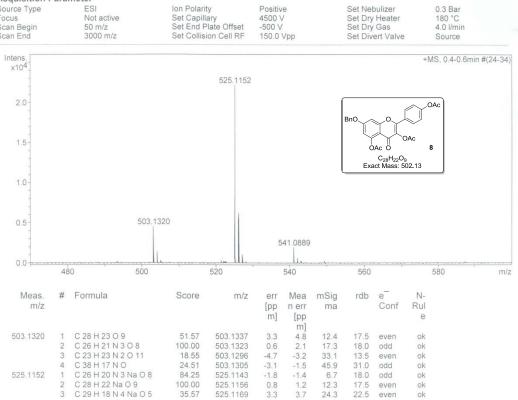
lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Source Type Focus Scan Begin Scan End ESI Not active 50 m/z 3000 m/z

C 29 H 18 N 4 Na O 5

35.57

Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.3 Bar 180 °C 4.0 l/min Source



Bruker Compass DataAnalysis 4.0

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525.1169

3/11/2013 4:59:03 PM

Mass Spectrum SmartFormula Report Analysis Info Acquisition Date 4/10/2014 3:25:02 PM D:\Data\USER-2014\MEITBTS.d WU_tune_low_20121222.m MEITBTS Analysis Name Method Operator Sample Name Instrument / Ser# micrOTOF-Q II 10203 Comment Acquisition Parameter ESI Not active 50 m/z 3000 m/z lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Source Type Focus Scan Begin Scan End Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve 0.3 Bar 180 °C 3.0 I/min Intens. x10⁴ +MS, 0.1-0.2min #(5-12) 419.1115 OAc 1.5 1.0 441.0937 0.5 437.1926 415.2111 432.2387

425

[pp n err

m]

2.5

Mea

[pp m] 2.3

420

419.1125

430

mSig

ma

14.4

435

15.5

Bruker Compass DataAnalysis 4.0

402.3921 406.3516

400

419.1115

405

1 C 24 H 19 O 7

410

415

100.00

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4/10/2014 3:32:24 PM

Page 1 of 1

445

440

N-R

e Conf

450 m/z

Analysis Info

Method

D:\Data\USER-2014\mei0716.d

Analysis Name WU_tune_low_20121222.m

Sample Name Comment

mei0716

Acquisition Date 9/18/2014 10:02:01 AM

Instrument / Ser# micrOTOF-Q II 10203

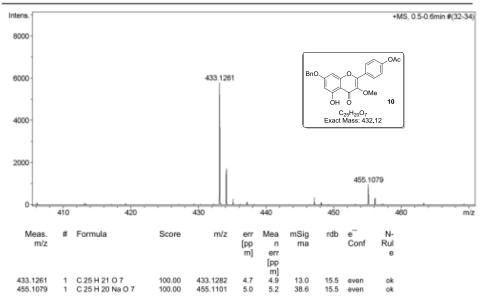
 Acquisition Parameter

 Source Type
 ESI

 Focus
 Not active

 Scan Begin
 30 m/z

 Scan End
 3000 m/z
 lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Set Nebulizer Set Dry Healer Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source Positive 4500 V -500 V 150.0 Vpp



Bruker Compass DataAnalysis 4.0

printed: 9/23/2014 9:17:17 AM

Analysis Info

Analysis Name Method D:\Data\USER-2014\mei37610.d WU_tune_low_20121222.m mei37610

Sample Name

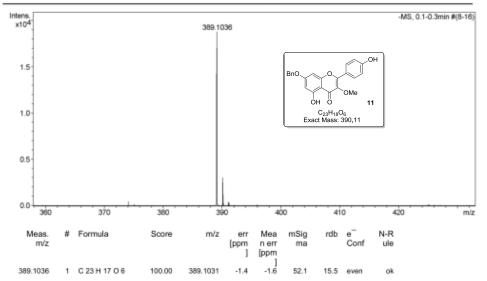
Comment

Acquisition Date 4/23/2014 4:48:20 PM

Operator Ma Instrument / Ser# micrOTOF-Q II 10203



Acquisition Parameter
Source Type ES
Focus Not
Scan Begin 50
Scan End 300 lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source ESI Not active 50 m/z 3000 m/z Negative 4000 V -500 V 150.0 Vpp



