

Recyclable fluorous cinchona alkaloid ester as a chiral promoter for asymmetric fluorination of β-ketoesters

Wen-Bin Yi^{*1}, Xin Huang², Zijuan Zhang², Dian-Rong Zhu², Chun Cai¹ and Wei Zhang^{*2}

Full Research Paper	Open Access
Address:	Beilstein J. Org. Chem. 2012, 8, 1233–1240.
¹ School of Chemical Engineering, Nanjing University of Science and Technology, Xiao Ling Wei Street, Nanjing 210094, People's Republic	doi:10.3762/bjoc.8.138
of China and ² Department of Chemistry, University of Massachusetts	Received: 17 May 2012
Boston, 100 Morrissey Boulevard, Boston, MA 02125, USA	Accepted: 10 July 2012
	Published: 03 August 2012
Email:	
Wen-Bin Yi [*] - yiwenbin@mail.njust.edu.cn; Zijuan Zhang - wei2.zhang@umb.edu; Wei Zhang [*] - wei2.zhang@umb.edu	This article is part of the Thematic Series "Organocatalysis".
	Guest Editor: B. List
* Corresponding author	
	© 2012 Yi et al; licensee Beilstein-Institut.
Keywords:	License and terms: see end of document.
asymmetric fluorination; β-ketoester; fluorous cinchona ester; organocatalysis; recyclable chiral promoter	

Abstract

A fluorous cinchona alkaloid ester has been developed as a chiral promoter for the asymmetric fluorination of β -ketoesters. It has comparable reactivity and selectivity to the nonfluorous versions of cinchona alkaloids and can be easily recovered from the reaction mixture by simple fluorous solid-phase extraction (F-SPE) and used for the next round of reaction without further purification.

Introduction

Fluorinated organic compounds have unique properties because fluorine forms a strong carbon–fluorine bond with a small covalent radius and high electronegativity. Other than fluorinated polymers in materials science, organofluorine compounds have gained increasing popularity in medical chemistry and agricultural chemistry. Introducing one or a few fluorine atoms to biologically interesting molecules can significantly change the physical, chemical and biological properties [1,2]. The significant amount of publications on fluorinated small molecules, amino acids, carbohydrates, steroids and nucleosides indicates that organofluorine chemistry plays an important role in the life sciences [3,4].

A fluorine atom has been introduced to the α -position of some biologically interesting β -ketoesters, such as erythromycin and sesquiterpenic drimane (Figure 1) [5,6]. The achiral fluorination of β -ketoesters can be achieved by electrophilic reaction with Selectfluor (F-TEDA-BF₄, 1-chloromethyl-4-fluoro-1,4diazoniabicyclo[2.2.2]octane bis(tetrafluoroborate)), as developed by Bank [7-9]. The Cahard [10-12] and Shibata [13,14] groups combined cinchona alkaloids and Selectfluor for asymmetric fluorination of substrates such as imido-protected phenylglycines (up to 94% ee), indanones and tetralones (up to 91% ee), ethyl a-cyanotolyl acetates (up to 87% ee), and cyclic β -ketoesters (up to 80% ee) [15]. A catalytic approach for the cinchona alkaloids and Selectfluor combinations has also been developed [16]. The Togni group employed chiral titanium Lewis acid TiCl₂(TADDOLate) for the asymmetric fluorination of β-ketoesters (up to 96% ee) [17-20]. Most Selectfluorpromoted asymmetric fluorinations require a stoichiometric amount of chiral promoters to suppress the competitively direct achiral fluorination. Different supported cinchona alkaloids have been developed as recyclable chiral promoters or organocatalysts. Among them, the Cahard group developed soluble polymer- and ionic-liquid-supported cinchona alkaloids for electrophilic fluorination [21,22]. The Fache and Soós groups developed fluorous tag-attached cinchona alkaloids for catalytic Diels-Alder reactions [23,24]. Introduced in this paper is a new fluorous cinchona alkaloid ester for flourination of β-ketoesters. It is part of our recent effort on the development of recyclable fluorous reagents and organocatalysts for asymmetric synthesis [25-27].



Results and Discussion

Cinchona alkaloids and their derivatives have been wellexplored in asymmetric synthesis [28]. We envisioned that the introduction of a fluorous tag could facilitate the recycling of cinchona alkaloids. The synthesis of fluorous quinine ester C-1 was accomplished by the reaction of quinine with a fluorous acid chloride (Scheme 1). This compound was easily purified by fluorous-solid phase extraction (F-SPE) with a cartridge charged with fluorous silica gels [29,30]. It is stable in air and soluble in solvents such as CH_2Cl_2 , CH_3OH , and CH_3CN .

