Supporting Information

for

Organocatalytic asymmetric allylic amination of Morita–Baylis–Hillman carbonates of isatins

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General procedures and analytical data

Table of Contents

1. General methods	S2
2. Preparation of modified β-ICD-type catalysts	S2
3. General procedure for assembly of MBH carbonates and <i>N</i> -silyloxycarbamates	S3
4. Synthetic transformations of multifunctional adduct 4d	S7
5. Crystal data and structure refinement for enantiopure 5	S9
6. NMR spectra and HPLC chromatograms	S 11
7. References	S39

1. General Methods

NMR spectra were recorded with tetramethylsilane as the internal standard. TLC was performed on glass-backed silica plates. Column chromatography was performed using silica gel (200–300 mesh) eluting with ethyl acetate and petroleum ether (PE) (EtOAc/PE). ¹H NMR spectra were recorded at 400 MHz (Varian) and ¹³C NMR spectra were recorded at 100 MHz (Varian). Chemical shifts are reported in ppm downfield from CDCl₃ ($\delta = 7.27$ ppm) for ¹H NMR and relative to the central CDCl₃ resonance ($\delta = 77.0$ ppm) for ¹³C NMR spectroscopy. Coupling constants are given in hertz (Hz). Optical rotations were measured at 589 nm at 20 °C. Enantiomeric excess was determined by HPLC analysis on Chiralpak IC and Chiralcel OD columns. Et₂O was distilled from sodium (Na) under an argon (Ar) atmosphere. Mesitylene was distilled from CaH₂. Chlorobenzene was dried by 4 Å. All other chemicals were used as commercially available, without purification. Cinchona alkaloids catalysts **1a**, **1b**, **1d** and **1e–1h** were prepared according to the literature procedure [1]. Catalysts **1c** was prepared according to the literature procedure [2]. Morita–Baylis–Hillman carbonates of isatins [3] and N-silyloxycarbamates were prepared according to the literature procedure [4].

2. Preparation of modified β-ICD-type catalysts



To a tube charged with $Pd(PPh_3)_4$ (21.9 mg, 0.02 mmol), K_3PO_4 (201.7 mg, 0.95 mmol,), **1i** (168.0 mg, 0.38 mmol) [1] and 4-tert-butylphenylboronic acid (101.5 mg, 0.57 mmol) was added a solution of H_2O (0.4 mL) and toluene (1.2 mL) via syringe under a nitrogen atmosphere. The mixture was heated at 80 °C until **1i** had been consumed according to TLC analysis. The mixture was cooled to room temperature, diluted with ethyl acetate, filtered, and concentrated. The crude material was purified by flash chromatography on silica gel (DCM/MeOH = 20:1) to give **1h** as

white solid (100.3 mg, 62%). $[\alpha]_D^{20} = +46.1$ (c = 0.8 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 8.92$ (d, J = 4.8 Hz, 1H), 8.18 (d, J = 9.2 Hz, 1H), 8.12 (s, 1H), 7.96 (dd, J = 8.8 Hz, 1.6 Hz, 1H), 7.78 (d, J = 4.4 Hz, 1H), 7.69 (d, J = 8.4 Hz, 2H), 7.52 (d, J = 8.4 Hz, 2H), 6.12 (s, 1H), 3.64–3.58 (m, 2H), 3.04–3.02 (m, 2H), 2.71 (d, J = 13.6 Hz, 1H), 2.18 (t, J = 4.8 Hz, 1H), 1.80 (qd, J = 6.8 Hz, 2.0 Hz, 1H), 1.73–1.67 (m, 3H), 1.65–1.53 (m, 1H), 1.37 (s, 9H), 1.32–1.26 (m, 1H), 1.05 (t, J = 7.6 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 150.9$, 149.9, 147.2, 144.4, 139.6, 130.7, 128.9, 127.4, 126.0, 125.7, 119.9, 119.5, 110.7, 77.1, 73.0, 56.8, 54.8, 46.7, 34.6, 32.8, 31.3, 27.4, 24.0, 23.4, 7.3 ppm; ESI–HRMS: calcd. for C₂₉H₃₄N₂O + H 427.2749, found 427.2749.