Bruker Compass DataAnalysis 4.0

printed: 4/23/2014 5:25:32 PM

Analysis Info

D:\Data\USER-2014\3300.d WU_tune_low_20121222.m Analysis Name Method

Operator

Sample Name 3300 Instrument / Ser# micrOTOF-Q II 10203

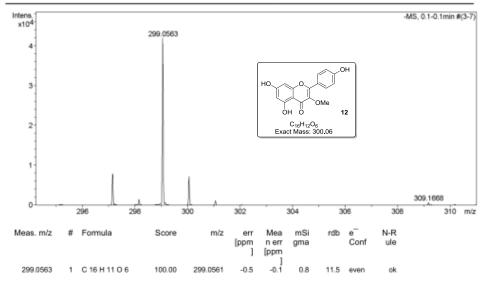
Acquisition Date 9/27/2014 1:04:19 PM

Comment

Acquisition Parameter

Source Type Focus Scan Begin Scan End ESI Not active 30 m/z 3000 m/z Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Negative 4000 V -500 V 150.0 Vpp Set Nebulizer Set Dry Healer Set Dry Gas Set Divert Valve

0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

printed: 9/27/2014 1:13:34 PM

Mass Spectrum SmartFormula Report Analysis Info 3/28/2013 4:28:26 PM Acquisition Date D:\Data\USER-2013\M418.d WU_tune_low_20121222.m Analysis Name Method Operator M418 Sample Name Instrument / Ser# micrOTOF-Q II 10203 Comment Acquisition Parameter lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF ESI Not active 50 m/z 3000 m/z Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve Source Type Focus Scan Begin Scan End Negative 4000 V -500 V 150.0 Vpp 0.3 Bar 180 °C 4.0 I/min -MS, 0.3min #(18) 417.0993 1.5 1.0-C₂₄H₁₈O₇ Exact Mass: 418.11 375.0861 375 350 400 475 500 525 425 450 m/z

Meas. m/z

417.0993

1 C 24 H 17 O 7

m/z

417.0980

100.00

err

-3.2

Mea

n err

[ppm

-3.0

mSi

gma

7.9

rdb

16.5

Analysis Info

Analysis Name Method Sample Name Comment D:\Data\USER-2014\mei51316.d WU_tune_low_20121222.m mei51316

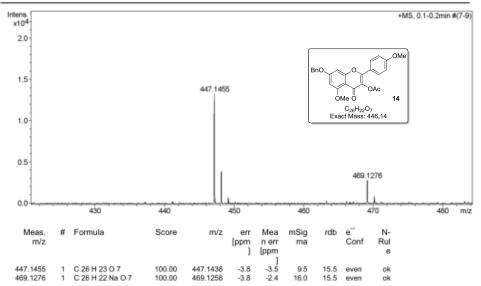
Acquisition Date 9/18/2014 10:17:32 AM

Operator Ma Instrument / Ser# micrOTOF-Q II 10203



Methou
Sample Name
Comment

Acquisition Parameter
Source Type
Sour Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF



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Analysis Info

D:\Data\USER-2014\8328.d WU_tune_low_20121222.m

Analysis Name Method

Sample Name Comment

Acquisition Parameter

Source Type Focus Scan Begin Scan End ESI Not active 30 m/z 3000 m/z

Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

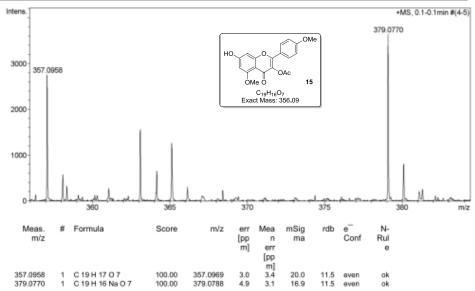
Operator

Acquisition Date 9/27/2014 12:58:26 PM

Ma

Instrument / Ser# micrOTOF-Q II 10203

0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

printed: 9/27/2014 1:22:59 PM

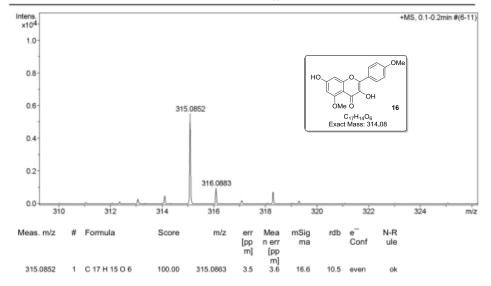
Acquisition Date 9/18/2014 10:08:49 AM

Analysis Info

D:\Data\USER-2014\mei0705.d WU_tune_low_20121222.m mei0705 Analysis Name Method

Operator Ma Instrument / Ser# micrOTOF-Q II 10203 Sample Name Comment

Acquisition Parameter						
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar	
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C	
Scan Begin	30 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min	
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source	