With the fluorous quinine ester C-1 in hand, we explored the fluorination reaction using ethyl 2-methyl-3-oxo-3-phenylpropanoate (1a) as a model compound. Nonfluorous quinine esters, such as C-2 and C-3, cinchona alkaloids C-4 and C-5, and fluorous pyrrolidine ester C-6, were also evaluated (Figure 2). The results of the fluorination of β -ketoester **1a** with Selectfluor and different promoters are listed in Table 1. It was found that using MeCN as a solvent with 1 equiv of C-1 gave fluorinated product 2a in 49% yield and 65% ee (Table 1, entry 1). Compared to other promoters (Table 1, entries 2-5), C-1 gave fluorinated products in a slightly low yield but better enantioselectivity. This may be attributed to the stereo and the electronic effect of the fluorous tag. Fluorous pyrrolidine C-6 (Table 1, entry 6) gave the lowest product yield and ee among all six promoters. Reducing the amount of C-1 from 1 equiv to 0.5 and 0.2 equiv significantly reduced the ee of the product (Table 1, entries 7 and 8). A control reaction without C-1 gave 2a in 35% yield as a racemic product (Table 1, entry 9). The results suggest that a stoichiometric amount of C-1 is required to minimize the formation of achiral fluorination product by direct fluorination. Solvent screening indicated that using 1:1 CH₃CN/CH₂Cl₂ gave product 2a in 51% yield and 70% ee (Table 1, entry 15), which is better than using CH₃CN alone. Other single or binary solvent systems containing toluene, THF, H₂O, and CF₃C₆H₅ did not afford better results (Table 1, entries 10-14). It was also found that lowering of the reaction temperature from 25 to 10 or 0 °C did not necessarily improve the enantioselectivity of the fluorination (Table 1, entries 16 and 17).

Recycling of promoter C-1 is an important part of this project. In our previous work we have demonstrated that fluorous organocatalysts and reagents can be readily recovered by F-SPE





Figure 2: Promoters for asymmetric fluorination.

able 1: Asy	mmetric fluorination of 1a . ^a						
$Ph \xrightarrow{O} OEt + \overbrace{F^{+} 2 BF_{4}^{-}}^{OCH_{2}CI} \xrightarrow{Cat.} Ph \xrightarrow{O} OF_{4}^{-} OEt$ $1a \qquad Selectfluor \qquad 2a$							
Entry	Cat. (equiv)	Solvent	<i>t</i> (h)	Yield (%)	ee (%)		
1	C-1 (1.0)	MeCN	72	49	65		
2	C-2 (1.0)	MeCN	72	52	56		
3	C-3 (1.0)	MeCN	72	54	51		
4	C-4 (1.0)	MeCN	72	62	46		
5	C-5 (1.0)	MeCN	72	65	48		
6	C-6 (1.0)	MeCN	96	41	18		
7	C-1 (0.5)	MeCN	60	51	26		
8	C-1 (0.2)	MeCN	60	41	<5		
9	-	MeCN	96	35	0		
10	C-1 (1.0)	Toluene	72	16	23		
11	C-1 (1.0)	THF	72	32	41		
12	C-1 (1.0)	H ₂ O	96	_	_		
13	C-1 (1.0)	MeCN/THF	60	38	45		
14	C-1 (1.0)	MeCN/CF ₃ C ₆ H ₅	60	43	59		
15	C-1 (1.0)	MeCN/CH ₂ Cl ₂	60	51	70		
16 ^b	C-1 (1.0)	MeCN/CH ₂ Cl ₂	72	46	69		
17 ^c	C-1 (1.0)	MeCN/CH ₂ Cl ₂	72	39	71		

[19,20]. In the current work, upon completion of the fluorination reaction, a base such as aqueous NaOH or KOH was added to the reaction mixture to convert the cinchona alkaloid/ Selectfluor complex to free cinchona alkaloid. The organic phase was loaded onto a fluorous silica gel cartridge for F-SPE. Promoter **C-1** was recovered in high yield (94%) and excellent purity (98%). It was used for five rounds without significant change of product yield and ee (Scheme 2).



