

# Synthesis of phenanthridines via palladium-catalyzed picolinamide-directed sequential C–H functionalization

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## Letter

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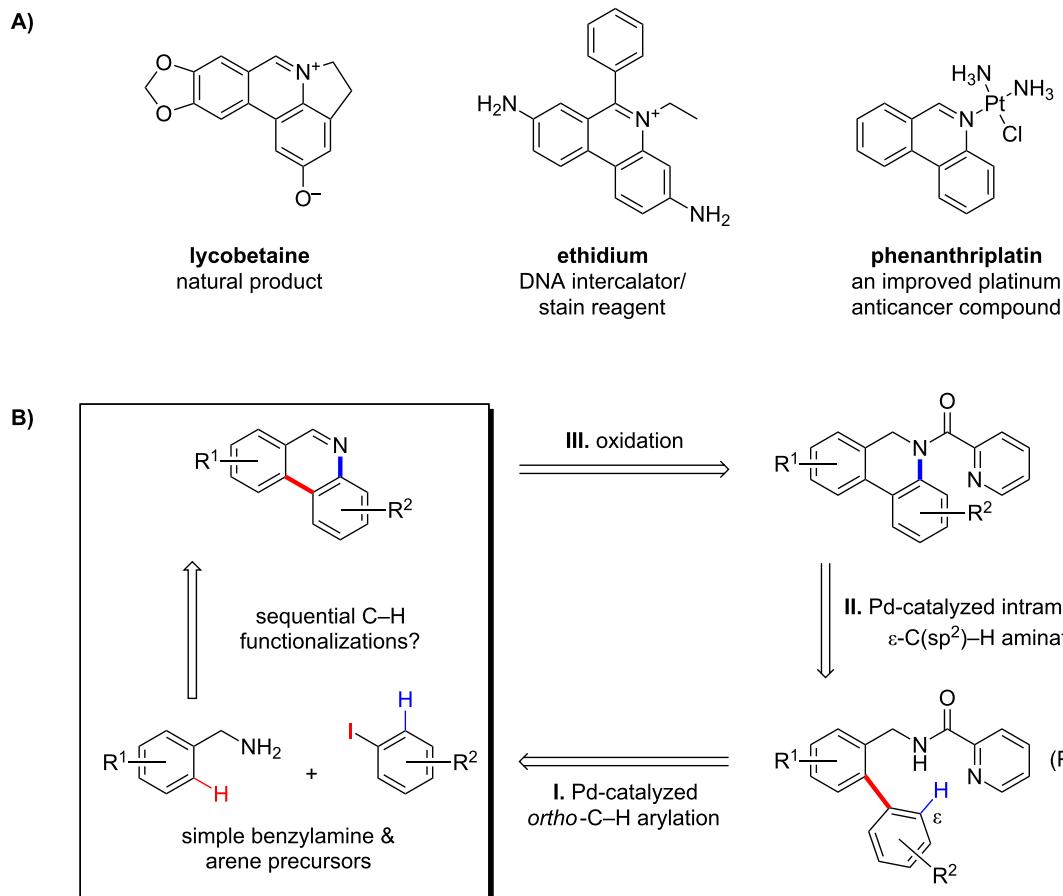
## Abstract

We report a new synthesis of phenanthridines based on palladium-catalyzed picolinamide-directed sequential C–H functionalization reactions starting from readily available benzylamine and aryl iodide precursors. Under the catalysis of Pd(OAc)<sub>2</sub>, the *ortho*-C–H bond of benzylpicolinamides is first arylated with an aryl iodide. The resulting biaryl compound is then subjected to palladium-catalyzed picolinamide-directed intramolecular dehydrogenative C–H amination with PhI(OAc)<sub>2</sub> oxidant to form the corresponding cyclized dihydrophenanthridines. The benzylic position of these dihydrophenanthridines could be further oxidized with Cu(OAc)<sub>2</sub>, removing the picolinamide group and providing phenanthridine products. The cyclization and oxidation could be carried out in a single step and afford phenanthridines in moderate to good yields.

## Introduction

Phenanthridines and 5,6-dihydro-phenanthridines are important core structures found in a variety of natural products and functional molecules (Scheme 1A) [1–8]. Synthetic methods for their preparation include the classical Pictet–Hubert condensation [9], radical-mediated reactions [10–13], metal-catalyzed cross-couplings [14–18], cycloadditions [19], and others [20–22]. More recently, methods based on the metal-catalyzed func-

tionalization of carbon–hydrogen (C–H) bonds have also emerged as viable strategies for synthesizing phenanthridines [23–25]. Despite these advances, construction of phenanthridines with complex substitution patterns remains difficult and often requires lengthy and inefficient synthetic sequences. Herein, we report a novel method for phenanthridine synthesis based on sequential palladium-catalyzed picolinamide



**Scheme 1:** Representative phenanthridine compounds and our synthetic strategy based on Pd-catalyzed sequential C–H functionalizations.

(PA)-directed C–H functionalization reactions beginning from easily accessible PA-protected benzylamine and aryl iodide precursors.

