

Supporting Information

for

Rasta resin–triphenylphosphine oxides and their use as recyclable heterogeneous reagent precursors in halogenation reactions

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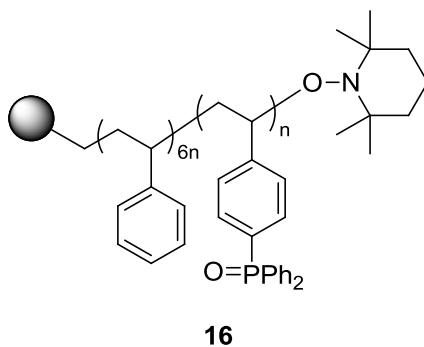
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Additional experimental details and characterization data of synthesized compounds

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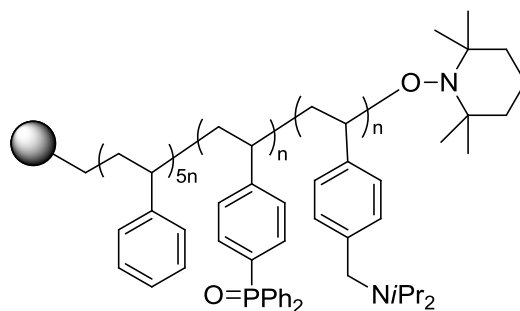
General experimental details

All reagents were obtained from the Acros, Aldrich, and Alfa Aesar companies, and were used as received. All reactions were carried out in dry glassware under a N₂ atmosphere, and were monitored by TLC analysis using GF₂₅₄ silica gel coated plates. ¹H and ¹³C NMR spectra were recorded in CDCl₃ on a Bruker DRX-300 or DRX-400 spectrometer operating at 300/400 MHz for ¹H and 75/100 MHz for ¹³C analysis. Gel-phase ³¹P NMR analysis was performed on a Bruker DRX-400 spectrometer operating at 162 MHz. Chemical shift data is expressed in ppm with reference to TMS. Elemental analysis was performed at the Shanghai Institute of Organic Chemistry. EIMS data was recorded on a Finnigan MAT 96 mass spectrometer. X-ray diffraction data was collected on a Bruker Smart Apex II CCD.



Synthesis of 16

Rasta resin-PPh₃ **14** [1] (25.0 g, 24.3 mmol) was added to dichloromethane (50 mL) and 50% hydrogen peroxide (50 mL). The reaction mixture was magnetically stirred at room temperature for 1 h. The resin was then filtered, washed sequentially with CH₂Cl₂ (3 × 100 mL), methanol (3 × 100 mL), THF (3 × 100 mL), diethyl ether (3 × 100 mL) and hexane (3 × 100 mL), and then dried under vacuum at 60 °C for 16 h to afford **16** as white free-flowing beads (100% yield). Elemental analysis was used to determine the phosphine content (3.0%), and thus a loading level of 0.97 mmol PPh₃/g. A peak at δ 29.4 was observed by gel-phase ³¹P NMR analysis.



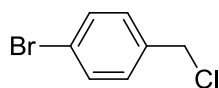
18

Synthesis of **18**

Rasta resin-NBnPr₂-PPh₃ **19** [2] (20.0 g, 18.8 mmol) was added to dichloromethane (50 mL) and 50% hydrogen peroxide (50 mL). The reaction mixture was magnetically stirred at room temperature for 1 h. The resin was then filtered, washed sequentially with CH₂Cl₂ (3 × 100 mL), methanol (3 × 100 mL), THF (3 × 100 mL), diethyl ether (3 × 100 mL) and hexane (3 × 100 mL) and then dried under vacuum at 60 °C for 16 h to afford **18** as white free-flowing beads (100% yield). Elemental analysis was used to determine the phosphine content (3.3%), and nitrogen content (1.5%), and thus loading levels of 1.07 mmol PPh₃/g, 1.06 mmol NiPr₂/g. A peak at δ 29.8 was observed by gel-phase ³¹P NMR analysis.

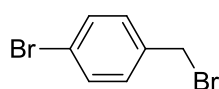
General procedure for Appel reactions using **16**

To **16** (0.6 g, 0.6 mmol) was added dichloromethane (5 mL) in a round-bottom flask. After 10 min, oxalyl chloride or oxalyl bromide was added (0.6 mmol). The reaction mixture was magnetically stirred at room temperature. Upon cessation of gas evolution, **4** was added (0.5 mmol), and the reaction mixture was heated to reflux. After the reaction was complete according to TLC analysis, the mixture was cooled to room temperature and filtered. The solid on the funnel was washed with dichloromethane (3 × 10 mL), and the filtrate was concentrated under reduced pressure to afford the desired product **5** in an essentially pure state based on ¹H and ¹³C NMR spectroscopic analyses.



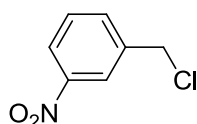
5Aa

1-Bromo-4-(chloromethyl)benzene (5Aa, Table 1, entry 1) [3]. Isolated yield: 98%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 4.52 (s, 2H), 7.25 (d, 2H, $J = 8.4$ Hz), 7.48 (d, 2H, $J = 8.4$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 45.5, 122.6, 130.4, 132.0, 136.6; MS for $\text{C}_7\text{H}_6\text{BrCl}$: calcd 204.0, found 204.0.



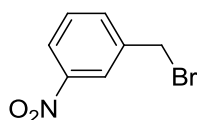
5Ab

1-Bromo-4-(bromomethyl)benzene (5Ab, Table 1, entry 2) [4]. Isolated yield: 92%. $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 4.44 (s, 2H), 7.27 (d, 2H, $J = 8.4$ Hz), 7.47 (d, 2H, $J = 8.4$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 32.5, 122.6, 130.8, 132.1, 136.9; MS for $\text{C}_7\text{H}_6\text{Br}_2$: calcd 247.9, found 247.9.



5Ba

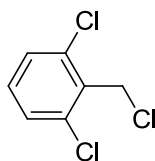
1-(Chloromethyl)-3-nitrobenzene (5Ba, Table 1, entry 3) [3]. Isolated yield: 93%. $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 4.67 (s, 2H), 7.56 (t, 1H, $J = 7.8$ Hz), 7.74 (d, 1H, $J = 7.6$ Hz), 8.20 (d, 1H, $J = 8.2$ Hz), 8.28 (s, 1H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 44.7, 123.5, 123.6, 130.0, 134.6, 148.5; MS for $\text{C}_7\text{H}_6\text{ClNO}_2$: calcd 171.0, found 171.0.



5Bb

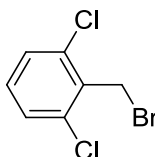
1-(Bromomethyl)-3-nitrobenzene (5Bb, Table 1, entry 4) [4]. Isolated yield: 91%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 4.55 (s, 2H), 7.55 (t, 1H, $J = 7.9$ Hz), 7.74 (d, 1H, $J = 7.7$ Hz), 8.16 (d, 1H, $J = 8.2$ Hz), 8.26 (s, 1H); $^{13}\text{C-NMR}$

(100 MHz, CDCl₃) δ 31.4, 123.4, 124.1, 130.1, 135.2, 139.9, 148.5; MS for C₇H₆BrNO₂: calcd 216.0, found 215.0.



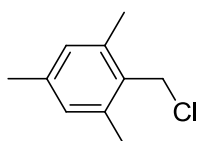
5Ca

1,3-Dichloro-2-(chloromethyl)benzene (5Ca, Table 1, entry 5) [5]. Isolated yield: 93%. ¹H-NMR (400 MHz, CDCl₃) δ 4.88 (s, 2H), 7.19-7.35 (m, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 40.9, 128.7, 130.4, 133.6, 136.3; MS for C₇H₅Cl₃: calcd 193.9, found 193.9.



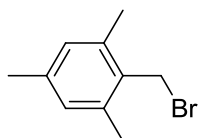
5Cb

2-(Bromomethyl)-1,3-dichlorobenzene (5Cb, Table 1, entry 6) [5]. Isolated yield: 90%. ¹H-NMR (400 MHz, CDCl₃) δ 4.76 (s, 2H), 7.17-7.34 (m, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 27.6, 128.7, 130.2, 133.8, 136.1; MS for C₇H₅BrCl₂: calcd 237.9, found 237.9.



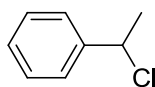
5Da

2-(Chloromethyl)-1,3,5-trimethylbenzene (5Da, Table 1, entry 7) [5]. Isolated yield: 98%. ¹H-NMR (300 MHz, CDCl₃) δ 2.26 (s, 3H), 2.39 (s, 6H), 4.65 (s, 2H), 6.87 (s, 2H); ¹³C-NMR (100 MHz, CDCl₃) δ 19.3, 21.2, 41.3, 129.4, 131.2, 137.6, 138.6; MS for C₁₀H₁₃Cl: calcd 168.1, found 168.1.



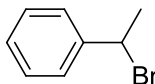
5Db

2-(Bromomethyl)-1,3,5-trimethylbenzene (5Db, Table 1, entry 8) [6]. Isolated yield: 93%. $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 2.26 (s, 3H), 2.38 (s, 6H), 4.56 (s, 2H), 6.85 (s, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 19.3, 21.2, 29.9, 129.4, 131.2, 137.6, 138.7; MS for $\text{C}_{10}\text{H}_{13}\text{Br}$: calcd 212.0, found 212.0.



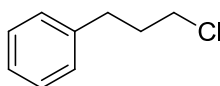
5Ea

(1-Chloroethyl)benzene (5Ea, Table 1, entry 9) [7]. Isolated yield: 85%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 1.86 (d, 3H, $J = 6.8$ Hz), 5.10 (q, 1H, $J = 6.8$ Hz), 7.30-7.43 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 26.6, 58.9, 126.6, 128.4, 128.7, 142.9; MS for C_8H_9 ($\text{C}_8\text{H}_9\text{Cl}$): calcd 105.1, found 105.1.



5Eb

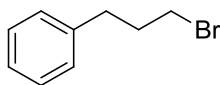
(1-Bromoethyl)benzene (5Eb, Table 1, entry 10) [5]. Isolated yield: 89%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 2.04 (d, 3H, $J = 6.9$ Hz), 5.21 (q, 1H, $J = 6.9$ Hz), 7.24-7.44 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 27.0, 49.8, 127.0, 128.5, 128.8, 143.4; MS for C_8H_9 ($\text{C}_8\text{H}_9\text{Br}$): calcd 105.1, found 105.1.



5Fa

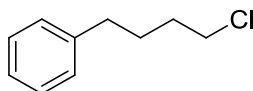
(3-Chloropropyl)benzene (5Fa, Table 1, entry 11) [5]. Isolated yield: 95%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 2.05-2.12 (m, 2H), 2.78 (t, 2H, $J = 7.2$ Hz), 3.53 (t, 2H, $J = 6.3$ Hz), 7.19-7.31 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz,

CDCl_3) δ 32.9, 34.1, 44.4, 126.3, 128.6, 128.7, 140.8; MS for $\text{C}_9\text{H}_{11}\text{Cl}$: calcd 154.1, found 154.1.



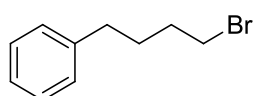
5Fb

(3-Bromopropyl)benzene (5Fb, Table 1, entry 12) [8]. Isolated yield: 90%. ^1H -NMR (300 MHz, CDCl_3) δ 2.12-2.21 (m, 2H), 2.78 (t, 2H, $J = 7.2$ Hz), 3.39 (t, 2H, $J = 6.6$ Hz), 7.18-7.32 (m, 5H); ^{13}C -NMR (100 MHz, CDCl_3) δ 33.3, 34.1, 34.3, 126.3, 128.6, 128.7, 140.7; MS for $\text{C}_9\text{H}_{11}\text{Br}$: calcd 198.0, found 198.0.



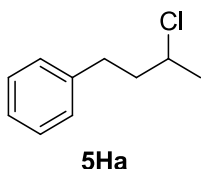
5Ga

(4-Chlorobutyl)benzene (5Ga, Table 1, entry 13) [9]. Isolated yield: 98%. ^1H -NMR (400 MHz, CDCl_3) δ 1.78-1.80 (m, 4H), 2.64 (t, 2H, $J = 7.0$ Hz), 3.54 (M, 2H), 7.17-7.20 (m, 3H), 7.26-7.30 (m, 2H); ^{13}C -NMR (100 MHz, CDCl_3) δ 28.8, 32.3, 35.3, 45.1, 126.1, 126.2, 128.6, 142.1; MS for $\text{C}_{10}\text{H}_{13}\text{Cl}$: calcd 168.1, found 168.1.

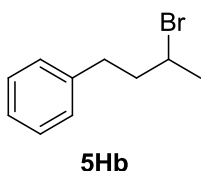


5Gb

(4-Bromobutyl)benzene (5Gb, Table 1, entry 14) [10]. Isolated yield: 92%. ^1H -NMR (400 MHz, CDCl_3) δ 1.75-1.81 (m, 2H), 1.86-1.91 (m, 2H), 2.64 (t, 2H, $J = 7.5$ Hz), 3.42 (t, 2H, $J = 6.7$ Hz), 7.17-7.30 (m, 5H); ^{13}C -NMR (100 MHz, CDCl_3) δ 30.0, 32.4, 33.8, 35.1, 126.1, 128.5, 128.5, 141.9; MS for $\text{C}_{10}\text{H}_{13}\text{Br}$: calcd 212.0, found 212.0.



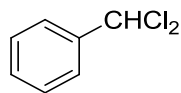
(3-Chlorobutyl)benzene (5Ha, Table 1, entry 15 and 17, Table 4) [11]. Isolated yields: 95% for entry 15 and 92% for entry 17. $^1\text{H-NMR}$ (300 MHz, CDCl_3) δ 1.53 (d, 3H, J = 6.5 Hz), 1.98-2.05 (m, 2H), 2.72-2.86 (m, 2H), 3.96-4.03 (m, 1H), 7.20-7.32 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 25.6, 33.1, 42.1, 58.1, 126.3, 128.6, 128.7, 141.3; MS for $\text{C}_{10}\text{H}_{13}\text{Cl}$: calcd 168.1, found 168.1.



(3-Bromobutyl)benzene (5Hb, Table 1, entry 16) [12]. Isolated yield: 91%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 1.73 (d, 3H, J = 6.6 Hz), 2.03-2.15 (m, 2H), 2.74-2.86 (m, 2H), 4.06-4.09 (m, 1H), 7.18-7.31 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 26.7, 34.1, 42.8, 51.1, 126.4, 128.6, 128.7, 141.1; MS for $\text{C}_{10}\text{H}_{13}\text{Br}$: calcd 212.0, found 212.0.

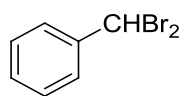
General procedure for aldehyde halogenation reactions using 16

To **16** (0.3 g, 0.3 mmol) was added chloroform (3 mL) in a round-bottom flask. After 10 min, oxalyl chloride or oxalyl bromide was added (0.3 mmol). The reaction mixture was magnetically stirred at room temperature. Upon cessation of gas evolution, **6** was added (0.1 mmol) and the reaction mixture was heated to reflux. After 72 h, the reaction mixture was cooled to room temperature and then filtered. The solid on funnel was washed with dichloromethane (3×5 mL). The filtrate was concentrated under reduced pressure to afford the crude product. The crude product was purified by flash silica gel column chromatography using 5% ethyl acetate in hexanes as the eluent.



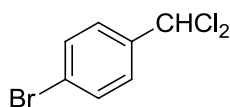
7Aa

(Dichloromethyl)benzene (7Aa, Table 2, entry 1) [13]. Isolated yield: 54%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 6.73 (s, 1H), 7.39-7.42 (m, 3H), 7.58-7.60 (m, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 71.9, 126.2, 128.9, 130.0, 140.5; MS for $\text{C}_7\text{H}_6\text{Cl}^+$ ($\text{C}_7\text{H}_6\text{Cl}_2$): calc 125.0, found 125.0.



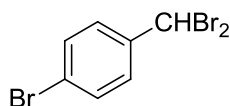
7Ab

(Dibromomethyl)benzene (7Ab, Table 2, entry 2) [14]. Isolated yield: 75%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 6.66 (s, 1H), 7.32-7.39 (m, 3H), 7.57 (d, 2H, $J = 7.3$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 41.2, 126.6, 128.7, 130.0, 142.0; MS for $\text{C}_7\text{H}_6\text{Br}^+$ ($\text{C}_7\text{H}_6\text{Br}_2$): calc 170.0, found 170.0.



7Ba

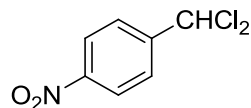
1-Bromo-4-(dichloromethyl)benzene (7Ba, Table 2, entry 3) [13]. Isolated yield: 65%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 6.66 (s, 1H), 7.44 (d, 2H, $J = 8.5$ Hz), 7.54 (d, 2H, $J = 8.5$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 71.0, 124.2, 127.9, 132.1, 139.5; MS for $\text{C}_7\text{H}_5\text{BrCl}_2$: calc 237.9, found 237.9.



7Bb

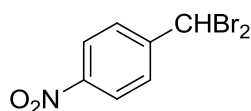
1-Bromo-4-(dibromomethyl)benzene (7Bb, Table 2, entry 4) [15]. Isolated yield: 83%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 6.59 (s, 1H), 7.45 (d, 2H, $J = 8.4$ Hz), 7.51 (d, 1H, $J = 8.4$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 39.7,

124.0, 128.3, 132.0, 141.1; MS for $C_7H_4Br_2$ ($C_7H_5Br_3$): calc 247.9, found 247.1.



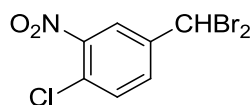
7Ca

1-(Dichloromethyl)-4-nitrobenzene (7Ca, Table 2, entry 5) [13]. Isolated yield: 64%. 1H -NMR (400 MHz, $CDCl_3$) δ 6.77 (s, 1H), 7.76 (d, 2H, J = 8.7 Hz), 8.26 (d, 2H, J = 8.7 Hz). ^{13}C -NMR (100 MHz, $CDCl_3$) δ 69.9, 124.2, 127.5, 146.3, 148.6; MS for $C_7H_5ClNO_2^+$ ($C_7H_5Cl_2NO_2$): calc 170.0, found 170.0.



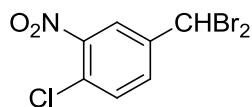
7Cb

1-(Dibromomethyl)-4-nitrobenzene (7Cb, Table 2, entry 6). Isolated yield: 89%. 1H -NMR (400 MHz, $CDCl_3$) δ 6.67 (s, 1H), 7.75 (d, 2H, J = 8.7 Hz), 8.25 (d, 2H, J = 8.7 Hz); ^{13}C -NMR (100 MHz, $CDCl_3$) δ 38.3, 124.2, 127.9, 148.0; HRMS for $C_7H_4BrNO_2^+$ ($C_7H_5Br_2NO_2$): calc 213.9504, found 214.0910.



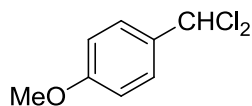
7Da

1-Chloro-4-(dibromomethyl)-2-nitrobenzene (7Da, Table 2, entry 7). Isolated yield: 61%. 1H -NMR (300 MHz, $CDCl_3$) δ 6.72 (s, 1H), 7.63 (d, 1H, J = 8.4 Hz), 7.76 (dd, 1H, J_1 = 8.4 Hz, J_2 = 2.3 Hz), 8.1 (d, 1H, J = 2.3 Hz); ^{13}C -NMR (100 MHz, $CDCl_3$) δ 69.1, 123.6, 128.7, 130.8, 132.6, 138.2, 140.3; MS for $C_7H_4Cl_2NO_2^+$ ($C_7H_4Cl_3NO_2$): calc 203.9, found 203.9.



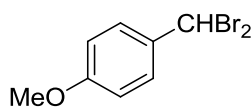
7Db

1-Chloro-4-(dibromomethyl)-2-nitrobenzene (7Db, Table 2, entry 8) [14]. Isolated yield: 93%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 6.63 (s, 1H), 7.59 (d, 1H, $J = 8.4$ Hz), 7.76 (d, 1H, $J = 8.4$ Hz), 8.09 (s, 1H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 36.9, 123.9, 128.4, 131.3, 132.6, 141.9, 147.6; MS for $\text{C}_7\text{H}_4\text{BrClNO}_2^+$ ($\text{C}_7\text{H}_4\text{Br}_2\text{ClNO}_2$): calc 247.9, found 248.1.



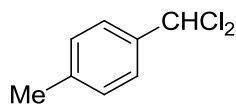
7Ea

1-(Dichloromethyl)-4-methoxybenzene (7Ea, Table 2, entry 9) [13]. Isolated yield: 94%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.83 (s, 3H), 6.71 (s, 1H), 6.92 (d, 2H, $J = 8.7$ Hz), 7.51 (d, 2H, $J = 8.7$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 55.5, 71.9, 114.1, 127.7, 132.8, 160.7; MS for CH_8ClO^+ ($\text{C}_8\text{H}_8\text{Cl}_2\text{O}$): calc 155.0, found 155.0.