The scope of fluorous quinine ester C-1-mediated fluorination was evaluated by carrying out the reactions with a number of α -substituted ethyl benzoylacetates **1a–e** and **1g–i** as well as ethyl 2-cyclohexanonecarboxylate (**1f**). Results summarized in Figure 3 indicate that benzoylacetates bearing R² such as Me, PhCH₂, Cl, and Br gave fluorination products **2a–d** in 43–71% yields and 60–70% ee. The nonsubstituted benzoylacetate **1e** gave product **2e** in good yield 69% but low ee (31%). Ethyl 2-cyclohexanonecarboxylate (**1f**) afforded product **2f** in 73% yield and 63% ee. Reactions of ethyl benzoylacetates with bigger substitution groups, such as phenylsulfonyl and maleimide derivatives, were also attempted and gave products **2g–i** in 74–83% yields and 78–81% ee.

Conclusion

A fluorous cinchona alkaloid-ester has been introduced as a promoter for Selectfluor-based asymmetric fluorination of β -ketoesters. The fluorous promoter has slightly lower reactivity but better enantioselectivity than the nonfluorous cinchona alkaloids. It can be easily recovered by simple fluorous solidphase extraction for reuse.

Experimental

General

Chemicals and solvents were purchased from commercial suppliers and used as received. ¹H and ¹³C NMR spectra were recorded on a 300 MHz Varian NMR spectrometer. Chemical



Figure 3: The asymmetric fluorination of various β-ketoesters.

shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference, i.e., proton (chloroform δ 7.26), carbon (chloroform δ 77.0). Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), br s (broad singlet). Coupling constants were reported in hertz (Hz). LC-MS were performed on an Agilent 2100 system. A C18 column (5.0 μ m, 6.0 \times 50 mm) was used for the separation. The mobile phases were methanol and water, both containing 0.05% trifluoroacetic acid. A linear gradient was used to increase from 25:75 v/v methanol/water to 100% methanol over 7.0 min at a flow rate of 0.7 mL/min. UV detections were conducted at 210, 254 and 365 nm. Low-resolution mass spectra were recorded in APCI (atmospheric pressure chemical ionization). The highresolution mass spectra were obtained on a Finnigan/MAT 95XL-T spectrometer. Sorbent silica gel XHL TLC plates (130815) were used for the thin-layer chromatography (TLC). Flash chromatography separations were performed on YAMAZEN AI-580 flash column system with Agela silica gel columns (230-400 µm mesh). The enantiomeric excesses of products were determined by chiral phase HPLC analysis on an SHIMADZU LC-20AD system.

Synthesis of fluorous quinine ester C-1

Thionyl chloride (1.19 g, 10 mmol) was added to a mixture of (1H,1H,2H,2H-perfluorooctyl)benzoic acid (0.468 g, 1 mmol) and pyridine (75 mg, 1 mmol). After stirring of the mixture for 4 h at 50 °C, the reaction container was flushed with nitrogen gas to remove unreacted thionyl chloride. Quinine (0.275 g, 0.85 mmol) and N,N-diisopropylethylamine (129 mg, 1 mmol) in CH₂Cl₂ (3 mL) was added, and the solution was stirred for 24 h under reflux. After the reaction had been quenched with H₂O (2 mL) for 1 h, aqueous K₂CO₃ (2 M, 10 mL) was added, and the mixture was extracted with CH_2Cl_2 (3 \times 10 mL). The CH₂Cl₂ layer was washed with aqueous HCl (ca. 2 M, 10 mL) and H₂O (20 mL). The combined extracts were dried over K₂CO₃ and evaporated. The slightly yellow residue was purified by a fluorous silica gel cartridge (5 g). It was first eluted with 80:20 MeOH/H₂O (20 mL) and then with 100% MeOH. The MeOH fraction was concentrated to give C-1 as a yellowish solid (0.625 g, 95%). Mp 175-177 °C; ¹H NMR (CDCl₃, 300 MHz) & 1.51-2.05 (m, 6H), 2.30-2.42 (m, 3H), 2.65–2.70 (m, 2H), 2.97–3.18 (m, 4H), 3.50 (q, J = 6.9 Hz, 1H), 3.98 (s, 3H), 5.02 (m, 2H), 5.83 (m, 1H), 6.72 (d, J = 6.9 Hz, 1H), 7.32-7.51 (m, 5H), 8.01-8.07 (m, 3H), 8.72-8.73 (d, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 24.2, 26.5, 27.6, 27.9, 32.4, 39.7, 42.6, 55.6, 56.7, 59.4, 74.5, 101.3, 114.6, 117.3, 118.6, 121.9, 126.9, 128.3, 128.63, 130.2, 131.9, 141.7, 143.6, 144.8, 145.0, 147.5, 156.0, 165.3; APCIMS m/z: 775.1 (M⁺ + 1); HRMS-ESI (m/z): $[M + H]^+$ calcd. for C₃₅H₃₂F₁₃N₂O₃, 775.2205; found, 775.2214.