## Results and Discussion

**New synthetic strategy for phenanthridine compounds.** The picolinamide (PA) group has been shown to be an excellent directing group for a range of Pd-catalyzed C–H functionalization reactions [26–35]. In 2005, the Daugulis laboratory first reported that the *ortho*-C(sp<sup>2</sup>)–H bond of benzylpicolinamides could be arylated with aryl iodides under Ag-promoted Pd-catalyzed conditions [26]. In 2012, our laboratory [28] as well as that of Daugulis [27] independently reported that picolinamide substrates can undergo intramolecular dehydrogenative C–H amination reactions to afford medium-sized N-heterocycles under the catalysis of Pd(OAc)<sub>2</sub> with PhI(OAc)<sub>2</sub> oxidant. These discoveries led us to explore whether we could develop a new strategy for synthesizing phenanthridines. As outlined in Scheme 1B, we envisioned that *ortho*-arylated benzylamine picolinamides could undergo an intramolecular amination at the *ortho*  $\epsilon$ -C–H position of the newly installed

arene group to form cyclized dihydropheanthridines, which could be further converted to phenanthridine products under oxidative conditions. Ideally, we hoped to perform both the intramolecular C–H amination and subsequent oxidation in a single step [36].

**Arylation of 2-methoxybenzyl picolinamide 1 with 4-iodoanisole (2) under various conditions.** We commenced the study by investigating the arylation of 2-methoxybenzyl picolinamide **1** with 4-iodoanisole (**2**) under various conditions (Table 1) to form our desired arylated product **3**. Our initial attempt under the original Pd(OAc)<sub>2</sub>-catalyzed AgOAc-promoted solvent-free condition afforded the desired arylated product **3** in good yield (Table 1, entry 1). This method, however, required the use of expensive silver salt as an additive and high reaction temperature (150 °C). We next sought to replace the silver salts with cheaper reagents and lower the reaction temperature [12]. Not surprisingly, the arylation yield dropped significantly when the reaction was performed in toluene solvent at 120 °C (Table 1, entry 2). Addition of PivOH (0.3 equiv) gave little improvement (Table 1, entries 3 and 4).

**Table 1:** Optimization of the Pd-catalyzed *ortho*-C–H arylation of benzylpicolinamide. All screening reactions were carried out in a 10 mL glass vial on a 0.2 mmol scale.

entry	additives (equiv)	temperature (°C)	solvent	yield of <b>3</b> (%) <sup>a</sup>	
				Pd(OAc) <sub>2</sub> (5 mol %)	(2 equiv)
1	AgOAc (1.5)	150	no solvent	76	
2	AgOAc (1.5)	120	toluene	6	
3	AgOAc (1.5), PivOH (0.3)	120	toluene	3	
4	PivOH (0.3)	120	toluene	5	
5	K <sub>2</sub> CO <sub>3</sub> (2)	120	toluene	57	
6	PivOH (0.3), K <sub>2</sub> CO <sub>3</sub> (2)	120	toluene	90	
7	PivOH (0.3), KHCO <sub>3</sub> (2)	120	toluene	95 (91) <sup>b</sup>	
8	AcOH (0.3), KHCO <sub>3</sub> (2)	120	toluene	78	
9	oPBA <sup>c</sup> (0.3), KHCO <sub>3</sub> (2)	120	toluene	84	
10	PivOH (0.3), KHCO <sub>3</sub> (2)	90	toluene	29	

<sup>a</sup>Yields are based on <sup>1</sup>H NMR analysis of the reaction mixture after workup; <sup>b</sup>Isolated yield; <sup>c</sup>oPBA: *ortho*-phenylbenzoic acid.

