

A novel family of (1-aminoalkyl)(trifluoromethyl)- and -(difluoromethyl)phosphinic acids – analogues of α -amino acids

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Abstract

A series of novel (1-aminoalkyl)(trifluoromethyl)- and -(difluoromethyl)phosphinic acids – analogues of proteinogenic and nonproteinogenic α -amino acids were prepared. The synthetic methodology was based on nucleophilic addition of (trifluoromethyl)phosphinic acid or (difluoromethyl)phosphinic acid or its ethyl ester to substrates with C=N or activated C=C double bonds. Analogues of glycine, phenylglycine, alanine, valine, proline, aminomalonic and aspartic acids were thus prepared. Three-component one-pot reactions of (trifluoromethyl)phosphinic acid and dibenzylamine with aldehydes were also tested to prepare the title compounds.

Introduction

For a long time aminophosphonic and aminophosphinic acids as isosters of aminocarboxylic acids have attracted a particular interest for the preparation of analogues of numerous natural products. Among the literature concerning various aspects of the chemistry and biological activity of aminophosphonic and aminophosphinic acids, several monographs and reviews have

appeared over the last decade [1-6]. The chemistry of fluorinated aminophosphonic and -phosphinic acids is a relatively new area of research. Incorporation of fluorine or fluorinated moieties can be used for the alteration of physiological properties of many biologically significant substances. The changes of their biological properties caused by this fluorination are

influenced by complex factors, however. The similarity of the diameters of fluorine and hydrogen atoms in organic compounds makes fluorine an obvious choice as a substituent for biologically active substances, frequently without disrupting the shape and geometry of the substituted molecules. Nevertheless fluorine influences the electronic properties of a compound drastically because of its strong electronegativity. This enables modulation of the lipophilicity profile, of electrostatic interactions with the target structure and inhibition of some metabolic pathways [7-9]. Data concerning the biological activity and synthetic approaches toward fluorinated aminophosphonates, bearing side chain C–F linkages are well documented in a review [10].

The isolation of phosphinothricin, a naturally occurring phosphorus analogue of glutamic acid and the discovery of its antibiotic, fungicidal and herbicidal properties [11] has led to an increased activity in the study of methylphosphinic acid analogues of the protein amino acids [12] and those of glycine [13], alanine [14], valine [14], leucine [15], proline [16], aspartic [17] and glutamic [11] acids and GABA [18] have been described. But almost nothing is known about phosphorus isosters of aminocarboxylic acids bearing a (trifluoromethyl)- or (difluoromethyl)phosphonyl moiety instead of the carboxylate function. To the best of our knowledge there is only one report on the application of ethyl (difluoromethyl)phosphinate $\text{CHF}_2(\text{H})\text{P}(\text{O})(\text{OEt})$ in the synthesis of a (difluoromethyl)phosphinic acid analogue of GABA, as a potent agonist of the GABA_B receptor [18].

In light of the above and in connection with our interest in the chemistry of fluorinated compounds of phosphorus we report here the preparation of a series of novel (1-aminoalkyl)phosphinic acids bearing CF_3 or CHF_2 groups at phosphorus.

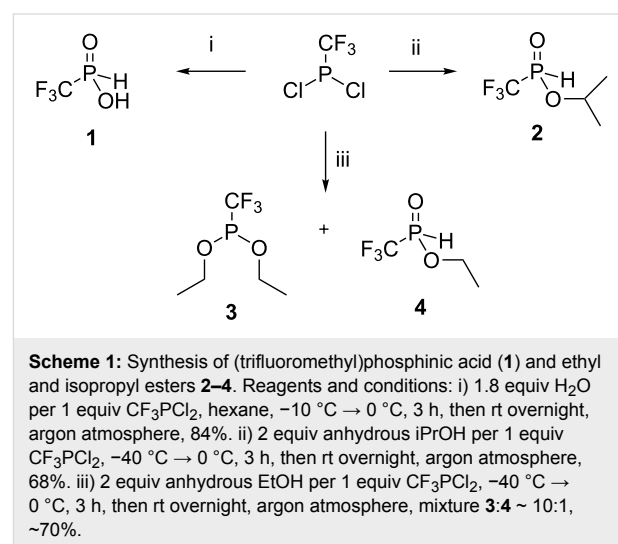
Results and Discussion

Research efforts have established H-phosphinates $\text{R}(\text{H})\text{P}(\text{O})(\text{OR}')$ or appropriate P(III) acids $\text{R}(\text{H})\text{P}(\text{O})(\text{OH})$ as appropriate starting materials for the preparation of aminophosphinic acids. The most typical route involves the three-component reaction of an aldehyde, an amine and a P–H substrate in a one-pot Mannich type protocol [19-21]. An alternative to this approach involves the simple addition of alkyl H-phosphinates or H-phosphinic acids to Schiff bases [5,22]. In this paper we exploit both routes to prepare (1-aminoalkyl)(trifluoromethyl)- and -(difluoromethyl)phosphinic acids using P–H compounds bearing CF_3 and CHF_2 groups attached to phosphorus.

