



Supporting Information

for

Supramolecular assembly of pentamidine and polymeric cyclodextrin bimetallic core–shell nanoarchitectures

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Beilstein J. Nanotechnol. **2022**, *13*, 1361–1369. doi:10.3762/bjnano.13.112

Additional experimental data

Materials and Methods

All the dispersions used for preparation and spectroscopic characterization of nanoassemblies were prepared in ultrapure water (Fresenius Kabi Italia) or in 10 mM phosphate buffer containing NaCl (137 mM) and KCl (2.7 mM) at pH 7.4 (PBS) at room temperature (rt \approx 25 °C).

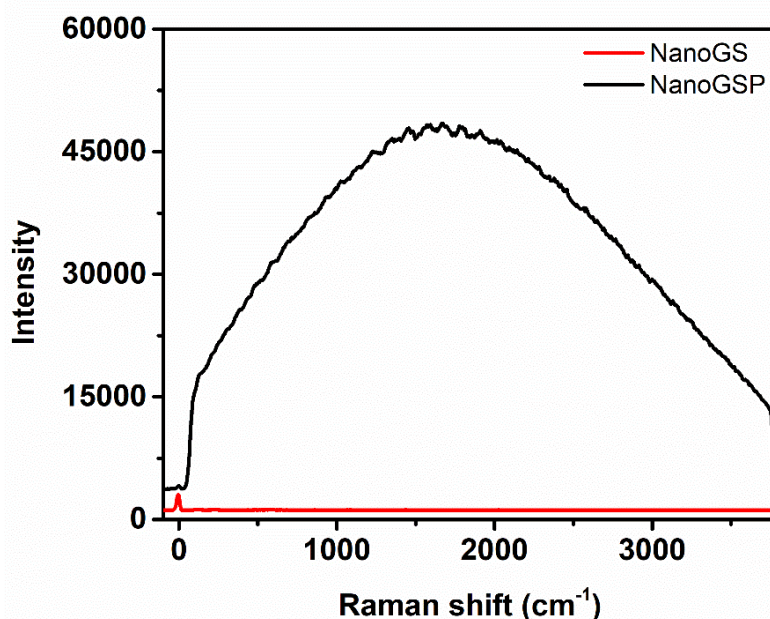


Figure S1: The Raman spectra of nanoGS and nanoGSP. The Raman spectra were recorded under an excitation of 532 nm. No specific Raman signal was detected.

Table S1: In vitro antileishmanial activity and cytotoxicity (IC₅₀ μ M and CC₅₀ μ M, respectively) of gold nanoparticles decorated with polycyclodextrins (nanoG), bimetallic Au–Ag core–shell nanoparticles (nanoGS), and bimetallic Au–Ag core–shell nanoparticles containing pentamidine (nanoGSP). Values represent the mean of two independent tests.

Compound	<i>L. infantum</i> ^a	PMM ^b	MRC-5 ^c
nanoG	> 64.00	> 64.00	> 64.00
nanoGS	> 64.00	> 64.00	> 64.00
nanoGSP	> 51.40	> 51.40	> 51.40
Pent ^d	5.66	32.00	24.87
Miltefosine	3.17	–	–
Tamoxifen	–	–	13.60

^a *Leishmania infantum* MHOM/MA/67/ITMAP263.

^b Primary peritoneal mouse macrophages.

^c Human fetal lung fibroblast (MRC-5) cell line toxicity.

^d Pentamidine

Preparation of the PolyCD@Pent complex

The complex was prepared at [Pent]:[PolyCD] 1:1 molar ratio ([Pent] = [PolyCD] = 1 mM). The PolyCD was dissolved in ultrapure water (1.71 mg/mL) and sonicated in an ultrasonic bath (5 min). A thin organic film of free low Pent (0.35 mg/mL) was prepared by slow evaporation of an ethanol solution (EtOH) and the latter was hydrated at room temperature, with PolyCD dispersion followed by sonication in an ultrasonic bath (30 minutes). The diluted dispersions (Pent = 52.5 $\mu\text{g/mL}$, [Pent] = [CD] = 154 μM) were characterized by UV/Vis. For comparison, Pent/PolyCD nanocomplexes at the same concentration were prepared in PBS and analyzed by UV/Vis.