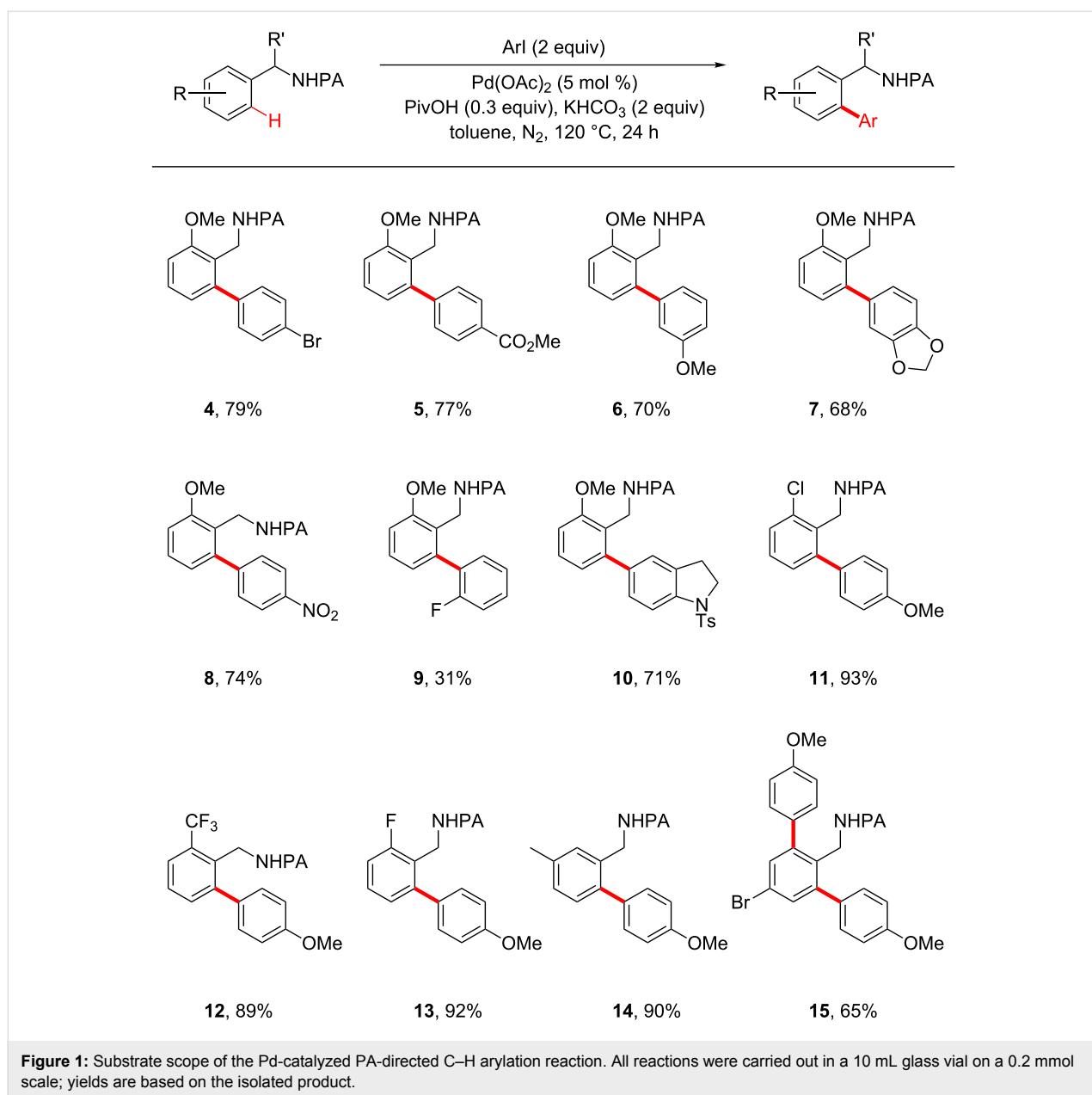
To our delight, the desired arylation reaction was largely restored with the application of 2 equiv of K<sub>2</sub>CO<sub>3</sub> at 120 °C for 24 h (Table 1, entry 5). Furthermore, an excellent yield was obtained when K<sub>2</sub>CO<sub>3</sub> was replaced with KHCO<sub>3</sub> and 0.3 equiv of PivOH was applied (Table 1, entry 7). The most effective carboxylate ligand and solvent was found to be PivOH and toluene, respectively.

**The determination of the scope of this reaction with benzylpicolinamide and aryl iodide substrates.** With the optimized conditions in hand, we next explored the scope of benzylpicolinamide and aryl iodide substrates (Figure 1). The electronic properties of benzylpicolinamide and aryl iodides had little influence on the reactivity, as benzylpicolinamide and aryl iodide substrates bearing electron-donating and withdrawing substituents react in good yields (**3**, **8**, and **12**). Significantly decreased arylation yield was observed for *ortho*-substituted aryl iodides (e.g., **9**). The sterics of the benzylpicolinamides is also important for the regioselectivity of the arylation reaction. For instance, the less hindered *ortho* position is preferentially arylated (e.g., **14**) when a *meta* substituent is present on the benzylpicolinamide. Aryl bromides are much less reactive compared with aryl iodide substrates **4**. This is in accordance with results on the Pd-catalyzed PA-directed arylation of more inert C(sp<sup>3</sup>)–H bonds [29].

**Cyclization of biaryl compounds to form dihydrophenanthridines.** Next, we investigated the cyclization of biaryl compounds to form dihydrophenanthridines via Pd-catalyzed