1f, 70% yield; $[\alpha]_D^{20} = +11.2$ (*c* = 1.0 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 8.94$ (d, *J* = 4.4 Hz, 1H), 8.24–8.20 (m, 2H), 7.91 (dd, *J* = 8.8 Hz, 2.0 Hz, 1H), 7.80–7.78 (m, 3H), 7.68 (d, *J* = 8.0 Hz, 2H), 6.18 (s, 1H), 3.60–3.57 (m, 2H), 3.10–3.00 (m, 2H), 2.66 (d, *J* = 13.6 Hz, 1H), 2.17 (s, 1H), 1.81–1.77 (m, 1H), 1.70–1.62 (m, 3H), 1.57–1.51 (m, 1H), 1.30–1.24 (m, 1H), 1.00 (t, *J* = 7.6 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta =$

150.5, 147.6, 144.6, 143.7, 138.1, 133.9, 131.1, 128.6, 128.0, 125.8, 125.8, 125.7, 124.0, 120.9, 119.8, 77.0, 72.8, 56.9, 54.5, 46.6, 32.8, 27.3, 23.9, 23.3, 7.2 ppm; ESI–HRMS: calcd. for $C_{26}H_{25}F_{3}N_{2}O + H$ 439.1997, found 439.1996.



1g, 50% yield; $[\alpha]_D^{20} = +9.1$ (*c* = 0.9 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 8.93$ (d, *J* = 4.4 Hz, 1H), 8.19–8.17 (m, 2H), 7.95 (dd, *J* = 8.8 Hz, 1.6 Hz, 1H), 7.78 (d, *J* = 4.0 Hz, 1H), 7.38 (s, 2H), 7.04 (s, 1H), 6.18 (s, 1H), 3.79–3.75 (m, 2H), 3.11 (s, 2H), 2.79 (d, *J* = 13.6 Hz, 1H), 2.43 (s, 6H), 2.24 (s, 1H), 1.90–1.86 (m, 1H), 1.78–1.69 (m, 3H), 1.65–1.62 (m,

1H), 1.38–1.33 (m, 1H), 1.07 (t, J = 7.6 Hz, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 149.8$, 147.3, 143.2, 140.5, 140.0, 138.5, 132.0, 130.7, 129.6, 129.5, 128.5, 125.9, 125.6, 119.9, 119.5, 77.2, 72.2, 57.1, 54.1, 46.1, 32.9, 27.3, 23.1, 22.7, 21.4, 7.3 ppm; ESI–HRMS: calcd. for $C_{27}H_{30}N_2O + H$ 399.2436, found 399.2437.

3. General procedure for assembly of MBH carbonates and N-silyloxycarbamates

To a solution of MBH carbonate 2a (41.6 mg, 0.12 mmol), N-silyloxycarbamates 3d (28.1 mg, 0.1

mmol) in chlorobenzene (0.5 mL) at 0 °C, catalyst **1h** (4.3 mg, 10 mol %) was added and the resulting mixture was kept at the temperature until the consumption of **3d**, as monitored by TLC analysis. Purification by flash chromatography on silica gel (AcOEt/petroleum ether = 1:15) gave **4d** as a reddish brown oil (47.0 mg, 92% yield).



4d, 92% yield; $[\alpha]_D^{20} = -100.8$ (c = 0.5 in CHCl₃); 91% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 9.463 min, *t* (minor) = 13.188 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.76$ (d, J = 6.8 Hz, 1H), 7.32–7.27 (m, 6H),

7.01 (td, J = 7.6 Hz, 0.8 Hz, 1H), 6.78 (d, J = 7.6 Hz, 1H), 6.08 (s, 1H), 5.57 (s, 1H), 5.19 (d, J = 11.6 Hz, 1H), 5.11 (d, J = 12.0 Hz, 1H), 3.67 (s, 3H), 3.20 (s, 3H), 0.59 (s, 9H), 0.04 (s, 3H), -0.15 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.2$, 165.1, 159.9, 143.8, 138.1, 135.1, 129.5, 128.8, 128.3, 128.2, 128.2, 126.9, 124.9, 122.6, 108.3, 68.5, 51.8, 26.4, 25.6, 17.9, -4.3, -4.6 ppm; ESI–HRMS: calcd. for C₂₇H₃₄N₂O₆Si + Na 533.2084, found 533.2089.



