

Olefin cross metathesis based de novo synthesis of a partially protected L-amicetose and a fully protected L-cinerulose derivative

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Abstract

Cross metathesis of a lactate derived allylic alcohol and acrolein is the entry point to a de novo synthesis of 4-benzoate protected L-amicetose and a cinerulose derivative protected at C5 and C1.

Introduction

Many drugs and bioactive natural products are glycoconjugates, which contain an aglycon part linked through glycosidic bonds to one or more oligosaccharide side chains [1]. While it was assumed for quite some time that the carbohydrate side chain merely influences the pharmacokinetics, more recent investigations led to the conclusion that the oligosaccharide moiety contributes essentially to the mechanisms of action, e.g., through molecular recognition of a preferred binding site [2–5], thereby ensuring the selectivity of a chemotherapeutic agent. Particularly common are side chains composed of deoxygenated sugars [6]. For example, the kigamicins are bacterial secondary metabolites isolated from *Amiclatopsis* sp. [7,8] and display both antibiotic and cytotoxic activity [9]. They have a polycyclic xanthone aglycone in common, which is glycosylated at the C14–OH group. In Figure 1 the structure of

kigamicin B, which carries a D-amicetose disaccharide unit, is shown as a representative example. Another antitumor antibiotic is aclacinomycin A, which has been used clinically under the name aclarubicin. It is an anthracycline [10] bearing a trisaccharide side chain consisting of L-rhodosamin, 2,6-dideoxy-L-lyxose, and L-cinerulose attached to the aglycon aclavinon [11,12]. It was found to be a potent antineoplastic agent with particular activity against different forms of leukemia [13] (Figure 1).

Desoxy-sugars are sometimes scarcely available from natural sources and therefore the chemical syntheses of the monosaccharides [6,14] and their assembly to oligosaccharides [15–17] has attracted constant attention for many years. For example, D-amicetose has been obtained from other carbohydrates

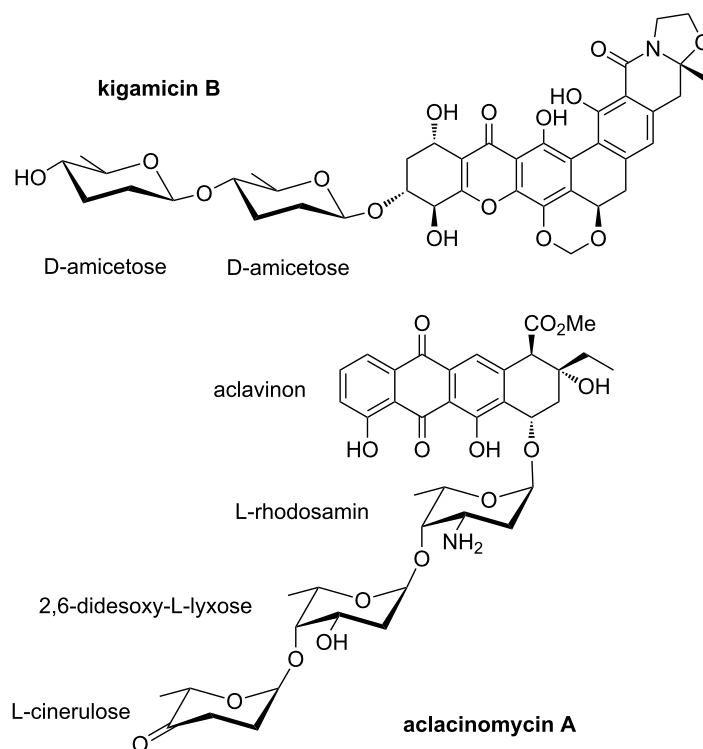
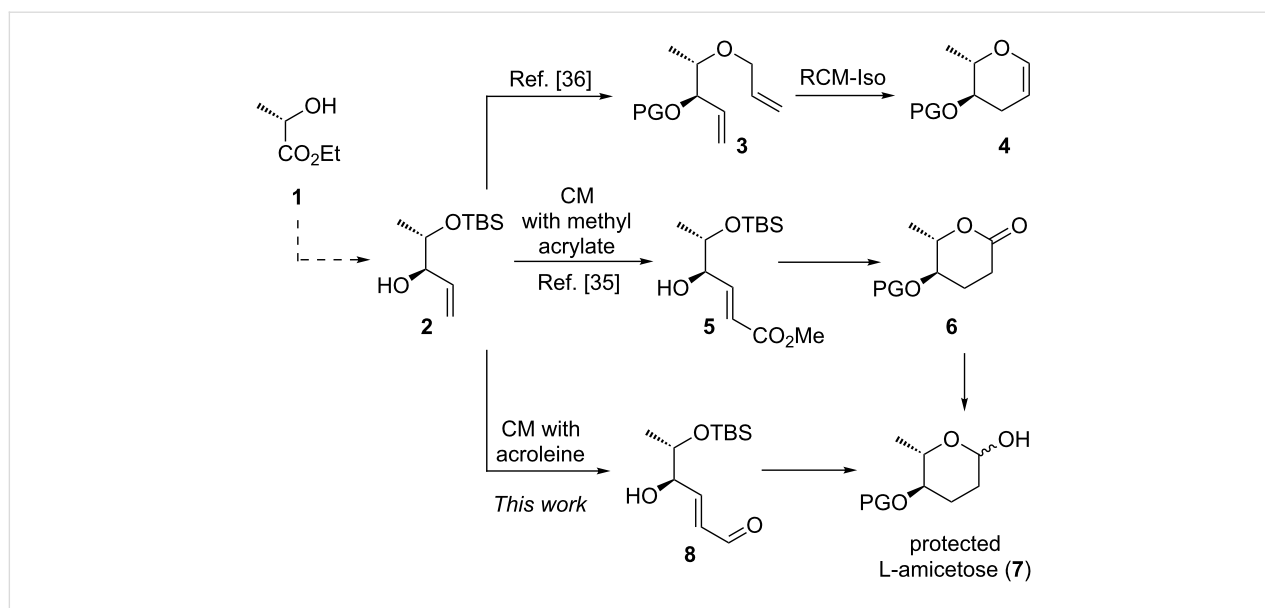