intramolecular dehydrogenative amination of ε-C(sp<sup>2</sup>)–H bonds [37–45]. To our delight, treatment of **3** in the presence of 5 mol % of Pd(OAc)<sub>2</sub> and 2 equiv of PhI(OAc)<sub>2</sub> in toluene at 120 °C for 24 h gave the desired dihydrophenanthridine **16** in good yield (Table 2, entry 1). In addition, a further oxidized phenanthridine **17** was obtained as a side product. Compound **17** is presumably generated through the PhI(OAc)<sub>2</sub>-mediated oxidation of the benzylic C–H bond to form a phenanthridinium intermediate **18**, which then undergoes a removal of the PA group. Encouraged by these observations, we proceeded to explore whether the cyclization and oxidation steps can be performed in one step to give the phenanthridines in a shorter procedure. A variety of oxidants, such as 1,4-benzoquinone (BQ), KMnO<sub>4</sub>, ceric ammonium nitrate (CAN), and copper salts were examined [46]. The combination of PhI(OAc)<sub>2</sub> (2 equiv) and Cu(OAc)<sub>2</sub> (2 equiv) afforded the phenanthridine product **17** in highest yield (Table 2, entry 7). The yield can be further improved using 10 mol % of Pd(OAc)<sub>2</sub> catalyst (Table 2, entry 8). In a control experiment, dihydrophenanthridine **16** was oxidized with the application of Cu(OAc)<sub>2</sub> (2 equiv) in toluene at 120 °C for 24 h, forming **17** in excellent yield. We believe that PhI(OAc)<sub>2</sub> serves as the oxidant for the initial Pd-catalyzed intramolecular C–H amination step, in which a Pd<sup>II/IV</sup> catalytic manifold might be operative. Cu(OAc)<sub>2</sub> is responsible for the subsequent oxidation of the benzylic C–H bond of dihydrophenanthridine.

**Extension of the cyclization–oxidation step to other arylated picolinamide substrates.** The coupled cyclization–oxidation



**Figure 1:** Substrate scope of the Pd-catalyzed PA-directed C–H arylation reaction. All reactions were carried out in a 10 mL glass vial on a 0.2 mmol scale; yields are based on the isolated product.

step detailed above was then used to synthesize phenanthridines from other arylated picolinamide substrates (Figure 2). In general, electron-rich arene motifs, installed by C–H arylation, gave a higher yield of phenanthridine products; electron-deficient substrates provide a lower yield. For instance, substrate **8** with a *para*-nitro group failed to give any cyclized product under the standard conditions. Substrates with moderately electron-withdrawing groups, such as **20** bearing a *para*-ester group, reacted in moderate yield. The electronic properties of the benzylpicolinamide scaffold had much less influence on the reaction. For example, product **22** bearing an *ortho*-CF<sub>3</sub> substituent was obtained in 51% yield. Finally, it is noteworthy that all of the above phenanthridine products

show intense blue fluorescence. We expect our synthetic strategy will afford access to phenanthridines bearing varied substitution patterns, enabling applications in biology and materials science.

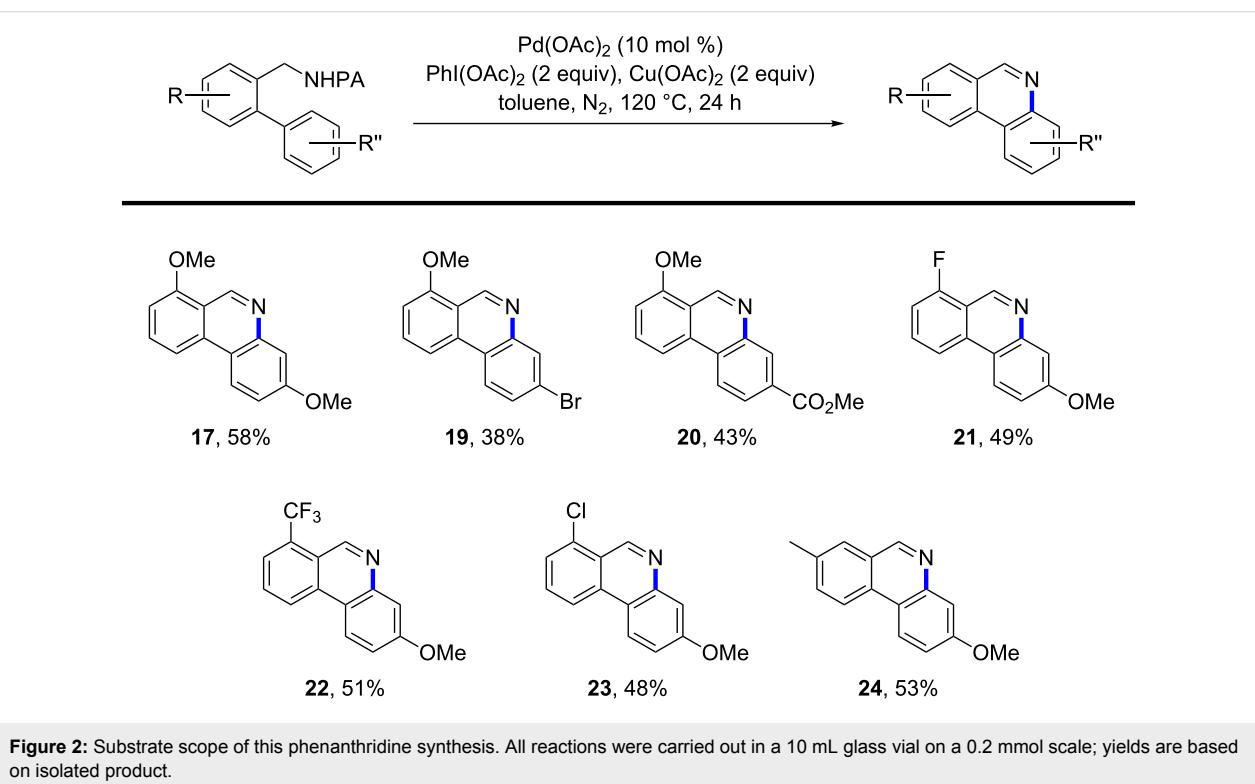
## Conclusion

In summary, we have developed a readily applicable two-step method for the synthesis of phenanthridines from easily accessible benzylamine picolinamides and aryl iodides. In the first step, an improved protocol allows us to carry out the Pd-catalyzed PA-directed C–H arylation reaction without the use of expensive silver additives. In the second step, application of PhI(OAc)<sub>2</sub> and Cu(OAc)<sub>2</sub> oxidant under the catalysis of

