

# Synthesis of skeletally diverse alkaloid-like molecules: exploitation of metathesis substrates assembled from triplets of building blocks

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## Full Research Paper

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## Abstract

A range of metathesis substrates was assembled from triplets of unsaturated building blocks. The approach involved the iterative attachment of a propagating and a terminating building block to a fluorine-tagged initiating building block. Metathesis cascade chemistry was used to “reprogram” the molecular scaffolds. Remarkably, in one case, a cyclopropanation reaction competed with the expected metathesis cascade process. Finally, it was demonstrated that the metathesis products could be derivatised to yield the final products. At each stage, purification was facilitated by the presence of a fluorine-tagged protecting group.

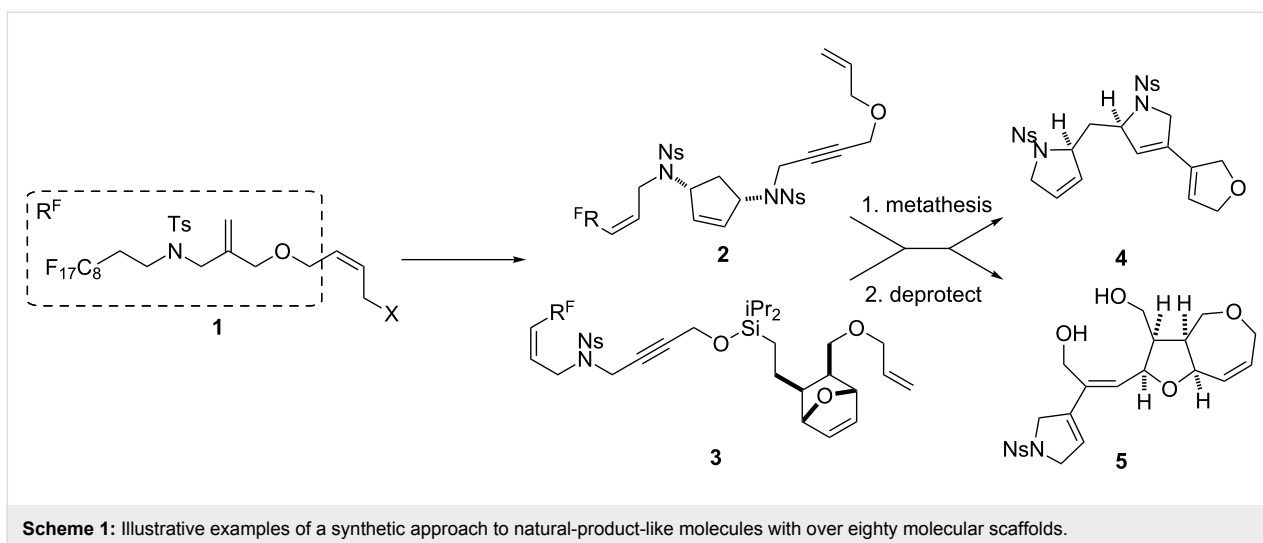
## Introduction

Our collective understanding of the biological relevance of chemical space has been shaped, in large part, by the historic exploration of chemical space by chemical synthesis (and biosynthesis) [1]. The scaffolds of known bioactive small molecules, in particular, play a key role in guiding the navigation of chemical space [2-4]. The field of biology-oriented synthesis (BIOS) [5], for example, uses biologically validated scaffolds [6-8] to inspire library design.

Known organic molecules populate chemical space unevenly and unsystematically. Around half of all known organic com-

pounds are based on only 0.25% of the known molecular scaffolds [9]! This uneven coverage of chemical space is also typical of small-molecule screening collections [7,10]. Consequently, the biological relevance of most known scaffolds has been poorly explored. The field of diversity-oriented synthesis [11-13] has emerged with the specific aim of populating screening collections with diverse and novel small molecules.

We have previously developed a robust approach for the synthesis of skeletally diverse small molecules (Scheme 1) [14]. The approach relied on the synthesis of metathesis substrates by



iterative attachment of simple unsaturated building blocks to a fluorine-tagged linker **1** (e.g., → **2** or **3**). Subsequently, metathesis cascade reactions were used to “reprogram” the molecular scaffolds, concomitantly releasing the products from the linker (e.g., → **4** or **5**) [14–17]. The approach enabled the combinatorial variation of molecular scaffolds, and was exploited in the synthesis of natural-product-like small molecules with unprecedented scaffold diversity (over 80 distinct scaffolds).

Although powerful, this general approach to skeletally diverse molecules had only been exemplified by varying pairs of unsaturated building blocks [14]. Thus, by exploiting the linker **1**, which is an allyl alcohol or allyl amine equivalent, all of the products were inevitably allylic alcohols or cyclic allylic amines. Here, we demonstrate that the approach is considerably more general, and that it is feasible to exploit triplets of building blocks, extending the range of diverse molecular scaffolds that may be prepared.