4f, 93% yield; $[\alpha]_D^{20} = -64.8$ (c = 1.1 in CHCl₃); 91% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 10.204 min, *t* (minor) = 14.245 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.57$ (s, 1H), 7.34–7.31 (s, 5H), 7.07

(d, J = 7.6 Hz, 1H), 6.67 (d, J = 7.6 Hz, 1H), 6.06 (s, 1H), 5.55 (s, 1H), 5.20 (d, J = 12.0 Hz, 1H), 5.12 (d, J = 12.0 Hz, 1H), 3.67 (s, 3H), 3.19 (s, 3H), 2.29 (s, 3H), 0.59 (s, 9H), 0.03 (s, 3H), -0.15 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.2$, 165.1, 159.9, 141.4, 138.3, 135.1, 132.1, 129.6, 128.8, 128.3, 128.2, 128.2, 127.8, 124.6, 108.0, 68.4, 51.8, 26.4, 25.6, 21.1, 17.9, -4.3, -4.7 ppm; ESI–HRMS: calcd. for C₂₈H₃₆N₂O₆Si + Na 547.2240, found 547.2239.



4g, 97% yield; $[\alpha]_D^{20} = -64.0$ (*c* = 1.0 in CHCl₃); 94% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 13.032 min, *t* (minor) = 18.202 min]; ¹H NMR (400 MHz, CDCl₃):

δ = 7.46 (d, *J* = 2.8 Hz, 1H), 7.31 (s, 5H), 6.81 (dd, *J* = 8.4 Hz, 2.4 Hz, 1H), 6.68 (d, *J* = 8.4 Hz, 1H), 6.06 (s, 1H), 5.56 (s, 3H), 5.19 (d, *J* = 11.6 Hz, 1H), 5.11 (d, *J* = 11.6 Hz, 1H), 3.74 (s, 3H),

3.66 (s, 3H), 3.17 (s, 3H), 0.60 (s, 9H), 0.05 (s, 3H), -0.10 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.0$, 165.0, 159.8, 155.8, 138.1, 137.2, 135.1, 129.4, 128.8, 128.2, 128.2, 124.9, 114.2, 114.0, 108.6, 75.6, 68.4, 55.7, 51.8, 26.4, 25.6, 17.9, -4.2, -4.5 ppm; ESI–HRMS: calcd. for C₂₈H₃₆N₂O₇Si + Na 563.2189, found 563.2187.



4h, 93% yield; $[\alpha]_D^{20} = -60.0$ (*c* = 0.8 in CHCl₃); 90% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 12.702 min, *t* (minor) = 15.971 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.41$ (s, 1H), 7.33 (s, 5H), 6.81 (s, 1H),

6.06 (s, 1H), 5.54 (s, 1H), 5.18 (d, J = 12.0 Hz, 1H), 5.12 (d, J = 11.6 Hz, 1H), 3.66 (s, 3H), 3.44 (s, 3H), 2.46 (s, 3H), 2.23 (s, 3H), 0.62 (s, 9H), 0.04 (s, 3H), -0.14 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.1$, 165.1, 160.0, 139.1, 138.5, 135.2, 133.6, 131.9, 129.1, 128.8, 128.2, 128.1, 125.5, 124.8, 119.4, 74.5, 68.4, 51.8, 29.9, 25.6, 20.8, 18.9, 17.9, -4.2, -4.7 ppm; ESI–HRMS: calcd. for C₂₉H₃₈N₂O₆Si + Na 561.2397, found 561.2394.



4i, 86% yield; $[\alpha]_D^{20} = -64.3$ (*c* = 1.1 in CHCl₃); 90% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 7.288 min, *t* (minor) = 8.890 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.58$ (dd, J = 8.4 Hz, 2.4 Hz, 1H), 7.32 (s,

5H), 6.99 (td, J = 8.4 Hz, 2.4 Hz, 1H), 6.70 (dd, J = 8.4 Hz, 4.0 Hz, 1H), 6.10 (s, 1H), 5.60 (s, 1H), 5.19 (d, J = 12.0 Hz, 1H), 5.11 (d, J = 12.0 Hz, 1H), 3.68 (s, 3H), 3.20 (s, 3H), 0.61 (s, 9H), 0.04 (s, 3H), -0.10 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.1$, 164.9, 160.0 (d, ${}^{I}J_{C,F} = 239.1$ Hz), 159.6, 139.8, 137.6, 134.9, 129.8, 128.9, 128.3, 128.3, 125.4, 115.6 (d, ${}^{2}J_{C,F} = 23.5$ Hz), 115.4 (d, ${}^{2}J_{C,F} = 26.1$ Hz), 108.6 (d, ${}^{3}J_{C,F} = 8.0$ Hz), 68.6, 52.0, 26.6, 25.6, 17.9, -4.3, -4.6 ppm; ESI–HRMS: calcd. for C₂₇H₃₃FN₂O₆Si + Na 551.1990, found 551.1993.