**Table 2:** Formation of phenanthridine **17** in a single step by Pd-catalyzed intramolecular C–H amination followed by oxidation. All screening reactions were carried out in a 10 mL glass vial on a 0.2 mmol scale.

entry	Pd(OAc) <sub>2</sub> (mol %)	additives (equiv)	yield (%) <sup>a</sup>	
			<b>16</b>	<b>17</b>
1	5	Phl(OAc) <sub>2</sub> (2)	40	5
2	5	Phl(OAc) <sub>2</sub> (2), AcOH (2)	23	8
3	5	Phl(OAc) <sub>2</sub> (2), BQ (2)	35	10
4	5	Phl(OAc) <sub>2</sub> (2), KMnO <sub>4</sub> (2)	56	3
5	5	Phl(OAc) <sub>2</sub> (2), CAN (2)	37	25
6	5	Phl(OAc) <sub>2</sub> (2), CuCl <sub>2</sub> (2)	29	34
7	5	Phl(OAc) <sub>2</sub> (2), Cu(OAc) <sub>2</sub> (2)	17	51
8	10	Phl(OAc) <sub>2</sub> (2), Cu(OAc) <sub>2</sub> (2)	15	62 (58) <sup>b</sup>

<sup>a</sup>Yields are based on <sup>1</sup>H NMR analysis of the reaction mixture after workup; <sup>b</sup>Isolated yield.



Pd(OAc)<sub>2</sub> affords phenanthridines in moderate to good yields. Applications of this method to the synthesis of more complex phenanthridines with novel photophysical properties are currently underway.

## Experimental

**General conditions:** All commercial materials were used as received unless otherwise noted. All solvents were obtained from a JC Meyer solvent dispensing system and used without

further purification. Flash chromatography was performed using 230–400 mesh SiliaFlash 60<sup>®</sup> silica gel (Silicycle Inc.). PhI(OAc)<sub>2</sub> (98%, Aldrich), Pd(OAc)<sub>2</sub> (98%, Aldrich) were used in the Pd-catalyzed reactions. NMR spectra were recorded on Bruker CDPX-300, DPX-300, DPX-400 instruments and calibrated by using residual solvent peaks as the internal reference. Multiplicities are recorded as: s = singlet, d = doublet, t = triplet, dd = doublet of doublets, m = multiplet. High-resolution ESI mass experiments were operated on a Waters LCT Premier instrument.

**Standard procedure for the Pd-catalyzed *ortho* C–H arylation reaction:** A mixture of picolinamide **1** [30] (48 mg, 0.2 mmol, 1 equiv), aryl iodide **2** (94 mg, 0.4 mmol, 2 equiv), Pd(OAc)<sub>2</sub> (2.2 mg, 0.01 mmol, 0.05 equiv), KHCO<sub>3</sub> (40 mg, 0.4 mmol, 2.0 equiv), and PivOH (6 mg, 0.06 mmol, 0.3 equiv) in anhydrous toluene (4 mL) in a 10 mL glass vial (purged with N<sub>2</sub>, sealed with PTFE cap) was heated at 120 °C for 24 h. The reaction mixture was filtered through a short pad of celite and concentrated in vacuo. The resulting residue was purified by silica-gel flash chromatography (hexanes/EtOAc 3:1) to give the product **3** as a pale white solid (64 mg, 91%). Compounds **4–15** were prepared from the known precursors [30] by using the standard C–H arylation procedure.

**Standard procedure for the Pd-catalyzed cyclization and oxidation reaction to form phenanthridines:** A mixture of picolinamide **3** (70 mg, 0.2 mmol, 1 equiv), Pd(OAc)<sub>2</sub> (4.4 mg, 0.02 mmol, 0.1 equiv), PhI(OAc)<sub>2</sub> (129 mg, 0.4 mmol, 2.0 equiv), and Cu(OAc)<sub>2</sub> (72 mg, 0.4 mmol, 2 equiv) in anhydrous toluene (4 mL) in a 10 mL glass vial (purged with N<sub>2</sub>, sealed with PTFE cap) was heated at 120 °C for 24 h. The reaction mixture was filtered through a short pad of celite and concentrated in vacuo. The resulting residue was purified by silica gel flash chromatography (hexanes/EtOAc 4:1) to give the product **17** as a pale white solid (28 mg, 58%). Compounds **19–24** were prepared by using the standard cyclization–oxidation procedure.