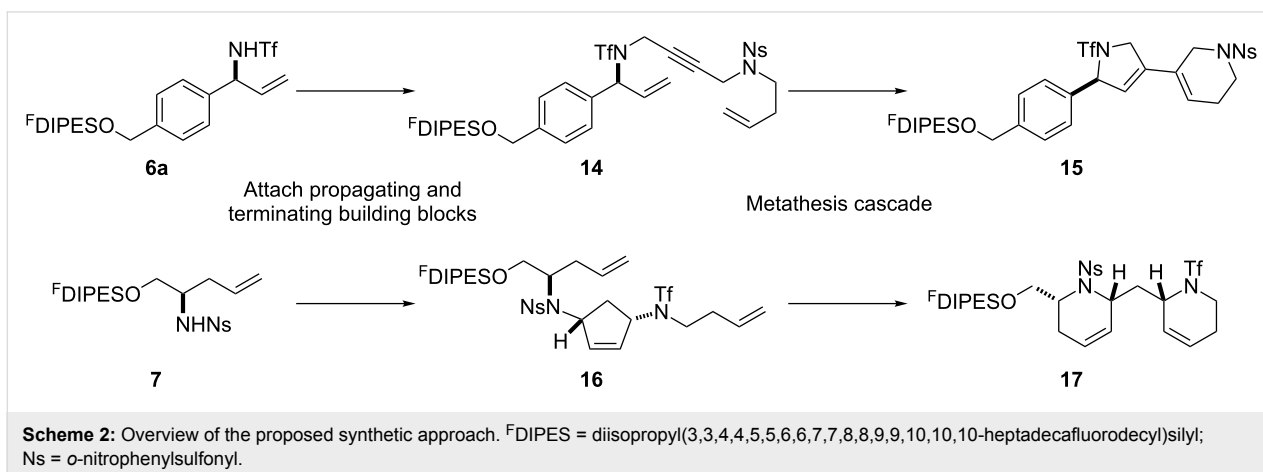
## Results and Discussion

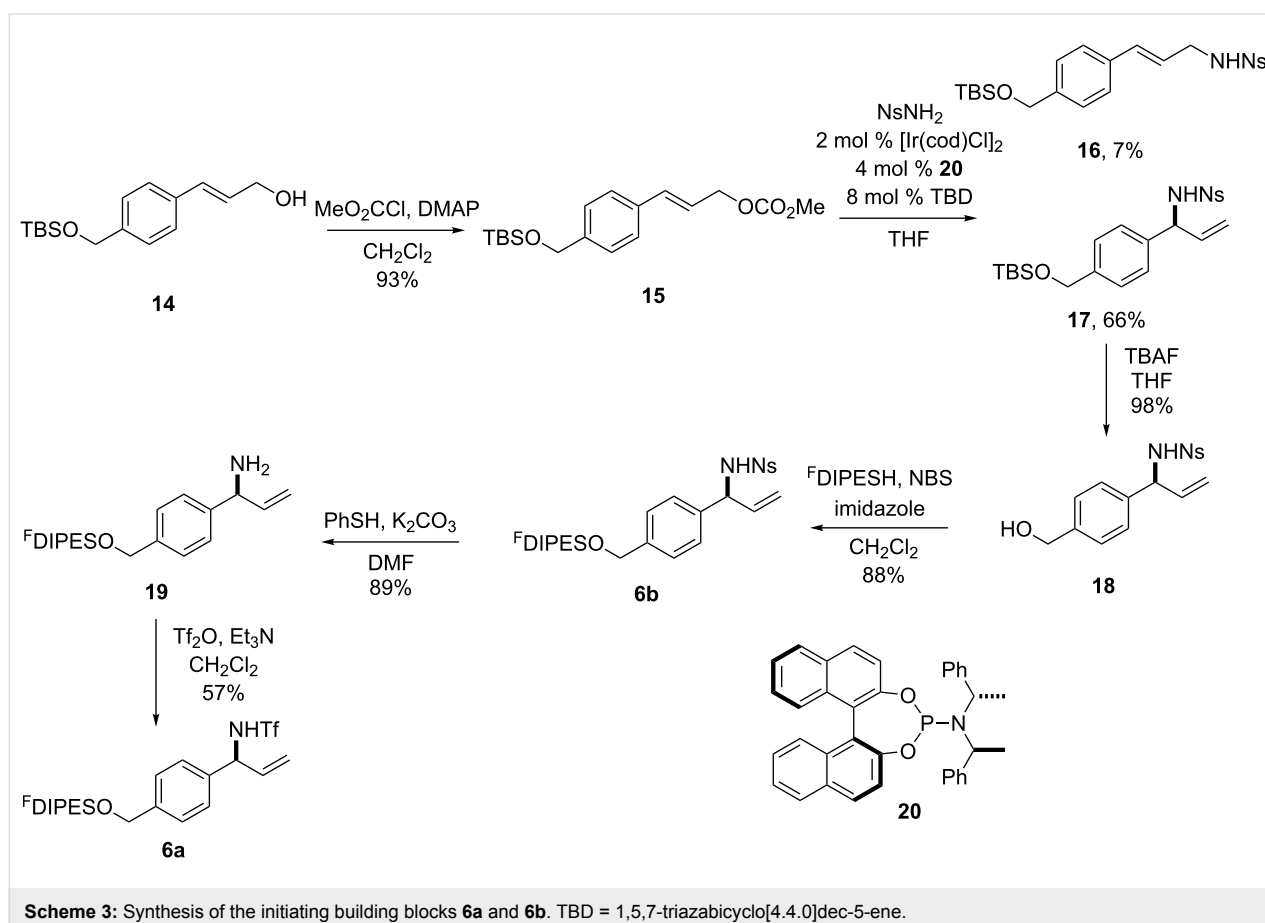
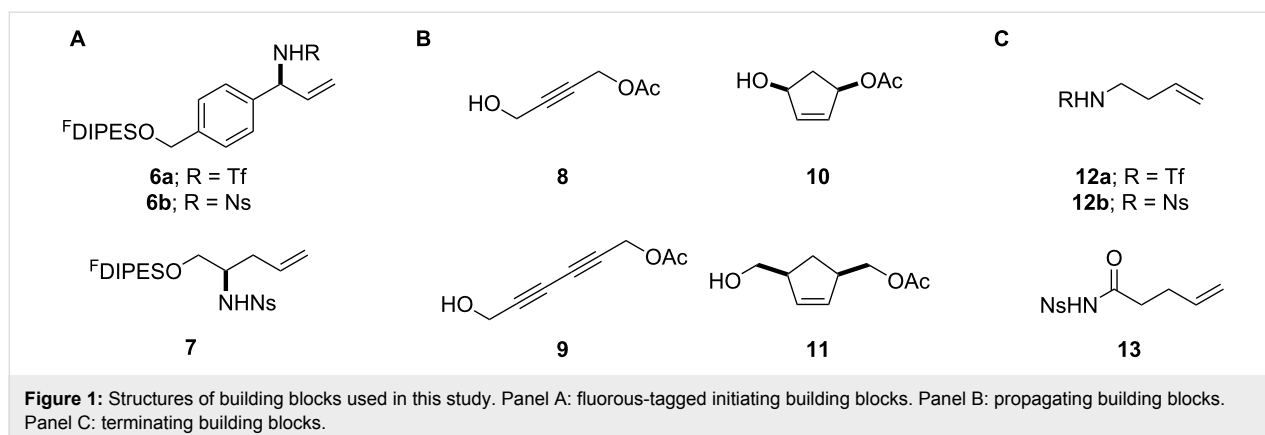
### Library design