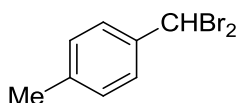
7Eb

1-(Dibromomethyl)-4-methoxybenzene (7Eb, Table 2, entry 10). NMR yield: 5%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.76 (s, 3H), 6.63 (s, 1H), 6.81 (d, 2H, $J = 8.5$ Hz), 7.46 (d, 2H, $J = 8.5$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 40.9, 55.4, 113.7, 121.2, 127.9, 160.4; MS for $\text{C}_8\text{H}_8\text{BrO}^+$ ($\text{C}_8\text{H}_8\text{Br}_2\text{O}$): calc 199.0, found 199.0



7Fa

1-(Dichloromethyl)-4-methylbenzene (7Fa, Table 2, entry 11) [13]. Isolated yield: 85%. ^1H -NMR (400 MHz, CDCl_3) δ 2.37 (s, 3H), 6.68 (s, 1H), 7.20 (d, 2H, $J = 8.0$ Hz), 7.45 (d, 2H, $J = 8.0$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.4, 72.0, 126.1, 129.5, 137.7, 140.2; MS for $\text{C}_8\text{H}_8\text{Cl}^+$ ($\text{C}_8\text{H}_8\text{Cl}_2$): calc 139.0, found 139.0.

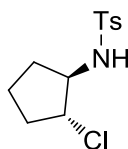


7Fb

1-(Dibromomethyl)-4-methylbenzene (7Fb, Table 2, entry 12). Isolated yield: 77%. ^1H -NMR (400 MHz, CDCl_3) δ 2.36 (s, 3H), 6.63 (s, 1H), 7.16 (d, 2H, $J = 8.0$ Hz), 7.45 (d, 2H, $J = 8.0$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.4, 41.3, 126.6, 129.4, 139.4, 140.2; MS for $\text{C}_8\text{H}_8\text{Br}^+$ ($\text{C}_8\text{H}_8\text{Br}_2$): calc 183.0, found 182.9.

General procedure for aziridine halogenation reactions using **16**

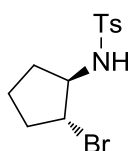
To **16** (0.6 g, 0.6 mmol) was added dichloromethane (5 mL) in round-bottom flask. After 10 min, oxalyl chloride or oxalyl bromide was added (0.6 mmol). The reaction mixture was magnetically stirred at room temperature. Upon cessation of gas evolution, **8** was added (0.5 mmol), and the reaction mixture was heated to reflux. After the reaction was complete according to TLC analysis, the mixture was cooled to room temperature and filtered. The solid on funnel was washed with dichloromethane (3×10 mL), and the filtrate was concentrated to afford the desired product **9** in an essentially pure state based on ^1H and ^{13}C NMR spectroscopic analyses.



9Aa

***N*-(2-Chlorocyclopentyl)-4-methylbenzenesulfonamide (9Aa, Table 3, entry 1)** [16]. Isolated yield: 95%.

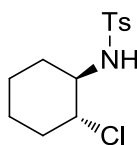
$^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 1.41-1.43 (m, 1H), 1.72-1.85 (m, 3H), 2.11-2.18 (m, 2H), 2.44 (s, 3H), 3.57-3.59 (m, 1H), 4.06-4.10 (m, 1H), 5.28 (d, 1H, $J = 5.9$ Hz), 7.33 (d, 2H, $J = 8.0$ Hz), 7.79 (d, 2H, $J = 8.0$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 20.9, 21.8, 30.7, 33.6, 62.8, 63.8, 127.4, 130.0, 137.1, 144.0; MS for $\text{C}_{12}\text{H}_{16}\text{ClNO}_2\text{S}$: calcd 273.1, found 273.1.



9Ab

***N*-(2-Bromocyclopentyl)-4-methylbenzenesulfonamide (9Ab, Table 3, entry 2)** [17]. Isolated yield: 93%.

$^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 1.26-1.42 (m, 1H), 1.63-1.82 (m, 2H), 1.93-1.96 (m, 1H), 2.16-2.28 (m, 2H), 2.45 (s, 3H), 3.65-3.80 (m, 1H), 4.10-4.11 (m, 1H), 5.25 (d, 1H, $J = 4.2$ Hz), 7.34 (d, 2H, $J = 7.4$ Hz), 7.80 (d, 2H, $J = 7.4$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 21.5, 21.7, 30.8, 34.2, 54.4, 63.1, 127.4, 130.0, 136.9, 143.9; MS for $\text{C}_{12}\text{H}_{16}\text{BrNO}_2\text{S}$: calcd 317.0, found 317.0.

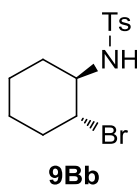


9Ba

***N*-(2-Chlorocyclohexyl)-4-methylbenzenesulfonamide (9Ba, Table 3, entry 3 and 5, Table 5)** [18]. Isolated

yields: 89% for entry 3, and 95% for entry 5. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 1.23-1.32 (m, 4H), 1.58-1.71 (m, 2H), 2.16-2.24 (m, 2H), 2.43 (s, 3H), 3.08-3.10 (m, 1H), 3.71 (dt, 1H, $J_1 = 9.6$ Hz, $J_2 = 4.1$ Hz), 4.98-5.10 (m, 1H), 7.31 (d, 2H, $J = 8.2$ Hz), 7.78 (d, 2H, $J = 8.2$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 21.4, 23.2, 24.0, 32.1, 34.7, 58.4, 61.9,

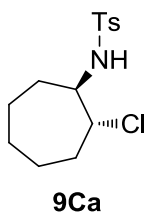
127.1, 129.5, 137.5, 143.2; MS for C₁₃H₁₆ClNO₂S: calcd 287.1, found 287.1.



***N*-(2-Bromocyclohexyl)-4-methylbenzenesulfonamide (9Bb, Table 3, entry 4)** [17]. Isolated yield: 92%.

¹H-NMR (300 MHz, CDCl₃) δ 1.25-1.44 (m, 3H), 1.65-1.82 (m, 3H), 2.27-2.32 (m, 2H), 2.43 (s, 3H), 3.14-3.17 (m, 1H), 3.84 (dt, 1H, *J*₁ = 9.6 Hz, *J*₂ = 5.7 Hz), 4.78-4.90 (m, 1H), 7.31 (d, 2H, *J* = 8.0 Hz), 7.78 (d, 2H, *J* = 8.3 Hz);

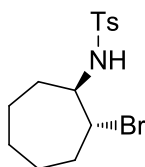
¹³C-NMR (100 MHz, CDCl₃) δ 21.7, 23.6, 25.5, 33.0, 35.9, 55.2, 58.8, 127.5, 129.7, 137.3, 143.6; MS for C₁₃H₁₈BrNO₂S: calcd 331.0, found 331.0.



***N*-(2-Chlorocycloheptyl)-4-methylbenzenesulfonamide (9Ca, Table 3, entry 6)**. Isolated yield: 96%. ¹H-NMR

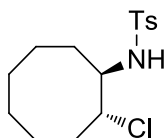
(400 MHz, CDCl₃) δ 1.45-1.60 (m, 7H), 2.00-2.05 (m, 3H), 2.43 (s, 3H), 3.36-3.38 (m, 1H), 3.92-3.94 (m, 1H), 5.03-5.08 (m, 1H), 7.31 (d, 2H, *J* = 8.1 Hz), 7.78 (d, 1H, *J* = 8.1 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ 21.8, 22.8,

22.9, 27.4, 31.0, 34.2, 61.9, 65.7, 127.6, 129.8, 137.1, 143.8; HRMS for C₁₄H₂₀ClNO₂S: calcd 301.0903, found 301.0899.



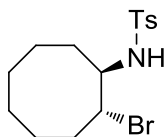
9Cb

***N*-(2-Bromocycloheptyl)-4-methylbenzenesulfonamide (9Cb, Table 3, entry 7).** Isolated yield: 98%. ^1H -NMR (400 MHz, CDCl_3) δ 1.48-1.61 (m, 7H), 2.03-2.15 (m, 3H), 2.43 (s, 3H), 3.48-3.50 (m, 1H), 4.04-4.09 (m, 1H), 4.92-4.99 (m, 1H), 7.31 (d, 2H, $J = 8.0$ Hz), 7.78 (d, 2H, $J = 8.2$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.8, 22.8, 24.0, 27.5, 31.3, 34.9, 58.8, 62.2, 127.6, 129.9, 137.0, 143.8; HRMS for $\text{C}_{14}\text{H}_{20}\text{BrNO}_2\text{S}$: calcd 345.0398, found 345.0393.



9Da

***N*-(2-Chlorocyclooctyl)-4-methylbenzenesulfonamide (9Da, Table 3, entry 8).** Isolated yield: 93%. ^1H -NMR (400 MHz, CDCl_3) δ 1.26-1.35 (m, 2H), 1.47-1.49 (m, 2H), 1.63-1.67 (m, 2H), 1.70-1.77 (m, 5H), 1.94-2.14 (m, 3H), 2.43 (s, 3H), 3.37-3.39 (m, 1H), 3.93-3.96 (m, 1H), 4.90-4.97 (m, 1H), 7.31 (d, 2H, $J = 8.1$ Hz), 7.77 (d, 2H, $J = 8.1$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.7, 24.0, 25.5, 25.7, 29.8, 31.3, 32.0, 60.7, 65.8, 127.6, 129.7, 136.7, 143.6; HRMS for $\text{C}_{15}\text{H}_{22}\text{ClNO}_2\text{S}$: calcd 315.1060, found 315.1053.

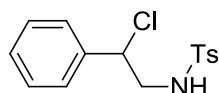


9Db

***N*-(2-Bromocyclooctyl)-4-methylbenzenesulfonamide (9Db, Table 3, entry 9) [19].** Isolated yield: 92%.

^1H -NMR (400 MHz, CDCl_3) δ 1.26-1.33 (m, 2H), 1.48 (s, 2H), 1.67-1.74 (m, 5H), 1.99-2.11 (m, 2H), 2.11-2.43 (m,

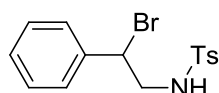
1H), 2.48 (s, 3H), 3.50 (s, 1H), 4.05-4.08 (m, 1H), 4.95-5.03 (m, 1H), 7.31 (d, 2H, $J = 7.8$ Hz), 7.78 (d, 2H, $J = 7.8$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.7, 25.0, 25.4, 25.5, 25.8, 31.8, 32.2, 59.7, 61.0, 127.7, 129.6, 136.5, 143.6; MS for $\text{C}_{15}\text{H}_{22}\text{BrNO}_2\text{S}$: calcd 360.0, found 360.0.



9Ea

***N*-(2-Chloro-2-phenylethyl)-4-methylbenzenesulfonamide (9Ea, Table 3, entry 10)** [20]. Isolated yield: 91%.

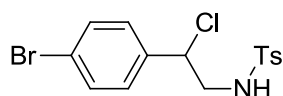
^1H -NMR (400 MHz, CDCl_3) δ 2.44 (s, 3H), 3.39-3.52 (m, 2H), 4.76 (t, 1H, $J = 6.0$ Hz), 4.87 (dd, 1H, $J_1 = 8.2$ Hz, $J_2 = 5.9$ Hz), 7.27-7.36 (m, 7H), 7.73 (d, 2H, $J = 8.3$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.6, 50.4, 61.7, 127.1, 127.3, 129.0, 129.1, 130.0, 137.0, 137.9, 143.9; MS for $\text{C}_{15}\text{H}_{16}\text{ClNO}_2\text{S}$: calcd 309.1, found 309.1.



9Eb

***N*-(2-Bromo-2-phenylethyl)-4-methylbenzenesulfonamide (9Eb, Table 3, entry 11)** [21]. Isolated yield: 93%.

^1H -NMR (400 MHz, CDCl_3) δ 2.45 (s, 3H), 3.54-3.60 (m, 2H), 4.83 (t, 1H, $J = 5.9$ Hz), 4.90 (t, 1H, $J = 7.0$ Hz), 7.27-7.33 (m, 7H), 7.73 (d, 2H, $J = 8.1$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.7, 50.2, 52.7, 127.2, 127.7, 129.2, 129.3, 130.0, 137.1, 138.3, 144.0; Ms for $\text{C}_{15}\text{H}_{16}\text{BrNO}_2\text{S}$: calcd 353.0, found 353.0.

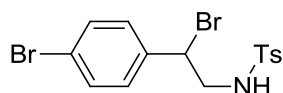


9Fa

***N*-(2-(4-Bromophenyl)-2-chloroethyl)-4-methylbenzenesulfonamide (9Fa, Table 3, entry 12)** [22]. Isolated

yield: 93%. ^1H -NMR (400 MHz, CDCl_3) δ 2.44 (s, 3H), 3.39-3.44 (m, 2H), 4.85 (t, 1H, $J = 7.7$ Hz), 4.92 (t, 1H, $J =$

6.5 Hz), 7.16 (d, 2H, $J = 8.3$ Hz), 7.31 (d, 2H, $J = 8.3$ Hz), 7.46 (d, 2H, $J = 8.3$ Hz), 7.70 (d, 2H, $J = 8.3$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.7, 50.4, 61.0, 123.3, 127.1, 129.0, 130.0, 132.2, 136.9, 137.0, 144.1; MS for $\text{C}_{15}\text{H}_{15}\text{BrClNO}_2\text{S}$: calcd 387.0, found 387.0.

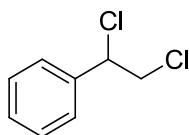


9Fb

***N*-(2-Bromo-2-(4-bromophenyl)ethyl)-4-methylbenzenesulfonamide (9Fb, Table 3, entry 13).** Isolated yield: 95%. ^1H -NMR (400 MHz, CDCl_3) δ 2.45 (s, 3H), 4.88 (t, 2H, $J = 7.3$ Hz), 7.16 (d, 2H, $J = 8.3$ Hz), 7.32 (d, 2H, $J = 8.3$ Hz), 7.44 (d, 2H, $J = 8.3$ Hz), 7.70 (d, 2H, $J = 8.3$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.7, 50.1, 51.6, 123.3, 127.1, 129.4, 130.0, 132.3, 137.0, 137.4, 144.1; HRMS for $\text{C}_{15}\text{H}_{15}\text{Br}_2\text{NO}_2\text{S}$: calcd 430.9190, found 430.9189.

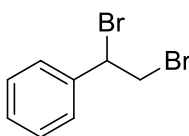
General procedure for epoxide halogenation reactions using **18**

To **18** (1.3 g, 1.2 mmol) was added chloroform (10 mL) in a round-bottom flask. After 10 min, oxalyl chloride or oxalyl bromide was added (1.1 mmol). The reaction mixture was magnetically stirred at room temperature. Upon cessation of gas evolution, **10** was added (0.5 mmol) and the reaction mixture was heated to reflux. After the reaction was completed as monitored by TLC, the mixture was cooled to room temperature and filtered. The solid on funnel was washed with dichloromethane (3×10 mL). The solvent of filtrate was removed to afford the desired product **11** in an essentially pure state based on ^1H and ^{13}C NMR spectroscopic analyses.



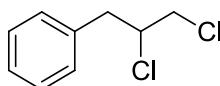
11Aa

(1,2-Dichloroethyl)benzene (11Aa, Table 6, entry 1) [23]. Isolated yield: 95%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.90-4.02 (m, 2H), 5.01 (t, 1H, $J = 7.3$ Hz), 7.35-7.42 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 48.5, 61.9, 127.5, 129.0, 129.3, 138.1; MS for $\text{C}_8\text{H}_8\text{Cl}_2$: calcd 174.0, found 174.0.



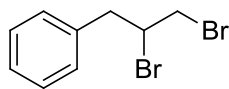
11Ab

(1,2-Dibromoethyl)benzene (11Ab, Table 6, entry 2) [24]. Isolated yield: 93%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.40-4.1 (m, 2H), 5.11-5.15 (dd, 1H, $J_1 = 10.6$ Hz, $J_2 = 5.5$ Hz), 7.24-7.32 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 35.1, 51.0, 127.7, 128.9, 129.3, 138.7; MS for $\text{C}_8\text{H}_8\text{Br}_2$: calcd 261.9, found 261.9.



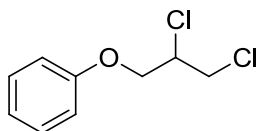
11Ba

(2,3-Dichloropropyl)benzene (11Ba, Table 6, entry 3) [23]. Isolated yield: 89%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.12 (dd, 1H, $J_1 = 14.2$ Hz, $J_2 = 7.3$ Hz), 3.37 (dd, 1H, $J_1 = 14.2$ Hz, $J_2 = 5.7$ Hz), 3.69-3.81 (m, 2H), 4.29-4.34 (m, 1H), 7.30-7.42 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 41.1, 47.6, 61.1, 127.3, 128.7, 129.7, 136.4; MS for $\text{C}_9\text{H}_{10}\text{Cl}_2$: calcd 188.0, found 188.0.



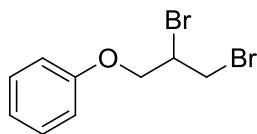
11Bb

(2,3-Dibromopropyl)benzene (11Bb, Table 6, entry 4) [25]. Isolated yield: 92%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.12 (dd, 1H, $J_1 = 14.5$ Hz, $J_2 = 7.8$ Hz), 3.38 (dd, 1H, $J_1 = 14.5$ Hz, $J_2 = 4.8$ Hz), 3.61 (dd, 1H, $J_1 = 10.4$ Hz, $J_2 = 9.0$ Hz), 3.81 (dd, 1H, $J_1 = 10.5$ Hz, $J_2 = 4.2$ Hz), 4.32-4.35 (m, 1H), 7.22-7.35 (m, 5H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 36.2, 42.1, 52.5, 127.3, 128.6, 129.6, 136.9; MS for $\text{C}_9\text{H}_{10}\text{Br}_2$: calcd 275.9, found 275.9.



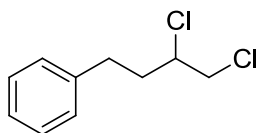
11Ca

(2,3-Dichloropropoxy)benzene (11Ca, Table 6, entry 5) [26]. Isolated yield: 95%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.88-3.99 (m, 2H), 4.27 (d, 2H, $J = 6.0$ Hz), 4.34-4.39 (m, 1H), 6.93 (d, 2H, $J = 7.9$ Hz), 6.94-7.01 (m, 1H), 7.28-7.32 (m, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 45.2, 57.5, 68.3, 114.9, 121.8, 129.8, 158.1; MS for $\text{C}_9\text{H}_{10}\text{Cl}_2\text{O}$: calcd 204.0, found 204.0.



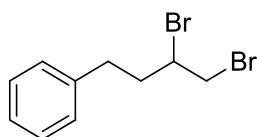
11Cb

(2,3-Dibromopropoxy)benzene (11Cb, Table 6, entry 6) [27]. Isolated yield: 96%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 3.88-3.97 (m, 2H), 4.35-4.40 (m, 2H), 4.42-4.46 (m, 1H), 6.95 (d, 2H, $J = 8.2$ Hz), 7.02 (t, 1H, $J = 7.4$ Hz), 7.29-7.34 (m, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 32.9, 47.9, 69.1, 115.0, 121.8, 129.7, 158.1; MS for $\text{C}_9\text{H}_{10}\text{Br}_2\text{O}$: calcd 291.9, found 291.9.



11Da

(3,4-Dichlorobutyl)benzene (11Da, Table 6, entry 7). Isolated yield: 98%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 2.01-2.05 (m, 1H), 2.28-2.33 (m, 1H), 2.74-2.80 (m, 1H), 2.88-2.92 (m, 1H), 3.66 (dd, 1H, $J_1 = 11.3$ Hz, $J_2 = 7.4$ Hz), 3.77 (dd, 1H, $J_1 = 11.3$ Hz, $J_2 = 5.1$ Hz), 3.98-4.00 (m, 1H), 7.20-7.22 (m, 3H), 7.28-7.32 (m, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 32.1, 36.8, 48.4, 60.3, 126.4, 128.6, 128.7, 140.5; HRMS for $\text{C}_{10}\text{H}_{12}\text{Cl}_2$: calcd 202.0316, found 202.0321.