**Compound 3.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.53 (d, *J* = 4.2 Hz, 1H), 8.40 (s, 1H), 8.21 (d, *J* = 7.5 Hz, 1H), 7.85–7.80 (m, 1H), 7.42–7.30 (m, 3H), 6.99–6.93 (m, 4H), 4.65 (d, *J* = 5.4 Hz, 2H), 3.95 (s, 3H), 3.85 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.3, 158.8, 158.6, 150.2, 147.9, 143.6, 137.1, 132.7, 130.3, 128.3, 125.8, 123.4, 122.7, 122.1, 113.6, 109.1, 55.7, 55.2, 36.8; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>, 349.1552; found, 349.1546.

**Compound 4.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.52 (d, *J* = 4.5 Hz, 1H), 8.36 (s, 1H), 8.19 (d, *J* = 7.8 Hz, 1H), 7.84 (td, *J* = 7.8, 1.4 Hz, 1H), 7.55 (d, *J* = 8.3 Hz, 2H), 7.41 (dd, *J* = 5.0,

6.8 Hz, 1H), 7.35 (t, *J* = 8.0 Hz, 1H), 7.25 (d, *J* = 8.3 Hz, 2H), 6.97 (d, *J* = 8.2 Hz, 1H), 6.89 (d, *J* = 7.7 Hz, 1H), 4.59 (d, *J* = 5.5 Hz, 2H), 3.95 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 163.3, 158.7, 150.1, 148.0, 142.5, 139.4, 137.1, 131.3, 130.9, 128.6, 125.9, 123.5, 122.4, 122.1, 121.6, 109.9, 55.8, 36.7; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>18</sub>BrN<sub>2</sub>O<sub>2</sub>, 397.0552; found, 397.0561.

**Compound 5.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 8.51 (d, *J* = 4.0 Hz, 1H), 8.33 (s, 1H), 8.15 (d, *J* = 7.6 Hz, 1H), 8.08 (d, *J* = 8.0 Hz, 1H), 7.80 (t, *J* = 7.6 Hz, 1H), 7.44–7.34 (m, 4H), 6.97 (d, *J* = 8.0 Hz, 1H), 6.90 (d, *J* = 7.6 Hz, 1H), 4.58 (d, *J* = 5.2 Hz, 2H), 3.95 (s, 3H), 3.92 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.9, 163.3, 158.7, 150.2, 148.0, 145.2, 142.9, 137.2, 129.5, 129.3, 129.0, 128.6, 125.9, 123.5, 122.3, 122.2, 110.0, 55.8, 52.1, 36.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>21</sub>N<sub>2</sub>O<sub>4</sub>, 377.1501; found, 377.1509.

**Compound 6.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.51 (d, *J* = 4.2 Hz, 1H), 8.34 (s, 1H), 8.19 (d, *J* = 7.8 Hz, 1H), 7.40–7.31 (m, 1H), 6.98–6.91 (m, 5H), 4.66 (d, *J* = 5.4 Hz, 2H), 3.94 (s, 3H), 3.79 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.8, 159.7, 159.0, 150.7, 148.4, 144.4, 142.3, 137.6, 129.7, 128.9, 126.3, 124.0, 123.0, 122.6, 122.1, 114.9, 113.8, 110.1, 55.3, 55.6, 37.2; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>, 349.1552; found, 349.1564.

**Compound 7.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.53 (d, *J* = 4.2 Hz, 1H), 8.34 (s, 1H), 8.19 (d, *J* = 7.8 Hz, 1H), 7.84–7.80 (m, 1H), 7.40–7.29 (m, 2H), 6.96–6.80 (m, 5H), 6.00 (s, 2H), 4.64 (d, *J* = 5.4 Hz, 1H), 3.96 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.4, 158.7, 150.3, 148.0, 147.4, 146.9, 143.7, 137.1, 134.4, 128.4, 125.8, 123.7, 122.8, 122.7, 122.2, 109.9, 109.5, 108.1, 101.0, 55.8, 36.8; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>20</sub>N<sub>2</sub>O<sub>4</sub>, 363.1345; found, 363.1355.

**Compound 8.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.53 (d, *J* = 4.1 Hz, 1H), 8.41 (s, 1H), 8.29 (d, *J* = 8.8 Hz, 2H), 8.16 (d, *J* = 7.8 Hz, 1H), 7.85 (td, *J* = 7.7, 1.6 Hz, 1H), 7.55 (d, *J* = 8.7 Hz, 2H), 7.42–7.34 (m, 2H), 7.03 (d, *J* = 8.2 Hz, 1H), 6.89 (d, *J* = 7.7 Hz, 1H), 4.58 (d, *J* = 5.7 Hz, 2H), 3.98 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 163.4, 158.8, 150.0, 148.1, 147.4, 147.1, 141.4, 137.3, 130.4, 128.9, 126.1, 123.7, 123.5, 122.2, 122.2, 110.7, 56.0, 36.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>18</sub>N<sub>3</sub>O<sub>4</sub>, 364.1297; found, 364.1292.