An overview of the proposed approach to the synthesis of diverse scaffolds is shown in Scheme 2. The building blocks used in this study are shown in Figure 1. It was planned to start with an “initiating” building block (e.g., **6a** or **7**) bearing a fluorine tag to facilitate the purification of synthetic intermediates [18]. Iterative attachment of a propagating and a terminating building block would yield a metathesis substrate (such as **14** or **16**). Finally, a metathesis cascade reaction would yield a product scaffold (such as **15** and **17**). It was planned that many of the product scaffolds would bear an *o*-nitrophenylsulfonyl protecting group. The combinations of building blocks were carefully chosen to ensure that, after deprotection, selective derivatisation of the product scaffolds would be possible.

### Synthesis of building blocks

The initiating building blocks **6a** and **6b** were prepared by using the approach outlined in Scheme 3. The allylic alcohol **14** [19]

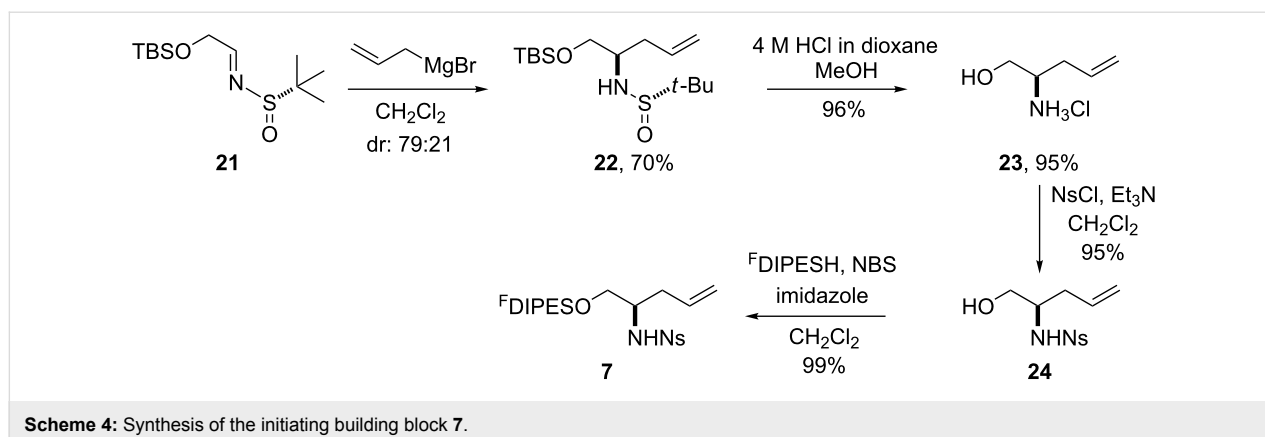




was converted into the allylic carbonate **15** by treatment with methyl chloroformate and DMAP. The allylic carbonate **15** underwent efficient asymmetric allylic amination [20] with *o*-nitrophenylsulfonamide as the nucleophile to give the allylic sulfonamide **17** in 66% yield; in addition, the linear product **16** was also obtained in 7% yield. Desilylation of **17** ( $\rightarrow$  **18**) and reaction with diisopropyl(3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-heptafluorodecyl)silyl (<sup>F</sup>DIPES) bromide, generated in situ from the corresponding silyl hydride, gave the fluororous-tagged

building block **6b**. Finally, desulfonation ( $\rightarrow$  **19**) and trifluoromethylsulfonation yielded the alternative initiating building block **6a**.