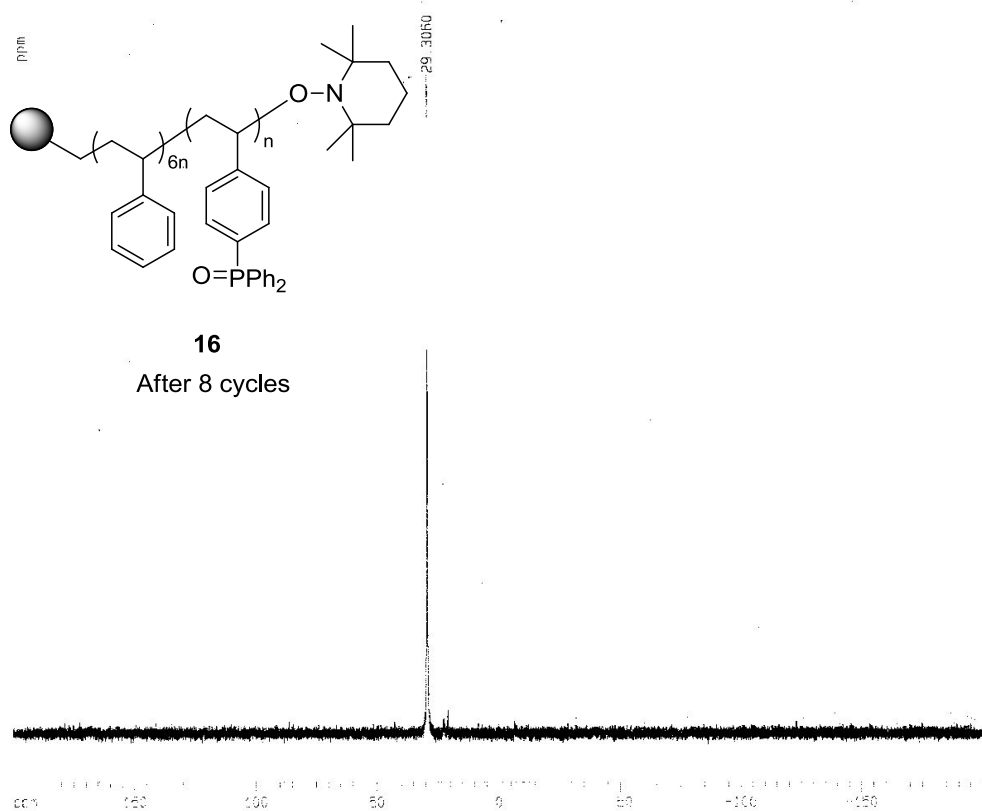
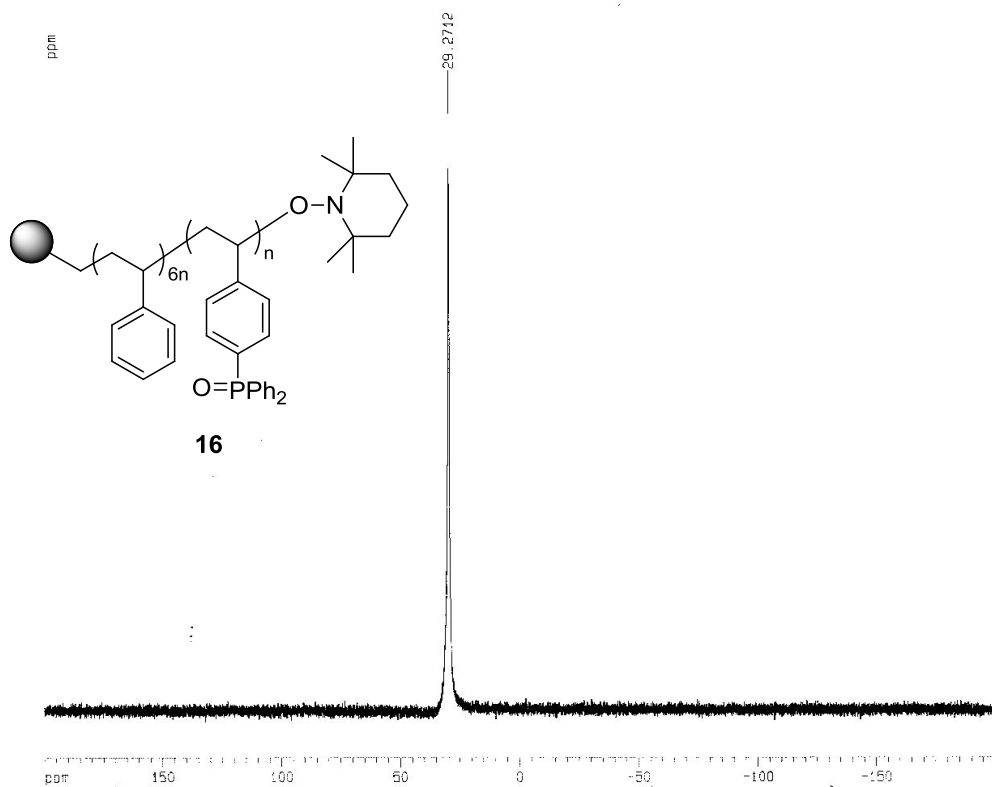


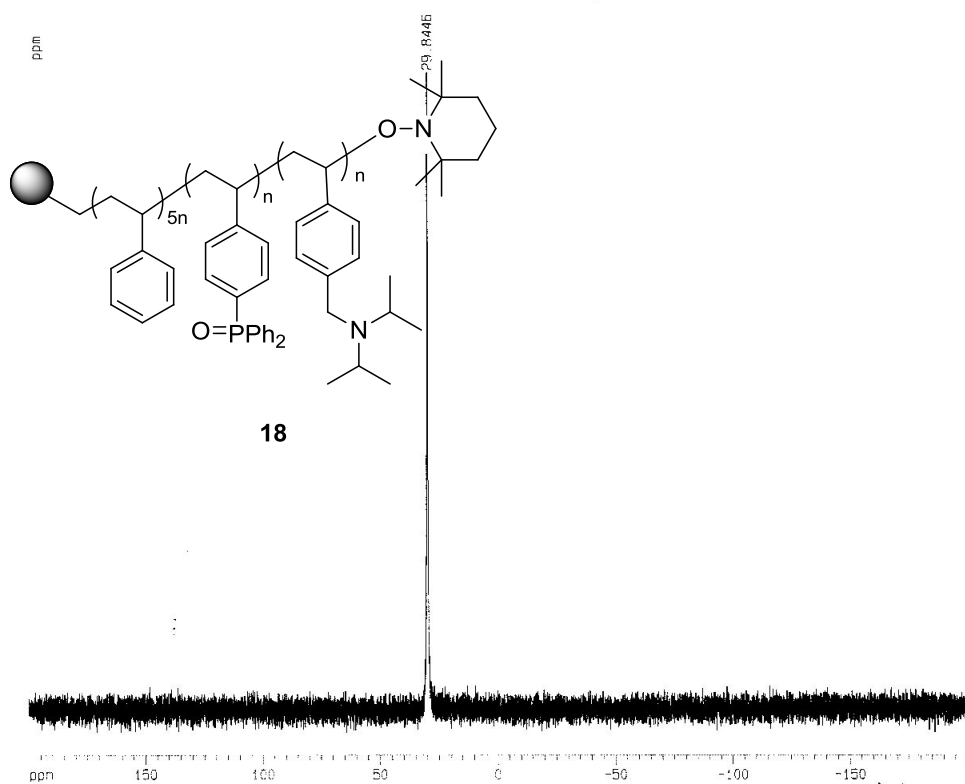
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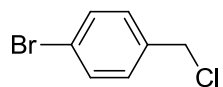
(3,4-Dibromobutyl)benzene (11Db, Table 6, entry 8). Isolated yield: 92%. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 2.08-2.11 (m, 1H), 2.46-2.50 (m, 1H), 2.74-2.79 (m, 1H), 2.91-2.94 (m, 1H), 3.65 (t, 1H, $J = 10.2$ Hz), 3.87 (dd, 1H, $J_1 = 10.3$ Hz, $J_2 = 4.4$ Hz), 4.09-4.13 (m, 1H), 7.20-7.24 (m, 3H), 7.30-7.33 (m, 2H); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 33.1, 36.4, 37.8, 52.2, 126.4, 128.7, 128.7, 140.4; HRMS for $\text{C}_{10}\text{H}_{12}\text{Br}_2$: calcd 289.9306, found 289.9300.

References

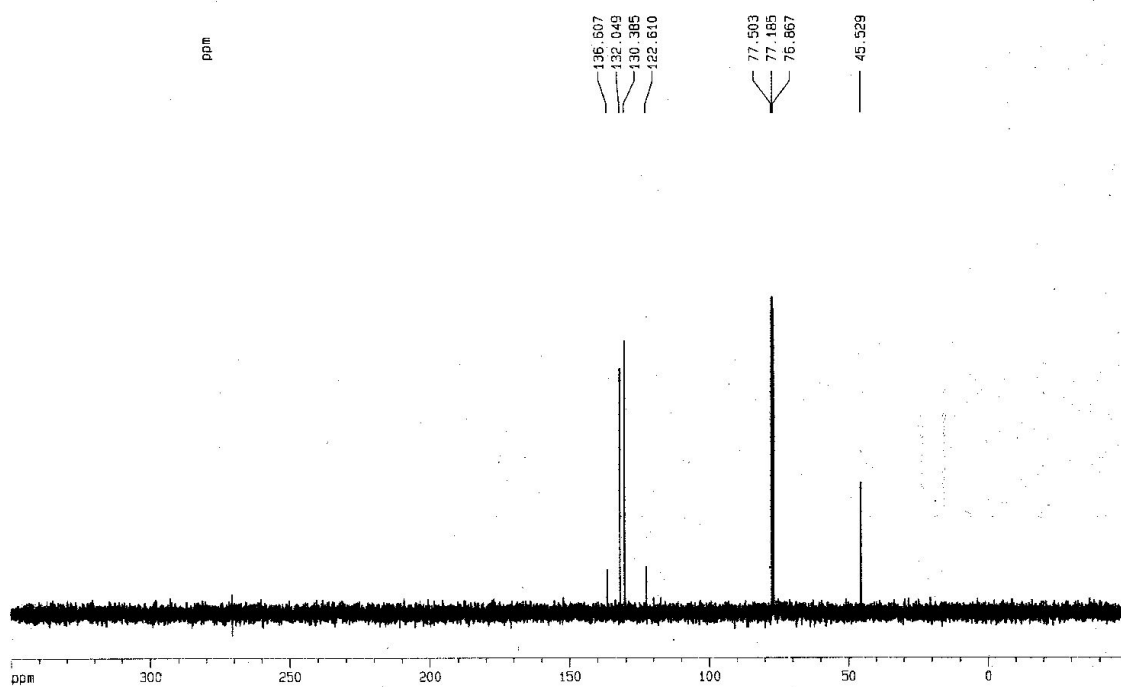
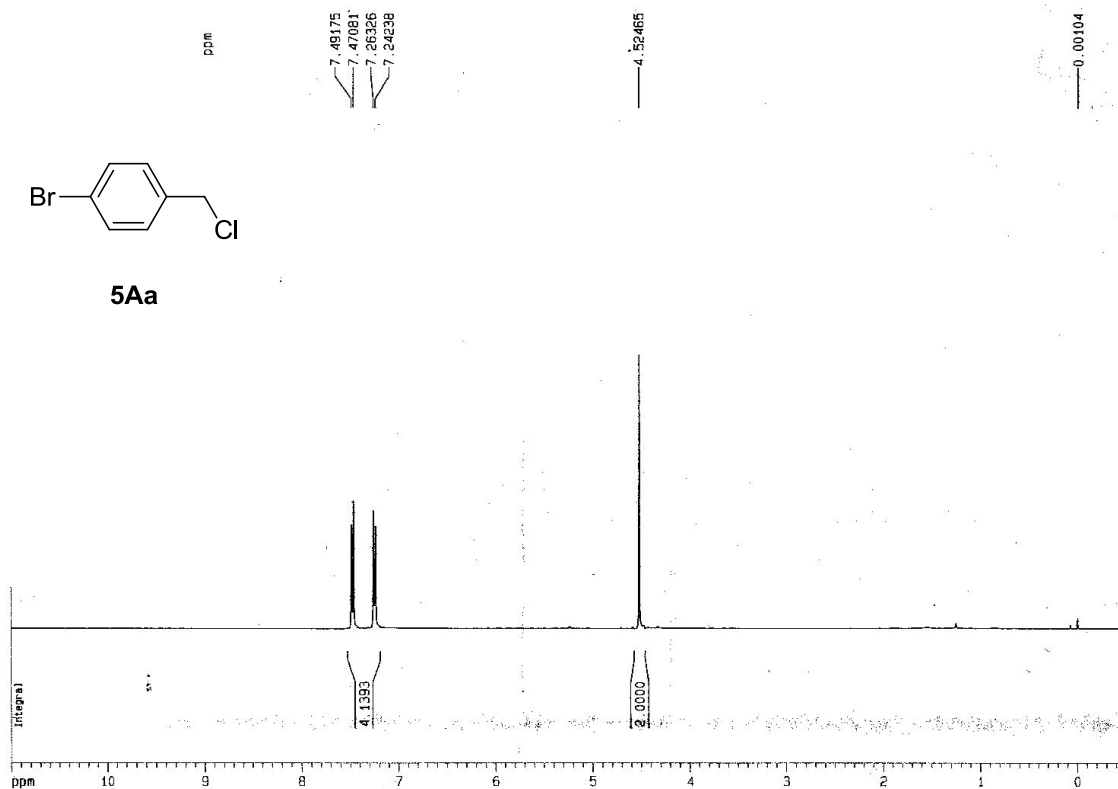
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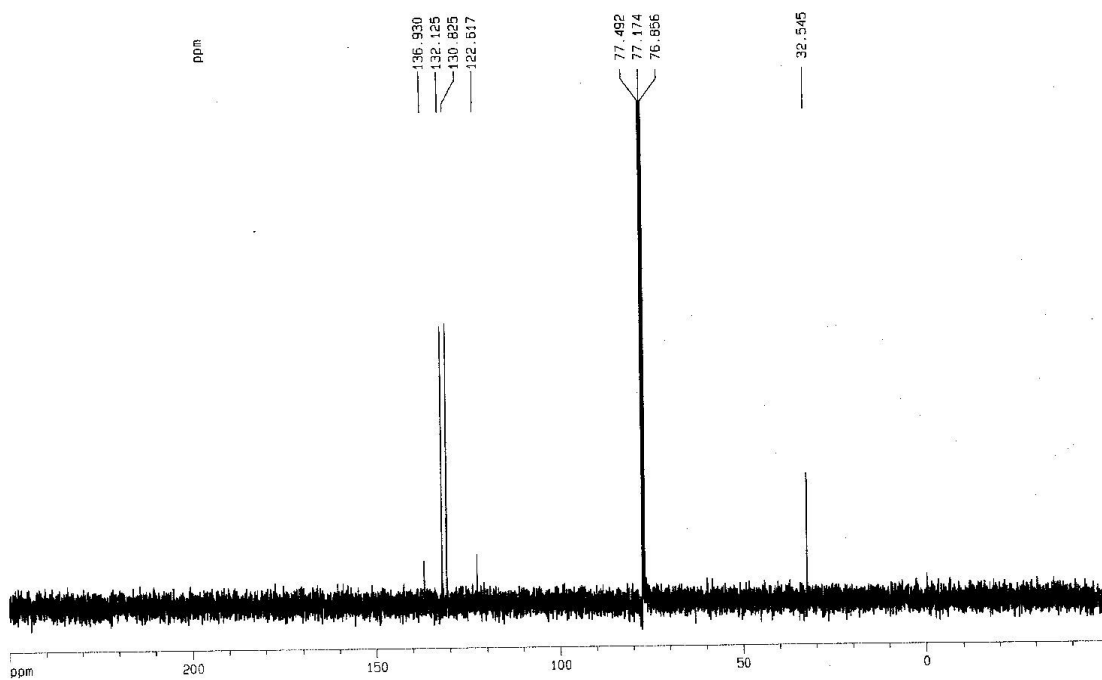
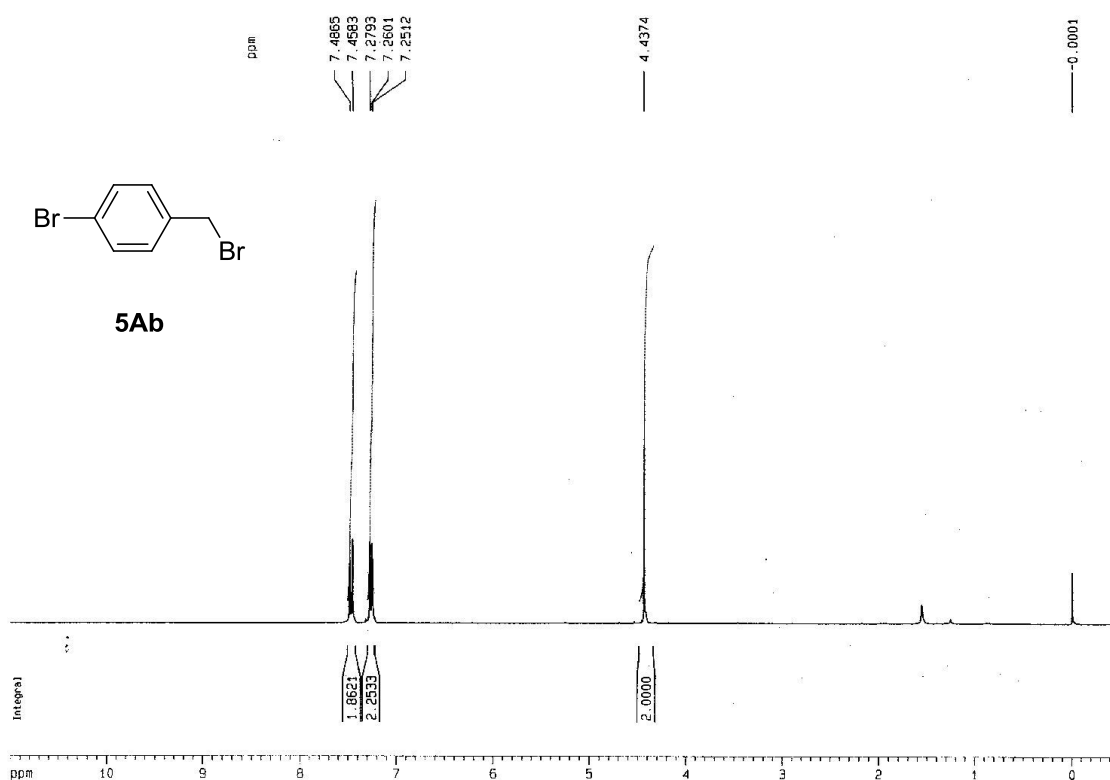


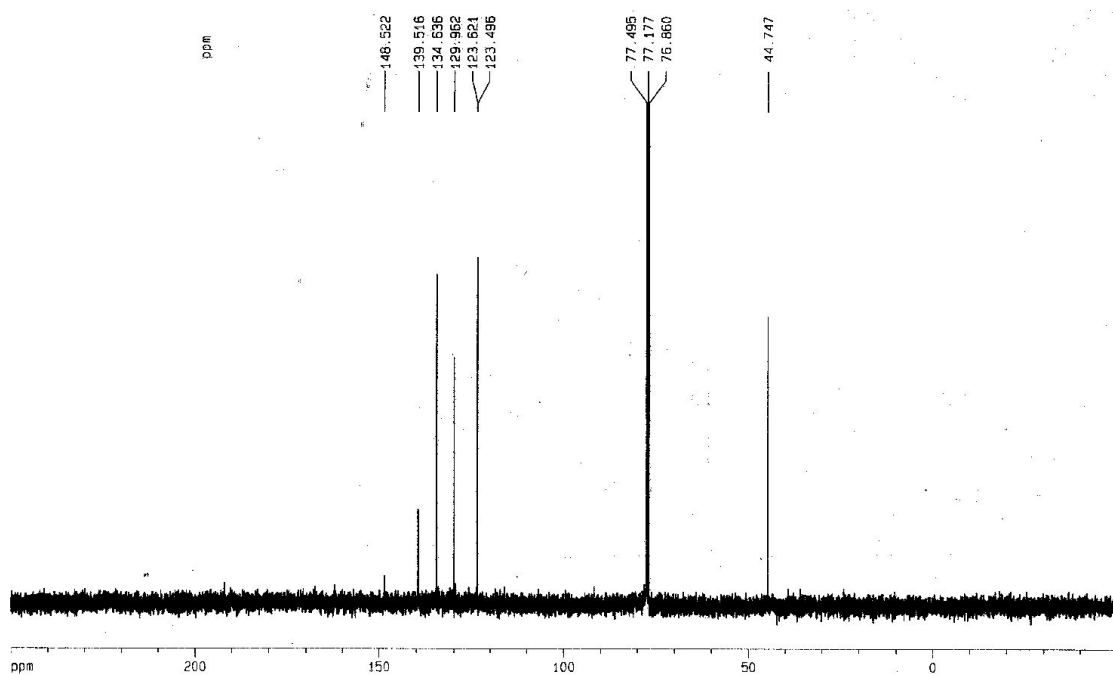
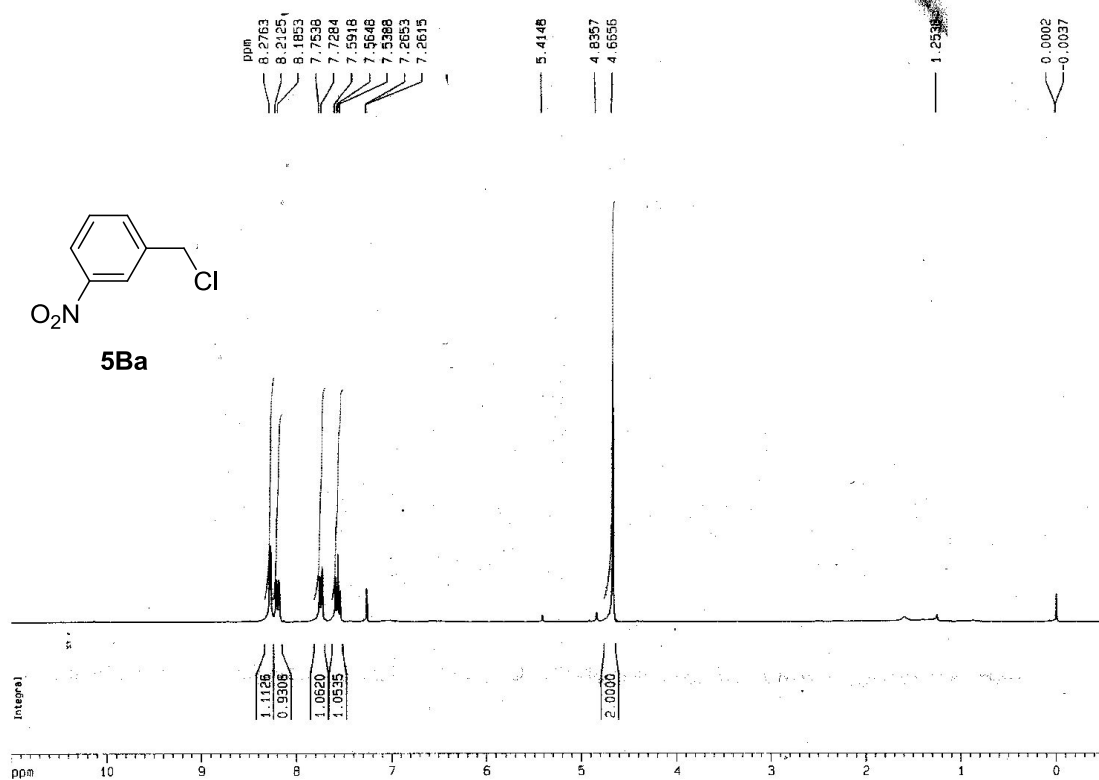