Bruker Compass DataAnalysis 4.0

9/23/2014 9:23:54 AM

Analysis Info Analysis Name

D:\Data\USER-2013\MEI390-2.d WU_tune_low_20121222-negative.m MEI390-2

Sample Name

Method

Acquisition Date 4/16/2013 4:50:18 PM

Ма Instrument / Ser# micrOTOF-Q II 10203

Comment

 Acquisition Parameter

 Source Type
 ESI

 Focus
 Not active

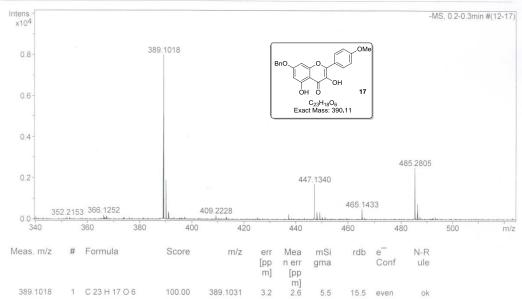
 Scan Begin
 50 m/z

 Scan End
 3000 m/z

Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Negative 4000 V -500 V 150.0 Vpp

Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.3 Bar 180 °C 4.0 l/min Source



Bruker Compass DataAnalysis 4.0

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4/16/2013 4:54:59 PM

Analysis Info

Analysis Name Method

D:\Data\USER-2013\mei300.d WU_tune_low_20121222.m

Sample Name mei300

Comment

Acquisition Date

10/15/2013 11:45:54 AM

Operator

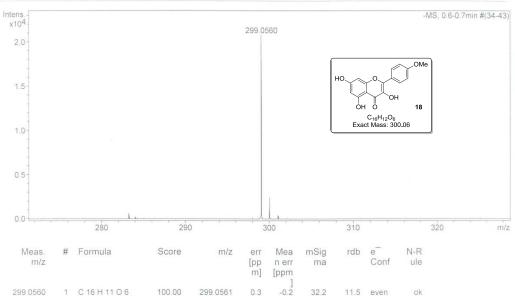
Instrument / Ser# micrOTOF-Q II 10203

Acquisition Parameter
Source Type ESI
Focus Not
Scan Begin 50
Scan End 300 ESI Not active 50 m/z 3000 m/z

Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp

Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.4 Bar 100 °C 3.0 l/min Source



Analysis Info

Analysis Name Method D:\Data\USER-2014\mei39045.d WU_tune_low_20121222.m mei39045

Sample Name

Comment

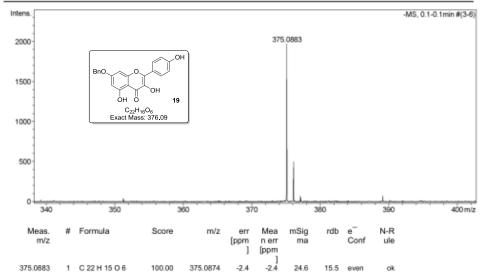
Operator Ma Instrument / Ser# micrOTOF-Q II 10203

Acquisition Date 4/23/2014 4:58:39 PM

Acquisition Parameter

lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Source Type Focus Scan Begin Scan End ESI Not active 50 m/z 3000 m/z Negative 4000 V -500 V 150.0 Vpp

Set Nebulizer Set Dry Healer Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

4/23/2014 5:26:32 PM

Mass Spectrum SmartFormula Report Acquisition Date 4/23/2014 5:21:10 PM

D:\Data\USER-2014\mei37603+.d WU_tune_low_20121222.m mei37603+ Analysis Name Method

Operator Ma

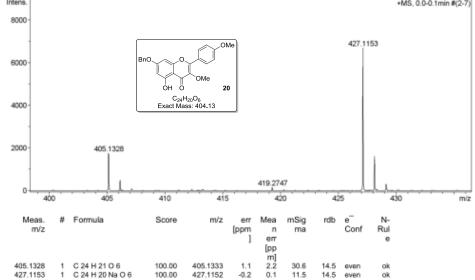
Sample Name

Instrument / Ser# micrOTOF-Q II 10203

Comment

Analysis Info

Acquisition Parameter Source Type Focus Scan Begin Scan End ESI Not active 50 m/z 3000 m/z Ion Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve 0.4 Bar 180 °C 3.0 l/min Source Intens. +MS, 0.0-0.1min #(2-7) 8000