P–H Substrates

(Trifluoromethyl)phosphinic acid $\text{CF}_3\text{P}(\text{O})\text{H}(\text{OH})$ (**1**) was first prepared in 1954 [23], but since then little chemistry has been

reported involving **1**. Also monoesters of **1** such as $\text{CF}_3\text{P}(\text{O})\text{H}(\text{OAlk})$ [24,25] have not been widely applied. These compounds, contain a labile P–H bond and synthetic problems underly their preparation. Emel us and Haszeldine were the first [26] to prepare $\text{CF}_3\text{P}(\text{III})$ compounds via the interaction of red phosphorus and CF_3I in an autoclave. This gave mixtures of CF_3 -containing phosphanes and phosphane iodides but in poor yields. More recently, Ruppert described the reaction between CF_3Br , $\text{P}(\text{NET}_2)_3$ and PCl_3 , which gave $\text{CF}_3\text{P}(\text{NET}_2)_2$ in a good yield [27]. We applied this procedure to prepare CF_3PCl_2 [27] by interaction of diamide $\text{CF}_3\text{P}(\text{NET}_2)_2$ with gaseous HCl and then the chlorine was replaced by neutral hydrolysis to give (trifluoromethyl)phosphinic acid (**1**) [23] or by alcoholysis with ethanol or isopropanol to reach the appropriate esters **2–4** [24,25] (Scheme 1).



In our hands CF_3PCl_2 was hydrolyzed by two equivalents of water in hexane over the temperature range $-10\text{ }^\circ\text{C} \rightarrow 0\text{ }^\circ\text{C}$, to give water-free (trifluoromethyl)phosphinic acid (**1**). Acid **1** proved easy to handle as a distillable liquid when prepared in this way and is stable under storage for months in contrast to that prepared by Emel us and Haszeldine [23]. Phosphinate **2** was prepared with anhydrous isopropanol. When stored under anhydrous conditions at room temperature, ester **2** was partially converted to acid **1** as determined by ^{31}P NMR. Alcoholysis of CF_3PCl_2 with ethanol under the same conditions produced diethyl phosphinate **3** admixed with monoester **4** (~10%), as previously described [25]. ^{31}P NMR of these products indicated that they were converted to esters **3** and **4** on storage in a ratio of ~ 3:2 with acid **1** as an impurity. The low stability of all three esters **2–4** can be attributed to the lability of the O–C ester bond, resulting from the electron-withdrawing effect of the CF_3 group attached to phosphorus, therefore esters **2**, **3** and **4** were used in the syntheses only when freshly prepared and distilled.

We next explored the CHF₂ group attached to phosphorus. Ethyl (difluoromethyl)phosphinate CHF₂P(O)H(OEt) (**5**) was prepared as previously described [18]. The appropriate (difluoromethyl)phosphinic acid CHF₂P(O)H(OH) (**6**) was obtained from **5** by ester deprotection with NaHCO₃, as a viscous undistillable liquid, which was stable for weeks on storage.

Three-component reactions

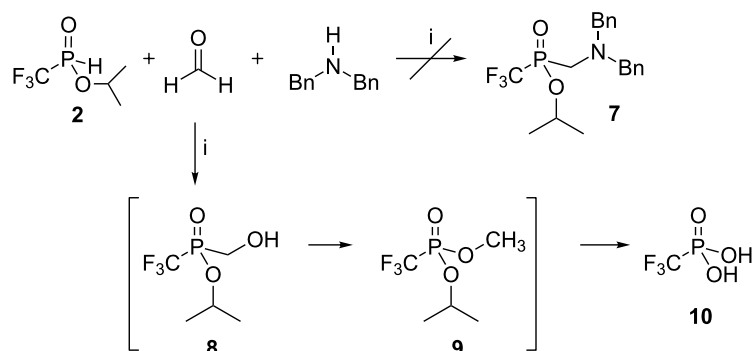
At the outset of our work three-component reactions of formaldehyde, dibenzylamine and the esters **2** or **5** were explored as model transformations to evaluate the feasibility of the Kabachnik–Fields procedure [19,20] to the synthesis of fluorinated (1-aminoalkyl)phosphinate **7** (Scheme 2).

It turned out that this method is unsuitable for the synthesis of phosphinate **7**. Formalin was added to an equimolar mixture of dibenzylamine and ester **2** at 80 °C under an oxygen-free atmosphere to give reaction mixtures with a low content of P–C products (³¹P NMR). A similar outcome was obtained when the reaction was run at room temperature or in dioxane with simul-

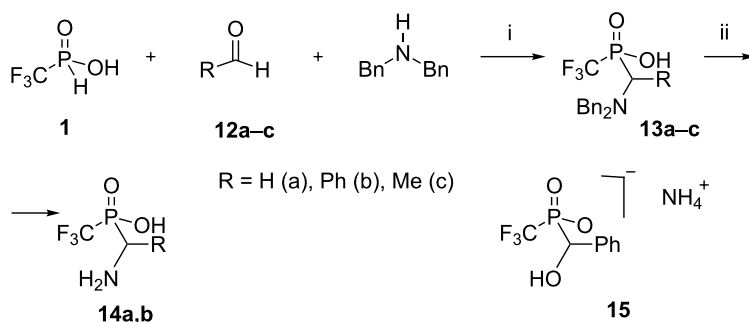
taneous water azeotropic distillation. Such a result might be explained by high reactivity of the starting ester, which readily reacted with formaldehyde, forming (α-hydroxymethyl)phosphinate **8**. Its further irreversible rearrangement [28] to the corresponding phosphonate **9** was accompanied with hydrolysis of the ester function and formation of (trifluoromethyl)phosphonic acid (**10**) [29] as the main product. Analogous results were obtained, when CHF₂ containing ester **5** was introduced into the reaction with formaline and dibenzylamine to give CHF₂P(O)(OH)₂ (**11**) as the major product [30].