4j, 88% yield; $[\alpha]_D{}^{20} = -62.8$ (*c* = 1.1 in CHCl₃); 89% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 7.504 min, *t* (minor) = 9.269 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.79$ (d, *J* = 2.0 Hz, 1H), 7.32

(s, 5H), 7.26 (dd, J = 8.4 Hz, 2.4 Hz, 1H), 6.70 (d, J = 8.4 Hz, 1H), 6.10 (s, 1H), 5.59 (s, 1H), 5.19

(d, J = 11.6 Hz, 1H), 5.11 (d, J = 11.6 Hz, 1H), 3.67 (s, 3H), 3.20 (s, 3H), 0.61 (s, 9H), 0.04 (s, 3H), -0.11 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.0$, 164.8, 159.6, 142.4, 137.6, 134.9, 129.9, 129.3, 128.9, 128.3, 128.3, 128.0, 127.5, 125.3, 109.1, 68.6, 52.0, 26.5, 25.6, 17.9, -4.3, -4.6 ppm; ESI–HRMS: calcd. for C₂₇H₃₃ClN₂O₆Si + Na 567.1694, found 567.1698.



4k, 81% yield; $[\alpha]_D{}^{20} = -54.6$ (*c* = 0.9 in CHCl₃); 88% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 7.186 min, *t* (minor) = 8.624 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.91$ (d, *J* = 2.0 Hz, 1H), 7.40

(dd, J = 8.0 Hz, 2.0 Hz, 1H), 7.32 (s, 5H), 6.65 (d, J = 8.4 Hz, 1H), 6.10 (s, 1H), 5.59 (s, 1H), 5.19 (d, J = 11.6 Hz, 1H), 5.10 (d, J = 11.6 Hz, 1H), 3.67 (s, 3H), 3.19 (s, 3H), 0.59 (s, 9H), 0.03 (s, 3H), -0.11 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.9$, 164.8, 159.6, 142.9, 137.6, 134.9, 132.2, 130.2, 128.9, 128.3, 128.3, 125.3, 115.4, 109.7, 74.8, 68.6, 52.0, 26.5, 25.6, 17.9, -4.3, -4.6 ppm; ESI–HRMS: calcd. for C₂₇H₃₃BrN₂O₆Si + Na 611.1189, found 611.1195.



4I, 85% yield; $[\alpha]_D^{20} = -57.5$ (*c* = 1.2 in CHCl₃); 86% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 7.946 min, *t* (minor) = 9.788 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.07$ (d, *J* = 1.6 Hz, 1H), 7.60 (dd, *J* = 8.4

Hz, 2.0 Hz, 1H), 7.32 (s, 5H), 6.56 (d, J = 8.4 Hz, 1H), 6.10 (s, 1H), 5.58 (s, 1H), 5.19 (d, J = 12.0 Hz, 1H), 5.11 (d, J = 11.6 Hz, 1H), 3.67 (s, 3H), 3.20 (s, 3H), 0.60 (s, 9H), 0.03 (s, 3H), -0.10 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.7$, 164.8, 159.6, 143.6, 138.2, 137.6, 135.6, 134.9, 130.6, 128.9, 128.3, 128.3, 125.3, 110.3, 68.6, 52.0, 26.5, 25.6, 17.9, -4.3, -4.6 ppm; ESI–HRMS: calcd. for C₂₇H₃₃IN₂O₆Si + Na 659.1050, found 659.1046.