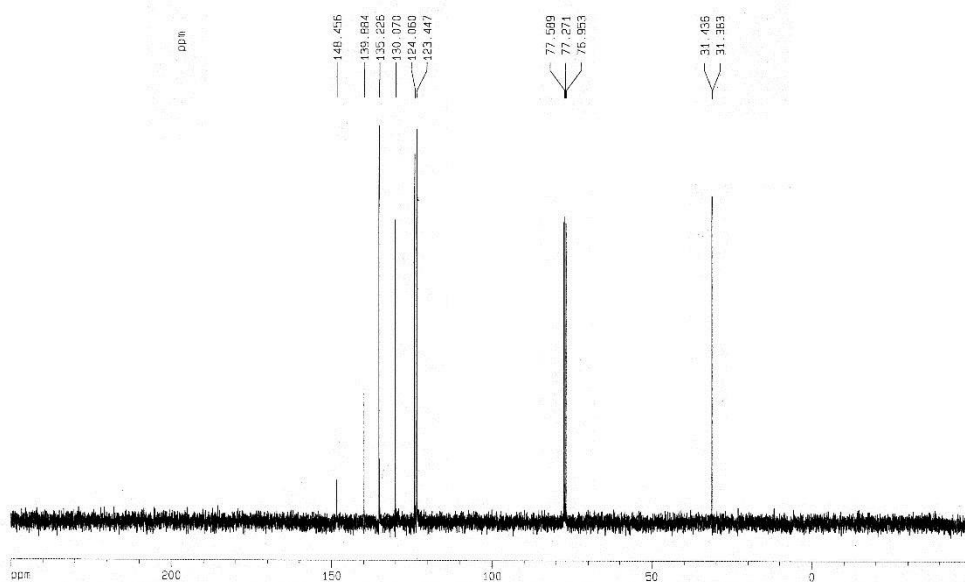
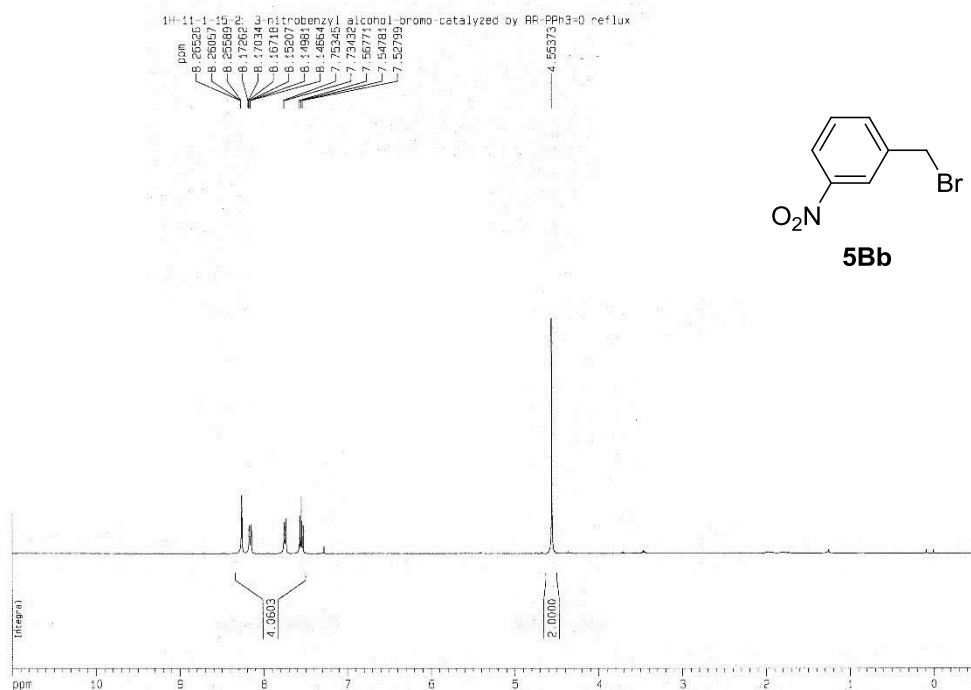


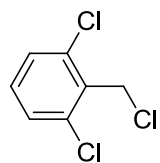
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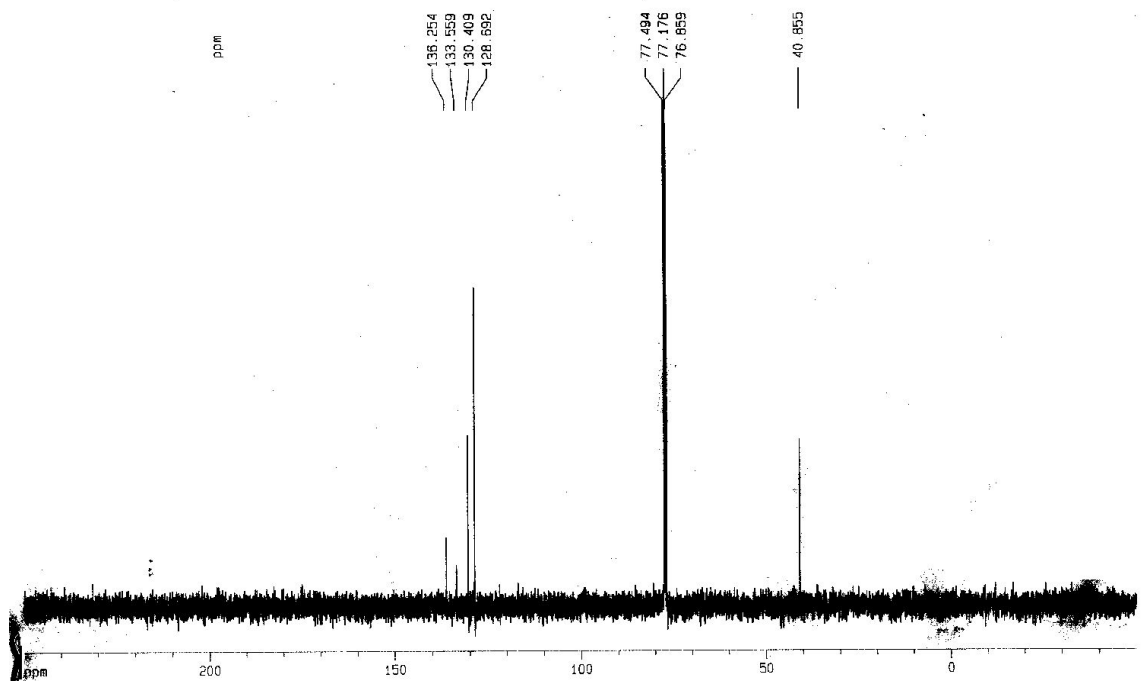
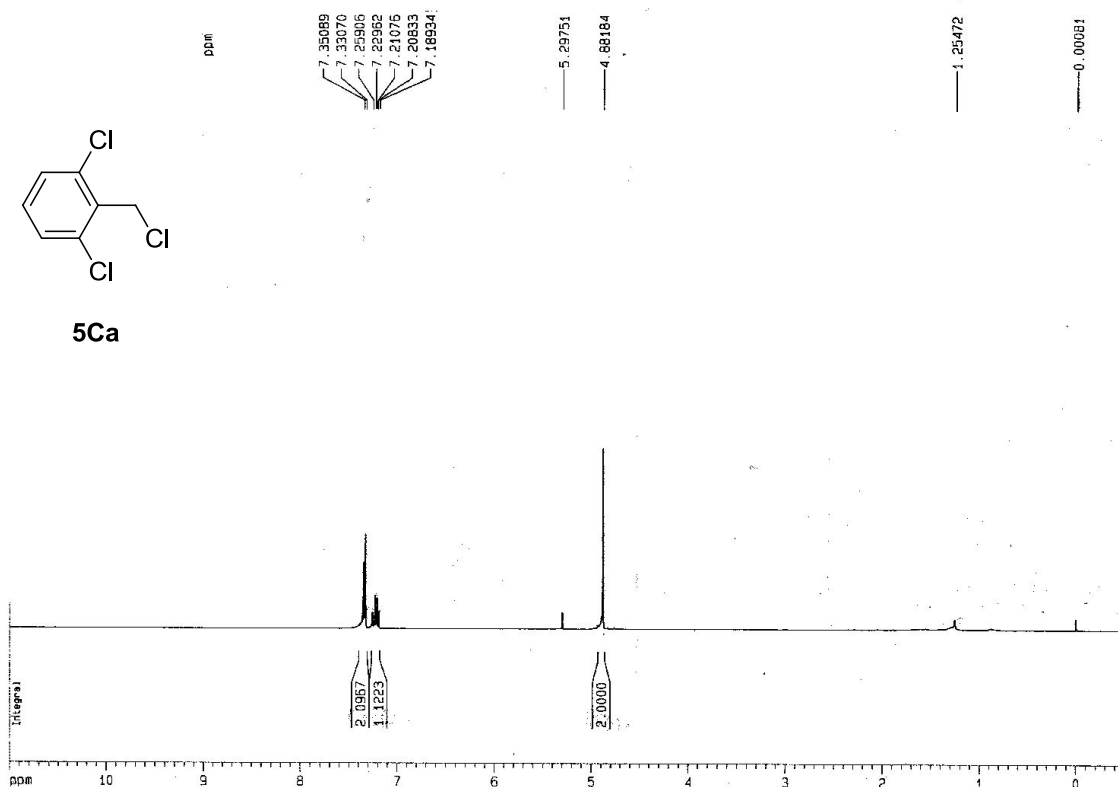


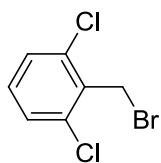




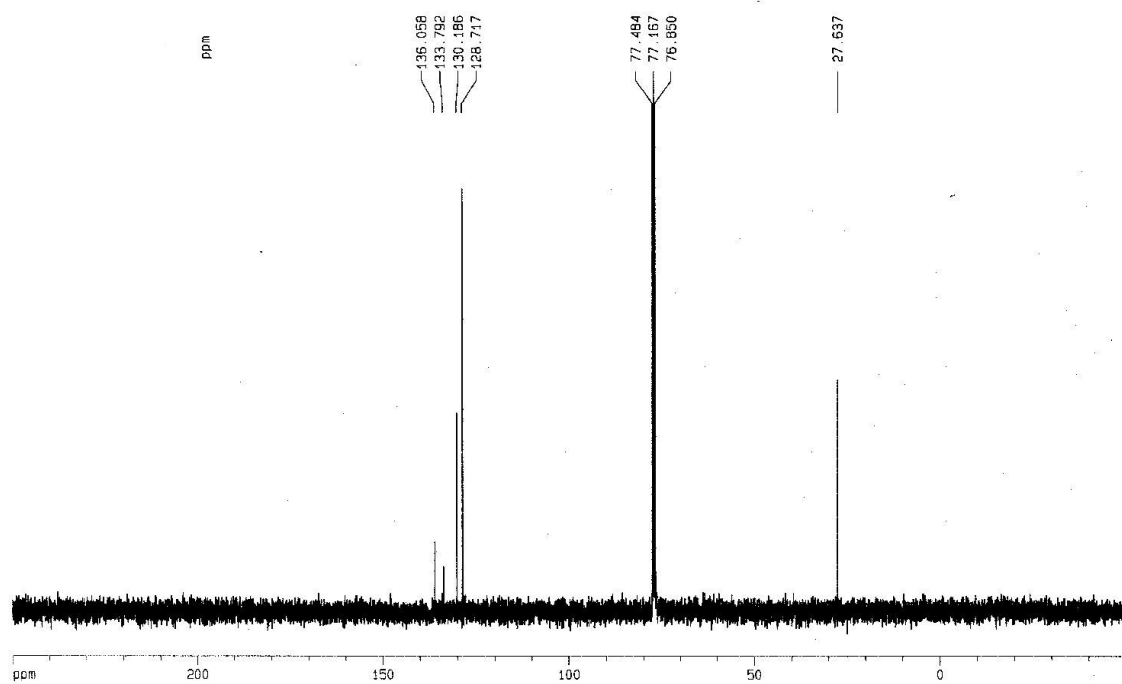
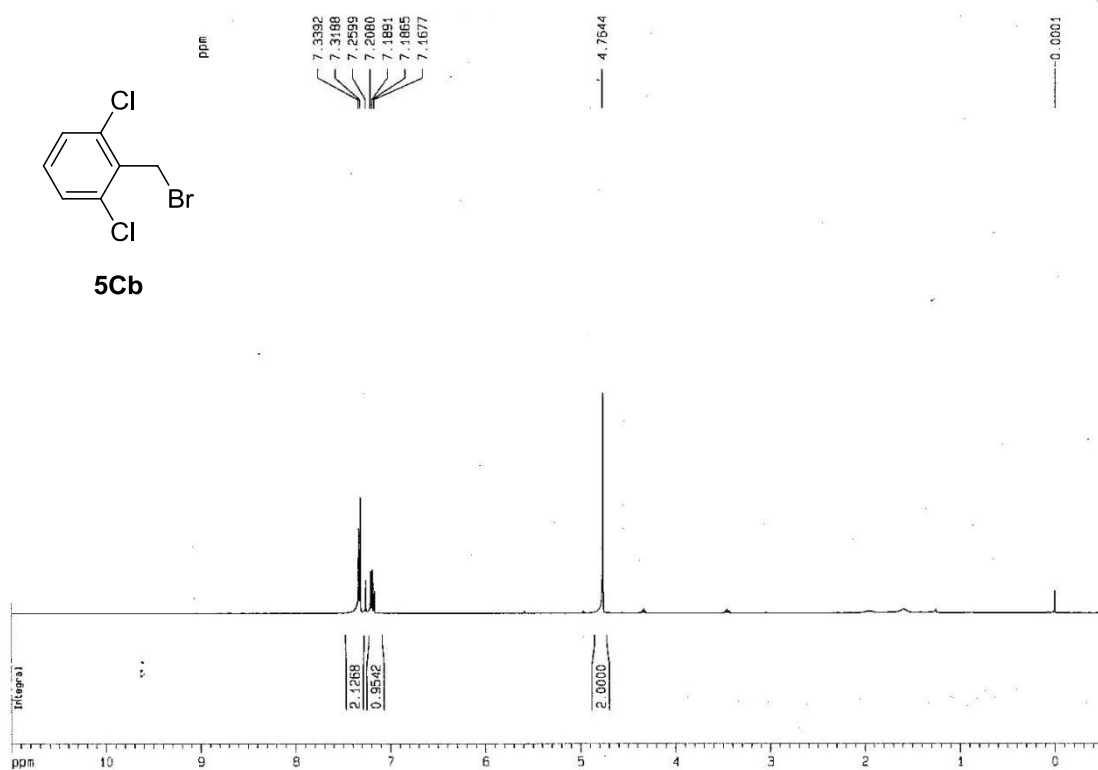


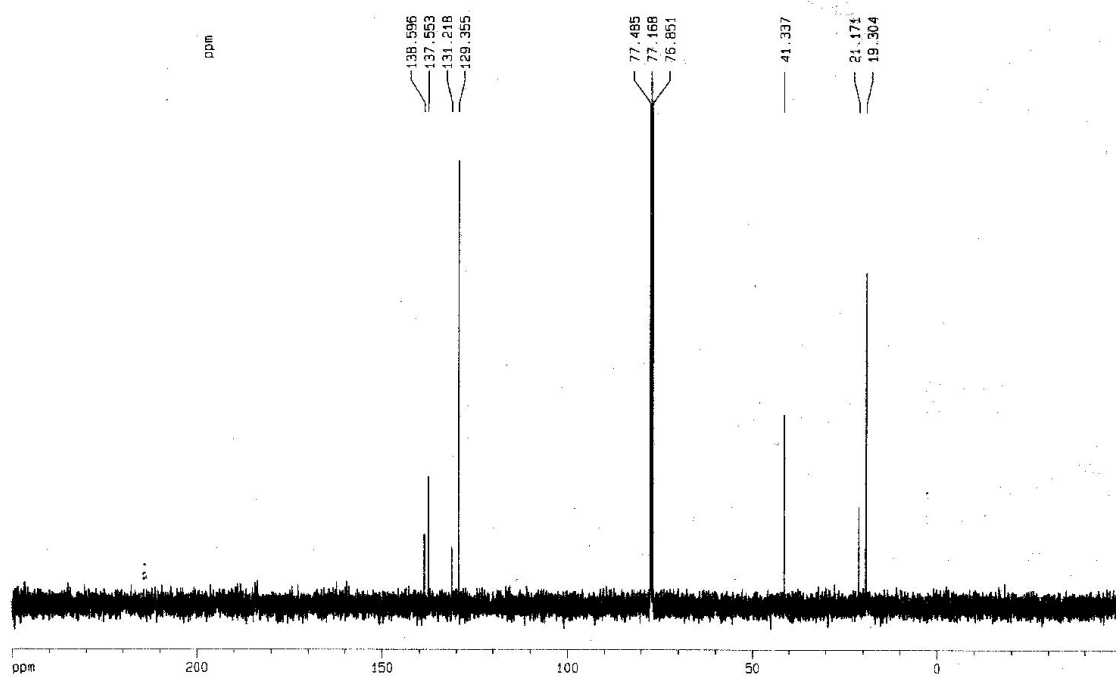
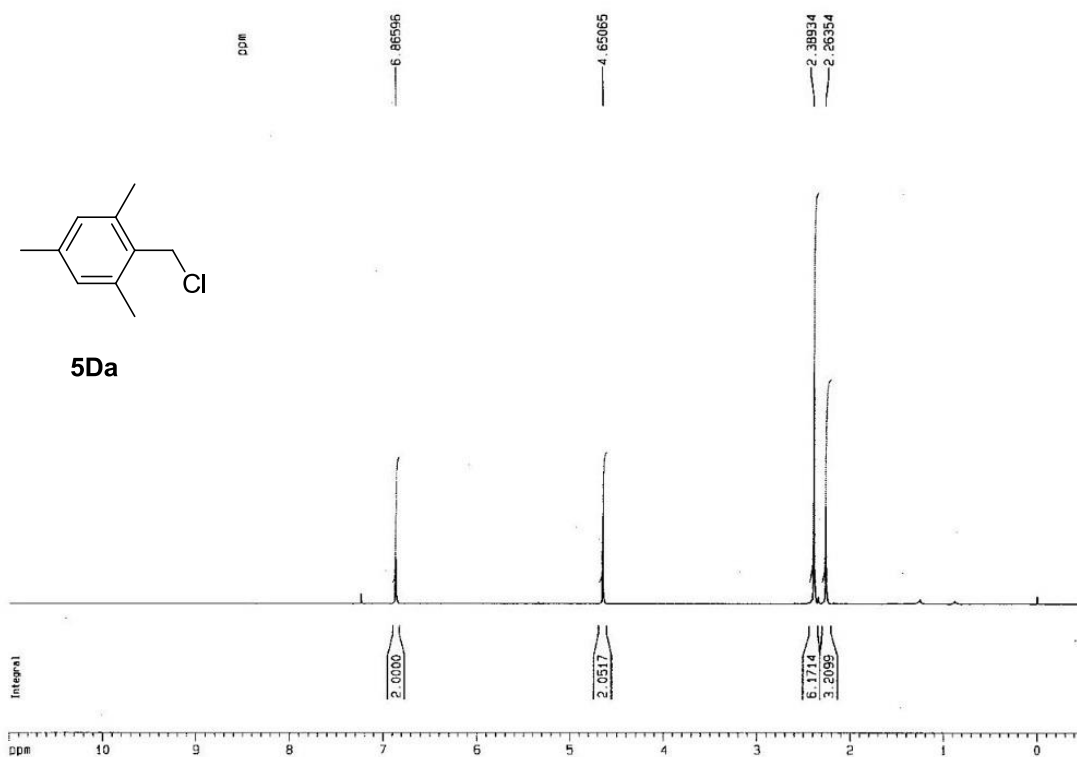
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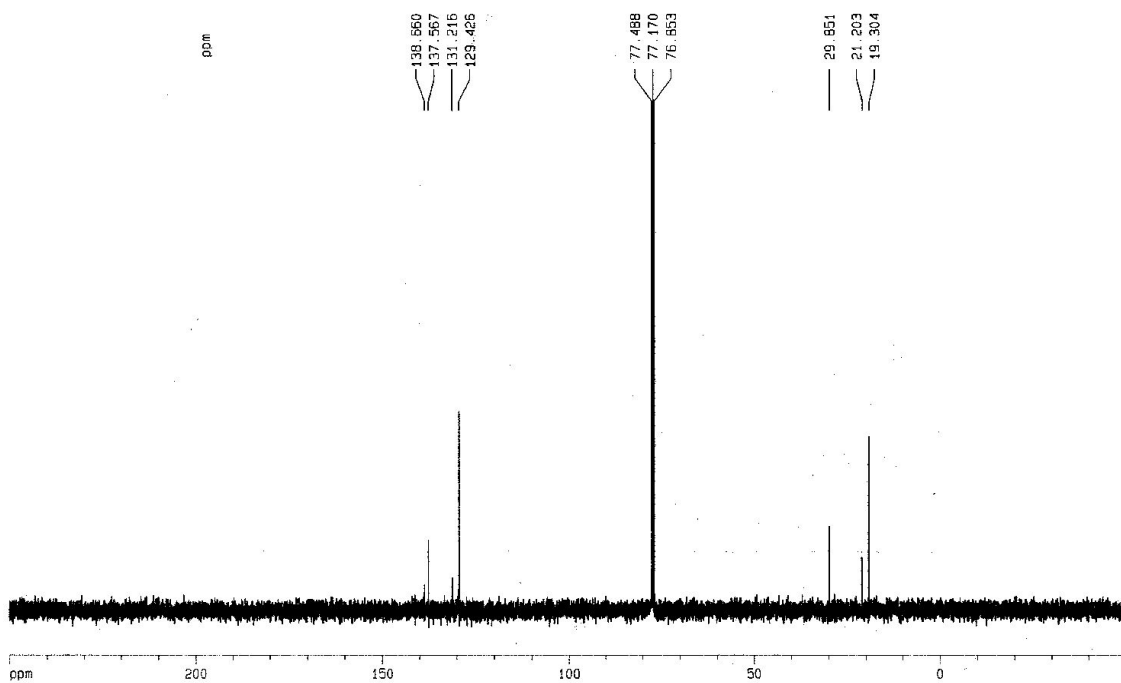
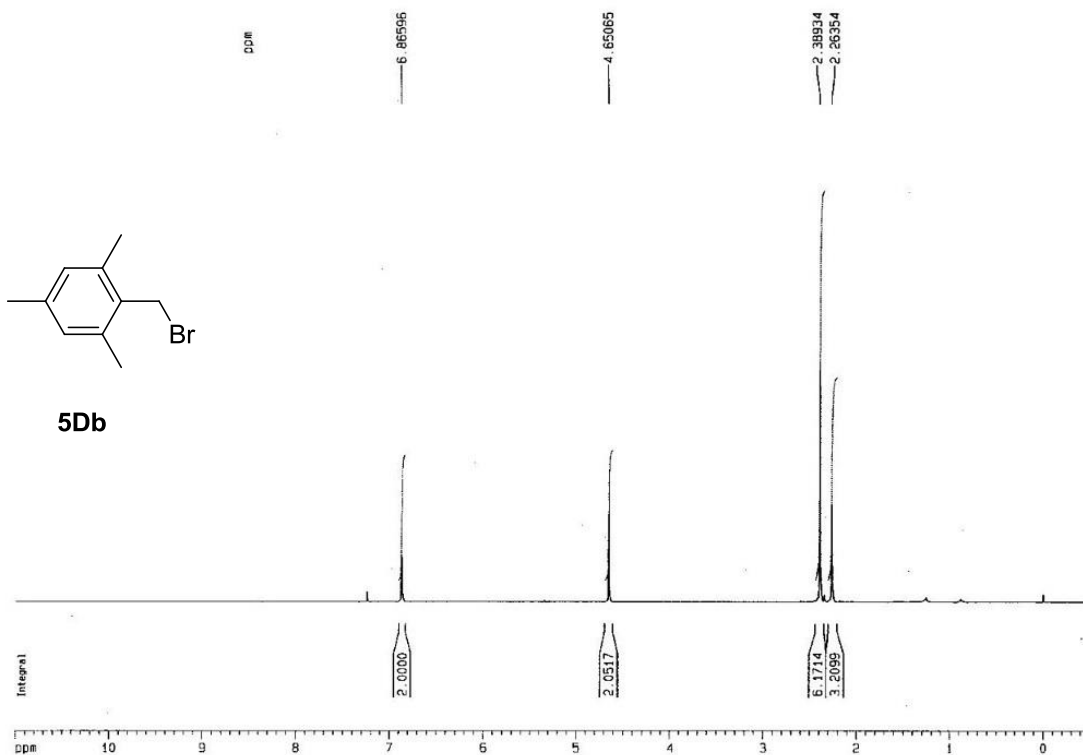