Figure 1: Structures of kigamicin B and aclacinomycin A as representative examples for antineoplastic glycoconjugates.

through ethanethiolysis and subsequent Ni-catalyzed desulfurization [18,19], by reduction of the corresponding aldolactone [20], or by conversion of a protected mannopyranoside into the 2,3-unsaturated enopyranoside and subsequent hydrogenation and reductive 6-deoxygenation [21,22]. Successful *de novo* approaches [14] to both enantiomers of amictose include an Achmatowicz rearrangement–hydrogenation sequence, starting from enantiomerically pure 2-(1-hydroxyethyl)furan [23–25], a sequence comprising perdeuteration of an alkynoate derived from L-threonine (giving a 2,2,3,3-tetradeuterated amictose) [26], enantioselective hydroboration of hetero Diels–Alder adducts [27], stereoconservative diazotation of L-glutamic acid [28,29], diastereoselective addition of methylmagnesium bromide to enantiopure 2-benzyloxyhex-5-enal [30], oxidative degradation of aromatics and subsequent lactonization [31], and a two-carbon homologation of an enantiopure C4-building block with a metallated sulfone [32]. A combinatorial biosynthetic approach to D- and L-amictose has also recently been established by combining different sugar biosynthesis genes [5].

Comparatively few transition metal mediated or -catalyzed reactions have hitherto been used for the synthesis of 2,3,6-trideoxy sugars such as amictose. An example is the W-promoted cycloisomerization of lactaldehyde derived

alkynols, which yields the glycal of amictose or its epimer rhodose, respectively [33]. An approach to amictose (and a few other 6-deoxy sugars) involves the formation of an enantiopure β,γ -unsaturated δ -valerolactone via ring closing metathesis (RCM). The RCM product is subsequently converted into L-amictose in four steps [34]. Unfortunately, the steps determining the configuration at C4 proceed in both cases with virtually no diastereoselectivity, although the resulting epimers were conveniently separated by chromatography. We have recently established two different metathesis based routes to L-amictose [35] or L-amictal [36], respectively, which both use enantiomerically pure L-ethyl lactate (**1**) as the starting material (Scheme 1). The synthetic routes rely on the highly diastereoselective two-step conversion of ethyl lactate to allylic alcohol **2** [36,37] and further to the RCM precursor **3**, which then undergoes RCM-isomerization to L-amictal **4** [36]. Alternatively, we have used **2** as a substrate for a cross metathesis reaction with methyl acrylate under isomerization-free conditions to furnish enoate **5**, which underwent cyclization to the lactone **6** after hydrogenation. Reduction of **6** with DIBAL-H eventually yields a protected L-amictose **7** [35]. To avoid the last step of the sequence, we envisaged a cross metathesis with acrolein, rather than methyl acrylate, to give enal **8** which should undergo spontaneous lactol formation after hydrogenation and desilylation. However, compared to cross metathesis



Scheme 1: RCM-isomerization approach to L-amicetal **4** and alternative CM approaches to L-amicetose.

reactions with methyl acrylate literature precedence for the use of acroleins as CM partner is significantly smaller [38–44].

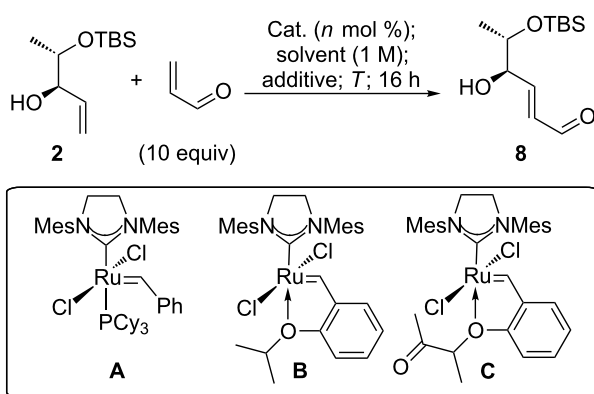
In this contribution we report a de novo synthesis of a protected L-amicetose, using a cross metathesis reaction of allyl alcohol **2** with acrolein, and the elaboration of the cross metathesis product **8** to a fully protected acyclic cinerulose derivative. We are aware of only one previous de novo synthesis of DL-cinerulose, which used an Achmatowicz rearrangement of 2-(1-hydroxyethyl)furan [45].