Such results prompted us to explore the Mannich-type procedure of Moedritzer and Irani [21] for the syntheses of the desired aminophosphinic acids starting from acid **1**. This resulted in the preparation of the analogues of glycine **14a** and phenylglycine **14b** (Scheme 3).

The three-component reaction with formaldehyde gave the best results and *N*-protected aminophosphinic acid **13a** was isolated in a moderate yield, alongside phosphonic acid **10**. The analo-



Scheme 2: Three-component Kabachnik–Fields reaction of CF₃(H)P(O)(OiPr) (**2**) with formaldehyde and dibenzylamine. Reagents and conditions: i) an equimolar mixture of reagents, H₂O, 80 °C, 3 h, argon atmosphere, yield **10** ~80% or an equimolar mixture of reagents, dioxane, 100 °C, 3 h, argon atmosphere, yield **10** ~90%.



Scheme 3: Three-component synthesis of CF₃ containing α-aminophosphinic acids **14a,b**. Reagents and conditions: i) An equimolar mixture of acid **1**, dibenzylamine, HCl and two fold excess of aldehyde, H₂O, 80 °C, 3 h; isolated yields: **13a** (52%), **13b** (28%); yields, determined by ³¹P and ¹⁹F NMR: **13c** (<10%). ii) H₂, ethanol, catalysis 10% Pd/C, rt, normal pressure, yields: **14a** (95%), **14b** (90%).

gous reaction with benzaldehyde provided acid **13b** in 28% yield and the main product of this reaction was an adduct of acid **1** with benzaldehyde which was isolated from the reaction mixture as ammonium salt **15** in 60% yield. Reaction with acetaldehyde was less successful and generated aminophosphinic acid **13c** in low yield (<10%). Attempts to improve conversions products **13a–c** by increasing the reaction temperature or varying the amino component (MeC(O)NH₂, BnOC(O)NH₂ or NH₄OAc instead of Bn₂NH) and molar equivalent of HCl were unsuccessful. It should be noted, that in contrast to the non-fluorinated counterparts the adducts **13a,b** did not form hydrochlorides under this procedure consistent with the strongly acidic nature of the CF₃ phosphinic acid

group. Catalytic hydrogenation of intermediates **13a,b** with Pd/C removed the benzyl groups and produced the corresponding acids **14a,b** in high yields.

The hydrophosphinylation of azomethines

The addition of the P–H functionality to C=N double bonds is a very general procedure for the formation of P–C–N systems. Based on our experience of these three-component reactions (Scheme 2 and Scheme 3) we investigated the scope and limitations of the addition of (trifluoromethyl)phosphinic acid (**1**) to a series of *N*-benzylimines **16a–e** in order to obtain fluorinated phosphorus analogues of glycine **14a**, phenylglycine **14b**, alanine **14c**, valine **14d** and proline **14e** (Table 1).

Table 1: The interaction of (trifluoromethyl)phosphinic acid (**1**) with Schiff bases.^a

Entry	Schiff base	R	17 (yield, %) ^b	14 (yield, %) ^b
1 ^c		H	17a (83)	14a (95)
2		Ph	17b (79)	14b (96)
3 ^d		Me	17c (59)	14c (96)
4		iPr	17d (92)	14d (98)
5 ^c			–	14e (66)

^aReagents and conditions: i) an equimolar mixture of acid **1** and Schiff base, DME, rt, ³¹P NMR control; ii) H₂, ethanol, catalysis 10% Pd/C, rt, normal pressure. ^bIsolated yields. ^cSymmetrical cyclic triazinanes (masked imines) were used to generate unstable imines. ^dThe best yield was obtained with 2 mol equivalents of imine.

The transformations were mildly exothermic and were monitored by ^{31}P NMR. Acid **1** undergoes the typical P–C bond forming reactions with Schiff bases to give adducts **17** in satisfactory yields and these were successfully transformed into the appropriate free acids **14**.

The same series of Schiff bases was used to explore the reactivity of ethyl (difluoromethyl)phosphinate (**5**) in reactions with C=N double bonds and the desired (α -aminoalkyl)phosphinic acids **20a–e** were accordingly prepared (Table 2).

The syntheses of compounds **20a–e** were performed with purification and characterization of the intermediates, wherever possible as summarized in Table 2.

Products **14b–e** and **20b–e** were obtained as racemic mixtures. As expected, the ^{31}P NMR spectra of **14** and **20** display characteristic signals around 11–15 ppm with $^2J_{\text{FP}}$ couplings for CF_3 bearing substrates. The CHF_2 bearing also possessed $^2J_{\text{FP}}$ couplings of 75–95 Hz. The ^{19}F NMR spectra were characterized by signals in the region –75 ppm for CF_3 derivatives and

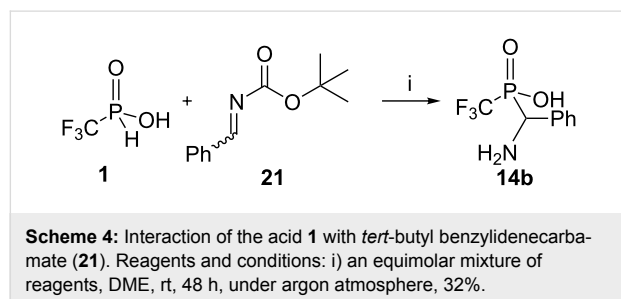
Table 2: The interaction of ethyl (difluoromethyl)phosphinate (**5**) with Schiff bases.^a