**Compound 9.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.53 (d, *J* = 4.2 Hz, 1H), 8.32 (s, 1H), 8.18–8.16 (m, 1H), 7.85–7.80 (m, 1H), 7.38–7.30 (m, 4H), 7.25–7.13 (m, 2H), 7.02–6.92 (m, 2H), 4.64–4.90 (m, 2H), 3.98 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ

163.8, 161.2, 158.9, 150.7, 148.4, 137.8, 137.6, 132.0, 129.9, 128.9, 126.2, 125.2, 124.5, 123.3, 122.6, 115.9, 115.8, 110.7, 56.22, 37.2; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub>, 337.1352; found, 337.1364.

**Compound 10.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.51 (d, *J* = 4.0 Hz, 1H), 8.25 (s, 1H), 8.17 (d, *J* = 7.8 Hz, 1H), 7.84 (td, *J* = 7.7, 1.7 Hz, 1H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.66 (d, *J* = 8.3 Hz, 1H), 7.41–7.36 (m, 1H), 7.32–7.24 (m, 3H), 7.17 (d, *J* = 8.3 Hz, 1H), 7.06 (s, 1H), 6.94 (d, *J* = 8.2 Hz, 1H), 6.88 (d, *J* = 7.4 Hz, 1H), 4.57 (d, *J* = 5.4 Hz, 2H), 3.95 (t, *J* = 8.8 Hz, 2H), 3.92 (s, 3H), 2.93 (t, *J* = 8.4 Hz, 2H), 2.37 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 163.3, 158.7, 150.2, 148.1, 144.1, 143.4, 141.3, 137.2, 136.1, 133.9, 131.8, 129.8, 128.7, 128.5, 127.3, 126.2, 125.9, 123.5, 122.6, 122.1, 114.5, 109.5, 55.8, 50.1, 36.7, 27.8, 21.5; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>29</sub>H<sub>18</sub>N<sub>3</sub>O<sub>4</sub>S, 514.1801; found: 514.1813.

**Compound 11.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.54 (d, *J* = 3.9 Hz, 1H), 8.21 (d, *J* = 7.5 Hz, 2H), 7.88–7.83 (m, 1H), 7.46–7.41 (m, 2H), 7.34–7.23 (m, 4H), 6.96 (d, *J* = 8.7 Hz, 1H), 4.72 (d, *J* = 5.4 Hz, 1H), 8.86 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.5, 159.1, 149.8, 148.0, 144.9, 137.2, 135.8, 132.9, 132.4, 130.1, 129.2, 128.7, 128.6, 126.0, 122.2, 113.8, 55.2, 39.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>18</sub>ClN<sub>2</sub>O<sub>2</sub>, 353.1057; found, 353.1067.

**Compound 12.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.50 (d, *J* = 4.8 Hz, 1H), 8.14 (d, *J* = 7.8 Hz, 1H), 7.93 (s, 1H), 7.83–7.74 (m, 2H), 7.50–7.39 (m, 3H), 7.28–7.22 (m, 2H), 6.89 (d, *J* = 8.7 Hz, 2H), 4.71 (d, *J* = 4.5 Hz, 1H), 3.82 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.0, 159.1, 149.6, 148.0, 145.6, 137.1, 134.6, 133.5, 131.9, 130.1, 129.9, 127.8, 126.0, 125.4, 122.1, 113.8, 55.2, 38.2; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>18</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub>, 387.1320; found, 387.1328.

**Compound 13.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.51 (d, *J* = 4.2 Hz, 1H), 8.18 (d, *J* = 7.8 Hz, 2H), 7.83–7.79 (m, 1H), 7.42–7.38 (m, 1H), 7.33–7.30 (m, 3H), 7.11 (d, *J* = 7.5 Hz, 2H), 6.97 (d, *J* = 8.7 Hz, 2H), 4.68 (d, *J* = 5.4 Hz, 2H), 3.84 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.7, 160.4, 159.1, 149.7, 147.9, 144.4, 137.1, 131.6, 130.1, 128.9, 128.7, 126.0, 122.6, 122.1, 114.3, 113.8, 55.2, 35.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub>, 337.1352; found, 337.1357.

**Compound 14.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.50 (d, *J* = 4.5 Hz, 1H), 8.27–8.21 (m, 2H), 7.82 (t, *J* = 7.5 Hz, 1H), 7.41–7.29 (m, 4H), 7.19 (dd, *J* = 14.1 and 7.8 Hz, 2H), 6.98 (d, *J* = 8.4 Hz, 2H), 4.64 (d, *J* = 6.0 Hz, 2H), 3.84 (s, 3H), 2.38 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 163.8, 158.5, 149.6, 147.8, 138.4, 137.0, 136.9, 135.2, 132.8, 130.0, 129.1, 127.9, 125.9,

122.0, 113.5, 55.0, 41.1, 20.9; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub>, 333.1603; found, 333.1609.