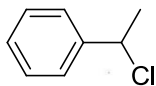


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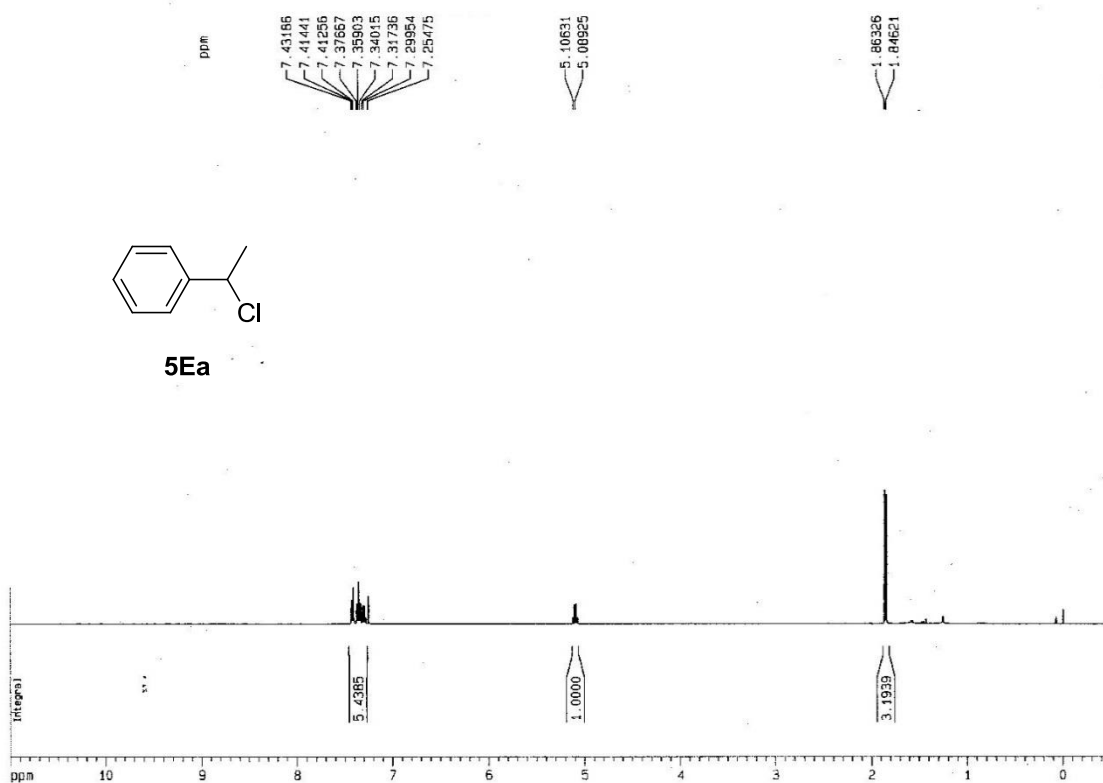






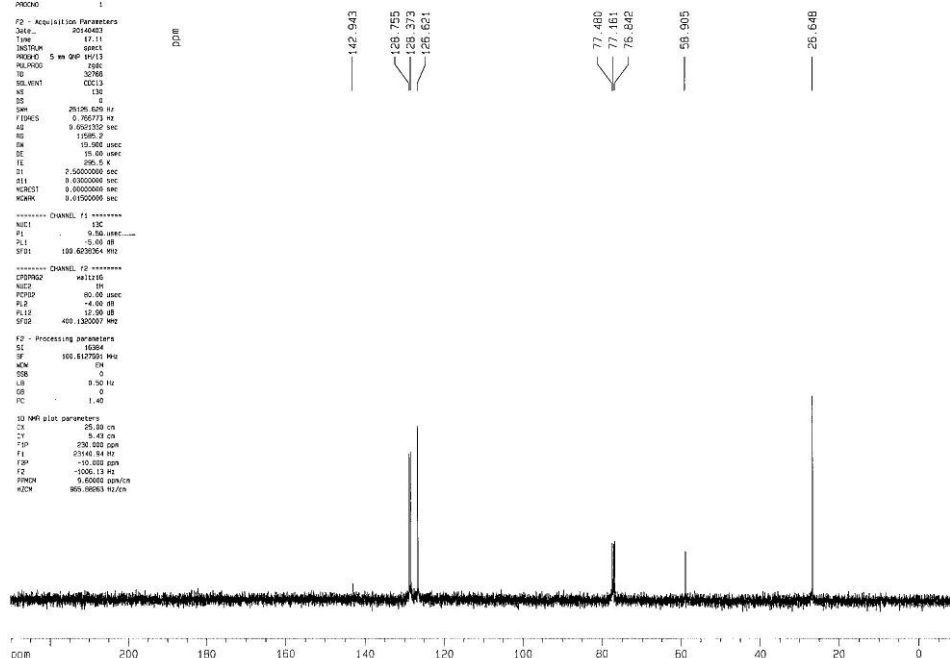


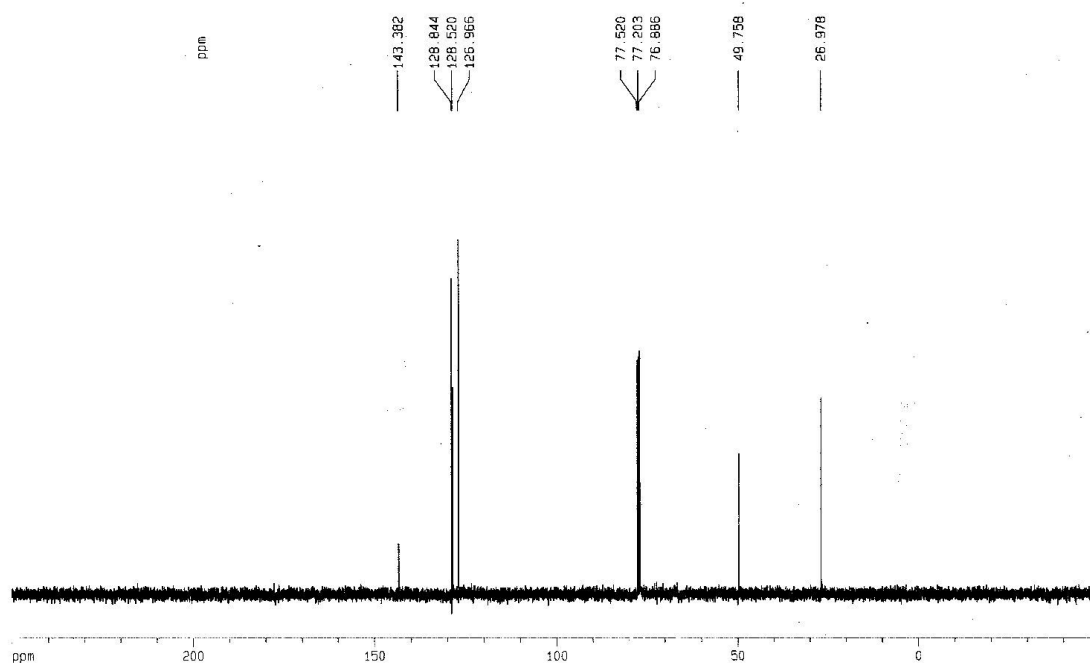
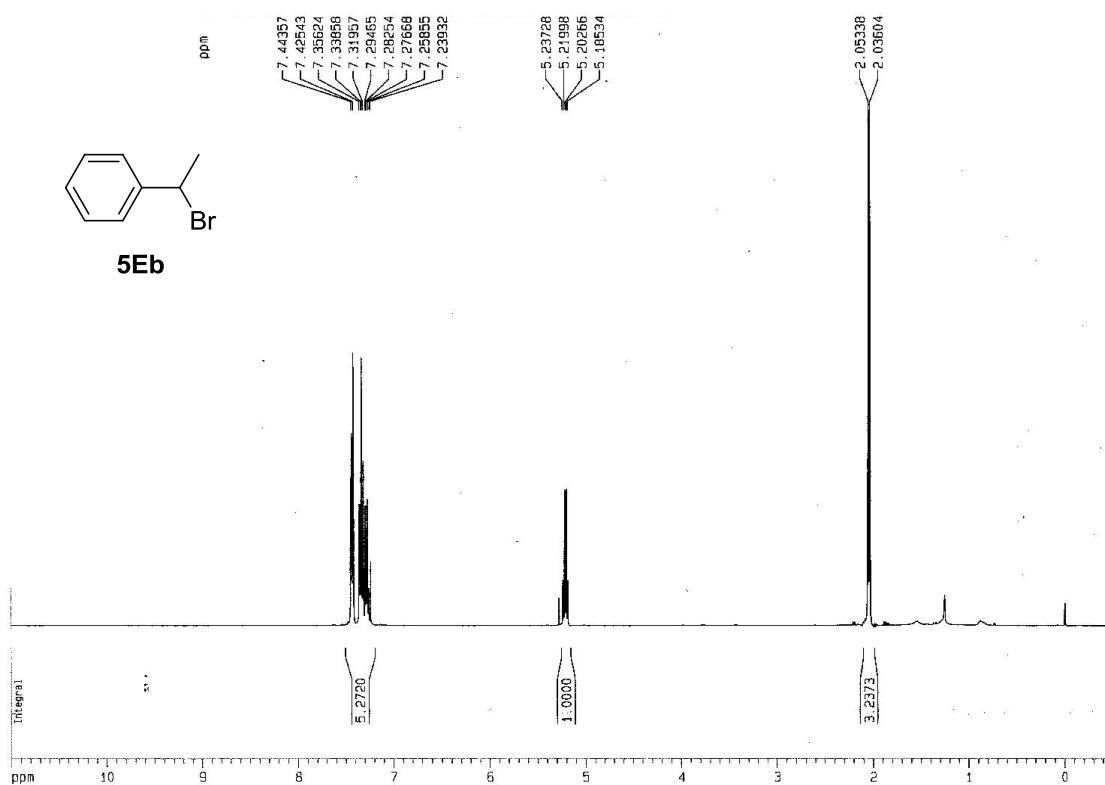
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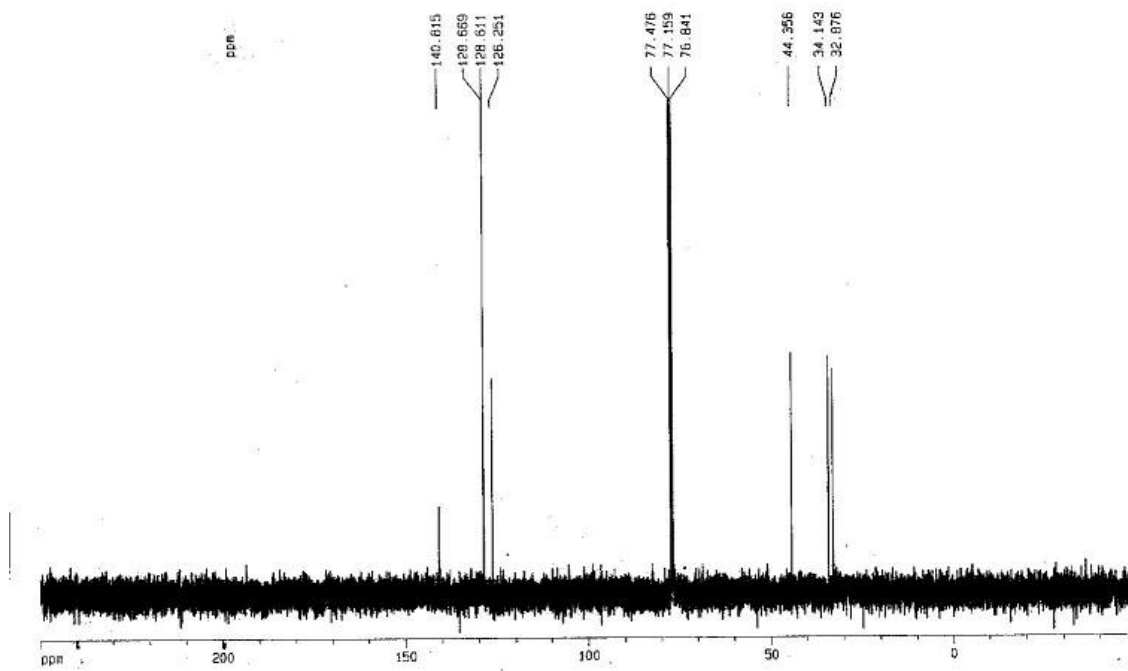
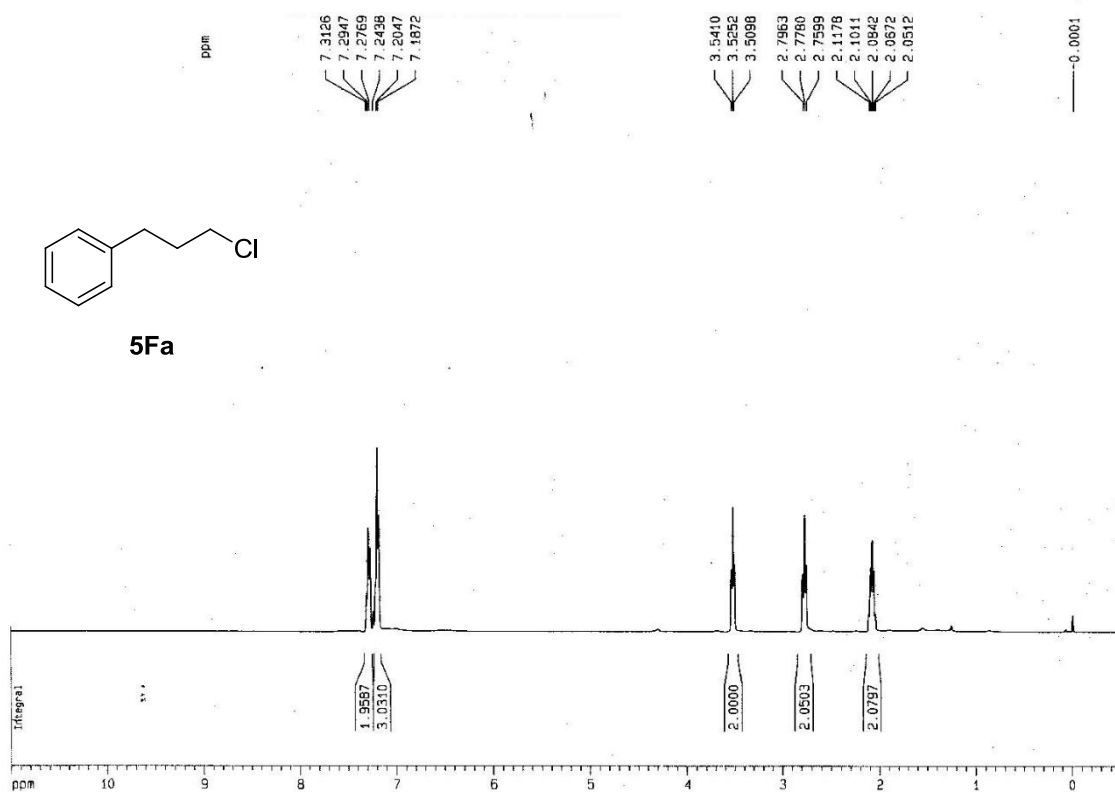