Entry	Schiff base	R	18 (yield, %) ^b	19 (yield, %) ^b	20 (yield, %) ^b
	$16\text{a} (=N\text{Bn})_3, 16\text{b-d}, 16\text{e} (\text{cyclic imine})_3$				
	R = H (a), Ph (b), Me (c), iPr (d), (e)				
1 ^c		H	18a (78) ^d	19a (68)	20a (91)
2		Ph	18b (58) ^e	19b (86) ^f	20b (96)
3 ^g		Me	18c (56) ^e	19c (80) ^f	20c (95)
4		iPr	18d (36) ^e	19d (82) ^f	20d (95)
5 ^c			–	–	20e (79)

^aReagents and conditions: i) an equimolar mixture of ester **5** and Schiff base, DME, rt, ^{31}P NMR control, under argon atmosphere; ii) 1 N HCl, rt, until clear solution; iii) H_2 , ethanol, catalysis 10% Pd/C, rt, normal pressure. ^bIsolated yields. ^cSymmetrical cyclic triazinanes (masked imines) were used to generate unstable imines. ^dYield was defined with ^{31}P NMR. ^eDiastereomeric ratio: **18b** (~7:2), **18c** (~3:2), **18d** (~3:2). ^fYields were defined as the sum of yields of compounds **19b–d**, isolated from the reaction mixture and obtained after hydrolysis of intermediates **18b–d**. ^gThe maximum yield was obtained with 2 mol equivalents of imine.

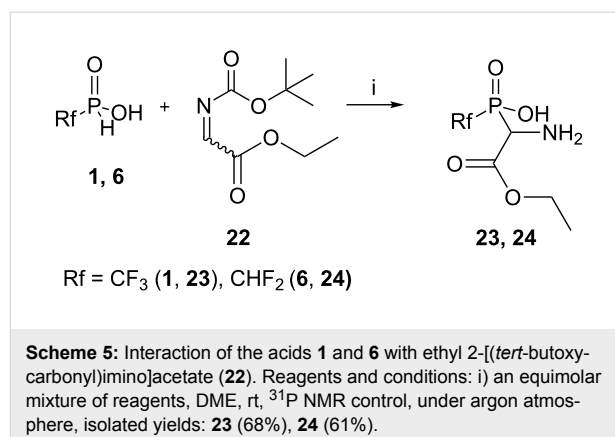
–137 ppm for CHF₂ ones. Some distinctive characteristics of the reactivity of ester **5** were observed. This ester readily reacted with azomethines, but in contrast to its nonfluorinated counterparts this ester generated mixtures of the adducts **18** and **19**. The interaction of ester **5** with imine **16a** gave adduct **18a** (Table 2, entry 1) in high conversion yield but after purification over silica gel only the appropriate acid **19a** was isolated. In the case of the reaction of ester **5** with imine **16e** no adduct was formed. Ethyl phosphinates **18b–d** were obtained as a mixture of two diastereoisomers, which were not separated but they are clearly observed by ¹H NMR as separate signals for the CHF₂ group. The adducts **18b–d** were hydrolyzed to acids **19b–d** in quantitative yields and did not form hydrochlorides similar to the CF₃ aminophosphinic acids **13a–b** and **17a–d**. Hydrogenolysis of the **17a–d** and **19a–d** efficiently gave free acids **14** and **20**, but required column ion-exchange chromatography to produce analytically pure products.

We then explored the addition of acid **1** to imine **21** [31], which is *N*-Boc protected, typically used for amino acid protection (Scheme 4). This produced acid **14b** in one step, but in only 32% yield. The *N*-*tert*-butoxycarbonyl group was removed during the reaction due to the high acidity of the CF₃ phosphinic acid group.



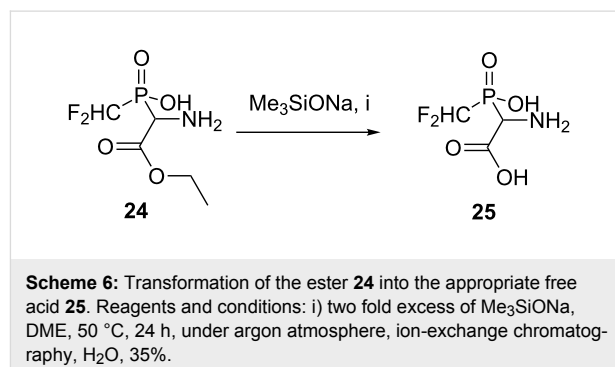
The variability of Schiff bases ensures access to a range of structurally diverse phosphinic acid analogues of amino acids in the relatively simple way. Thus, we investigated the hydrophosphinylation of some Schiff bases bearing a carboxylate functionality to obtain aminocarboxylic acids, containing pendant CF₃ or CHF₂ phosphinic acid linkages. Thus, phosphinic acids **1** and **6** reacted with the *N*-Boc-protected Schiff base of ethyl glyoxalate **22** [32] under mild conditions to produce the *N*-deprotected phosphinylglycines **23** and **24** in satisfactory yields (Scheme 5).