Synthesis of quinine benzoate catalyst C-2

Benzoyl chloride (28 mg, 0.2 mmol) was added to a mixture of quinine (65 mg, 0.2 mmol) in CH₂Cl₂ (0.5 mL). After stirring at rt for 4 h, aqueous K2CO3 (2 M, 1 mL) was added. The reaction mixture was extracted with CH_2Cl_2 (2 × 3 mL). The CH₂Cl₂ layer was washed with aqueous HCl (2 M, 2 mL) and H₂O (3 mL). The combined organic extracts were dried (K₂CO₃) and evaporated. The white residue was purified by flash column chromatography (18:1 CH₂Cl₂/MeOH) to give quinine benzoate C-2 (77 mg, 90%) as a colorless solid. ¹H NMR (CDCl₃, 300 MHz) δ 1.69–2.00 (m, 5H), 2.42 (m, 1H), 2.82 (m, 2H), 3.19–3.40 (m, 2H), 3.49–3.56 (q, J = 7.2 Hz, 1H), 4.00 (s, 3H), 5.04 (m, 2H), 5.82 (m, 1H), 6.97 (d, *J* = 7.2, 1H), 7.40-7.65 (m, 6H), 8.01-8.13 (m, 3H), 8.73 (d, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 23.1, 27.2, 27.5, 39.0, 42.5, 56.0, 56.1, 59.0, 73.5, 101.2, 115.2, 117.2, 122.3, 126.6, 127.9, 128.7, 129.6, 129.6, 131.6, 131.8, 133.6, 140.6, 144.7, 147.2, 158.3, 165.1, 200.2; APCIMS m/z: 429.2 (M⁺ + 1).

Synthesis of quinine acetate C-3

Acetic anhydride (30 mg, 0.3 mmol) was added to a mixture of quinine (65 mg, 0.2 mmol) in CH₂Cl₂ (0.5 mL). After stirring at rt for 8 h, aqueous K₂CO₃ (2 M, 1 mL) was added, and the mixture was extracted with CH_2Cl_2 (2 × 3 mL). The CH_2Cl_2 layer was washed with aqueous HCl (2 M, 2 mL) and H₂O (3 mL). The combined organic extracts were dried (K₂CO₃) and evaporated. The white residue was purified by flash column chromatography (18:1 CH₂Cl₂/MeOH) to give quinine acetate C-3 (67 mg, 92%) as a colorless oil. ¹H NMR (CDCl₃, 300 MHz) δ 1.26-1.89 (m, 5H), 2.42 (m, 1H), 2.12 (s, 3H), 2.23-2.36 (m, 2H), 2.37–2.70 (m, 2H), 3.00–3.16 (m, 2H), 3.34–3.42 (q, J = 7.2 Hz, 1H), 3.96 (s, 3H), 5.03 (m, 2H), 5.86 (m, 1H), 6.50 (d, J = 7.2 Hz, 1H), 7.35–7.44 (m, 3H), 8.02 (d, J = 9.0 Hz, 3H), 8.74 (d, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 21.1, 24.3, 27.5, 27.7, 39.6, 42.4, 55.6, 56.5, 59.0, 73.7, 101.4, 114.5, 118.9, 121.8, 127.0, 131.8, 141.7, 143.5, 144.8, 147.4, 149.6, 157.9, 170.0, 199.5, 200.2; ACPIMS m/z: 367.2 (M⁺ + 1).