Results and Discussion

The three second generation catalysts **A** [46], **B** [47,48] and **C** [49,50] were initially tested for the cross metathesis of **2** and acrolein (Table 1). We started with the most common catalyst, the second generation Grubbs' catalyst. In order to suppress undesired double bond isomerization reactions [51,52] we tried to avoid high reaction temperatures, which may cause catalyst decomposition to isomerization active species. This phenomenon has often been observed for second generation catalysts [53]. For these reasons, phenol was used as a rate accelerating additive. Phenols are believed to coordinate to the Ru and stabilize the catalytically active 14-electron species, resulting in a retarded catalyst decomposition [54,55]. In the presence of 5 mol % of **A** and 0.5 equiv of phenol, however, the yield of cross metathesis product **8** was very low at ambient temperature (Table 1, entry 1). Heating the mixture to reflux in dichloromethane resulted in a comparably low yield (Table 1, entry 2), whereas a significant improvement could be observed in toluene at 80 °C (Table 1, entry 3). We thought that the yield could not be further increased with catalyst **A** and therefore

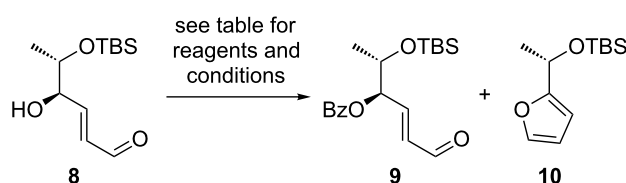
decided to test the phosphine free catalyst **B**. This and related catalysts [56], comprising a hemilabile *ortho*-isopropoxy substituted alkylidene ligand, are particularly suited for cross metathesis reactions [57,58]. Indeed, even at ambient temperature and without any rate accelerating additive, a good yield of 86% was obtained (Table 1, entry 4), which could be improved to quantitative by increasing the reaction temperature to 40 °C (Table 1, entry 5). Reducing the catalyst loading to 2.5 mol % is possible, but at the expense of isolated yield of product (Table 1, entry 6). We have previously used the Umicore M5₁ catalyst (**C**) [49] for cross metathesis reactions and discovered that high selectivities and rates of conversion can be accomplished even with comparatively low catalyst loadings [59,60]. For the cross metathesis of **2** and acrolein, **C** gave significantly better yields than **A**, but was found to be inferior to **B** (Table 1, entries 7–9).

In the following step we decided to protect the hydroxy group at C4 first to avoid any complications arising from the formation of a furanose after hydrogenation of the C–C double bond. Benzoyl was chosen as a protecting group, because of its UV-activity, its orthogonality to the TBS group at C5–OH and its stability under hydrogenation conditions. Interestingly, attempted benzylation in pyridine resulted exclusively in the formation of furan **10** (Table 2, entry 1). We reasoned that pyridine initiates an *E/Z*-isomerization of the enal through nucleophilic attack at the β-position, followed by lactol formation, benzylation of the lactol and finally elimination of benzoic acid. Replacing pyridine as a solvent by dichloromethane and using NEt(iPr)₂ as an HCl-scavenger led indeed to a suppression of the formation of furan **10**, but the desired benzoate **9**

Table 1: Catalyst screening for CM of allyl alcohol **2** and acrolein.

entry	catalyst	cat. loading	additive (equiv)	solvent	T /°C	yield of 8
1 ^a	A	5 mol %	phenol (0.5)	CH ₂ Cl ₂	25 °C	25%
2 ^a	A	5 mol %	phenol (0.5)	CH ₂ Cl ₂	40 °C	20%
3	A	5 mol %	phenol (0.5)	toluene	80 °C	68%
4	B	5 mol %	–	CH ₂ Cl ₂	25 °C	86%
5	B	5 mol %	–	CH ₂ Cl ₂	40 °C	99%
6	B	2.5 mol %	–	CH ₂ Cl ₂	40 °C	76%
7	C	5 mol %	–	CH ₂ Cl ₂	40 °C	65%
8	C	5 mol %	–	toluene	80 °C	80%
9	C	2.5 mol %	–	toluene	80 °C	65%

^aAllylic alcohol **2** was recovered in 21% yield (entry 1) and in 12% yield (entry 2), respectively.

Table 2: C4–OH protection as benzoate.

entry	reagents and conditions	product	yield
1	benzoyl chloride (1.5 equiv), pyridine (0.4 M), 65 °C, 12 h	10	47%
2 ^a	benzoyl chloride (2.6 equiv), NEt(iPr) ₂ (3.0 equiv), CH ₂ Cl ₂ , 40 °C, 12 h	9	<20%
3	benzoic acid (1.8 equiv), DCC (1.8 equiv), DMAP (0.1 equiv), CH ₂ Cl ₂ , 20 °C, 12 h	9	82%

^aStarting material **8** was recovered in ca. 80% yield.

was obtained in yields lower than 20%, along with ca. 80% of recovered starting material **8** (Table 2, entry 2). Benzoate **9** was eventually obtained in high yields from benzoic acid using Steglich's esterification [61] (Table 2, entry 3).

With enal **9** in hands, selective hydrogenation of the C–C double bond had to be accomplished in the next step. Lipshutz'

modification [62] of Stryker's reagent [63], which we had previously used successfully for the conjugate reduction of a related enoate [35], failed completely in this case and resulted only in the isolation of unreacted starting material. For these reasons we resumed to a hydrogenation catalyzed by Pd/C, in spite of the well-known capricious nature of these transformations [64].