Attempts to convert ester **23** to the free acid failed. Removal of the ester group from **23** by acidolysis with HCl or HI was accompanied by cleavage of the P–C bond to give only (trifluoromethyl)phosphonic acid (**10**) after an ion-exchange chromatography. Attempts to remove the ester group in anhydrous base



with 1 equivalent of sodium silanolate (Me₃SiONa) at room temperature efficiently produced the highly stable sodium salt of acid **23**. With an excess of Me₃SiONa and heating to 50 °C, fluoroform liberation from **23** was observed to give fluorine-free products.

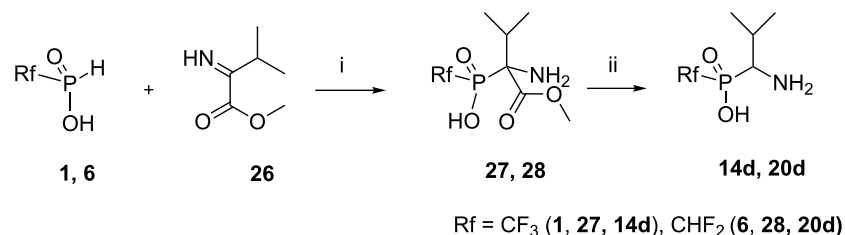
In contrast the CHF₂-containing ester **24** was stable toward acidic hydrolysis under mild conditions, but in the presence of an excess of sodium silanolate, free phosphinylglycine **25** was obtained, but in a poor yield (Scheme 6).



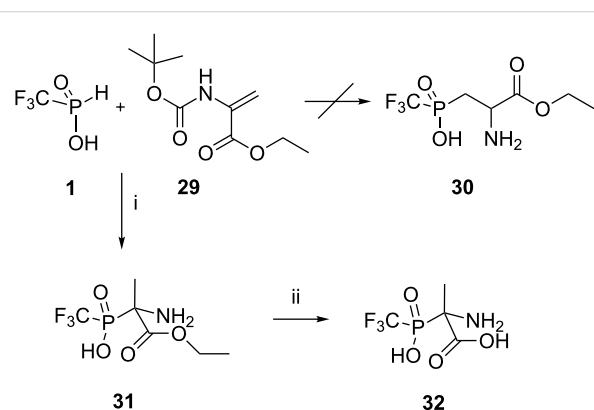
The reactions of substrates **1** and **6** with imine **26**, which is available from valine [33], readily gave adducts **27** and **28**, which were decarboxylated under acidolysis to afford the phosphinic acid analogues of valine **14d** and **20d** (Scheme 7).

The hydrophosphinylation of substrates with activated C=C double bonds

The high reactivity of (trifluoromethyl)phosphinic acid (**1**) with C=N double bonds prompted us to explore its reactivity towards activated C=C double bonds. By analogy with the synthesis of the phosphonic acid analogue of aspartic acid, developed by Chambers and Isbell [34], the P–H substrate **1** was reacted with *N*-Boc-protected aminoacrylate **29** [35], and this gave the precursor of the aspartic acid analogue **30** (Scheme 8).



Scheme 7: Reaction of the acids (**1**) and (**6**) with methyl 2-imino-3-methylbutanoate (**26**). Reagents and conditions: i) an equimolar mixture of reagents, DME, rt, ³¹P NMR control, under argon atmosphere, isolated yields: **27** (63%), **28** (40%).

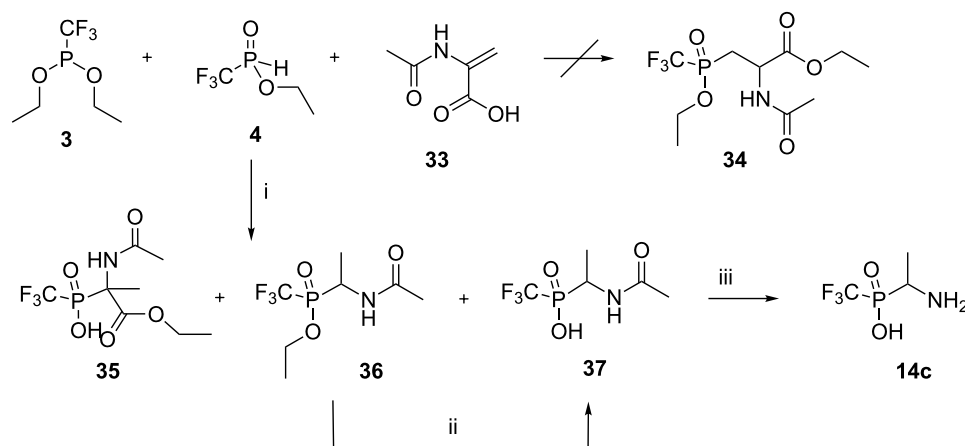


Scheme 8: Interaction of the acid **1** with ethyl 2-(*tert*-butoxycarbonylamino)acrylate (**29**). Reagents and conditions: i) an equimolar mixture of reagents, DME, rt, ³¹P NMR control, under argon atmosphere, 59%. ii) 5 N HCl, rt, 48 h, ion-exchange chromatography, H₂O, 54%.

Surprisingly, under the mild conditions of our experiment only the addition of acid **1** to the C=N double bond of the acrylic ester **29**, occurred to produce the *tertiary* phosphinyl derivative of alanine **31** in a satisfactory yield. Ester **31** was then hydrolyzed to give the free acid **32** in a moderate yield.