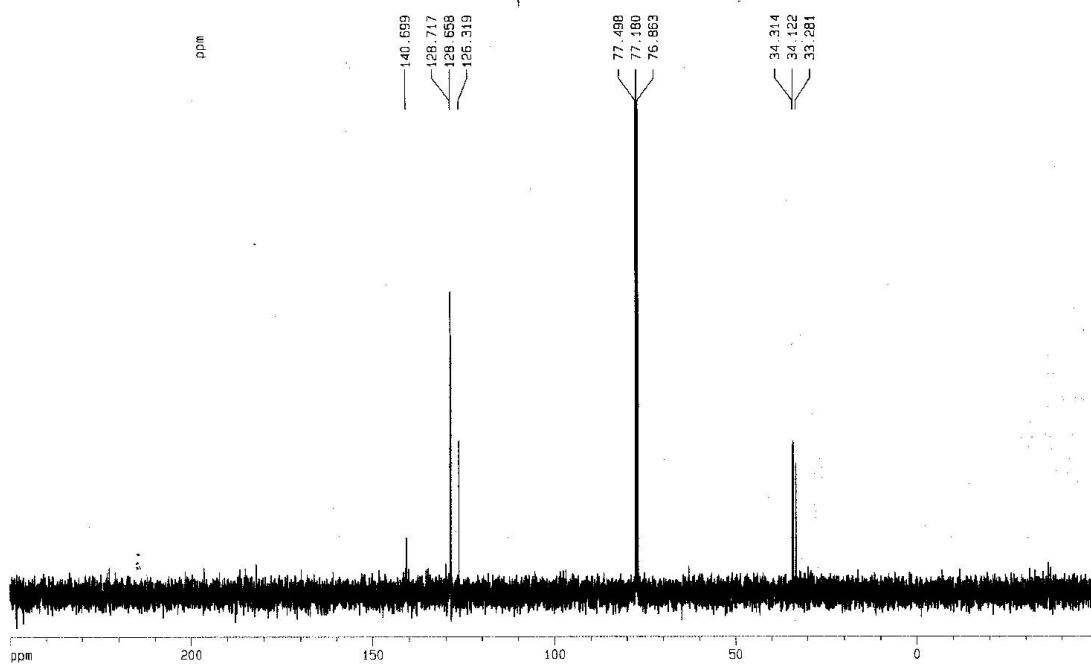
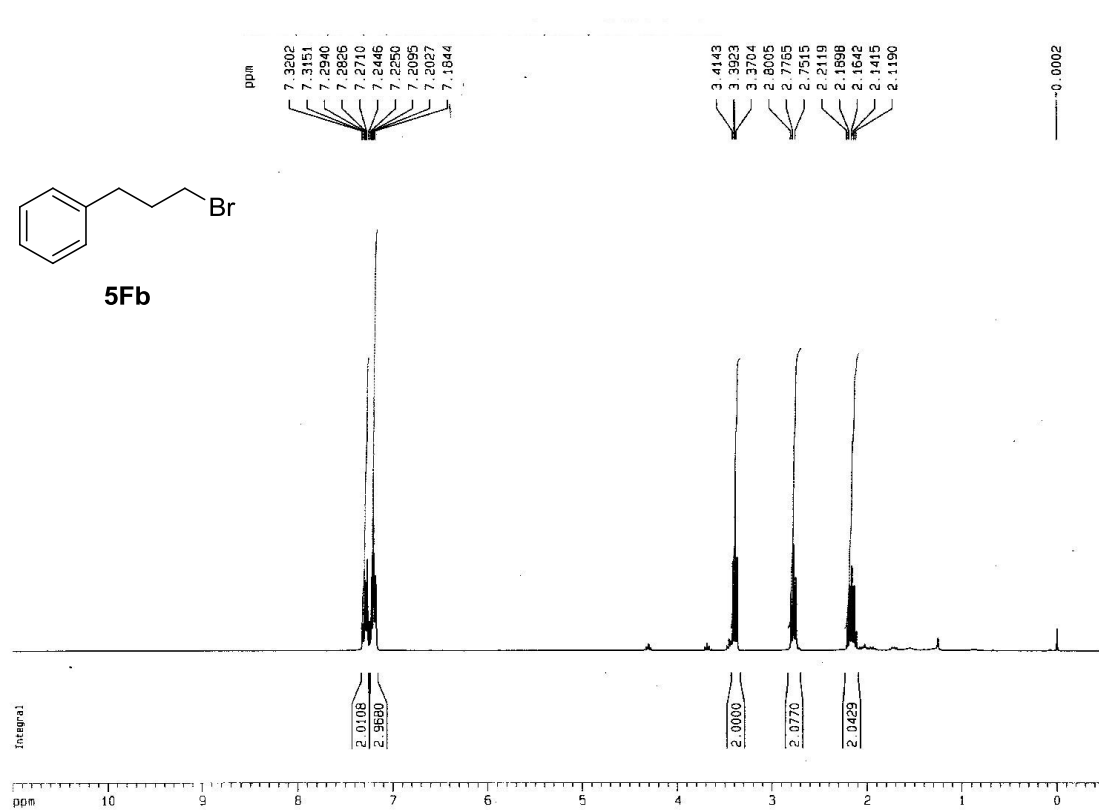
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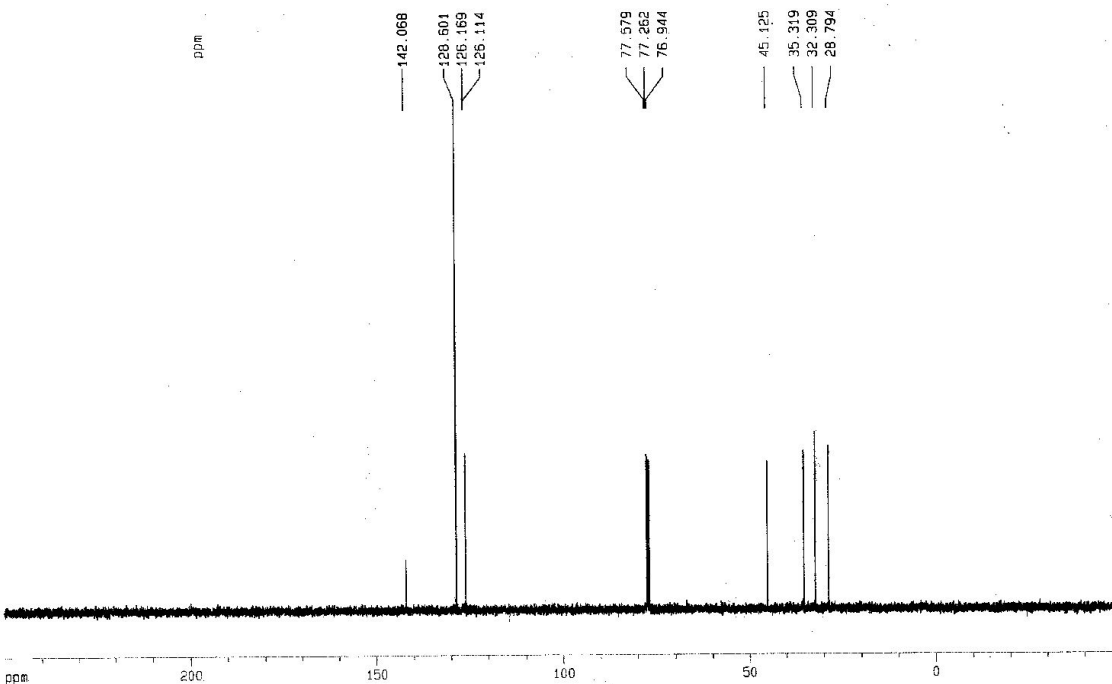
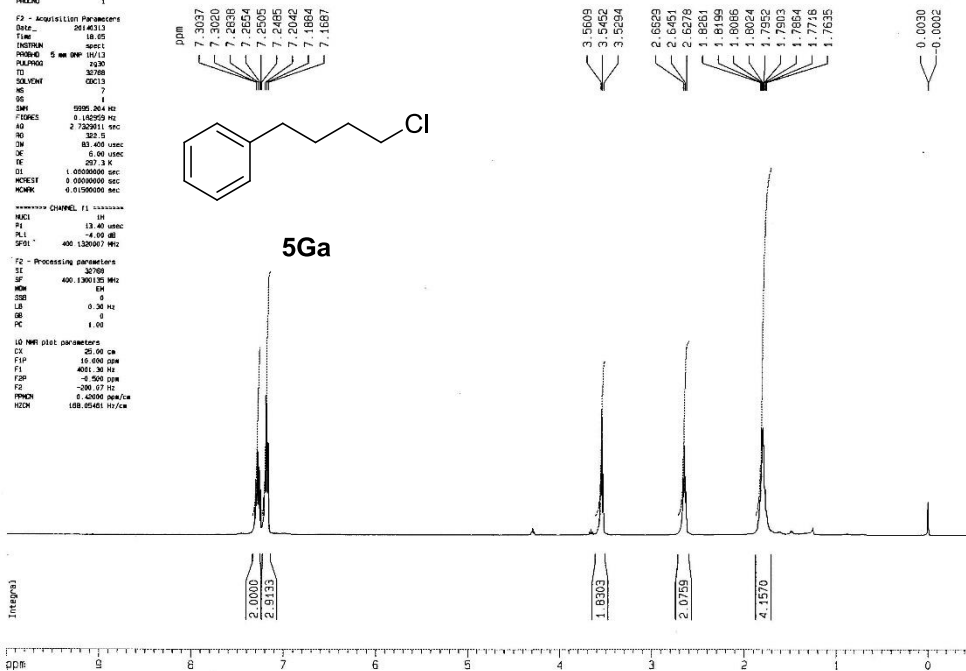
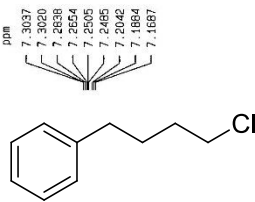


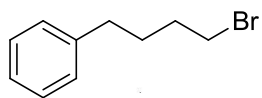




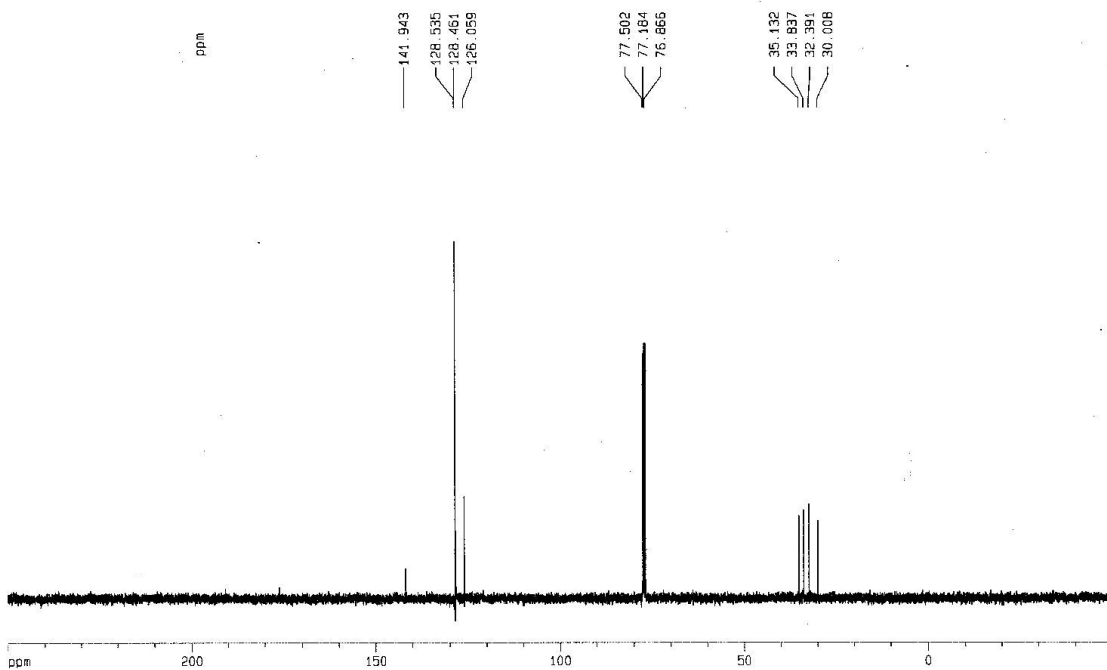
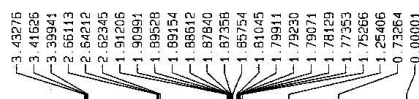
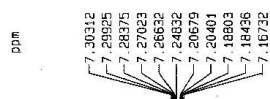
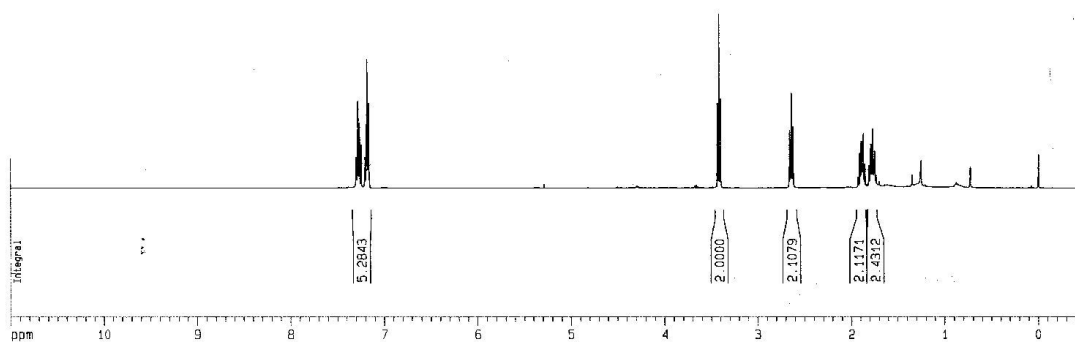


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5Gb



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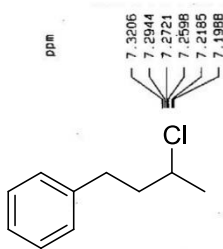
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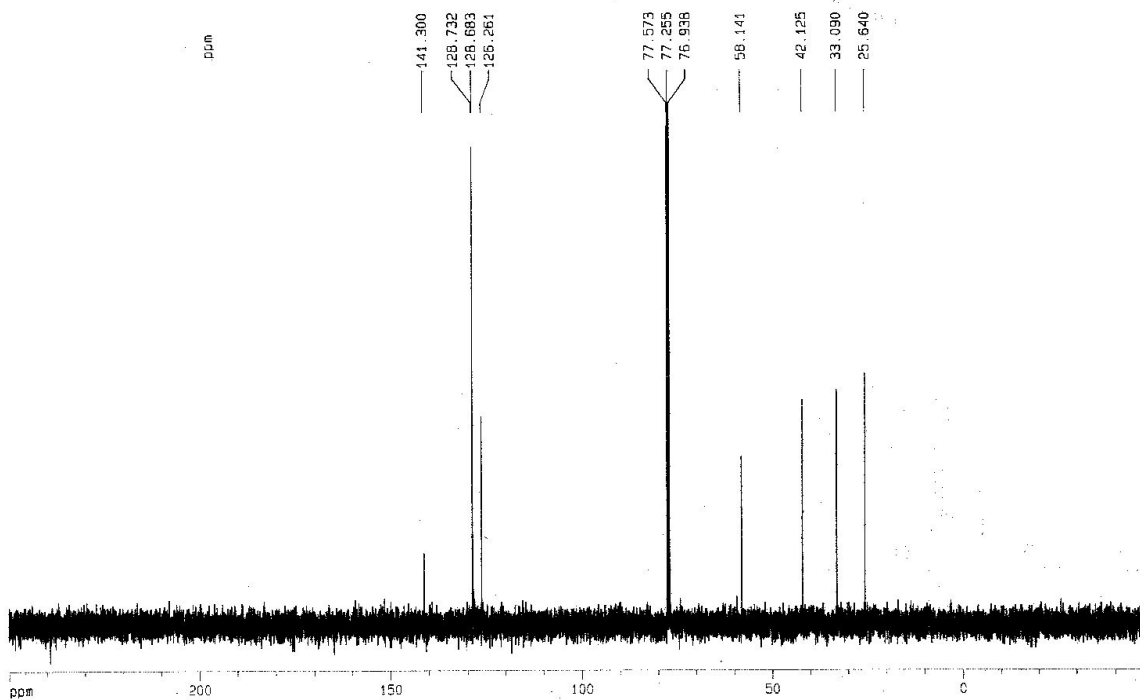
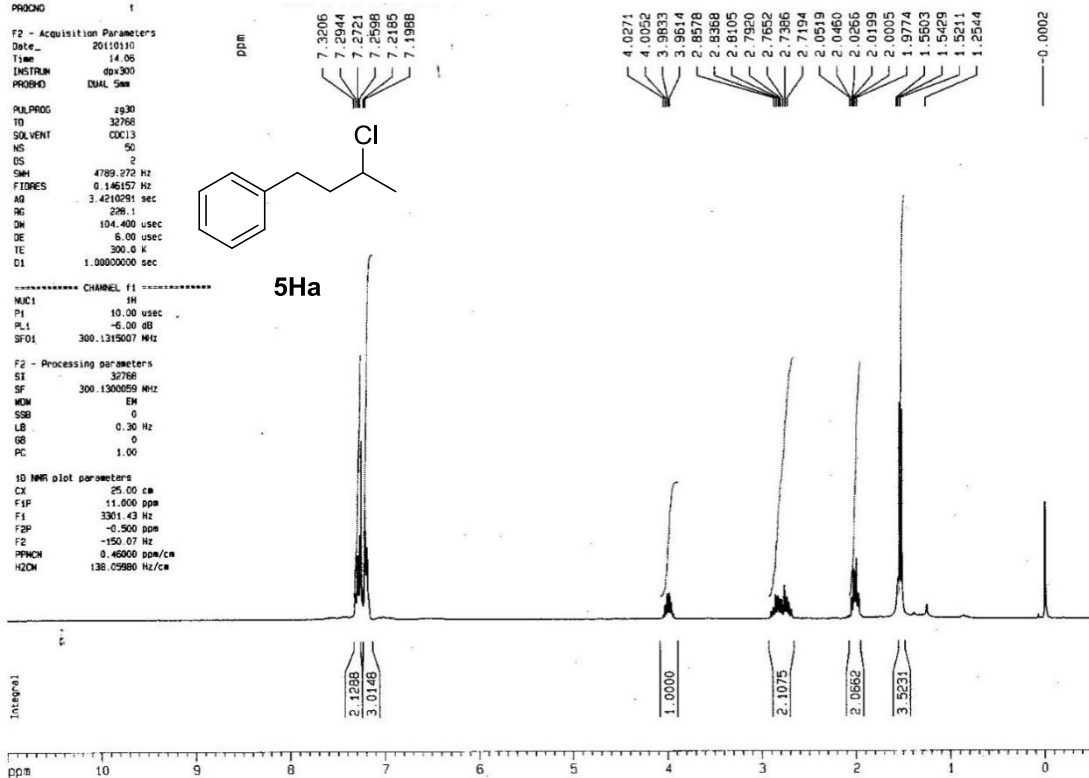
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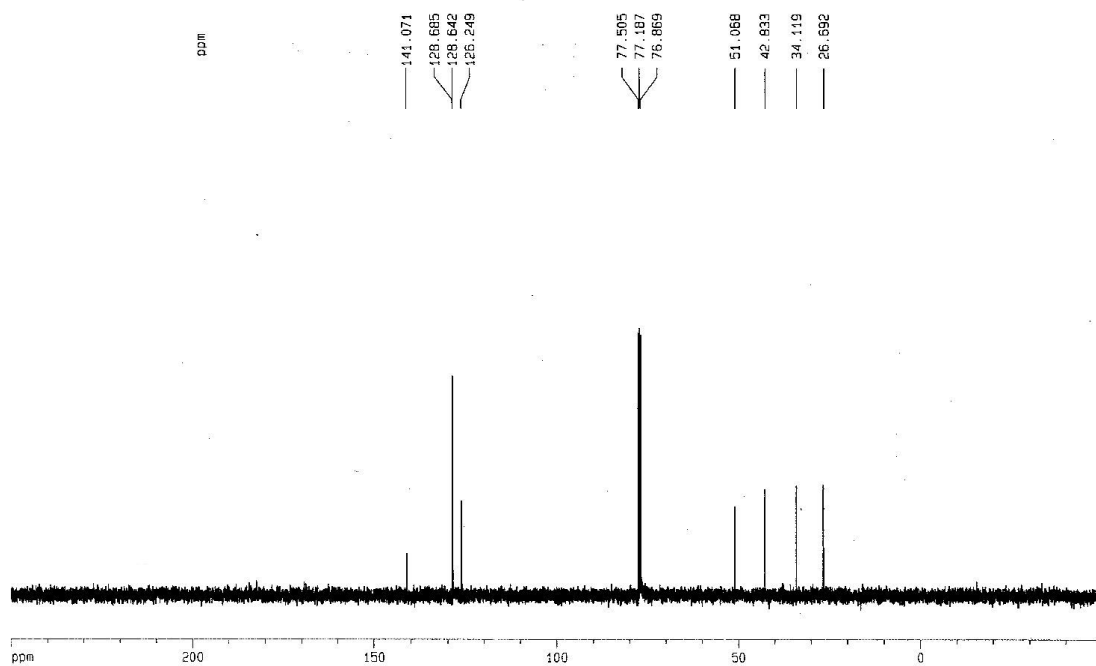
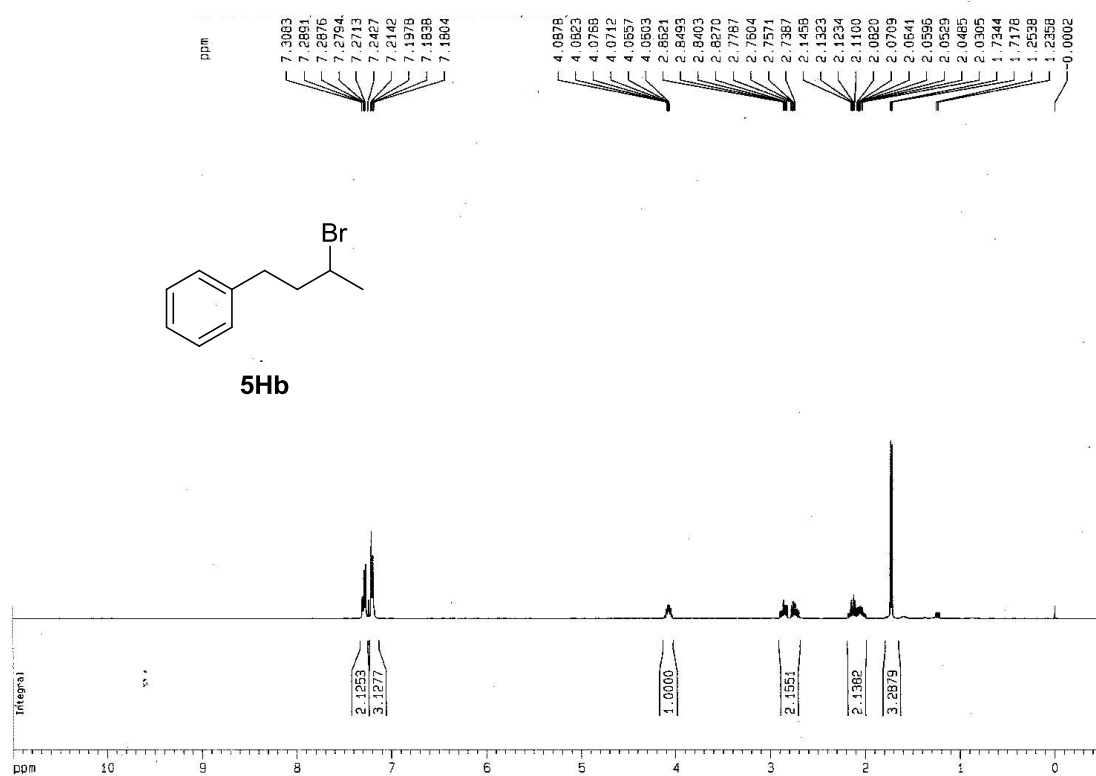
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F2 -150.07 Hz
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5Ha





