## **Supporting Information**

## for

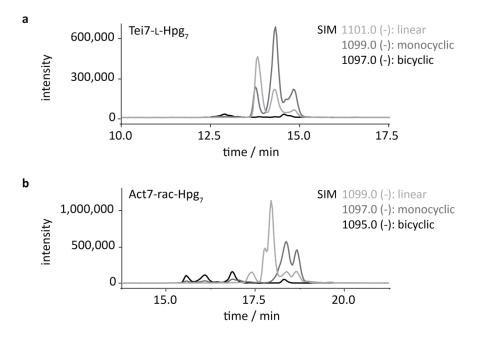
## Biochemical and structural characterisation of the second oxidative crosslinking step during the biosynthesis of the glycopeptide antibiotic A47934 Veronika Ulrich<sup>1</sup>, Clara Brieke<sup>1</sup> and Max J. Cryle\*<sup>1,2,3</sup>

Address: <sup>1</sup>Department of Biomolecular Mechanisms, Max Planck Institute for Medical Research, Jahnstrasse 29, 69120 Heidelberg, Germany <sup>2</sup>EMBL Australia, Monash University, Clayton, Victoria 3800 and <sup>3</sup>The Monash Biomedicine Discovery Institute, Department of Biochemistry and Molecular Biology and ARC Centre of Excellence in Advanced Molecular Imaging, Monash University, Clayton, Victoria 3800.

Email: Max Cryle - max.cryle@monash.edu

\*Corresponding author

## HPLC–MS analysis of StaF turnover activity of Tei7-L-Hpg7 (a) and Act7-rac-Hpg7 (b) bound to MBP-PCP-X<sub>tei</sub>



**Figure S1**: HPLC-MS analysis of StaF turnover activity of Tei7-L-Hpg<sub>7</sub> (a) and Act7-rac-Hpg<sub>7</sub> (b) bound to MBP-PCP-X<sub>tei</sub>. a) Ions corresponding to singly charged, linear (methylamine m/z 1101.0; depicted in light grey), monocyclic (methylamine m/z 1099.0; depicted in grey) and bicyclic Tei7-L-Hpg<sub>7</sub> (methylamine m/z 1097.0; depicted in black) were recorded using single-ion monitoring (SIM) in negative mode. The minor peak of the monocyclic peptide represents Tei7-D-Hpg<sub>7</sub>, which could not be separated completely from Tei7-L-Hpg<sub>7</sub> by preparative HPLC during peptide synthesis. b) Ions corresponding to singly charged, linear (methylamine m/z 1099.0; depicted in light grey), monocyclic (methylamine m/z 1097.0; depicted in grey) and bicyclic Act7-rac-Hpg<sub>7</sub> (methylamine m/z 1095.0; depicted in black) were recorded using single-ion monitoring (SIM) in negative mode. Major peaks for each m/z represent diastereomers due to racemisation of Hpg<sub>7</sub>. a) and b) Smaller peaks are caused through overlapping mass signal detection.