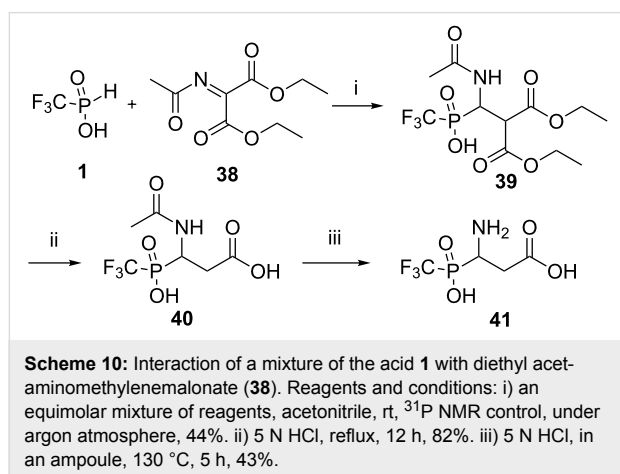
For the synthesis of the aspartic acid analogue **34**, a reaction between a mixture of the freshly prepared esters **3** and **4** and *N*-acetyl-protected aminoacrylic acid **33** was carried out [**34**] (Scheme 9).

It was thought that diester **3** might esterify aminoacrylic acid **33** [17,34] to produce ethyl 2-acetamidoacrylate and this compound in turn might add to monoester **4** to give the protected phosphinic acid analogue of aspartic acid **34**. Unfortunately, the only addition of **4** to the C=N double bond occurred similar to the previous transformation illustrated in Scheme 8. Insoluble in the reaction mixture phosphinic acid **37** was filtered off and characterized. ³¹P NMR analysis of filtrate showed the presence of adduct **35** and the decarboxylation product **36** in an approximate 1:10 ratio along with starting esters **3** and **4** and (trifluoromethyl)phosphonic acid (**10**) (<5%). Products **35** and **36** were separated by chromatography and characterized. Ester **36** was obtained as a mixture of two diastereoisomers, which are clearly seen by ¹H- and ¹⁹F NMR. This ester was then readily converted by acidolysis into the phosphinic acid analogue **37**, of *N*-acetylalanine, which was isolated in the 56% from acid **33**, and then transformed into free amino acid **14c**.



Scheme 9: Interaction of a mixture of the esters **3** and **4** with 2-acetamidoacrylic acid (**33**). Reagents and conditions: i) an equimolar mixture of **4** and **33** and 1.5 equiv of **3**, rt, ³¹P NMR control, under argon atmosphere, isolated yields: **35** (7%), **36** (40%) (diastereomeric ratio ~8:7), **37** (18%). ii) 5 N HCl, rt, 24 h, 95%. iii) 5 N HCl, in an ampoule, 130 °C, 8 h, 34%.

We have been able to prepare the isomeric aspartic acid analogue **41** with phosphorous α - to the amino group by the analogy with the published method [17,36]. The synthesis of phosphinic acid **41** was accomplished by addition of acid **1** to the activated C=C double bond of malonate **38** followed by hydrolysis and decarboxylation to generate adduct **39** in two steps (Scheme 10).



Conclusion

In conclusion, we have presented a variety of approaches to novel fluorinated (1-aminoalkyl)phosphinic acids starting from the appropriate fluorinated P–H compounds with CF_3 or CHF_2 groups attached to phosphorus. Three-component one pot Mannich-type reactions of $\text{CF}_3(\text{H})\text{P}(\text{O})(\text{OH})$ with dibenzylamine and aldehydes were investigated. Also nucleophilic addition of $\text{CF}_3(\text{H})\text{P}(\text{O})(\text{OH})$ or $\text{CHF}_2(\text{H})\text{P}(\text{O})(\text{OEt})$ to Schiff bases, aminoacrylates and acetaminomethylenemalonate have been used to prepare the title compounds.

Experimental

All reactions with P–H compounds were performed under an argon atmosphere. Flash chromatography was carried out using Merck silica gel 60 (230–400 mesh ASTM) and Aldrich ion-exchange resin Dowex WX-50. The NMR spectra were recorded on Varian VXR-300 or Bruker Avance DRX-500 spectrometers for ^1H (TMS); on a Bruker Avance DRX-500 spectrometer for ^{13}C {H} (TMS); on Varian Gemini-200 or Varian VXR-300 spectrometers for ^{19}F (CFCl_3) and for ^{31}P (H_3PO_4).

Synthesis of starting materials

(Trifluoromethyl)phosphinic acid (**1**). To an emulsion of water (3.2 g, 180 mmol) in anhydrous hexane (20 mL), cooled to -78 °C CF_3PCl_2 [27] (16.5 g, 96.5 mmol) was added under stirring and the temperature was slowly raised to -10 °C, when hydrolysis started. The reaction mixture was allowed to come to

0 °C at such a rate to avoid a vigorous reaction (~ 3 h) and then to room temperature and stirring was continued overnight. Hexane was evaporated under reduced pressure and the residue was distilled to give **1** as a colorless liquid (10.84 g, 84%), bp 35 °C (0.05 mm Hg); ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ_{H} 7.05 (dq, $^1J_{\text{HP}} = 638.9$ Hz, 1H, $^3J_{\text{HF}} = 4.2$ Hz, PH), 13.6 (1H, s, OH); ^{31}P NMR (121 MHz) δ_{P} 6.1 (dq, $^1J_{\text{PH}} = 639$ Hz, $^2J_{\text{PF}} = 82$ Hz); ^{19}F NMR (188 MHz) δ_{F} -76.8 (dd, $^2J_{\text{FP}} = 82$ MHz, $^3J_{\text{FH}} = 4$ Hz). **Caution:** Safety precautions are necessary, because CF_3PCl_2 reacts violently with air. Care must be taken not to warm the reaction system rapidly, because rapid volatilization of gaseous HCl will be accompanied by carrying off CF_3PCl_2 , which can inflame.