**Compound 15.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 8.45 (d, *J* = 4.7 Hz, 1H), 8.01 (d, *J* = 7.8 Hz, 1H), 7.78 (m, 2H), 7.42 (s, 2H), 7.37 (m, 1H), 7.30 (d, *J* = 8.7 Hz, 4H), 6.90 (d, *J* = 8.7 Hz, 4H), 4.47 (d, *J* = 5.1 Hz, 2H), 3.79 (s, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 162.9, 159.1, 149.5, 147.8, 145.2, 137.0, 132.3, 132.2, 132.1, 129.9, 125.9, 121.8, 121.0, 113.8, 55.2, 39.0; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>27</sub>H<sub>24</sub>Br<sub>3</sub>N<sub>2</sub>O<sub>3</sub>, 503.0970; found, 503.0975.

**Compound 17.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.72 (s, 1H), 8.45 (d, *J* = 9.0 Hz, 1H), 8.08 (d, *J* = 8.2 Hz, 1H), 7.75 (t, *J* = 8.1 Hz, 1H), 7.61 (d, *J* = 2.6 Hz, 1H), 7.33–7.29 (m, 1H), 6.99 (d, *J* = 7.8 Hz, 1H), 4.09 (s, 3H), 4.00 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 160.5, 157.5, 149.1, 147.2, 134.6, 132.1, 124.3, 118.5, 117.1, 113.8, 110.1, 105.9, 56.2, 56.0; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>14</sub>NO<sub>2</sub>, 240.1025; found, 240.1030.

**Compound 19.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 9.68 (s, 1H), 8.33 (m, 2H), 8.04 (d, *J* = 8.3 Hz, 1H), 7.75 (m, 2H), 7.04 (d, *J* = 7.9 Hz, 1H), 4.05 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 157.6, 149.5, 145.7, 133.8, 132.5, 132.2, 130.1, 124.3, 122.5, 117.4, 113.7, 107.2, 55.9; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>11</sub>BrNO, 228.0024; found, 228.0032.

**Compound 20.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.75 (s, 1H), 8.84 (s, 1H), 8.54 (d, *J* = 8.7 Hz, 1H), 8.24 (d, *J* = 7.5 Hz, 1H), 8.14 (d, *J* = 8.4 Hz, 1H), 7.79 (d, *J* = 8.1 Hz, 1H), 7.09 (d, *J* = 8.1 Hz, 1H), 4.06 (s, 3H), 4.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 166.8, 157.4, 149.2, 144.0, 133.2, 132.1, 132.0, 130.1, 127.0, 126.7, 122.9, 117.9, 114.2, 107.8, 55.8, 52.4; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>16</sub>H<sub>13</sub>NO<sub>3</sub>, 268.0974; found, 268.0970.

**Compound 21.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.59 (s, 1H), 8.46 (d, *J* = 9.0 Hz, 1H), 8.29 (d, *J* = 8.4 Hz, 1H), 7.82–7.74 (m, 1H), 7.64 (s, 1H), 7.37–7.26 (m, 2H), 4.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 160.6, 147.4, 146.6, 134.6, 131.7, 131.6, 123.8, 118.7, 117.4, 117.3, 111.1, 110.8, 110.1, 55.7; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>10</sub>FNO, 228.0825; found, 228.0830.

**Compound 22.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.62 (s, 1H), 8.72 (d, *J* = 8.4 Hz, 1H), 8.47 (d, *J* = 9.0 Hz, 1H), 7.95 (d, *J* = 7.2 Hz, 1H), 7.85 (t, *J* = 8.1 Hz, 1H), 7.62 (d, *J* = 2.4 Hz, 1H), 7.36 (dd, *J* = 9.0 Hz and 2.7 Hz, 1H), 4.01 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 160.7, 149.3, 145.8, 133.8, 129.5, 125.8, 124.4, 124.3, 123.4, 122.2, 119.1, 117.6, 109.8,

55.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>12</sub>F<sub>3</sub>NO, 278.0793; found, 278.0797.

**Compound 23.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.72 (s, 1H), 8.46–8.42 (m, 2H), 7.72–7.62 (m, 3H), 7.35 (dd, *J* = 9.0 and 2.4 Hz, 1H), 4.02 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz, ppm) δ 160.5, 150.1, 146.1, 134.5, 133.9, 131.0, 126.9, 123.6, 122.3, 120.4, 118.8, 117.1, 109.9, 55.6; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>11</sub>CINO, 244.0529; found, 244.0534.

**Compound 24.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 9.23 (s, 1H), 8.48–8.42 (m, 2H), 7.82 (s, 1H), 7.69 (d, *J* = 8.1 Hz, 1H), 7.62 (s, 1H), 7.34–7.30 (m, 1H), 4.02 (s, 3H), 2.62 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) δ 160.2, 154.2, 146.3, 137.4, 136.7, 133.4, 131.1, 128.5, 123.6, 121.7, 118.2, 118.4, 110.3, 56.0, 21.9; HRMS (*m/z*): [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>13</sub>NO, 244.1075; found, 244.1079.

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