Supporting Information

for

Characterization of non-heme iron aliphatic halogenase WeIO5* from Hapalosiphon welwitschii IC-52-3: Identification of a minimal protein sequence motif that confers enzymatic chlorination specificity in the biosynthesis of welwitindolelinones

Qin Zhu and Xinyu Liu*

Address: Department of Chemistry, University of Pittsburgh, 219 Parkman Avenue, Pittsburgh, PA 15260, USA

Email: Xinyu Liu - xinyuliu@pitt.edu

* Corresponding author

Additional figures



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hapalindoles and we	elwitindolinones isolated from H. welwitschii IC-52	-3 (47g dry tissue)	
Compound	Name	Quantity isolated	
1 (R=none)	12-epi-Hapalindole C isonitrile	11 mg	
1 (R=S)	12-epi-Hapalindole C isothiocyanate	1 mg	
1a (R=none)	12-epi-Hapalindole E isonitrile	93 mg	
1a (R=S)	12-epi-Hapalindole E isothiocyanate	4 mg	
2 (R=none)	12-epi-Fischerindole U isonitrile	4 mg	
2 (R=S)	12-epi-Fischerindole U isothiocyanate	2 mg	
2a (R=none)	12-epi-Fischerindole G isonitrile	4 mg	
3 (R=none)	12-epi-Fischerindole I isonitrile	10 mg	
4 (R=none)	welwitindolinone A isonitirle	2 mg	
5 (R=S, R'=H)	welwitindolinone B isothiocyanate	10 mg	
5 (R=S, R'=Me)	N-methylwelwitindolinone B isothiocyanate	5+12 mg	
6 (R=S, R'=H)	welwitindolinone C isothiocyanate	14 mg	
6 (R=none, R'=Me)	N-methylwelwitindolinone C isonitrile	47mg	
6 (R=S, R'=Me)	N-methylwelwitindolinone C isothiocyanate	110mg	

Figure S1: Summary of relative and absolute quantities of **1** and its biogenetic derivatives **1a** versus **2** and its biogenetic derivatives **2a**, **3–6** from (b) *H. welwitschii* UTEX B1830 and (c) IC-52-3. The relative quantity shown in (b) was derived from the HPLC analysis of *H. welwitschii* UTEX B1830 crude metabolites and the absolute quantity shown in (c) was derived from what was reported in ref [1]. These data were used to generate the relative molar ratio of each metabolite in the two different producers and the comparison graphs shown in Figure 1b/1c in the main text.



<i>wel</i> Gene	Size (aa)	Protein sequence identity to homolog from <i>H. welwitschii</i> UTEX B1830	<i>wel</i> Gene	
welO5	290	95%	<i>wel</i> C3	
<i>wel</i> M	329	99%	we/T5	
<i>wel</i> U2	226	99%	we/T4	
<i>wel</i> O4	359	99%	<i>we/</i> T3	
welO3	362	99%	we/T2	
welO2	357	99%	we/T1	
<i>wel</i> O1	360	99%	welC2	
<i>wel</i> U1	228	97%	we/D3	
welU3	228	Unique to IC523	welD2	
<i>wel</i> D4	400	95%	<i>wel</i> P1	
<i>wel</i> P2	334	99%	we/l3	
<i>wel</i> R1	234	100%	we/l2	
<i>wel</i> R2	243	99%	<i>wel</i> l1	
<i>wel</i> R3	368	100%	<i>wel</i> S1	
			14/0/00	-

<i>wel</i> Gene	Size (aa)	Protein sequence identity to homolog from <i>H.</i> <i>welwitschii</i> UTEX B1830
<i>wel</i> C3	214	100%
welT5	365	99%
<i>weI</i> Т4	415	99%
we/T3	274	99%
welT2	283	99%
<i>wel</i> ∏1	734	99%
<i>wel</i> C2	301	99%
<i>wel</i> D3	408	99%
<i>wel</i> D2	647	99%
<i>wel</i> P1	308	100%
we/l3	272	100%
well2	331	99%
<i>wel</i> l1	319	99%
<i>wel</i> S1	169	Unique to 1830
<i>wel</i> S2	156	Unique to 1830
<i>wel</i> D1	406	99%
<i>wel</i> C1	183	99%

Figure S2: Sequence identity comparison of proteins encoded in the welwitindolinone BGCs from *H. welwitschii* IC-52-3 and UTEX B1830. The names for some of the genes in the BGC from *H. welwitschii* IC-52-3 (including *wel*U1-3, *wel*O1-5 and *wel*M) (ref [2]) were renamed according to those in the BGC in *H. welwitschii* UTEX B1830 (ref [3]).

Ladd (Ka	er)	WelO5*	WelO5- <i>var</i>	
80 60				
50				
40				
30			-	
25				
20				
15				

Figure S3: SDS-PAGE of purified NHis7-tagged WelO5* and WelO5-var.

References

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