With a commercial sample of Pd on charcoal (10 wt %) a plethora of products was detected, four of which could be isolated and identified as the hydrogenated acetal **11**, the methyl ether **13**, and the two desilylated products **14** and **15** (Table 3, entry 1). Literature precedence exists for the Pd/C-induced dealkoxylation of acetals [65] and for desilylation reactions of silyl ethers [66]. With in situ generated Pd/C (obtained from Pd(OAc)₂ and charcoal according to Felpin's protocol) [67] an improved selectivity was observed as the cleavage of the silyl groups could be suppressed. However, reductive dealkoxylation of the acetal remained a problem and the combined yield of the desired products **11** and **12** was still unsatisfactory (Table 3, entry 2). This situation changed when we used Pd(OH)₂ on charcoal (10 wt %) as hydrogenation catalyst, which resulted in the exclusive formation of acetal **11** and aldehyde **12** in 83% combined yield (Table 3, entry 3).

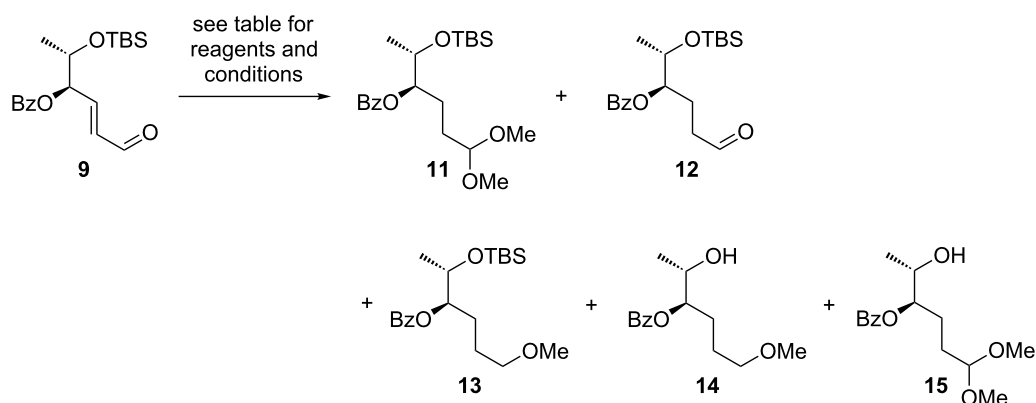
For the deprotection of **11** and **12** a desilylation initiated by TBAF, followed by acidic hydrolysis was investigated first. With isolated acetal **11**, these conditions induced a scrambling of the benzoate and led to a mixture of the desired pyranose **16** and the furanose **17**. Facile migration of carboxylates upon TBAF-mediated desilylation of a vicinal alcohol has previously been observed by us in a different context [68]. We assume that this process starts with a nucleophilic attack of the alkoxyate **18** at the ester carbon, giving a five membered intermediate **19** (Scheme 2).

The benzoate scrambling was completely suppressed by avoiding highly nucleophilic alkoxide intermediates, which was achieved with acidic deprotection conditions. Thus, treating the mixture of **11** and **12** with trifluoroacetic acid in dichloromethane at ambient temperature resulted in desilylation and acetal cleavage, giving 4-benzoyl protected L-amictose **16** as an anomeric mixture in 65% yield (Scheme 3).

With a view towards L-cinerulose, formally the C4-oxidation product of L-amictose, we started from cross metathesis product **8** which was first oxidized using Dess–Martin periodinane [69]. The α,β -unsaturated γ -keto aldehyde **21** was obtained in a very high yield of 90%, and subsequently subjected to hydrogenation conditions using a commercial sample of Pd/C (10 wt %, "batch 1"). We were pleased to find that the expected cinerulose derivative **22** could be isolated in analytically pure form in 85% yield (Scheme 4).

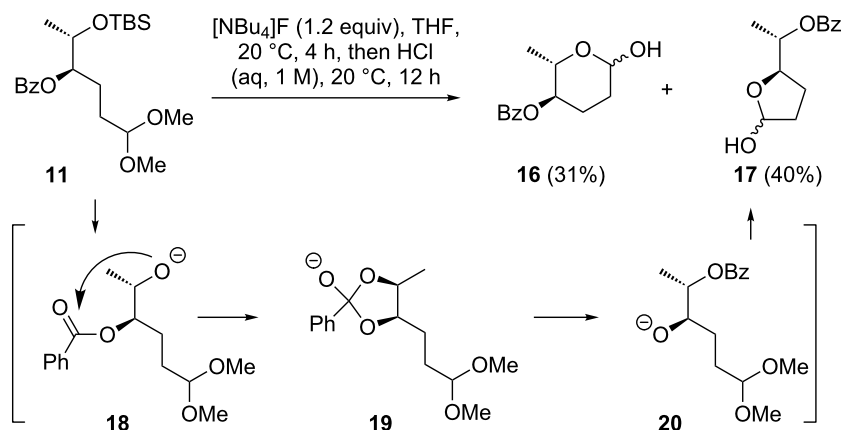
To our great dismay, we found that this result could not be reproduced with a different batch of commercial Pd/C (10 wt %, "batch 2"), purchased from the same supplier. As can be seen from Table 4, entry 1, the second batch of Pd/C catalyzes under otherwise identical conditions a chemoselective acetalization of the aldehyde, leading to **23** which was isolated in 52% yield. A similar result was obtained with Pd(OH)₂/C (Table 4, entry 2). In situ formed Pd on charcoal [67], prepared from 0.5 mol % of Pd(OAc)₂ as a precursor, gave an insepa-

Table 3: Hydrogenation of benzoyl protected enal **9**.