Synthesis of fluorous pyrrolidine ester C-6

N,*N*⁻Dicyclohexylcarbodiimide (DCC) (0.206 g, 1 mmol) was added to a mixture of (1*H*,1*H*,2*H*,2*H*-perfluorooctyl)benzoic acid (0.468 g, 1 mmol), *N*-Boc-L-prolinol (0.221 g, 1.1 mmol), 4-dimethylaminopyridine (DMAP) (0.122 g, 1 mmol) in THF. After being stirred for 24 h at rt, the mixture was directly loaded onto a fluorous silica-gel cartridge (5 g; eluted by 100% methanol) to give the *N*-Boc-L-prolinyl (1*H*,1*H*,2*H*,2*H*-perfluorooctyl)benzoate (0.618 g, 95%). The *N*-Boc ester was then added to a mixture of TFA in CH₂Cl₂. After being stirred for 12 h at 0 °C, the reaction mixture was loaded onto a fluorous silica-gel cartridge (5 g) again to give the title compound L-prolinyl (1*H*,1*H*,2*H*,2*H*-perfluorooctyl)benzoate (0.496 g, 90%). ¹H NMR (CDCl₃, 300 MHz) δ 1.62–1.89 (m, 3H), 2.16–2.21 (m, 1H), 2.29–2.47 (m, 2H), 2.92–2.98 (m, 2H), 3.48–3.54 (m, 2H), 3.71–3.84 (m, 2H), 4.38–4.42 (m, 1H), 4.95–4.97 (m, 2H), 7.27–7.31 (d, 2H), 7.48–7.50 (d, 2H); ¹³C NMR (CDCl₃, 75 MHz) δ 25.0, 26.2, 26.3, 26.3, 28.5, 32.2, 32.6, 51.1, 61.6, 67.2, 127.3, 127.6, 128.3, 128.3, 128.4, 130.1, 135.1, 141.4, 171.9; APCIMS *m/z*: 552.1 (M⁺ + 1).

General procedure for fluorination reaction

A mixture of Selectfluor (0.057 g, 0.16 mmol) and fluorous quinine ester C-1 (0.124 g, 0.16 mmol) in CH₃CN and CH₂Cl₂ was stirred at rt for 1 h. Ethyl 2-methyl-3-oxo-3-phenyl-propanoate (1a) (0.033 g, 0.16 mmol) was added. After stirring of the mixture at rt for 32 h, the reaction was quenched with H₂O. After F-SPE, the mixture was extracted with EtOAc. The organic layer was washed with aqueous HCl (2 M, 5 mL) and H₂O (5 mL), and then dried over Na₂SO₄. After evaporation of the solvent, the residue was purified by flash column chromatography (8:1 hexane/EtOAc) to give (S)-ethyl 2-methyl-2-fluoro-3-oxo-3-phenylpropanoate (2a) as a colorless oil.

(S)-Ethyl 2-methyl-2-fluoro-3-oxo-3-phenylpropanoate (2a)

51% yield, 70% ee. The enantiomeric excess was determined by HPLC on (R,R)-WHELK-O1 with hexane/iPrOH (92:8) as the eluent. Flow rate: 0.6 mL/min, $\lambda = 254$ nm; $t_{minor} = 20.132$ min, $t_{major} = 17.924$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.00 (t, J = 7.2 Hz, 3H), 1.93 (s, 1H), 4.11 (q, J = 7.2 Hz, 2H), 7.33–7.38 (m, 2H), 7.46 (m, 1H), 7.90–7.92 (m, 2H); APCIMS m/z: 225.2 (M⁺ + 1).

(S)-Ethyl 2-benzyl-2-fluoro-3-oxo-3-phenylpropanoate (**2b**)

59% yield, 60% ee. The enantiomeric excess was determined by HPLC on Regis Chiral 5 Micron with hexane/iPrOH (90:10) as the eluent. Flow rate: 0.8 mL/min, $\lambda = 254$ nm; $t_{minor} =$ 8.732 min, $t_{major} = 10.352$ min; ¹H NMR (CDCl₃, 300 MHz) δ 0.92 (t, J = 7.2 Hz, 3H), 3.48 (d, J = 14.1 Hz, 1H), 3.67 (d, J =14.1 Hz, 1H), 4.01 (q, J = 7.2 Hz, 2H), 7.14–7.23 (m, 5H), 7.26 (m, 2H), 7.36 (m, 1H), 7.91 (d, 2H); APCIMS *m/z*: 301.1 (M⁺ + 1).

(*R*)-Ethyl 2-chloro-2-fluoro-3-oxo-3-phenylpropanoate (**2c**)

71% yield, 66% ee. The enantiomeric excess was determined by HPLC on Regis Chiral 5 Micron with hexane/iPrOH (90:10) as the eluent. Flow rate: 0.8 mL/min, $\lambda = 254$ nm; $t_{minor} =$ 12.220 min, $t_{major} = 14.492$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.18 (t, J = 7.2 Hz, 3H), 4.32 (q, J = 7.2 Hz, 2H), 7.47–7.49 (m, 2H), 7.60 (m, 1H), 8.02–8.05 (m, 2H); APCIMS *m/z*: 245.0 (M⁺ + 1).