4m, 71% yield; $[\alpha]_D^{20} = -59.4$ (c = 0.6 in CHCl₃); 85% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 5.957 min, *t* (minor) = 6.527 min]; ¹H NMR (400 MHz, CDCl₃): δ

= 7.74 (s, 1H), 7.31 (s, 5H), 7.16 (d, J = 8.4 Hz, 1H), 6.75 (d, J = 8.4 Hz, 1H), 6.11 (s, 1H), 5.60 (s,

1H), 5.18 (d, J = 11.6 Hz, 1H), 5.09 (d, J = 11.6 Hz, 1H), 3.67 (s, 3H), 3.21 (s, 3H), 0.60 (s, 9H), 0.04 (s, 3H), -0.13 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.1$, 164.9, 159.5, 144.6, 142.5, 137.5, 134.9, 129.7, 128.9, 128.3, 128.3, 125.6, 122.5, 121.1, 119.2, 108.6, 75.0, 68.7, 52.0, 26.6, 25.6, 17.9, -4.2, -4.9 ppm; ESI–HRMS: calcd. for C₂₈H₃₃F₃N₂O₇Si + Na 617.1907, found 617.1912.

Cbz TBSON COOMe **4n**, 85% yield; $[\alpha]_D{}^{20} = -49.5$ (*c* = 0.5 in CHCl₃); 90% ee, determined by HPLC analysis [Daicel Chiralpak IC, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (major) = 6.892 min, *t* (minor) = 8.365 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.55$ (dd, *J* = 7.6 Hz, 0.8 Hz, 1H), 7.34-7.30

(m, 5H), 7.04–7.00 (m, 1H), 6.95–6.90 (m, 1H), 6.12 (s, 1H), 5.61 (s, 1H), 5.17 (d, J = 12.0 Hz, 1H), 5.11 (d, J = 12.0 Hz, 1H), 3.66 (s, 3H), 3.40 (d, J = 2.8 Hz, 3H), 0.64 (s, 9H), 0.05 (s, 3H), -0.13 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.0$, 165.0, 159.6, 147.8 (d, ${}^{I}J_{C,F} = 241.7$ Hz), 137.8, 135.0, 131.1, 130.6, 128.9, 128.3, 128.3, 125.7, 123.0 (d, ${}^{3}J_{C,F} = 6.3$ Hz), 122.7 (d, ${}^{4}J_{C,F} = 3.5$ Hz), 117.5 (d, ${}^{2}J_{C,F} = 19.0$ Hz), 77.2, 68.6, 51.9, 29.0 (d, ${}^{3}J_{C,F} = 6.3$ Hz), 25.7, 18.0, -4.2, -4.5 ppm; ESI–HRMS: calcd. for C₂₇H₃₃FN₂O₆Si + Na 551.1990, found 551.1996.

4. Synthetic transformations of multifunctional adduct 4d

Zinc powder (84.5 mg, 1.3 mmol) was added to a stirred solution of **4d** (25.5 mg, 0.05 mmol) in AcOH/H₂O/THF (3:1:1) (0.5 mL) under a nitrogen atmosphere. The mixture was stirred at room temperature for 0.5h, and then stirred at 60 °C until the consumption of **4d**. The mixture was diluted with water and filtered. The filtrate was extracted with Et₂O (3 × 5 mL), and the aqueous layer was washed with aq NaOH (6.0 M) until pH 7, and the aqueous layer was extracted with Et₂O (3 × 5 mL). The combined organic layers were dried with anhydrous Na₂SO₄ and concentrated. Flash chromatography on silica gel (petroleum ether/ethyl acetate = 4:1) afforded pure product **5** as a white solid (6.7 mg, 35%). $[\alpha]_D^{20} = -64.7$ (c = 0.9 in CHCl₃); 90% ee, determined by HPLC analysis [Daicel Chiralcel OD, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, *t* (minor) = 13.183 min, *t* (major) = 19.307 min]; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.44$ (d, J = 7.6 Hz, 1H), 6.57 (s, 1H), 6.32 (s, 1H), 5.87 (s, 1H), 4.97 (s, 2H), 3.73 (s, 3H), 3.23 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.2$, 166.0, 154.6, 143.8, 136.3, 135.7, 129.6, 128.7, 128.4, 128.1, 128.1, 128.1,

124.6, 122.9 108.5, 67.2, 64.1, 52.5, 26.7 ppm; ESI–HRMS: calcd. for $C_{21}H_{20}N_2O_5$ + Na 403.1270, found 403.1271.