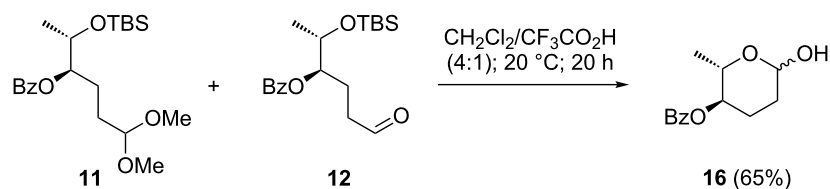
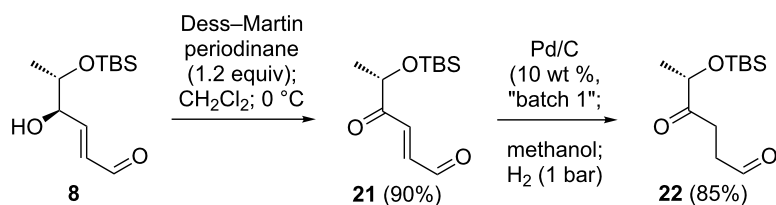
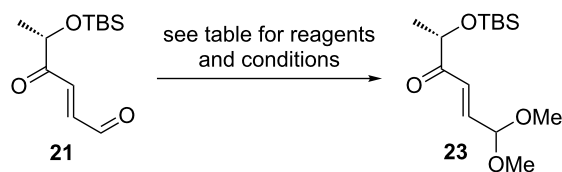


entry	reagents and conditions	11	12	13	14	15
1	Pd/C (10 wt %; 1.6 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	31%	–	16%	16%	8%
2	Pd(OAc) ₂ (1 mol %), activated charcoal (9 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	28%	9%	trace	–	–
3 ^a	Pd(OH) ₂ /C (10 wt %; 1.2 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	51%	32%	–	–	–

^aProducts **11** and **12** were obtained as an inseparable mixture, yields are estimated from ¹H NMR spectrum.



Scheme 2: Two step desilylation–acetal hydrolysis.

Scheme 3: Deprotection of **11** and **12** to L-amicetose derivative **16**.Scheme 4: Synthesis of a cinerulose-TBS ether **22**.Table 4: Chemoselective acetalization of oxidation product **21**.

entry	reagents and conditions	yield of 23
1	Pd/C ("batch 2", 10 wt %; 2.2 mol %), H_2 (1 bar), methanol, 20 °C, 12 h	52%
2	Pd(OH) ₂ /C (10 wt %; 1.7 mol %), H_2 (1 bar), methanol, 20 °C, 12 h	43%
3	Pd(OAc) ₂ (0.5 mol %), activated charcoal (9 mol %), H_2 (1 bar), methanol, 20 °C, 12 h	34% ^a
4	Pd(OAc) ₂ (1.0 mol %), activated charcoal (9 mol %), H_2 (1 bar), methanol, 20 °C, 12 h	82%

^aEstimated from ¹H NMR spectrum (**21** and **23** were obtained as an inseparable mixture in ca. 90% combined yield).

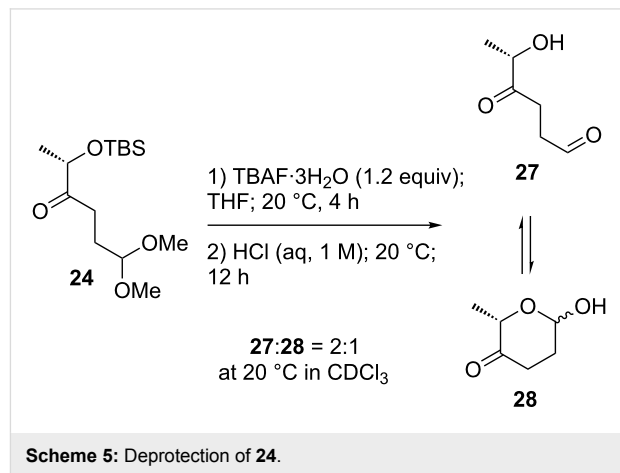
rable mixture of starting material **21** and acetal **23** in a 1.7:1 ratio (Table 4, entry 3). By increasing the amount of catalyst precursor to 1 mol %, conversion to acetal **23** was complete, and this product could be isolated in 82% yield (Table 4, entry 4).

Although the Pd(II)-catalyzed acetalization of aldehydes, including enals, is known [70], we are not aware of examples where Pd-catalysts immobilized on charcoal have been used for this transformation. Most likely the acetalization is catalyzed by residual Pd²⁺ via Lewis-acidic carbonyl activation. It is known that the reduction to Pd(0) is almost never complete, even if hydrogen is present [71,72].

At this stage we decided to reinvestigate the hydrogenation step, starting from acetal **23**, using fresh samples of catalyst which have not been used previously for the acetalization (Table 5). With in situ prepared Pd/C (Table 5, entry 1) a major amount of starting material **23** was recovered and minor quantities of hydrogenated products **24** and **26**, as well as desilylated starting material **25** were detected. With commercial Pd/C (10 wt %, “batch 2”) (Table 5, entry 2) the starting material was fully consumed and only hydrogenated products **24** and **26** could be detected in a 1:2 ratio, but unfortunately the isolated yields were only mediocre. As for the hydrogenation of benzoate **9** (Table 3), best results were reproducibly obtained with Pd(OH)₂ on charcoal (Table 5, entry 3), leading to an isolated yield of 84% of a double protected cinerulose derivative **24**.