(Difluoromethyl)phosphinic acid (**6**). A mixture of **5** [18] (8 g, 56 mmol) and NaHCO_3 (7 g, 83 mmol) in ether (50 mL) was stirred overnight at room temperature to produce a bulky precipitate of $\text{CHF}_2\text{P}(\text{O})\text{H}(\text{O})^-\text{Na}^+$ [^{31}P NMR (121 MHz, H_2O): δ_{P} 11.9 (dtd, $^1J_{\text{PH}} = 570$ Hz, $^2J_{\text{PF}} = 87$ Hz, $^2J_{\text{PH}} = 25$ Hz)]. This precipitate was filtered, thoroughly washed with ether, solved in water (25 mL) and passed down an ion-exchange column. Water from the resulting solution was evaporated under reduced pressure and the residue was kept in vacuo (0.05 mmHg) for 24 h at room temperature to give **6** as a viscous colorless undistillable liquid (7.19 g, 78%); Anal. calcd for $\text{CH}_3\text{F}_2\text{O}_2\text{P}$: C, 10.35; H, 2.61; P, 26.71; found: C, 10.48; H, 2.70; P, 26.59; ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ_{H} 6.15 (tdd, $^2J_{\text{HF}} = 48.6$ Hz, $^2J_{\text{HP}} = 24.5$ Hz, $^3J_{\text{HH}} = 1.5$ Hz, 1H, CHF_2), 6.9 (dm, $^1J_{\text{HP}} = 566.5$ Hz, 1H, PH), 12.7 (s, 1H, OH); ^{31}P NMR (121 MHz) δ_{P} 12.1 (dtd, $^1J_{\text{PH}} = 566$ Hz, $^2J_{\text{PF}} = 86$ Hz, $^2J_{\text{PH}} = 25$ Hz); ^{19}F NMR (188 MHz) δ_{F} 9.6 (dd, $^2J_{\text{FP}} = 86$ Hz, $^2J_{\text{FH}} = 49$ Hz).

Three-component reactions

The general procedure for the condensation of the acid 1 with dibenzylamine and aldehydes (I). An equimolar mixture of **1** (2.68 g, 20 mmol) and dibenzylamine (3.94 g, 20 mmol) in 1 N HCl (20 mL) was heated at 80 °C under stirring. In the course of ~ 1 h aldehydes **12a,b** were added with a syringe and the reaction mixture was kept at this temperature for additional 1 h. The resulting mixture was left overnight at room temperature to produce the precipitate, which was filtered, washed with acetone–water (10:1) and dried to afford **13a** or **13b**. The filtrate was evaporated to the dryness, the residue was triturated with acetone–water (10:1) to give an additional quantity of **13a,b**.

[(Dibenzylamino)methyl](trifluoromethyl)phosphinic acid (**13a**). Following the general procedure (I) using 3.2 mL of 37% aqueous formaldehyde solution (20 mmol) **13a** was obtained as a white solid (3.57 g, 52%), mp 229 °C; Anal. calcd for

$C_{16}H_{17}F_3NO_2P$: C, 55.98; H, 4.99; N, 4.08; found: C, 55.69; H, 5.28; N, 4.15; 1H NMR (300 MHz, DMSO- d_6) δ_H 2.94 (d, $^2J_{HP}$ = 9.3 Hz, 2H, CH_2P), 4.45 (s, 4H, CH_2Ph), 7.47–7.59 (m, 10H, $H_{arom.}$); ^{31}P NMR (121 MHz) δ_P 3.8 (qt, $^2J_{PF}$ = 81 Hz, $^2J_{PH}$ = 9 Hz); ^{19}F NMR (188 MHz) δ_F -73.8 (d, $^2J_{FP}$ = 81 Hz).

The general procedure for N-deprotection of compounds with N-Bn function under the catalytic hydrogenation conditions (II). To a solution of compounds, containing N-Bn fragment (5 mmol) in ethanol (10 mL) 10% Pd/C (0.05 g) was added, and the mixture was hydrogenated at room temperature and normal pressure. After ~3 h the precipitation commenced, and water (5 mL) was added to dissolve this precipitate. The hydrogenation was then continued with a fresh portion of the catalyst (0.05 g) for a further 3 h. Last procedure was repeated whenever necessary and the reaction was left overnight. To the resulting mixture water was added until a white solid was fully dissolved, and the catalyst was then filtered off. The filtrate was evaporated to dryness; the residue was dissolved in acetone and allowed to stand at 5 °C until complete precipitation. The formed solid was filtered, washed with acetone and dried to give compounds with the free NH_2 function.