(*R*)-Ethyl 2-bromo-2-fluoro-3-oxo-3-phenylpropanoate (**2d**)

43% yield, 66% ee. The enantiomeric excess was determined by HPLC on Regis Chiral 5 Micron with hexane/iPrOH (90:10) as the eluent. Flow rate: 1.2 mL/min, $\lambda = 254$ nm; $t_{minor} =$ 5.912 min, $t_{major} = 7.004$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.28 (t, J = 7.2 Hz, 3H), 4.38 (q, J = 7.2 Hz, 2H), 7.48–7.53 (m, 2H), 7.64 (m, 1H), 8.06–8.10 (m, 2H); APCIMS *m/z*: 289.0 (M⁺ + 1).

(S)-Ethyl 2-fluoro-3-oxo-3-phenylpropanoate (2e)

69% yield, 31% ee. The enantiomeric excess was determined by HPLC on (R,R)-WHELK-O1with hexane/iPrOH (95:5) as the eluent. Flow rate: 1.0 mL/min, λ = 254 nm; t_{minor} = 5.904 min, t_{major} = 5.380 min; ¹H NMR (CDCl₃, 300 MHz) δ 1.15 (t, *J* = 7.2 Hz, 3H), 4.23 (q, *J* = 7.2 Hz, 2H), 5.70–5.87 (s, *J* = 48.9 Hz, 1H), 7.18–7.45 (m, 2H), 7.53 (m, 1H), 7.94–7.98 (m, 2H); APCIMS *m/z*: 211.1 (M⁺ + 1).

(R)-Ethyl 2-fluoro-2-cyclohexanonecarboxylate (2f)

73% yield, 63% ee. The enantiomeric excess was determined by HPLC on (R,R)-WHELK-O1 with hexane/iPrOH (90:10) as the eluent. Flow rate: 0.8 mL/min, $\lambda = 210$ nm; $t_{minor} = 10.848$ min, $t_{major} = 12.440$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.32 (t, J = 7.2 Hz, 3H), 1.61–1.89 (m, 2H), 2.06–2.10 (m, 1H), 2.51–2.73 (m, 3H), 4.30 (q, J = 7.2 Hz, 2H).

(S)-Ethyl 2-(4'-methylbenzenesulfonyl)-2-fluoro-3oxo-3-phenylpropanoate (**2g**)

74% yield, 78% ee. The enantiomeric excess was determined by HPLC on Venusil Chiral OD-H with hexane/iPrOH (92:8) as the eluent. Flow rate: 0.3 mL/min, $\lambda = 254$ nm; $t_{minor} =$ 27.176 min, $t_{major} = 24.288$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.41 (t, J = 7.2 Hz, 3H), 2.36 (s, 3H), 4.38 (q, J = 7.2 Hz, 2H), 7.09–7.18 (m, 4H), 7.20–7.35 (m, 3H), 7.54 (s, 2H); APCIMS m/z: 365.1 (M⁺ + 1).

(S)-Ethyl 2-(*N*-ethylmaleimide)-2-fluoro-3-oxo-3phenylpropanoate (**2h**)

78% yield, 82% ee. The enantiomeric excess was determined by HPLC on Venusil Chiral OD-H with hexane/iPrOH (94:6) as the eluent. Flow rate: 0.5 mL/min, $\lambda = 254$ nm; $t_{minor} =$ 13.832 min, $t_{major} = 12.432$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.16 (t, J = 7.2 Hz, 3H), 1.28 (t, J = 7.2 Hz, 3H), 2.58 (dd, J =18.3 Hz, 1H), 3.05 (dd, J = 18.3 Hz, 1H), 3.61 (q, J = 7.2 Hz, 2H), 4.14 (m, 1H), 4.42 (m, J = 7.2 Hz, 2H), 7.46–7.51 (m, 2H), 7.62 (m, 1H), 8.12 (m, 2H); ¹³C NMR (CDCl₃, 75 MHz) δ 12.9, 13.8, 30.8, 34.1, 44.9, 63.5, 128.5, 128.8, 130.1, 130.2, 134.8, 168.5, 174.8; APCIMS *m/z*: 336.1 (M⁺ + 1).