All attempts to isolate cinerulose were hampered by the high volatility of the product. To accomplish complete deprotection of the precursor, compound **24** was first treated with TBAF trihydrate in THF, followed by treatment with diluted aqueous HCl. Although only small amounts of material were obtained

via this procedure, ¹H NMR spectroscopical analysis was possible. The data suggest that cinerulose exists in CDCl₃ as a 2:1 mixture of acyclic aldose **27** and lactol **28** (Scheme 5).



Conclusion

In summary, we showed that an allylic alcohol, available diastereoselectively from enantiomerically pure L-lactate in few steps, is a useful starting material for a metathesis based synthesis of protected L-amictose and L-cinerulose. It undergoes a clean and high yielding cross metathesis reaction with acrolein, however, the most commonly used second generation Grubbs' catalyst is not the ideal initiator for this particular transformation. Significantly better results were obtained with two different phosphine free catalysts comprising a hemilabile alkoxy substituted benzylidene ligand, even at only moderately elevated temperatures. The acrolein cross metathesis product can be converted into the 4-benzoate of L-amictose via benzylation, Pd-catalyzed hydrogenation and global deprotection, whereas a cinerulose derivative protected at C1 as a

Table 5: Hydrogenation of acetal **23**.

entry	reagents and conditions	24	25	26
1 ^a	Pd(OAc) ₂ (1.0 mol %), activated charcoal (9 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	7%	16%	7%
2	Pd/C (10 wt %; 2.2 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	19%	n.d.	38%
3	Pd(OH) ₂ /C (10 wt %; 1.0 mol %), H ₂ (1 bar), methanol, 20 °C, 12 h	84%	n.d.	n.d.

^aStarting material **23** (49%) was recovered.

dimethyl acetal and at C5–OH as a TBS ether becomes available via Dess–Martin oxidation, Pd-catalyzed acetalization and Pd-catalyzed hydrogenation. Notably, our results underline once again that apparently trivial olefin hydrogenation reactions catalyzed by commercial “Pd/C” can be capricious and very difficult to reproduce, unless the specific properties of the catalyst used and the conditions for its preparation are documented by the supplier. Unfortunately, this is very often not the case and the catalytic activities of apparently identical catalysts which are often designated just as “10 wt % Pd on carbon” may vary dramatically.

Supporting Information

Supporting Information File 1

Experimental procedures and analytical data.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-102-S1.pdf>]

Supporting Information File 2

Copies of ¹H and ¹³C NMR spectra.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-102-S2.pdf>]