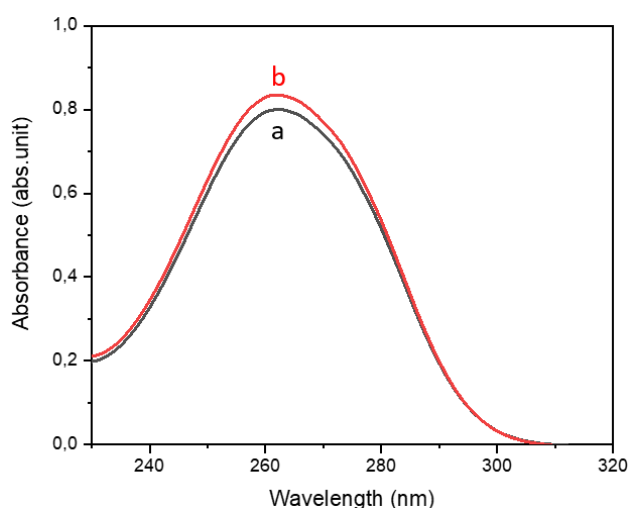


Figure S2: UV/Vis spectra of Pent (trace a) and of PolyCD@Pent in ultrapure water (trace b). Experimental conditions: [Pent] = 154 μM ; $d = 0.2$ cm; $T = 25$ °C.

The Pent absorption profile shows a main band centered at 262 nm in ultrapure water which is also maintained in PolyCD@Pent even if in the latter there is an increase in absorbance at the same concentration. This was ascribed to an increased solubility of the drug upon interaction with PolyCD.

Release study of PolyCD@Pent

The Pent release profile from the PolyCD@Pent was evaluated in PBS at pH 7.4 by a dialysis method. PolyCD@Pent (Pent = 0.35 mg/mL), PolyCD concentrated solution

3.42 mg/mL (500 μ L), and 500 μ L of 10 mM PBS to a final volume of 1 mL were placed in a dialysis tube (spectra dialysis bags/Por®, MWCO 3500 kDa) and immersed in 5 mL of PBS (sink condition) under continuous stirring (250 rpm) at 37 ± 0.5 °C. At predetermined times, 1 mL of release medium was removed and replaced with an equal volume of fresh aqueous solution of PBS. The quantity of Pent released was evaluated by UV–vis spectroscopy (at $\lambda = 262$ nm) by using the extinction coefficient of Pent salt in PBS ($\varepsilon \sim 26000$ mol⁻¹ cm⁻¹) and was expressed as a percentage ratio between the weight of Pent released and the total quantity of the drug.

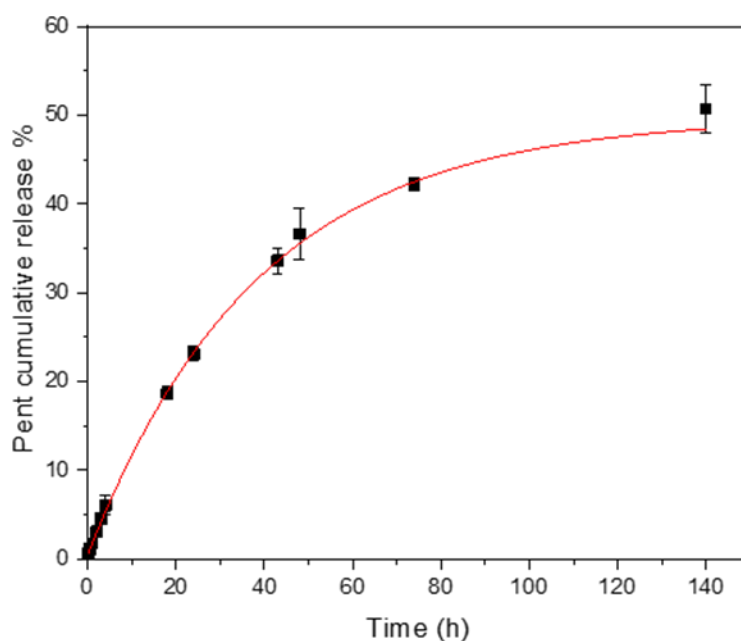


Figure S3: Release profile of Pent from PolyCD@Pent in PBS (pH 7.4, at 37 °C).

The amount of released Pent was determined in PBS by using UV–vis. No initial burst release was observed from PolyCD@Pent, with a slow and controlled release in time, leading to a final 53% of Pent in the external medium after 140 h (Figure S3).