The initiating building block **7** was prepared from the sulfinimine **21** by adapting a synthesis previously reported by Ellman (Scheme 4) [21]. Treatment of the sulfinimine **21** in dichloromethane with allylmagnesium bromide yielded the corresponding sulfinimides as a 79:21 mixture of diastereoisomers;



following column chromatography, the major diastereomer **22** was obtained in 70% yield, and was converted into the corresponding amino alcohol **23**. The configuration of the amino alcohol **23** was determined by conversion into the corresponding benzamide and comparison with racemic and enantiomerically enriched samples (prepared from the commercially available amino acid). Analysis by chiral HPLC indicated that the amino alcohol **23** had (*R*)-configuration. It was concluded that the sense of diastereoselectivity in the addition **21** → **22** contrasted with that reported by Ellman [21]. However, the sense of diastereoselectivity was the same as that reported for the addition of allylmagnesium bromide in dichloromethane to a similar sulfinimine [22]. The amino alcohol **23** was converted into the corresponding *o*-nitrophenylsulfonamide **24** and, hence, the fluorinated building block **7**.

The propagating building blocks **8–11**, and the terminating building block **12b**, were prepared by using established methods [14]. The enantiomeric excess (68% ee) of the hydroxy alcohol **11** was determined by conversion into the corre-

sponding diastereomeric *O*-methyl mandelate esters. The terminating building blocks **12a** and **13** were prepared by straightforward derivatisation of commercially available starting materials (see Supporting Information File 1).

### Synthesis of metathesis substrates

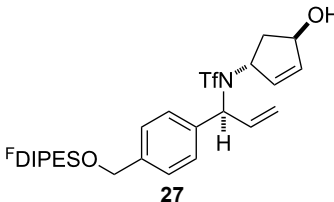
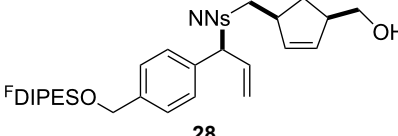
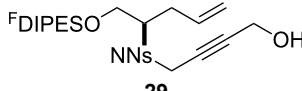
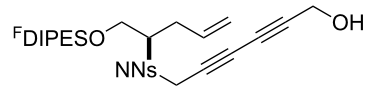
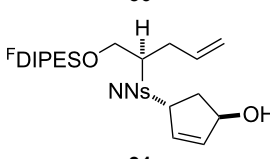
Initially, the propagating building blocks **8–11** were attached to the fluorinated initiating building blocks (**6a**, **6b** or **7**). In each case, an excess of the propagating building block, DEAD and triphenylphosphine was used. In general, the crude product was directly deacetylated. At each stage, the required fluorinated product was isolated by fluorinated-solid-phase extraction (F-SPE), and its purity determined by analysis by 500 MHz <sup>1</sup>H NMR spectroscopy. These results are summarised in Table 1.

The metathesis substrates were prepared by subsequent attachment of a terminating building block (**12a**, **12b** or **13**) (see Table 2). In each case, an excess of the terminating building block, DEAD and triphenylphosphine was used; the required

**Table 1:** Attachment of propagating building blocks to the fluorinated initiating building blocks.

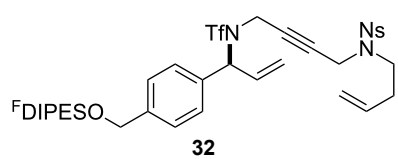
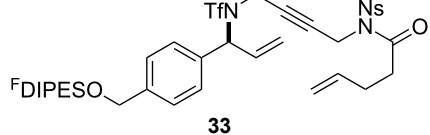
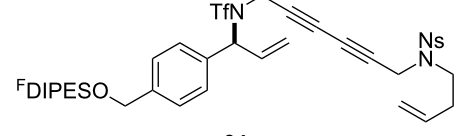
Building blocks	Attachment	Deacetylation <sup>a</sup>	Product
	Method <sup>a</sup> (mass recovery / %) {Purity <sup>b</sup> / %}	Mass recovery / % (Purity <sup>b</sup> / %)	
<b>6a, 8</b>	A1 (70) {>98}	92 (92)	
<b>6a, 9</b>	A2 (97) {92}	87 (93)	

**Table 1:** Attachment of propagating building blocks to the fluororous-tagged initiating building blocks. (continued)

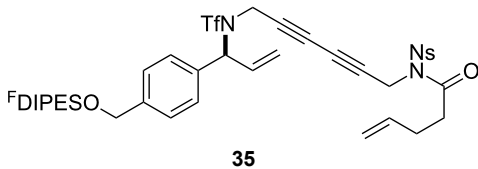
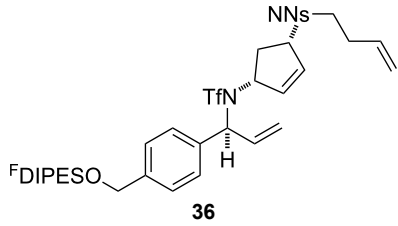
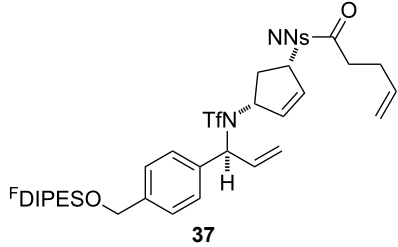
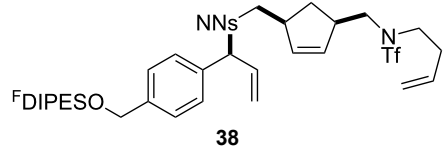
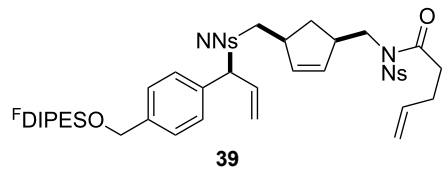
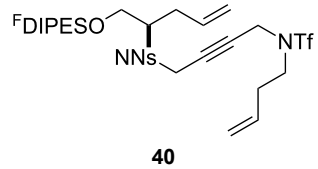
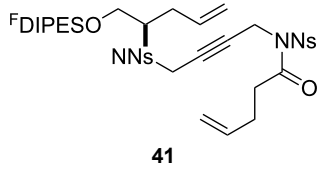
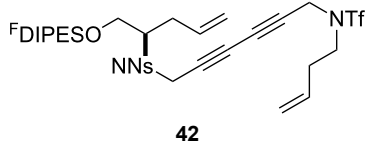
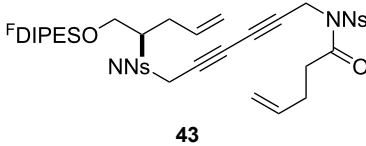
<b>6a, 10<sup>c</sup></b>	A3 (85) {>98}	87 (98)	
<b>6b, 11<sup>d</sup></b>	A3 (85) {76}	85 <sup>e</sup> (72)	
<b>7, 8</b>	A3 (92) {91}	94 <sup>f</sup>	
<b>7, 9</b>	A3 (74 <sup>f</sup> )	97 (98)	
<b>7, 10</b>	A3 (97) {91}	80 <sup>f</sup>	