HF pyridine (5.4 µL, 0.06 mmol) was added to a solution of 4d (25.5 mg, 0.05 mmol) in THF (0.3 mL) at 0 °C. The mixture was stirred at room temperature until the consumption of 4d. Then saturated NaHCO₃ was added until pH 7, and the aqueous layer was extracted with Et₂O (3×3.0 mL). The combined organic layers were dried with anhydrous Na₂SO₄ and concentrated. Flash chromatography on silica gel (petroleum ether/ethyl acetate = 3:1) afforded pure product 6 as a colorless oil (17.8 mg, 90%). K₂CO₃ (1.4 mg, 0.01 mmol) was added to a solution of 6 in THF (0.3 mL) at 0 °C. The mixture was stirred at room temperature until 6 was consumed. Then the reaction mixture was concentrated in vacuo at low temperature and purified by flash chromatography on silica gel (petroleum ether/ethyl acetate = 5:1) afforded pure product 7 as a white semisolid (9.5 mg, 53%). $[\alpha]_D^{20} = +8.8$ (c = 0.5 in CHCl₃); 90% ee, determined by HPLC analysis [Daicel Chiralcel OD, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, λ = 254 nm, *t* (minor) = 16.310 min, *t* (major) = 19.092 min]; ¹H NMR (400 MHz, CDCl₃): δ = 7.41 (t, J = 7.2 Hz, 1H), 7.39–7.26 (m, 4H), 7.17–7.13 (m, 3H), 6.82 (d, J = 7.6 Hz, 1H), 6.43 (s, 1H), 5.40 (s, 1H), 5.18 (d, J = 12.0 Hz, 1H), 5.07 (d, J = 12. Hz, 1H), 3.02 (s, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 172.0, 162.6, 153.6, 143.8, 135.0, 134.1, 130.9, 128.6, 128.5, 128.2, 126.7, 126.2, 124.1, 124.0, 109.1, 72.0, 69.1, 26.5 ppm; ESI-HRMS: calcd. for $C_{20}H_{16}N_2O_5 + Na 387.0957$, found 387.0959.

5. Crystal data and structure refinement for enantiopure 5



a/Å, b/Å, c/Å 8.6032(2), 9.40760(10), 23.8035(4)

α/°, *β*/°, *γ*/°, 90.00, 90.00, 90.00

Volume/Å³ 1926.55(6)

Z	4
$ ho_{calc} mg/mm^3$	1.311
m/mm ⁻¹	0.782
F(000)	800
Crystal size	$0.42\times0.36\times0.30$
Theta range for data collection	5.06 to 69.79°
Index ranges	$-10 \le h \le 10, -11 \le k \le 11, -28 \le l \le 28$
Reflections collected	15142
Independent reflections	3601[R(int) = 0.0224]
Data/restraints/parameters	3601/0/256
Goodness-of-fit on F ²	1.041
Final R indexes [I>2σ (I)]	$R_1 = 0.0279, wR_2 = 0.0782$

Final R indexes [all data] $R_1 = 0.0289$, $wR_2 = 0.0793$

Largest diff. peak/hole 0.141/-0.109



6. NMR spectra and HPLC chromatograms













		(min)	(µV*sec)	% Area	(µV)	Height
I	1	9.463	8818105	95.54	353863	96.46
I	2	13.188	411939	4.46	12985	3.54





























	Peak Name	RT (min)	Area (µV*sec)	% Area	Height (µV)	% Height
1	Peak1	7.174	975497	50.76	51 4 01	55.65
2	Peak2	8.672	946167	49.24	40970	44.35







	(mm)	(µv sec)		(µv)	Height
1	7.143	2177297	50.08	111955	54.68
2	8.601	2170090	49.92	92795	45.32















	RT (min)	Area (µV*sec)	% Area	Height (µV)	% Height
1	5.957	5838881	92.30	353402	93.05
2	6.527	486900	7.70	26398	6.95





	(min)	(µV*sec)	% Area	μV)	Height
1	6.750	1708260	51.24	103140	55.66
2	8.017	1625556	48.76	82170	44.34







*	[min]	[min]	mAU	*3	[mAU]	3
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1	13.113	VB 0.5677	5527	.01123	148,	65988	49.8333
2	19.613	BB 0,8043	5563	,98584	107.	06664	50.1667

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1	13.183	VВ	0.5804	604.95105	15,80855	4.9305
2	19.307	BВ	0,7887	1.16646e4	228.14536	95.0695





## 7. References

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