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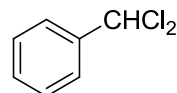
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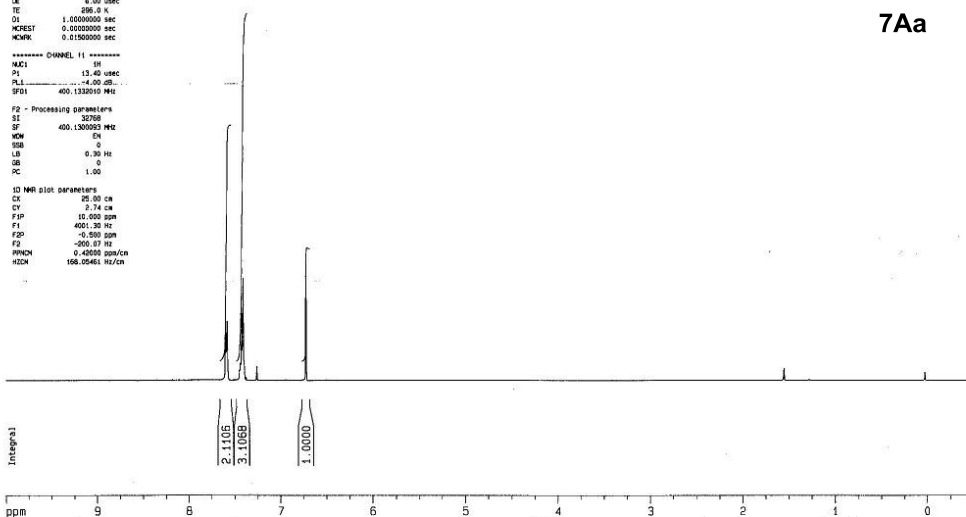
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H2ON 166.05461 Hz/cm

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7.59703
7.59214
7.58624
7.58134
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7.48915
6.72596



7Aa



Current Data Parameters
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PROCNO 1

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PL12 12.00 dB
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DDM

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130.046

128.890

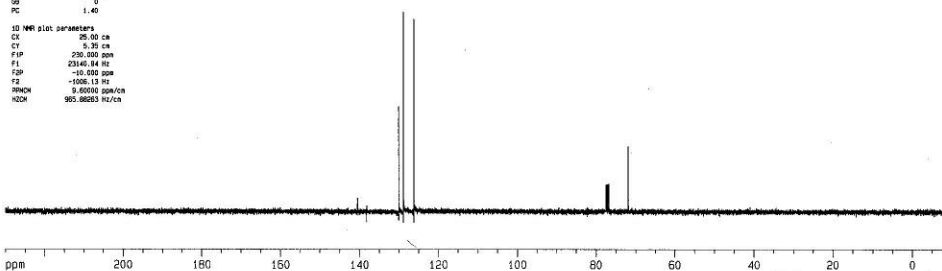
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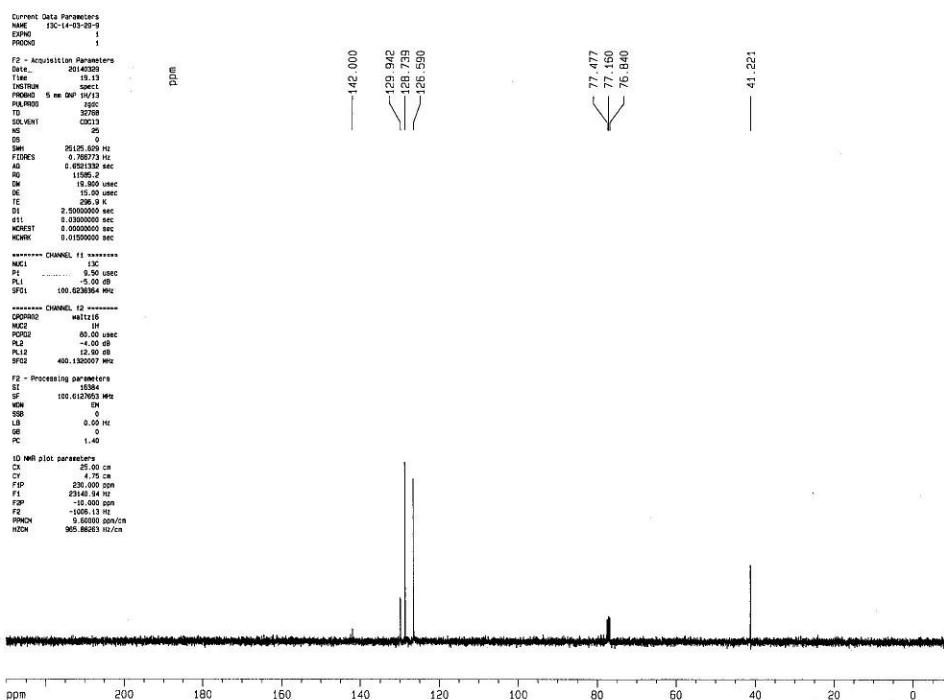
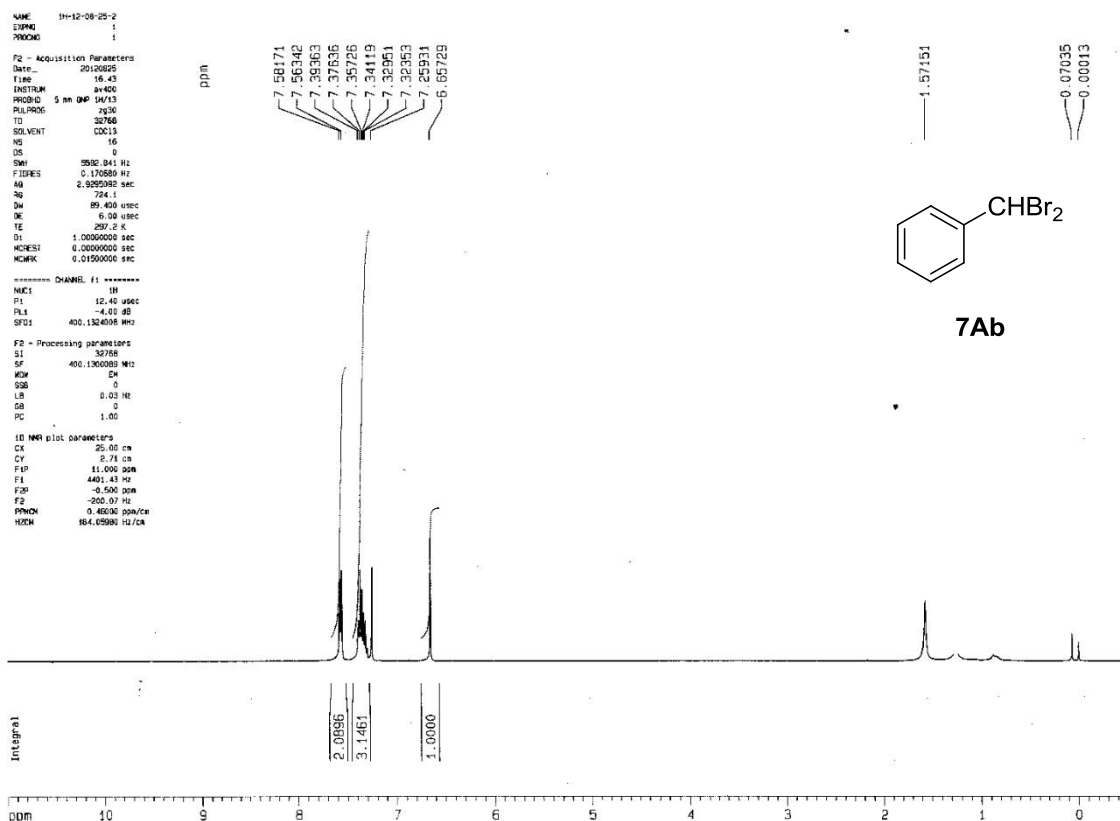
77.477

77.156

76.840

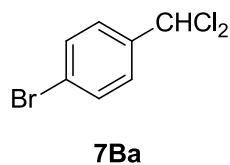
71.921





Current Data Parameters
 NAME 10-14-03-29-7
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20140329
 Time 18:54
 INSTRUM spect
 PROBRG 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 17
 DS 4
 SWH 7183.800 Hz
 FIDRES 0.215205 Hz
 AQ 2.2807025 sec
 RG 396
 DW 69.600 usec
 DE 6.00 usec
 TE 298.2 K
 IC 1.0000000 sec
 MCNEN 0.0000000 sec
 MCNEN 0.0100000 sec
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 13.40 usec
 PL1 -4.00 dB
 SFO1 400.132010 MHz
 F2 - Processing parameters
 SI 32768
 SF 400.132010 MHz
 MD 64
 SB 0
 LA 0.00 Hz
 GB 0
 PC 1.00
 IS NMR plot parameters
 CA 25.00 cm
 CY 3.69 cm
 FID 15.000 ppm
 FI 4001.30 Hz
 F2P -5.500 ppm
 F2 -260.87 Hz
 PPMH 6.45000 ppm/cx
 HZCX 166.01461 Hz/cx

ppm: 7.50041, 7.50813, 7.45363, 7.43240, 6.65994



Integral: 2.0596, 2.0761, 1.0000



Current Data Parameters
 NAME 13C-14-03-29-7
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20140329
 Time 18:56
 INSTRUM spect
 PROBRG 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 17
 DS 4
 SWH 25126.600 Hz
 FIDRES 0.768773 Hz
 AQ 0.662132 sec
 RG 11555.2
 DW 19.000 usec
 DE 15.00 usec
 TE 298.2 K
 IC 2.0000000 sec
 MCNEN 0.0300000 sec
 MCNEN 0.0000000 sec
 MCNEN 0.0100000 sec
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 19.50 usec
 PL1 0.00 dB
 SFO1 100.6261854 MHz
 ===== CHANNEL f2 =====
 NUC2 1H
 P2 86.00 usec
 PL2 -4.00 dB
 PL12 12.00 dB
 SFO2 400.1320007 MHz
 F2 - Processing parameters
 SI 32768
 SF 100.6261854 MHz
 MD 64
 SB 0
 LA 0.00 Hz
 GB 0
 PC 1.40
 IS NMR plot parameters
 CA 25.00 cm
 CY 9.39 cm
 FID 235.000 ppm
 FI 23140.94 Hz
 F2P -15.000 ppm
 F2 -1505.13 Hz
 PPMH 5.80000 ppm/cx
 HZCX 965.88067 Hz/cx

ppm: 139.485, 132.124, 127.516, 124.196, 77.468, 77.153, 76.837, 71.016



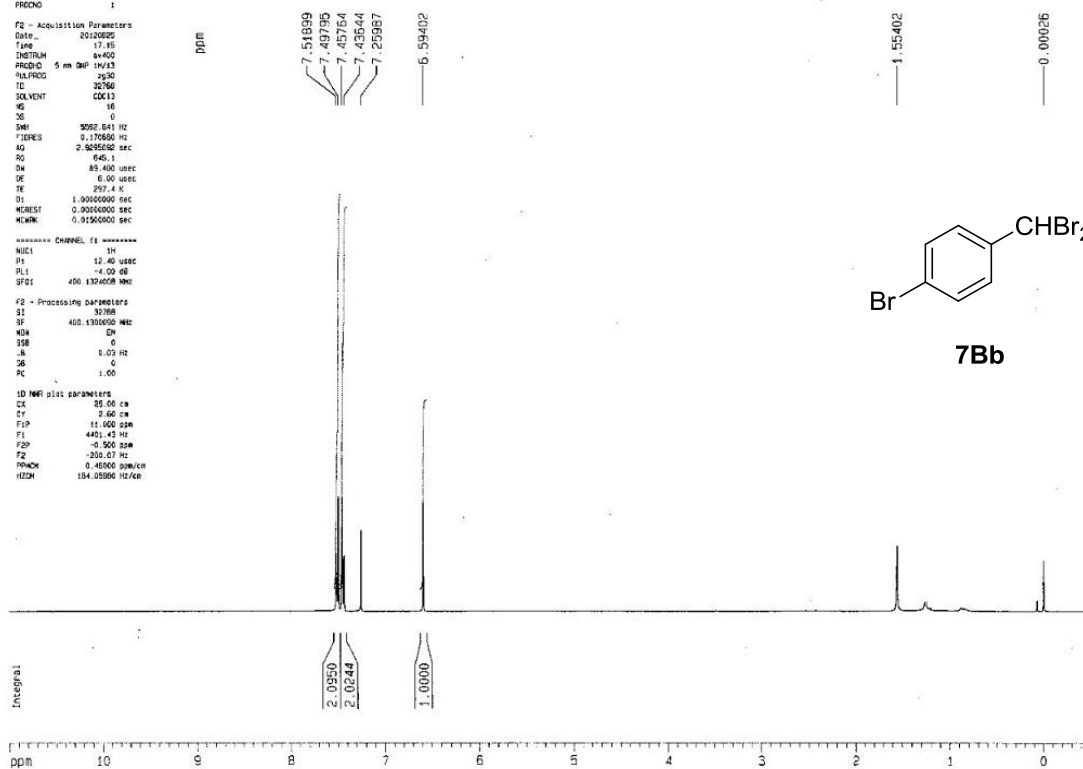
Current Data Parameters
 NAME 13C-12-08-25-3
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120825
 Time 17.15
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5062.541 Hz
 FIDRES 0.170856 Hz
 AQ 2.505556 sec
 RG 645.1
 DW 85.400 usec
 DE 6.00 usec
 TE 297.4 K
 D1 1.30000000 sec
 DECST 0.30000000 sec
 MCHNK 0.91360000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.40 usec
 PL1 -4.00 dB
 SFO1 400.1374000 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1374000 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 25.00 cm
 CY 2.00 cm
 FIDP 11.000 ppm
 F1 4401.43 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCH 0.40000000 Hz/cm
 HZCH 164.05800 Hz/cm



Current Data Parameters
 NAME 13C-12-08-25-3
 EXPNO 1
 PROCNO 1

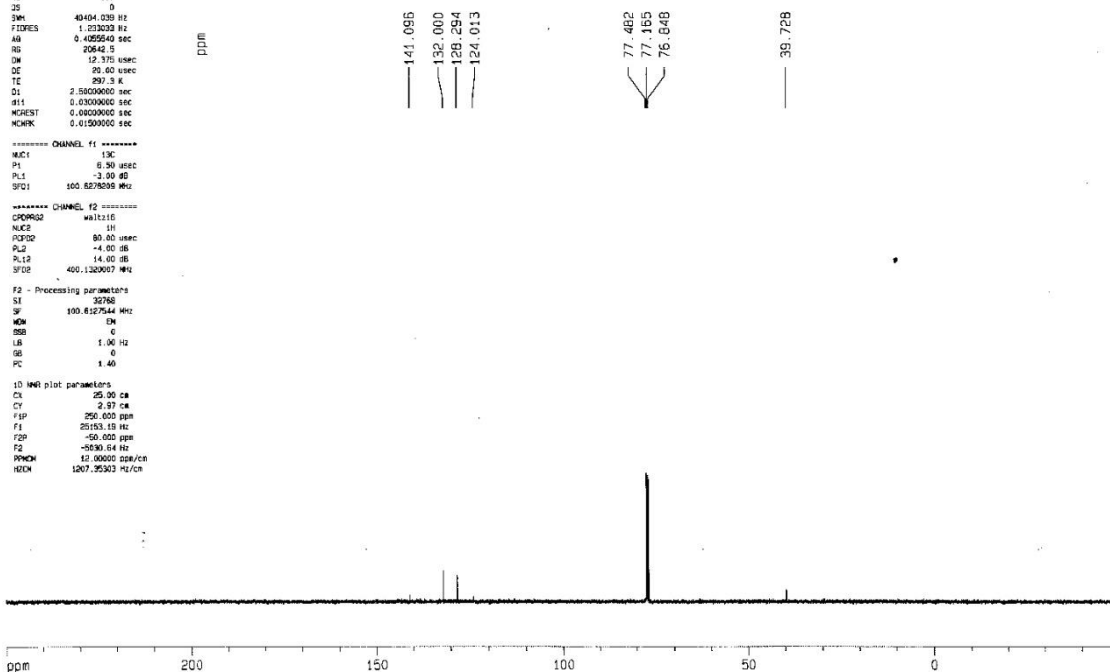
F2 - Acquisition Parameters
 Date_ 20120825
 Time 17.06
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 160
 DS 0
 SWH 49104.000 Hz
 FIDRES 1.233033 Hz
 AQ 0.4055640 sec
 RG 20642.5
 DW 12.370 usec
 DE 95.80 usec
 TE 297.3 K
 D1 2.50000000 sec
 D11 0.03000000 sec
 DECST 0.00000000 sec
 MCHNK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 6.30 usec
 PL1 -3.00 dB
 SFO1 100.6278399 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 P2P2 80.40 usec
 PL2 -4.00 dB
 PL12 14.00 dB
 SFO2 400.1320007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.627844 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 25.00 cm
 CY 2.07 cm
 FIDP 260.000 ppm
 F1 25153.19 Hz
 F2P -50.000 ppm
 F2 -50.9014 Hz
 PPMCH 12.00000 ppm/cm
 HZCH 1207.95303 Hz/cm



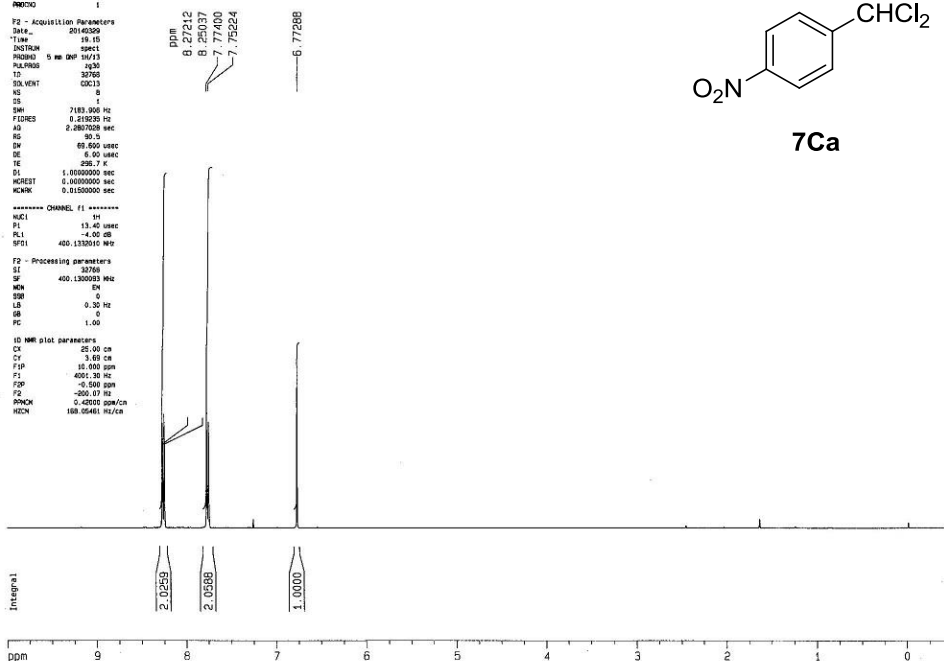
Current Data Parameters
NAME 10-14-03-29-10
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20140329
Time 19.15
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 8
DS 1
SWH 7189.800 Hz
FIDRES 0.218230 Hz
AQ 2.2867206 sec
RG 30.7
DM 69.650 usec
DE 6.90 usec
TE 299.7 K
D1 1.6000000 sec
MCREST 6.0000000 sec
MCNMR 0.0150000 sec

===== CHANNEL f1 =====
NUC1 13
P1 13.40 usec
PL1 -4.00 dB
SFO1 400.1320010 MHz

F2 - Processing parameters
SI 32768
SF 400.1320003 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CH 25.00 cm
CY 3.69 cm
FIDP 16.690 usec
F1 4001.30 Hz
F2 -6.500 usec
F3 -200.07 Hz
F4 0.0000000 sec
F5 0.0000000 sec
F6 0.0000000 sec
F7 0.0000000 sec
F8 0.0000000 sec
F9 0.0000000 sec
F10 0.0000000 sec
F11 0.0000000 sec
F12 0.0000000 sec
F13 0.0000000 sec
F14 0.0000000 sec
F15 0.0000000 sec
F16 0.0000000 sec
F17 0.0000000 sec
F18 0.0000000 sec
F19 0.0000000 sec
F20 0.0000000 sec
F21 0.0000000 sec
F22 0.0000000 sec
F23 0.0000000 sec
F24 0.0000000 sec
F25 0.0000000 sec
F26 0.0000000 sec
F27 0.0000000 sec
F28 0.0000000 sec
F29 0.0000000 sec
F30 0.0000000 sec
F31 0.0000000 sec
F32 0.0000000 sec
F33 0.0000000 sec
F34 0.0000000 sec
F35 0.0000000 sec
F36 0.0000000 sec
F37 0.0000000 sec
F38 0.0000000 sec
F39 0.0000000 sec
F40 0.0000000 sec
F41 0.0000000 sec
F42 0.0000000 sec
F43 0.0000000 sec
F44 0.0000000 sec
F45 0.0000000 sec
F46 0.0000000 sec
F47 0.0000000 sec
F48 0.0000000 sec
F49 0.0000000 sec
F50 0.0000000 sec
F51 0.0000000 sec
F52 0.0000000 sec
F53 0.0000000 sec
F54 0.0000000 sec
F55 0.0000000 sec
F56 0.0000000 sec
F57 0.0000000 sec
F58 0.0000000 sec
F59 0.0000000 sec
F60 0.0000000 sec
F61 0.0000000 sec
F62 0.0000000 sec
F63 0.0000000 sec
F64 0.0000000 sec
F65 0.0000000 sec
F66 0.0000000 sec
F67 0.0000000 sec
F68 0.0000000 sec
F69 0.0000000 sec
F70 0.0000000 sec
F71 0.0000000 sec
F72 0.0000000 sec
F73 0.0000000 sec
F74 0.0000000 sec
F75 0.0000000 sec
F76 0.0000000 sec
F77 0.0000000 sec
F78 0.0000000 sec
F79 0.0000000 sec
F80 0.0000000 sec
F81 0.0000000 sec
F82 0.0000000 sec
F83 0.0000000 sec
F84 0.0000000 sec
F85 0.0000000 sec
F86 0.0000000 sec
F87 0.0000000 sec
F88 0.0000000 sec
F89 0.0000000 sec
F90 0.0000000 sec
F91 0.0000000 sec
F92 0.0000000 sec
F93 0.0000000 sec
F94 0.0000000 sec
F95 0.0000000 sec
F96 0.0000000 sec
F97 0.0000000 sec
F98 0.0000000 sec
F99 0.0000000 sec
F100 0.0000000 sec