<sup>a</sup>Methods: A1: Initiating building block (1.0 equiv), propagating building block (4.0 equiv), DEAD (4.0 equiv), PPh<sub>3</sub> (4.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0 °C → rt then F–SPE; A2: Initiating building block (1.0 equiv), propagating building block (4.0 equiv), DEAD (2.0 equiv), PPh<sub>3</sub> (2.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0 °C → rt then F–SPE; A3: Initiating building block (1.0 equiv), propagating building block (4.0 equiv), DEAD (2.0 equiv), PPh<sub>3</sub> (2.0 equiv), THF, 0 °C → rt then F–SPE; Deacetylation: 0.025 M NH<sub>3</sub> in MeOH. <sup>b</sup>Determined by analysis of the 500 MHz <sup>1</sup>H NMR spectrum. <sup>c</sup>The building block had >98% ee. <sup>d</sup>The building block had 68% ee. <sup>e</sup>Isolated as a ca. 75:25 mixture of diastereoisomers. <sup>f</sup>Isolated yield of purified product (see Supporting Information File 1).

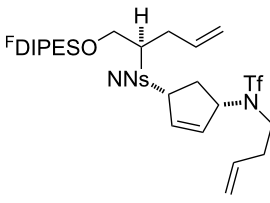
**Table 2:** Attachment of propagating building blocks to the fluororous-tagged initiating building blocks.

Substrate	Terminating building block	Attachment Method <sup>a</sup> (mass recovery / %) {Purity <sup>b</sup> / %}	Product
<b>25</b>	<b>12b</b>	A4 (89) {83}	
<b>25</b>	<b>13</b>	A4 (89) {86}	
<b>26</b>	<b>12b</b>	A4 (76) {93}	

**Table 2:** Attachment of propagating building blocks to the fluoros-tagged initiating building blocks. (continued)

26	13	A4 (75) {97}	 <p style="text-align: center;"><b>35</b></p>
27	12b	A5 (62 <sup>c</sup> )	 <p style="text-align: center;"><b>36</b></p>
27	13	A5 (54 <sup>c</sup> )	 <p style="text-align: center;"><b>37</b></p>
28 <sup>d</sup>	12a	A5 (86 <sup>c,e</sup> )	 <p style="text-align: center;"><b>38</b></p>
28 <sup>d</sup>	13	A5 (77 <sup>c,e</sup> )	 <p style="text-align: center;"><b>39</b></p>
29	12a	A6 (86 <sup>c</sup> )	 <p style="text-align: center;"><b>40</b></p>
29	13	A6 (77 <sup>c</sup> )	 <p style="text-align: center;"><b>41</b></p>
30	12a	A6 (92 <sup>c</sup> )	 <p style="text-align: center;"><b>42</b></p>
30	13	A6 (55 <sup>c</sup> )	 <p style="text-align: center;"><b>43</b></p>

**Table 2:** Attachment of propagating building blocks to the fluoros-tagged initiating building blocks. (continued)

<b>31</b>	<b>12a</b>	A6 (85 <sup>c</sup> )	 <p style="text-align: center;"><b>44</b></p>
<p><sup>a</sup>Methods: A4: Substrate (1.0 equiv), propagating building block (4.0 equiv), DEAD (2.0 equiv), PPh<sub>3</sub> (2.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0 °C → rt then F-SPE; A5: Substrate (1.0 equiv), propagating building block (4.0 equiv), DEAD (4.0 equiv), PPh<sub>3</sub> (4.0 equiv), THF, 0 °C → rt then F-SPE; A6: Substrate (1.0 equiv), propagating building block (4.0 equiv), DEAD (2.0 equiv), PPh<sub>3</sub> (2.0 equiv), THF, 0 °C → rt then F-SPE. <sup>b</sup>Determined by analysis of the 500 MHz <sup>1</sup>H NMR spectrum. <sup>c</sup>Isolated yield of purified product. <sup>d</sup>The starting material was a ca. 75:25 mixture of diastereoisomers. <sup>e</sup>Isolated as a ca. 75:25 mixture of diastereomers.</p>			

fluorous-tagged product was isolated by solid-fluorous phase extraction (F-SPE), and its purity was determined by analysis by 500 MHz <sup>1</sup>H NMR spectroscopy.