Bruker Compass DataAnalysis 4.0

printed: 4/23/2014 5:27:08 PM

Analysis Info Analysis Name

Method

D:\Data\USER-2014\mei418sj.d

WU_tune_low_20121222.m

Sample Name Comment

mei418sj

Acquisition Date 9/18/2014 10:14:43 AM

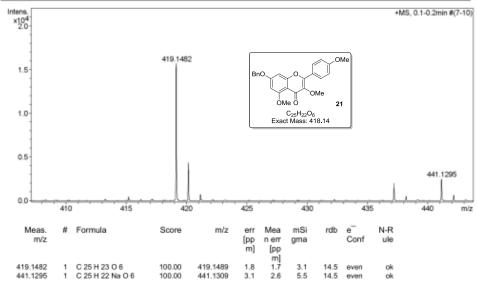
Operator

Instrument / Ser# micrOTOF-Q II 10203

Acquisition Parameter

Source Type Focus Scan Begin Scan End ESI Not active 30 m/z 3000 m/z lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

0.4 Bar 180 °C 3.0 l/min Source



Bruker Compass DataAnalysis 4.0

9/23/2014 9:25:43 AM

Analysis Info

D:\Data\USER-2014\mei3314.d WU_tune_low_20121222.m mei3314

Analysis Name Method

Sample Name Comment

Acquisition Date 9/17/2014 2:58:17 PM

Operator Ma Instrument / Ser# micrOTOF-Q II 10203



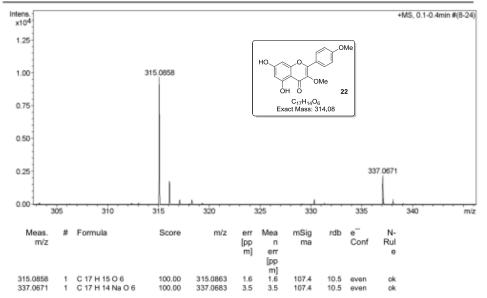
 Acquisition Parameter

 Source Type
 ESI

 Focus
 Not active

 Scan Begin
 30 m/z

 Scan End
 3000 m/z
 Positive 4500 V -500 V 150.0 Vpp 0.4 Bar 180 °C 3.0 l/min Source Ion Polarity Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve Set Capillary Set End Plate Offset Set Collision Cell RF



Bruker Compass DataAnalysis 4.0

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Analysis Info

Analysis Name Method Sample Name Comment

D:\Data\USER-2014\mei9328.d WU_tune_low_20121222.m mei9328

Operator Ma Instrument / Ser# micrOTOF-Q II 10203

Acquisition Date 9/17/2014 2:43:47 PM

 Acquisition Parameter

 Source Type
 ESI

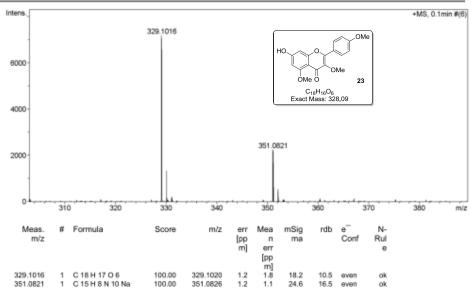
 Focus
 Not active

 Scan Begin
 30 m/z

 Scan End
 3000 m/z

lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF Positive 4500 V -500 V 150.0 Vpp Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve

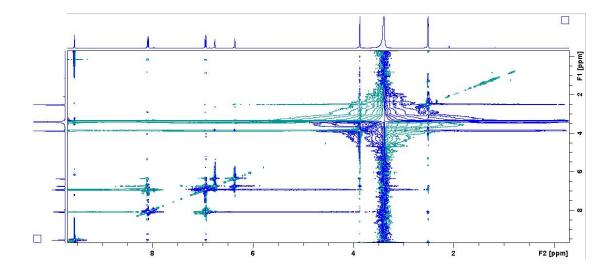
0.4 Bar 180 °C 3.0 l/min Source

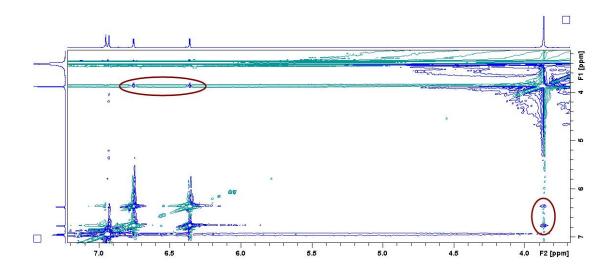


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NOESY Spectrum of Compound 4





NOESY Spectrum of Compound 13