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References

- La Ferla, B.; Airoidi, C.; Zona, C.; Orsato, A.; Cardona, F.; Merlo, S.; Sironi, E.; D'Orazio, G.; Nicotra, F. *Nat. Prod. Rep.* **2011**, *28*, 630–648. doi:10.1039/c0np00055h
- Gräfe, U. *Biochemie der Antibiotika*; Spektrum Verlag: Heidelberg, Germany, 1992.
- Hoffmeister, D.; Dräger, G.; Ichinose, K.; Rohr, J.; Bechthold, A. *J. Am. Chem. Soc.* **2003**, *125*, 4678–4679. doi:10.1021/ja029645k
- Hoffmeister, D.; Weber, M.; Dräger, G.; Ichinose, K.; Dürr, C.; Bechthold, A. *ChemBioChem* **2004**, *5*, 369–371. doi:10.1002/cbic.200300793
- Pérez, M.; Lombó, F.; Zhu, L.; Gibson, M.; Braña, A. F.; Rohr, J.; Salas, J. A.; Méndez, C. *Chem. Commun.* **2005**, 1604–1606. doi:10.1039/b417815g
- Kirschning, A.; Bechthold, A. F.-W.; Rohr, J. *Top. Curr. Chem.* **1997**, *188*, 1–84. doi:10.1007/BFb0119234
- Kunimoto, S.; Someno, T.; Yamazaki, Y.; Lu, J.; Esumi, H.; Naganawa, H. *J. Antibiot.* **2003**, *56*, 1012–1017. doi:10.7164/antibiotics.56.1012
- Someno, T.; Kunimoto, S.; Nakamura, H.; Naganawa, H.; Ikeda, D. *J. Antibiot.* **2005**, *58*, 56–60. doi:10.1038/ja.2005.6
- Lu, J.; Kunimoto, S.; Yamazaki, Y.; Kaminishi, M.; Esumi, H. *Cancer Sci.* **2004**, *95*, 547–552. doi:10.1111/j.1349-7006.2004.tb03247.x
- Arcamone, F.; Cassinelli, G. *Curr. Med. Chem.* **1998**, *5*, 391–419.
- Okii, T.; Matsuzawa, Y.; Yoshimoto, A.; Numata, K.; Kitamura, I.; Hori, S.; Takamatsu, A.; Umezawa, H.; Ishizuka, M.; Naganawa, H.; Suda, H.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1975**, *28*, 830–834. doi:10.7164/antibiotics.28.830
- Komiyama, T.; Okii, T.; Inui, T.; Takeuchi, T.; Umezawa, H. *Gann* **1979**, *70*, 395–401.
- Lee, Y.-L.; Chen, C.-W.; Liu, F.-H.; Huang, Y.-W.; Huang, H.-M. *PLoS One* **2013**, *8*, e61939. doi:10.1371/journal.pone.0061939
- Kirschning, A.; Jesberger, M.; Schöning, K.-U. *Synthesis* **2001**, 507–540. doi:10.1055/s-2001-12342
- Thiem, J.; Klaffke, W. *Top. Curr. Chem.* **1990**, *154*, 285–332. doi:10.1007/BFb0111564
- Seeberger, P. H.; Danishefsky, S. J. *Acc. Chem. Res.* **1998**, *31*, 685–695. doi:10.1021/ar9600648
- Seeberger, P. H.; Haase, W.-C. *Chem. Rev.* **2000**, *100*, 4349–4394. doi:10.1021/cr9903104
- Lajšič, S.; Miljković, D.; Četković, G. *Carbohydr. Res.* **1992**, *233*, 261–264. doi:10.1016/S0008-6215(00)90940-6
- Bethell, G. S.; Ferrier, R. J. *J. Chem. Soc., Perkin Trans. 1* **1973**, 1400–1405. doi:10.1039/p19730001400
- Pedersen, C.; Jensen, H. S. *Acta Chem. Scand.* **1994**, *48*, 222–227. doi:10.3891/acta.chem.scand.48-0222
- Spohr, U.; Paszkiewicz-Hnatiw, E.; Morishima, N.; Lemieux, R. U. *Can. J. Chem.* **1992**, *70*, 254–271. doi:10.1139/v92-036
- Kjølberg, O.; Neumann, K. *Acta Chem. Scand.* **1992**, *46*, 877–882. doi:10.3891/acta.chem.scand.46-0877
- Wang, H.-Y. L.; Wu, B.; Zhang, Q.; Kang, S.-W.; Rojanasakul, Y.; O'Doherty, G. A. *ACS Med. Chem. Lett.* **2011**, *2*, 259–263. doi:10.1021/ml100291n
- Wang, H.-Y. L.; Xin, W.; Zhou, M.; Stueckle, T. A.; Rojanasakul, Y.; O'Doherty, G. A. *ACS Med. Chem. Lett.* **2011**, *2*, 73–78. doi:10.1021/ml100219d
- Zhu, L.; Talukdar, A.; Zhang, G.; Kedenburg, J. P.; Wang, P. G. *Synlett* **2005**, 1547–1550. doi:10.1055/s-2005-869846
- Kirschning, A.; Hary, U.; Ries, M. *Tetrahedron* **1995**, *51*, 2297–2304. doi:10.1016/0040-4020(94)01081-A
- Noecker, L. A.; Martino, J. A.; Foley, P. J.; Rush, D. M.; Giuliano, R. M.; Villani, F. J., Jr. *Tetrahedron: Asymmetry* **1998**, *9*, 203–212. doi:10.1016/S0957-4166(97)00631-9
- Berti, G.; Caroti, P.; Catelani, G.; Monti, L. *Carbohydr. Res.* **1983**, *124*, 35–42. doi:10.1016/0008-6215(83)88353-0
- Marco, J. L. *Synth. Commun.* **1989**, *19*, 485–490. doi:10.1080/00397918908050690
- Itoh, T.; Yoshinaka, A.; Sato, T.; Fujisawa, T. *Chem. Lett.* **1985**, *14*, 1679–1680. doi:10.1246/cl.1985.1679
- Fuganti, C.; Grasselli, P. J. *J. Chem. Soc., Chem. Commun.* **1978**, 299–300. doi:10.1039/c39780000299
- Aidhen, I. S.; Satyamurthi, N. *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.* **2008**, *47*, 1851–1857.
- McDonald, F. E.; Zhu, H. Y. H. *J. Am. Chem. Soc.* **1998**, *120*, 4246–4247. doi:10.1021/ja980196r
- Zhu, L.; Kedenburg, J. P.; Xian, M.; Wang, P. G. *Tetrahedron Lett.* **2005**, *46*, 811–813. doi:10.1016/j.tetlet.2004.12.014
- Schmidt, B.; Hauke, S. *Org. Biomol. Chem.* **2013**, *11*, 4194–4206. doi:10.1039/c3ob40167g