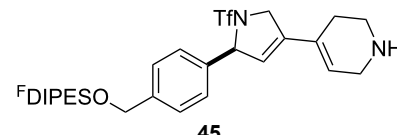
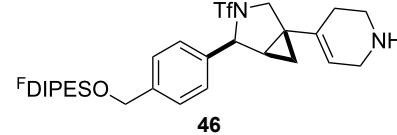
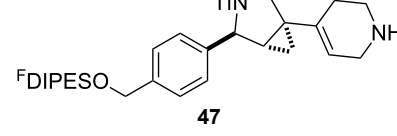
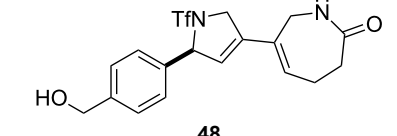
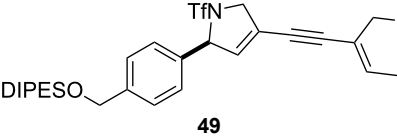
### Metathesis cascade reactions

The scaffolds of the metathesis substrates were “reprogrammed” by treatment with Hoveyda–Grubbs second-generation catalyst in either dichloromethane or *tert*-butyl methyl ether [23] (TBME). Many of the metathesis reactions were rather sluggish, and the catalyst was added portionwise until the reac-

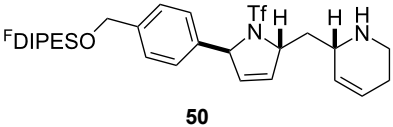
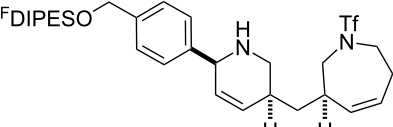
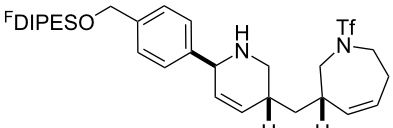
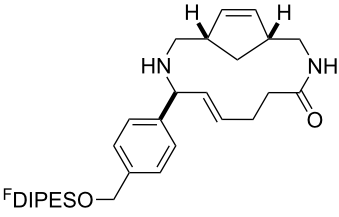
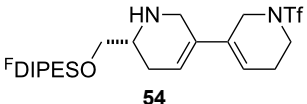
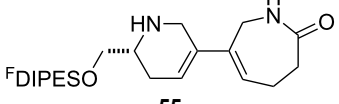
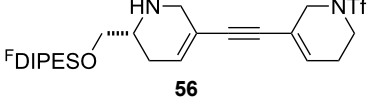
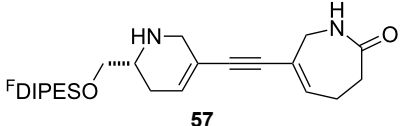
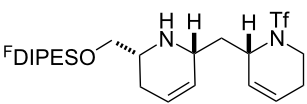
tions were judged to be complete by TLC analysis. After removal [24] of the catalyst by using tris(hydroxymethyl)phosphine, the metathesis products were generally purified by flash column chromatography. Finally, the *o*-nitrophenylsulfonyl groups were removed from the products. The results are summarised in Table 3.

In general, the metathesis reactions proceeded smoothly to give the expected metathesis cascade products. In the case of **39**, however, the cyclopentene did not participate in the metathesis

**Table 3:** Application of cascade metathesis reactions in the synthesis of diverse scaffolds and subsequent desulfonation.

Substrate	Method <sup>a</sup> (mol %; time)	Product	Yield / %
<b>32</b>	B1 (5 + 2.5; 3 d) then C1	 <p style="text-align: center;"><b>45</b></p>	<b>45</b> , 37% <sup>b</sup>
		 <p style="text-align: center;"><b>46</b></p>	<b>46</b> , 11% <sup>b</sup>
		 <p style="text-align: center;"><b>47</b></p>	<b>47</b> , 5% <sup>b</sup>
<b>33</b>	B1 (2 × 5; 4 d) then C1 then D	 <p style="text-align: center;"><b>48</b></p>	43% <sup>b</sup>
<b>34</b>	B1 (5 + 5 + 2.5; 10 d) then C1	 <p style="text-align: center;"><b>49</b></p>	49% <sup>b</sup> (86% <sup>c</sup> )

**Table 3:** Application of cascade metathesis reactions in the synthesis of diverse scaffolds and subsequent desulfonylation. (continued)