(S)-Ethyl 2-(*N*-benzylmaleimide)-2-fluoro-3-oxo-3phenylpropanoate (**2i**)

83% yield, 81% ee. The enantiomeric excess was determined by HPLC on Venusil Chiral OD-H with hexane/iPrOH (92:8) as the eluent. Flow rate: 0.3 mL/min, $\lambda = 254$ nm: $t_{minor} =$ 27.176 min, $t_{major} = 24.288$ min; ¹H NMR (CDCl₃, 300 MHz) δ 1.27 (t, J = 7.2 Hz, 3H), 2.58 (dd, J = 18.3 Hz, 1H), 3.05 (dd, J =18.3 Hz, 1H), 4.14 (m, 1H), 4.42 (m, J = 7.2 Hz, 2H), 7.67 (q, J = 15 Hz, 2H), 7.25–7.37 (m, 5H), 7.46–7.51 (m, 2H), 7.62 (m, 1H), 8.12 (m, 2H); ¹³C NMR (CDCl₃, 75 MHz) δ 13.8, 30.8, 42.7, 45.0, 45.3, 63.5, 128.0, 128.6, 128.7, 128.8, 130.1, 130.2, 134.9, 174.6; APCIMS *m/z*: 398.1 (M⁺ + 1).

Synthesis of racemic samples

The mixture of Selectfluor (0.057 g, 0.16 mmol) and ethyl benzoylacetate (0.031 g, 0.16 mmol) in CH_3CN (1 mL) was stirred at 90 °C under microwave irradiation for 40 min. The reaction was quenched by water. The mixture was extracted with ethyl acetate (3 mL). The organic layer was washed with aqueous HCl (2 M, 5 mL) and water (5 mL), and dried over Na₂SO₄. After evaporation of the solvent, the residue was purified by flash column chromatography (8:1 hexane/EtOAc) to give ethyl 2-methyl-2-fluoro-3-oxo-3-phenylpropanoate (0.032 g, 94%) as a colorless oil.

General procedure for recycling of C-1

The reaction mixture was loaded onto a fluorous silica-gel cartridge (5 g) and eluted by $80:20 \text{ MeOH/H}_2\text{O}$ to collect nonfluorous components, including the fluorinated product. The cartridge was eluted with MeOH to collect C-1. After concentration of the MeOH fraction and drying at 60 °C for 8 h, the recovered promoter was ready for the next round of reactions.

Supporting Information

Supporting Information File 1

Chiral HPLC chromatograms for fluorination products **2a–i**. LC–MS, NMR spectra for fluorination products **2a–i** and cinchona alkaloid derivatives **C-1**, **C-2**, **C-3** and **C-6**. LC–MS spectra for **2h** and HRMS spectra for **C-1**. [http://www.beilstein-journals.org/bjoc/content/ supplementary/1860-5397-8-138-S1.pdf]

Acknowledgements

W. Z. thanks the University of Massachusetts Boston for a grant support. W. Y. thanks for support from "NUST Excellence Initiative", NUST Research Funding (2011ZDJH07), Jiangsu Provincial Natural Science Foundation of China for Key Projects (BK2010070) and National Natural Science Foundation of China (20902047).