36. Schmidt, B.; Biernat, A. *Chem.–Eur. J.* **2008**, *14*, 6135–6141. doi:10.1002/chem.200800567
37. Ley, S. V.; Armstrong, A.; Diez-Martin, D.; Ford, M. J.; Grice, P.; Knight, J. G.; Kolb, H. C.; Madin, A.; Marby, C. A.; Mukherjee, S.; Shaw, A. N.; Slawin, A. M. Z.; Vile, S.; White, A. D.; Williams, D. J.; Woods, M. *J. Chem. Soc., Perkin Trans. 1* **1991**, 667–692. doi:10.1039/p19910000667
38. Chatterjee, A. K.; Morgan, J. P.; Scholl, M.; Grubbs, R. H. *J. Am. Chem. Soc.* **2000**, *122*, 3783–3784. doi:10.1021/ja9939744
39. Cossy, J.; BouzBouz, S.; Hoveyda, A. H. *J. Organomet. Chem.* **2001**, *624*, 327–332. doi:10.1016/S0022-328X(00)00932-3
40. Dinh, M.-T.; BouzBouz, S.; Peglion, J.-L.; Cossy, J. *Synlett* **2005**, 2851–2853. doi:10.1055/s-2005-918943
41. Chandra Rao, D.; Kumar Reddy, D.; Shekhar, V.; Venkateswarlu, Y. *Tetrahedron Lett.* **2013**, *54*, 828–829. doi:10.1016/j.tetlet.2012.11.061
42. Miao, X.; Fischmeister, C.; Bruneau, C.; Dixneuf, P. H. *ChemSusChem* **2009**, *2*, 542–545. doi:10.1002/cssc.200900028
43. Mohapatra, D. K.; Maity, S.; Rao, T. S.; Yadav, J. S.; Sridhar, B. *Eur. J. Org. Chem.* **2013**, 2859–2863. doi:10.1002/ejoc.201300053
44. Paul, T.; Andrade, R. B. *Tetrahedron Lett.* **2007**, *48*, 5367–5370. doi:10.1016/j.tetlet.2007.06.031
45. Achmatowicz, O.; Szechner, B. *Bull. Acad. Pol. Sci., Ser. Sci. Chim.* **1971**, *19*, 309–311.
46. Scholl, M.; Ding, S.; Lee, C. W.; Grubbs, R. H. *Org. Lett.* **1999**, *1*, 953–956. doi:10.1021/ol990909q
47. Garber, S. B.; Kingsbury, J. S.; Gray, B. L.; Hoveyda, A. H. *J. Am. Chem. Soc.* **2000**, *122*, 8168–8179. doi:10.1021/ja001179g
48. Gessler, S.; Randl, S.; Blechert, S. *Tetrahedron Lett.* **2000**, *41*, 9973–9976. doi:10.1016/S0040-4039(00)01808-6
49. Arlt, D.; Bieniek, M.; Karch, R. Novel Metathesis Catalysts. WO Patent Application WO2008/034552 A1, March 27, 2008.
50. Mutlu, H.; Montero de Espinosa, L.; Türünc, O.; Meier, M. A. R. *Beilstein J. Org. Chem.* **2010**, *6*, 1149–1158. doi:10.3762/bjoc.6.131
51. Schmidt, B. *Eur. J. Org. Chem.* **2004**, 1865–1880. doi:10.1002/ejoc.200300714
52. Schmidt, B. *J. Mol. Catal. A* **2006**, *254*, 53–57. doi:10.1016/j.molcata.2006.03.026
53. Higman, C. S.; Plais, L.; Fogg, D. E. *ChemCatChem* **2013**, *5*, 3548–3551. doi:10.1002/cctc.201300886
54. Forman, G. S.; McConnell, A. E.; Tooze, R. P.; van Rensburg, W. J.; Meyer, W. H.; Kirk, M. M.; Dwyer, C. L.; Serfontein, D. W. *Organometallics* **2005**, *24*, 4528–4542. doi:10.1021/om0503848
55. Forman, G. S.; Tooze, R. P. *J. Organomet. Chem.* **2005**, *690*, 5863–5866. doi:10.1016/j.jorganchem.2005.07.107
56. Samoǳowicz, C.; Bieniek, M.; Grell, K. *Chem. Rev.* **2009**, *109*, 3708–3742. doi:10.1021/cr800524f
57. Connon, S. J.; Blechert, S. *Angew. Chem., Int. Ed.* **2003**, *42*, 1900–1923. doi:10.1002/anie.200200556
58. Connon, S. J.; Blechert, S. *Top. Organomet. Chem.* **2004**, *11*, 93–124.
59. Schmidt, B.; Kunz, O.; Petersen, M. H. *J. Org. Chem.* **2012**, *77*, 10897–10906. doi:10.1021/jo302359h
60. Schmidt, B.; Kunz, O. *Beilstein J. Org. Chem.* **2013**, *9*, 2544–2555. doi:10.3762/bjoc.9.289
61. Neises, B.; Steglich, W. *Angew. Chem., Int. Ed. Engl.* **1978**, *17*, 522–524. doi:10.1002/anie.197805221
62. Baker, B. A.; Bošković, Z. V.; Lipshutz, B. H. *Org. Lett.* **2008**, *10*, 289–292. doi:10.1021/ol702689v
63. Mahoney, W. S.; Brestensky, D. M.; Stryker, J. M. *J. Am. Chem. Soc.* **1988**, *110*, 291–293. doi:10.1021/ja00209a048
64. Young, J. G.; Hartung, W. H.; Daniels, H. H. *J. Org. Chem.* **1953**, *18*, 229–234. doi:10.1021/jo01130a015
65. Haukaas, M. H.; O'Doherty, G. A. *Org. Lett.* **2002**, *4*, 1771–1774. doi:10.1021/ol025844x
66. Kim, S.; Jacobo, S. M.; Chang, C.-T.; Bellone, S.; Powell, W. S.; Rokach, J. *Tetrahedron Lett.* **2004**, *45*, 1973–1976. doi:10.1016/j.tetlet.2003.12.145
67. Felpin, F.-X.; Fouquet, E. *Chem.–Eur. J.* **2010**, *16*, 12440–12445. doi:10.1002/chem.201001377
68. Schmidt, B.; Kunz, O.; Biernat, A. *J. Org. Chem.* **2010**, *75*, 2389–2394. doi:10.1021/jo1002642
69. Zhdankin, V. V.; Stang, P. J. *Chem. Rev.* **2008**, *108*, 5299–5358. doi:10.1021/cr800332c
70. Gorla, F.; Venanzi, L. M. *Helv. Chim. Acta* **1990**, *73*, 690–697. doi:10.1002/hlca.19900730317
71. Seki, M. *Synthesis* **2006**, 2975–2992. doi:10.1055/s-2006-950197
72. Felpin, F.-X.; Ayad, T.; Mitra, S. *Eur. J. Org. Chem.* **2006**, 2679–2690. doi:10.1002/ejoc.200501004

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