(Aminomethyl)(trifluoromethyl)phosphinic acid (**14a**). Following the general procedure (II) **14a** was obtained as a white powder (0.78 g, 95%); mp 192 °C; Anal. calcd for $C_2H_5F_3NO_2P$: C, 14.73; H, 3.09; N, 8.59; found: C, 14.69; H, 2.89; N, 8.42; 1H NMR (300 MHz, D_2O) δ_H 3.14 (d, $^2J_{HP}$ = 11.4 Hz); ^{31}P NMR (121 MHz) δ_P 12.2 (qt, $^2J_{PF}$ = 96 Hz, $^2J_{PH}$ = 11 Hz); ^{19}F NMR (188 MHz) δ_F -76.1 (d, $^2J_{FP}$ = 96 Hz); ^{13}C NMR (125 MHz) δ_C 34.8 (d, $^1J_{CP}$ = 105.6 Hz, CH_2), 122,1 (qd, $^1J_{CF}$ = 316.0 Hz, $^1J_{CP}$ = 179.9 Hz, CF_3).

Hydrophosphinylation of azomethines

The general procedure for the addition of acid 1 and ester 5 to substrates with the C=N double bond (III). An equimolar mixture of an imine and **1** or **5** in DME (10 mL for 5 mmol) was stirred at room temperature under ^{31}P NMR control until the ^{31}P signals of starting P-H compounds disappeared. Sometimes an appropriate adduct precipitated and this was filtered off. The reaction mixture or the filtrate was then evaporated to dryness and the residue was worked up as described below for the individual substances.

Ethyl [(benzylamino)(phenyl)methyl](difluoromethyl)phosphinate (**18b**, Table 2, entry 2). Following the general procedure (III) a crude solid, obtained from **5** (0.49 g, 3.4 mmol) and **16b** (0.68g, 3.4 mmol) was extracted with boiling hexane (3 × 30 mL), this extract was evaporated to the dryness to afford **18b** as a yellowish solid (0.67 g, 58%); mp 85–93 °C, as a mixture of two diastereoisomers in an approximately 1:3.5 ratio due

to 1H NMR (300 MHz, $CDCl_3$) δ_H 0.97 (t, $^3J_{HH}$ = 7.6 Hz, 0.7H, CH_3 , minor isomer), 1.25 (t, $^3J_{HH}$ = 7.6 Hz, 2.3H, CH_3 , major isomer), 2.17 (br s, 1H, NH), 3.46 (d, J_{AB} = 12.6 Hz, 0.22H, CH_2Ph , minor isomer), 3.51 (d, J_{AB} = 12.6 Hz, 0.77H, CH_2Ph , major isomer), 3.78 (d, J_{AB} = 12.6 Hz, 1H, CH_2Ph), 3.9 (dm, $^2J_{HP}$ = 16.9 Hz, 0.22H, PCH, minor isomer), 4.08 (d, $^2J_{HP}$ = 17.1 Hz, 0.8H, PCH, major isomer), 4.1–4.25 (m, 2H, OCH_2), 5.88 (td, $^2J_{HF}$ = 49.2 Hz, $^2J_{HP}$ = 27.8 Hz, 0.8H, CHF_2 , major isomer), 6.25 (td, $^2J_{HF}$ = 49.3 Hz, $^2J_{HP}$ 27.6 Hz, 0.2H, CHF_2 , minor isomer), 7.15–7.40 (m, 10H, $H_{arom.}$); ^{31}P NMR (81 MHz) δ_P 30.8 (m); ^{19}F NMR (188 MHz) δ_F -132 to -140.5 (complex multiplet). To the viscous residue after extraction of **18b** water (20 mL) was added, resulting solution was decolorized with activated charcoal, filtrated and allowed to stand at 5 °C until crystallization completed, producing [(benzylamino)-(phenyl)methyl](difluoromethyl)phosphinic acid (**19b**) as a white solid (0.32 g, 30%); mp 247 °C; Anal. calcd for $C_{15}H_{16}F_2NO_2P$: C, 57.88; H, 5.18; N, 4.50; found: C, 57.91; H, 5.04; N, 4.48; 1H NMR (300 MHz, DMSO- d_6) δ_H 3.95 (d, J_{AB} = 12.9 Hz, 1H, CH_2Ph), 4.04 (d, $^2J_{HP}$ = 10.2 Hz, 1H, PCH), 4.10 (d, J_{AB} = 12.9 Hz, 1H, CH_2Ph), 5.63 (td, $^2J_{HF}$ = 49.2 Hz, $^2J_{HP}$ = 21.9 Hz, 1H, CHF_2), 7.30–7.42 (m, 10H, $H_{arom.}$); ^{31}P NMR (81 MHz) δ_P 13.4 (tm, $^2J_{PF}$ 68 Hz). An additional quantity of **19b** was obtained by hydrolysis of **18b** (0.67 g, 2 mmol) with 1N HCl (15 mL) at room temperature until the starting ester has dissolved. The resulting solution was evaporated to dryness at reduced pressure and the residue was recrystallized from water to give **19b** (0.6 g, 97%). The overall yield of **19b** is 0.92 g (87%).

See Supporting Information for details of the syntheses, characteristics and NMR spectra of all new compounds.

Supporting Information

Experimental procedures and full characterization data for all new compounds including elemental analysis and 1H , ^{31}P , ^{19}F and ^{13}C NMR are provided in the Supporting Information.

Supporting Information File 1

Experimental procedures, elemental analysis and NMR data.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-66-S1.pdf>]

Supporting Information File 2

NMR spectra of the most typical compounds.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-66-S2.pdf>]

Supporting Information File 3

NMR spectra of the most typical compounds
(continuation).[\[http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-66-S3.pdf\]](http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-66-S3.pdf)

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