References

- Banks, R. E.; Tatlow, J. C., Eds. Organofluorine Chemistry: Principles and Commercial Applications; Topics in Applied Chemistry; Plenum Press: New York, 1994.
- Ojima, I.; McCarthy, J. R.; Welch, J. T., Eds. *Biomedical Frontiers of Fluorine chemistry*; ACS Symposium Series, Vol. 639; American Chemical Society: Washington, 1996. doi:10.1021/bk-1996-0639
- Ojima, I. Fluorine in Medicinal Chemistry and Chemical Biology; Wiley-Blackwell: Hoboken, 2009.
- Qiu, X.-L.; Xu, X.-H.; Qing, F.-L. *Tetrahedron* 2010, 66, 789–843. doi:10.1016/j.tet.2009.11.001
- Phan, L. T.; Clark, R. F.; Rupp, M.; Or, Y. S.; Chu, D. T. W.; Ma, Z. Org. Lett. 2000, 2, 2951–2954. doi:10.1021/ol006226o
- Abad, A.; Agulló, C.; Cuñat, A. C.; González-Coloma, A.; Pardo, D. Eur. J. Org. Chem. 2010, 2182–2198. doi:10.1002/ejoc.200901499
- Banks, R. E. J. Fluorine Chem. 1998, 87, 1–17. doi:10.1016/S0022-1139(97)00127-9
- Nyffeler, P. T.; Gonzalez Durón, S.; Burkart, M. D.; Vincent, S. P.; Wong, C.-H. *Angew. Chem., Int. Ed.* **2005**, *44*, 192–212. doi:10.1002/anie.200400648
- Singh, R. P.; Shreeve, J. M. Acc. Chem. Res. 2004, 37, 31–44. doi:10.1021/ar030043v
- 10. Cahard, D.; Audouard, C.; Plaquevent, J.-C.; Roques, N. Org. Lett. 2000, 2, 3699–3701. doi:10.1021/ol006610l
- 11. Mohar, B.; Baudoux, J.; Plaquevent, J.-C.; Cahard, D. Angew. Chem., Int. Ed. 2001, 40, 4214–4216. doi:10.1002/1521-3773(20011119)40:22<4214::AID-ANIE4214>3.0.CO ;2-B
- Cahard, D.; Audouard, C.; Plaquevent, J.-C.; Toupet, L.; Roques, N. *Tetrahedron Lett.* 2001, *42*, 1867–1869. doi:10.1016/S0040-4039(01)00017-X
- Shibata, N.; Suzuki, E.; Takeuchi, Y. J. Am. Chem. Soc. 2000, 122, 10728–10729. doi:10.1021/ja002732x
- 14. Shibata, N.; Suzuki, E.; Asahi, T.; Shiro, M. J. Am. Chem. Soc. 2001, 123, 7001–7009. doi:10.1021/ja010789t
- 15. Ma, J.-A.; Cahard, D. *Chem. Rev.* **2008**, *108*, PR1–PR43. doi:10.1021/cr800221v
- Fukuzumi, T.; Shibata, N.; Sugiura, M.; Nakamura, S.; Toru, T. J. Fluorine Chem. 2006, 127, 548–551. doi:10.1016/j.jfluchem.2006.01.004
- 17. Hintermann, L.; Togni, A. Angew. Chem., Int. Ed. 2000, 39, 4359–4362.
 doi:10.1002/1521-3773(20001201)39:23<4359::AID-ANIE4359>3.0.CO
 :2-P
- Hintermann, L.; Togni, A. Catalytic halogenation of activated methylene and methane compound. Eur. Pat. Appl. EP1151980 A1, Nov 7, 2001.
- Muñiz, K. Angew. Chem., Int. Ed. 2001, 40, 1653–1656.
 doi:10.1002/1521-3773(20010504)40:9<1653::AID-ANIE16530>3.0.CO
 :2-W
- Hintermann, L.; Perseghini, M.; Togni, A. Beilstein J. Org. Chem. 2011, 7, 1421–1435. doi:10.3762/bjoc.7.166
- Thierry, B.; Audouard, C.; Plaquevent, J.-C.; Cahard, D. Synlett 2004, 856–860. doi:10.1055/s-2004-817781
- 22. Baudequin, C.; Plaquevent, J.-C.; Audouard, C.; Cahard, D. *Green Chem.* **2002**, *4*, 584–586. doi:10.1039/b208817g
- Fache, F.; Piva, O. *Tetrahedron Lett.* 2001, 42, 5655–5657. doi:10.1016/S0040-4039(01)01036-X
- 24. Kaleta, Z.; Egyed, O.; Soós, T. Org. Biomol. Chem. 2005, 3, 2228–2230. doi:10.1039/b504973c

- Wang, L.; Cai, C.; Curran, D. P.; Zhang, W. Synlett 2010, 433–436. doi:10.1055/s-0029-1219198
- 26. Chu, Q.; Zhang, W.; Curran, D. P. Tetrahedron Lett. 2006, 47, 9287–9290. doi:10.1016/j.tetlet.2006.10.101
- 27. Zhang, W. Top. Curr. Chem. 2012, 308, 175–190. doi:10.1007/128_2011_257
- Song, C. E., Ed. Cinchona Alkaloids in Synthesis and Catalysis: Ligands, Immobilization and Organocatalysis; Wiley-VCH: Weinheim, Germany, 2009.
- 29. Zhang, W.; Curran, D. P. *Tetrahedron* **2006**, *62*, 11837–11865. doi:10.1016/j.tet.2006.08.051
- 30. Fluorous SPE cartridges are available from Fluorous Technologies, Inc. http://www.fluorous.com and http://www.silicycle.com

License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/2.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions: (http://www.beilstein-journals.org/bjoc)

The definitive version of this article is the electronic one which can be found at: doi:10.3762/bjoc.8.138