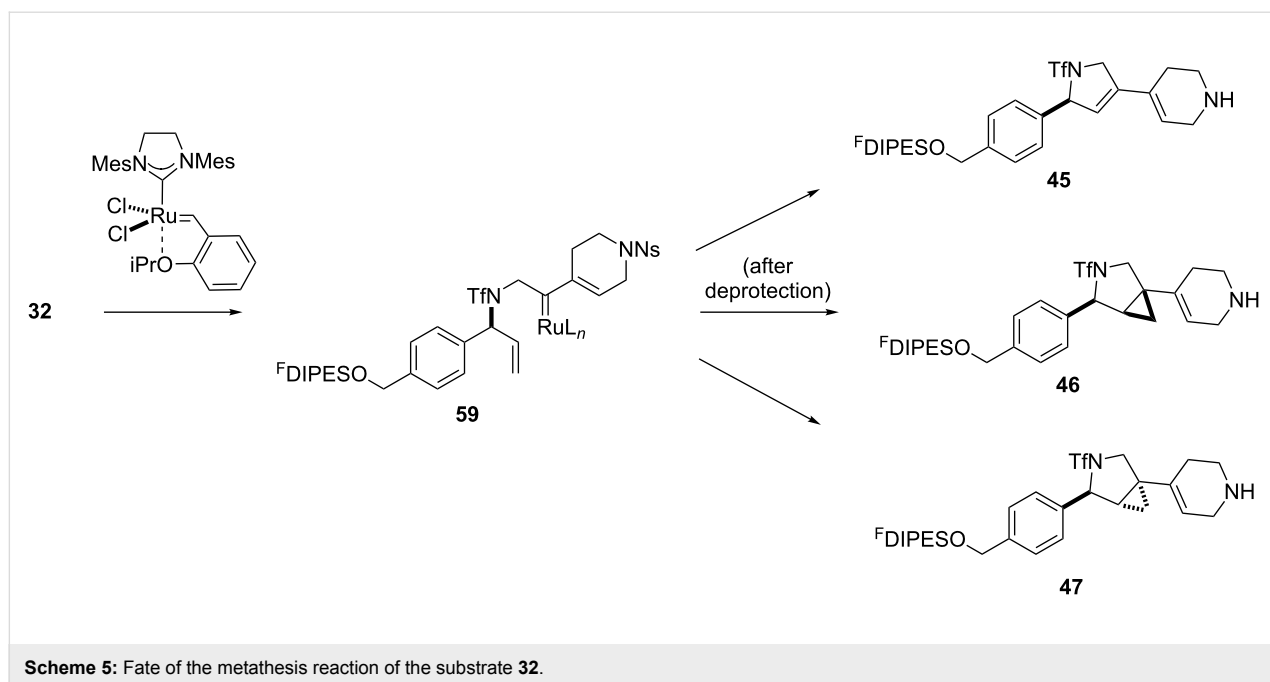
36	B1 (4 × 5; 20 d) then C1	 <p style="text-align: center;"><b>50</b></p>	77 then 81 (93% <sup>c</sup> )
38 <sup>d</sup>	B1 (3 × 5; 14 d) then C1	 <p style="text-align: center;"><b>51</b></p>	63 then 85 ( <b>51</b> )
		 <p style="text-align: center;"><b>52</b></p>	29 then 94 ( <b>52</b> )
39	B1 (4 × 5; 20 d) then C2	 <p style="text-align: center;"><b>53</b></p>	8% <sup>b</sup>
40	B2 (5; 24 h) then C1	 <p style="text-align: center;"><b>54</b></p>	93 then 96 (87% <sup>c</sup> )
41	B2 (5; 24 h) then C2	 <p style="text-align: center;"><b>55</b></p>	93 then 80 (93% <sup>c</sup> )
42	B2 (3 × 5; 7 d) then C1	 <p style="text-align: center;"><b>56</b></p>	76 then 92 (92% <sup>c</sup> )
43	B2 (2 × 5; 3 d) then C2	 <p style="text-align: center;"><b>57</b></p>	54 then 77 (86% <sup>c</sup> )
44	B2 (5; 24 h) then C1	 <p style="text-align: center;"><b>58</b></p>	53 then 99 (98% <sup>c</sup> )

<sup>a</sup>Methods: B1: Hoveyda–Grubbs second-generation catalyst, CH<sub>2</sub>Cl<sub>2</sub>, 50 °C then Et<sub>3</sub>N (86 equiv), P(CH<sub>2</sub>OH)<sub>3</sub> (86 equiv) then silica; B2: Hoveyda–Grubbs second-generation catalyst, MTBE, 50 °C then Et<sub>3</sub>N (86 equiv), P(CH<sub>2</sub>OH)<sub>3</sub> (86 equiv) then silica; C1: PhSH (1.2 equiv), K<sub>2</sub>CO<sub>3</sub> (3.0 equiv), DMF; C2: PhSH (2.4 equiv), K<sub>2</sub>CO<sub>3</sub> (6.0 equiv), DMF; E: aq HF, MeCN–CH<sub>2</sub>Cl<sub>2</sub>. <sup>b</sup>Yield over more than one step. <sup>c</sup>Purity of the product determined by 500 MHz <sup>1</sup>H NMR spectroscopy. <sup>d</sup>The starting material was a ca. 75:25 mixture of diastereoisomers.

reaction, and the bridged macrocycle **53** was obtained in low yield. We have previously observed the formation of macrocyclic metathesis products in similar metathesis cascade reactions [14]. The formation of the cyclopropanes **46** and **47** as

byproducts in the metathesis cascade reaction of **32** was remarkable [25]. Presumably, in this case, the metathesis cascade leads to the generation of the intermediate **59** (Scheme 5); the intermediate could then react to conclude the





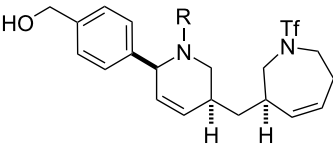
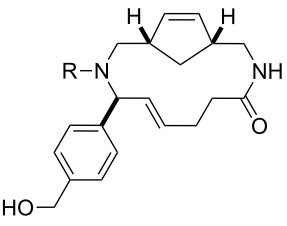
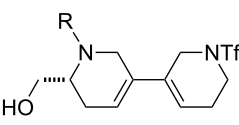
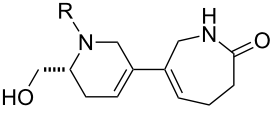
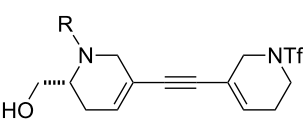
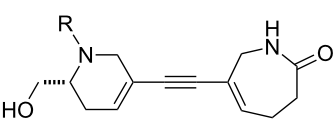
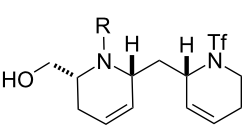
metathesis cascade (to give **45** after deprotection), or cyclopropanate [25] the terminal alkene (to give **46** or **47** after deprotection) (Scheme 5).

Finally, a selection of fluoruous-tagged products was derivatised (typically on a 50  $\mu\text{mol}$  scale) to yield a range of amides and ureas (Table 4). The fluoruous tag facilitated the purification of

**Table 4:** Derivatisation and deprotection of final products.

Substrate (purity <sup>a</sup> / %)	Product <sup>b</sup>	Method <sup>c</sup>	Yield / %	
<b>45</b>		<b>60a</b>	D	51
		<b>60b</b>	E1 then D	81 then 60
		<b>60c</b>	E2 then D	67 then 98
		<b>60d</b>	E3 then D	94 then 81
<b>46</b>		<b>61b</b>	E1 then D	39 then 70
<b>47</b>		<b>62b</b>	E1 then D	43 then 63
<b>49</b> (86)		<b>63a</b>	D	83
		<b>63b</b>	E1 then D	32 then 64
		<b>63c</b>	E2 then D	83 then 58
		<b>63d</b>	E3 then D	84 then 79
<b>50</b> (93)		<b>64a</b>	D	87
		<b>64b</b>	E1 then D	29 <sup>d</sup>
		<b>64c</b>	E2 then D	43 <sup>d</sup>
		<b>64d</b>	E3 then D	34 <sup>d</sup>

**Table 4:** Derivatisation and deprotection of final products. (continued)

<b>51</b> (85)		<b>65a</b>	D	70
		<b>65b</b>	E1 then D	40 <sup>d</sup>
		<b>65c</b>	E2 then D	82 <sup>d</sup>
		<b>65d</b>	E3 then D	74 <sup>d</sup>
<b>53</b>		<b>66a</b>	D	52
<b>54</b> (87)		<b>67a</b>	D	91
		<b>67b</b>	E4 then D	77 <sup>d</sup>
		<b>67c</b>	E2 then D	83 <sup>d</sup>
		<b>67d</b>	E3 then D	42 <sup>d</sup>
<b>55</b> (93)		<b>68a</b>	D	94
		<b>68c</b>	E2 then D	67 <sup>d</sup>
		<b>68d</b>	E3 then D	40 <sup>d</sup>
		<b>68b</b>	E4 then D	67 <sup>d</sup>
<b>56</b> (92)		<b>69a</b>	D	91
		<b>69b</b>	E4 then D	67 <sup>d</sup>
		<b>69c</b>	E2 then D	67 <sup>d</sup>
<b>57</b> (86)		<b>70a</b>	D	47
		<b>70c</b>	E2 then D	59 <sup>d</sup>
<b>58</b> (98)		<b>71a</b>	D	91
		<b>71b</b>	E4 then D	64 <sup>d</sup>
		<b>71c</b>	E2 then D	53 <sup>d</sup>
		<b>71d</b>	E3 then D	29 <sup>d</sup>

<sup>a</sup>Determined by analysis of the product by 500 MHz <sup>1</sup>H NMR spectroscopy. <sup>b</sup>The suffix refers to the identity of the R substituent: a, R = H; b, R = isoxazole-5-carbonyl; c, R = pyridine-3-carbonylamino; d, R = morpholine-4-carbonyl; <sup>c</sup>Methods: D: aq HF, MeCN–CH<sub>2</sub>Cl<sub>2</sub>; E1: isoxazole-5-carbonyl chloride (2.0 equiv), Et<sub>3</sub>N (3.0 equiv), DMAP (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>; E2: pyridine-3-isocyanate (2.0 equiv), Et<sub>3</sub>N (3.0 equiv), DMAP (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>; E3: morpholine-4-carbonyl chloride (2.0 equiv), Et<sub>3</sub>N (3.0 equiv), DMAP (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>; E4: isoxazole-5-carbonyl chloride (2.0 equiv), pyridine; <sup>d</sup>Yield over two steps.

the derivatised products by F-SPE. The final products **60–71** (Table 4) were obtained after removal of the fluororous tag by desilylation.

## Conclusion

Metathesis is an extremely powerful reaction for diversity-oriented synthesis. It was demonstrated that metathesis substrates could be assembled efficiently from triplets of building blocks. Thereafter, metathesis cascades yielded a

diverse range of molecular scaffolds. The diversity of the products was increased through variation of all three of the building blocks used: the initiating, the propagating, and the terminating building block.

The overall approach was facilitated by fluororous tagging of the initiating building block, allowing easy purification (by F-SPE) of synthetic intermediates and metathesis products. The presence of a fluororous tag also facilitated the purification of the

functionalised products. Evaluation of the biological activity of the final products will be reported in due course.

## Supporting Information

### Supporting Information File 1

Experimental and compound characterisation.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-9-88-S1.pdf>]

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See for a reaction with a similar outcome.

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