Current Data Parameters
NAME 10-14-03-29-10
EXPNO 1
PROCNO 1

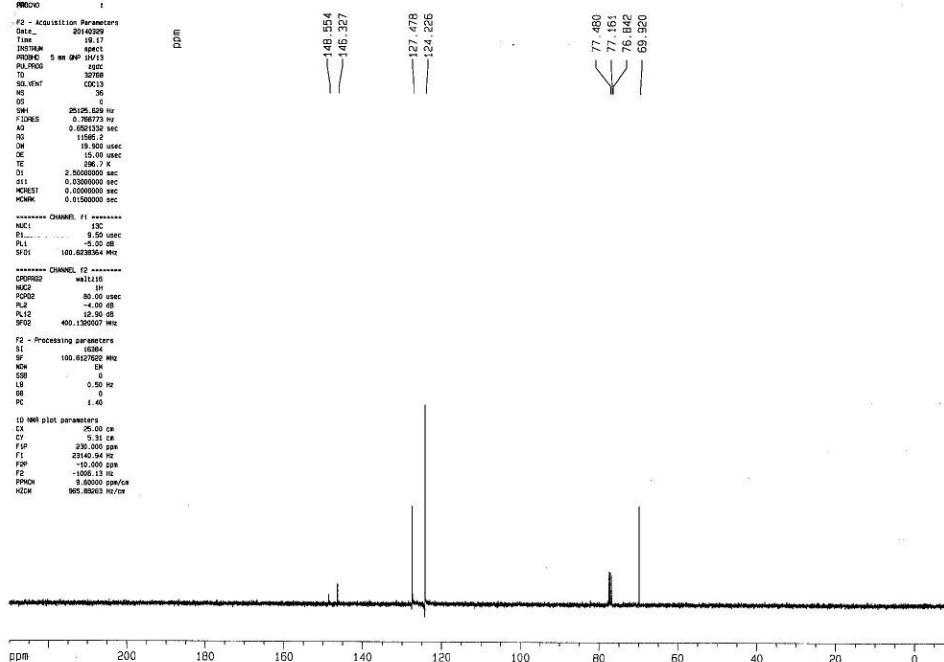
F2 - Acquisition Parameters
Date_ 20140329
Time 19.17
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 8
DS 1
SWH 25126.620 Hz
FIDRES 0.786773 Hz
AQ 0.0001330 sec
RG 11566.2
DM 19.960 usec
DE 15.00 usec
TE 299.7 K
D1 2.5000000 sec
MCREST 0.0000000 sec
MCNMR 0.0150000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -5.00 dB
SFO1 100.6261804 MHz

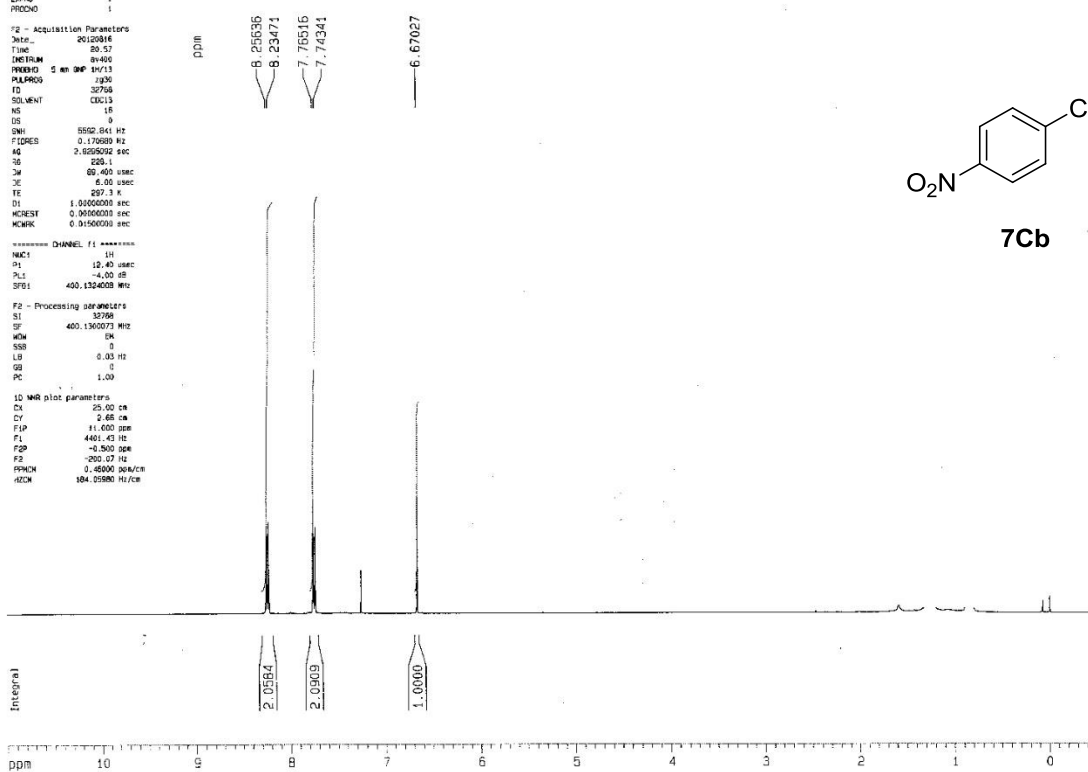
===== CHANNEL f2 =====
GPROG2 wa11116
NUC2 1H
P2P2 80.00 usec
PL2 -4.00 dB
PL12 12.50 dB
SFO2 400.1320007 MHz

F2 - Processing parameters
SI 32768
SF 100.6261804 MHz
WDW EM
SSB 0
LB 0.50 Hz
GB 0
PC 1.40

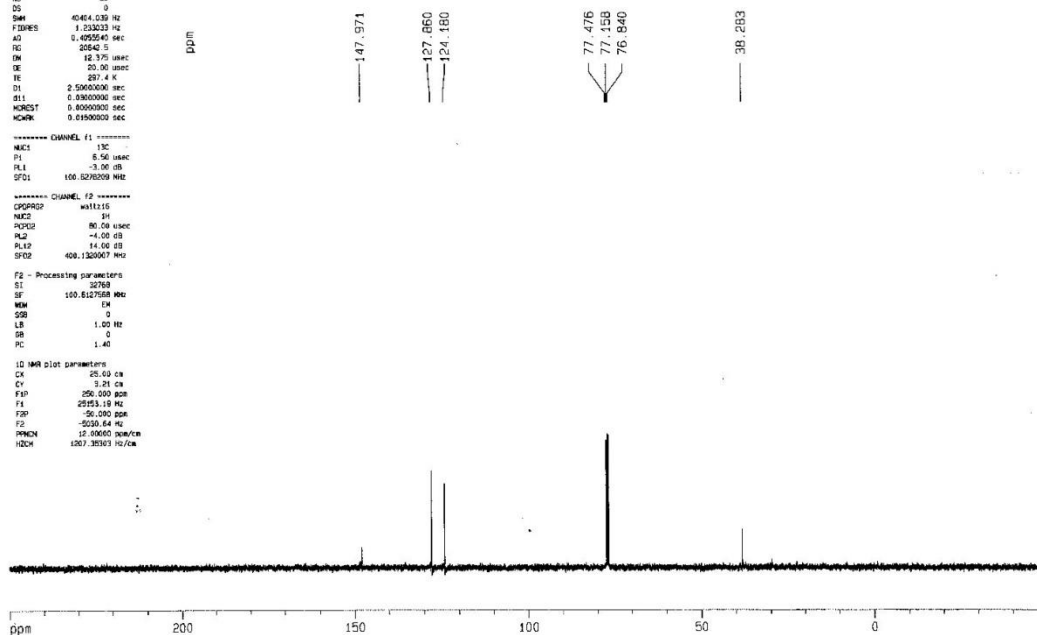
1D NMR plot parameters
CH 25.00 cm
CY 5.31 cm
FIDP 230.000 usec
F1 23140.84 Hz
F2 -155.000 usec
F3 -1006.13 Hz
F4 0.0000000 sec
F5 0.0000000 sec
F6 0.0000000 sec
F7 0.0000000 sec
F8 0.0000000 sec
F9 0.0000000 sec
F10 0.0000000 sec
F11 0.0000000 sec
F12 0.0000000 sec
F13 0.0000000 sec
F14 0.0000000 sec
F15 0.0000000 sec
F16 0.0000000 sec
F17 0.0000000 sec
F18 0.0000000 sec
F19 0.0000000 sec
F20 0.0000000 sec
F21 0.0000000 sec
F22 0.0000000 sec
F23 0.0000000 sec
F24 0.0000000 sec
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F26 0.0000000 sec
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F28 0.0000000 sec
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F50 0.0000000 sec
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F64 0.0000000 sec
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F66 0.0000000 sec
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F68 0.0000000 sec
F69 0.0000000 sec
F70 0.0000000 sec
F71 0.0000000 sec
F72 0.0000000 sec
F73 0.0000000 sec
F74 0.0000000 sec
F75 0.0000000 sec
F76 0.0000000 sec
F77 0.0000000 sec
F78 0.0000000 sec
F79 0.0000000 sec
F80 0.0000000 sec
F81 0.0000000 sec
F82 0.0000000 sec
F83 0.0000000 sec
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F88 0.0000000 sec
F89 0.0000000 sec
F90 0.0000000 sec
F91 0.0000000 sec
F92 0.0000000 sec
F93 0.0000000 sec
F94 0.0000000 sec
F95 0.0000000 sec
F96 0.0000000 sec
F97 0.0000000 sec
F98 0.0000000 sec
F99 0.0000000 sec
F100 0.0000000 sec

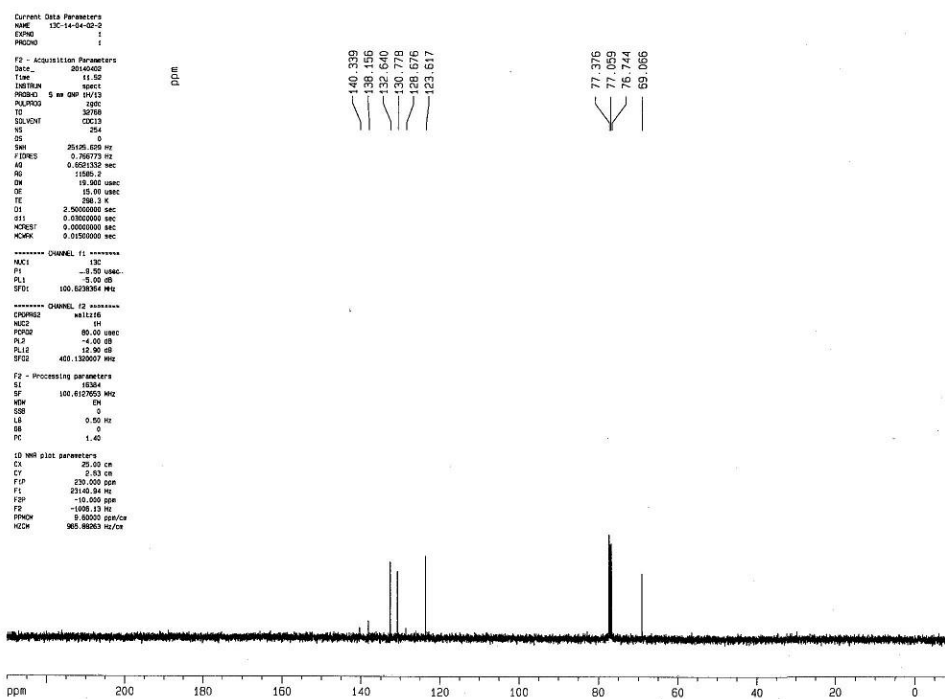
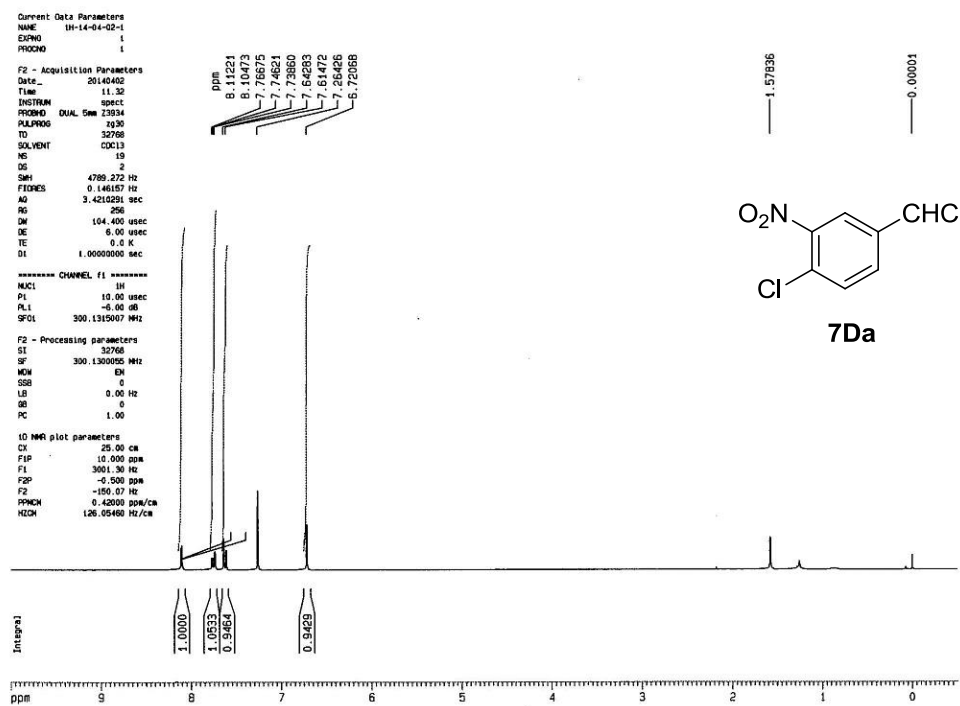


Current Data Parameters
 NAME 1H-12-08-15-1
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20120816
 TIME 06:57
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5502.841 Hz
 FIDRES 0.170889 Hz
 AQ 2.609000 sec
 RG 220.1
 JW 60.400 usec
 JE 0.00 usec
 TE 297.3 K
 DT 1.01000000 sec
 WDWT 0.30100000 sec
 WDECT 0.31500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 12.40 usec
 PL1 -4.00 dB
 SFO1 400.1324000 MHz
 F2 - Processing parameters
 SI 32768
 SF 400.1324000 MHz
 WDM 0.000000
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00
 ID - NMR plot parameters
 CX 25.00 cm
 CY 2.65 cm
 FID 11.000 usec
 FI 4461.41 Hz
 F2P -0.500 ppm
 F2 -0.000 Hz
 FWHM 0.45000 ppm/cm
 AQCM 104.05580 Hz/cm



Current Data Parameters
 NAME 13C-12-08-15-1
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20120816
 TIME 07:41
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 40484.130 Hz
 FIDRES 1.233033 Hz
 AQ 0.492540 sec
 RG 204.0
 JW 12.375 usec
 JE 0.00 usec
 TE 297.3 K
 DT 2.50000000 sec
 WDWT 0.30100000 sec
 WDECT 0.31500000 sec
 WDECT 0.31500000 sec
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -3.00 dB
 SFO1 100.6250000 MHz
 ===== CHANNEL f2 =====
 NUC2 1H
 P2 12.40 usec
 PL2 -4.00 dB
 SFO2 400.1324000 MHz
 F2 - Processing parameters
 SI 32768
 SF 100.6250000 MHz
 WDM 0.000000
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.40
 ID - NMR plot parameters
 CX 25.00 cm
 CY 2.65 cm
 FID 11.000 usec
 FI 25000.00 Hz
 F2P -0.500 ppm
 F2 -0.000 Hz
 FWHM 0.45000 ppm/cm
 AQCM 107.35363 Hz/cm





NAME 13C-12-08-02-1
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120802
Time 13.12
INSTRUM av400
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 16
DS 4
SWH 5592.841 Hz
FIDRES 0.170500 Hz
AQ 2.9266000 sec
RG 329.5
OW 89.400 usec
DE 6.00 usec
TE 297.2 K
D1 1.00000000 sec
NOEST 0.00000000 sec
NOHRC 0.01000000 sec

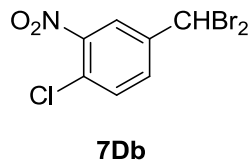
===== CHANNEL f1 =====
NUC1 13C
P1 19.40 usec
PL1 -4.00 dB
SFO1 400.1324000 MHz

F2 - Processing parameters
SI 32768
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.03 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 25.00 cm
CY 3.00 cm
FIP 11.000 ppm
F1 4401.43 Hz
F2 -50.000 MHz
F2 -200.37 Hz
PRNCH 0.45000 ppm/cm
H2CH 184.00000 Hz/cm

ppm

8.03372
8.08954
7.76951
7.75313
7.74754
7.74209
7.60397
7.59287
7.26722
6.62509



Integrat

1.0000
1.0493
1.0282
1.0200

ppm 10 9 8 7 6 5 4 3 2 1 0

===== CHANNEL f1 =====
NAME 13C-12-08-02-1
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120802
Time 13.12
INSTRUM av400
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 16
DS 4
SWH 40464.0750 Hz
FIDRES 1.230000 Hz
AQ 0.4009000 sec
RG 268.40
OW 12.375 usec
DE 20.00 usec
TE 297.2 K
D1 2.50000000 sec
D11 0.00000000 sec
NOEST 0.00000000 sec
NOHRC 0.01000000 sec

===== CHANNEL f2 =====
NAME 13C
P1 6.50 usec
PL1 -3.00 dB
SFO1 100.6270000 MHz

===== CHANNEL f2 =====
NAME 13C
P1 6.50 usec
PL1 -3.00 dB
SFO1 100.6270000 MHz

F2 - Processing parameters
SI 32768
SF 100.6270000 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 25.00 cm
CY 3.00 cm
FIP 250.000 ppm
F1 25153.19 Hz
F2 -50.000 MHz
PRNCH 12.00000 ppm/cm
H2CH 1207.50000 Hz/cm

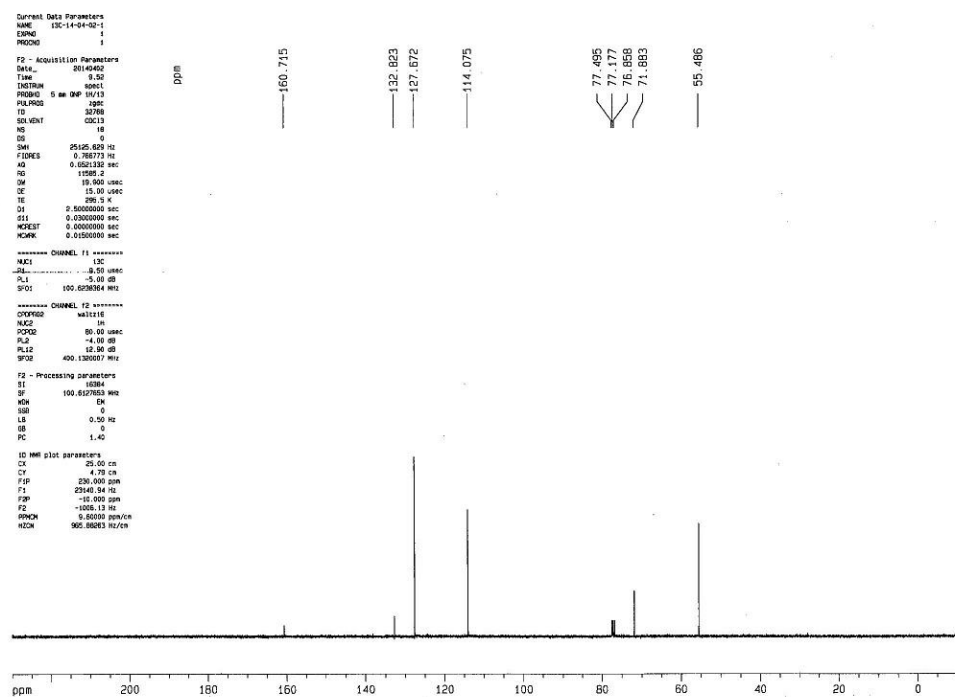
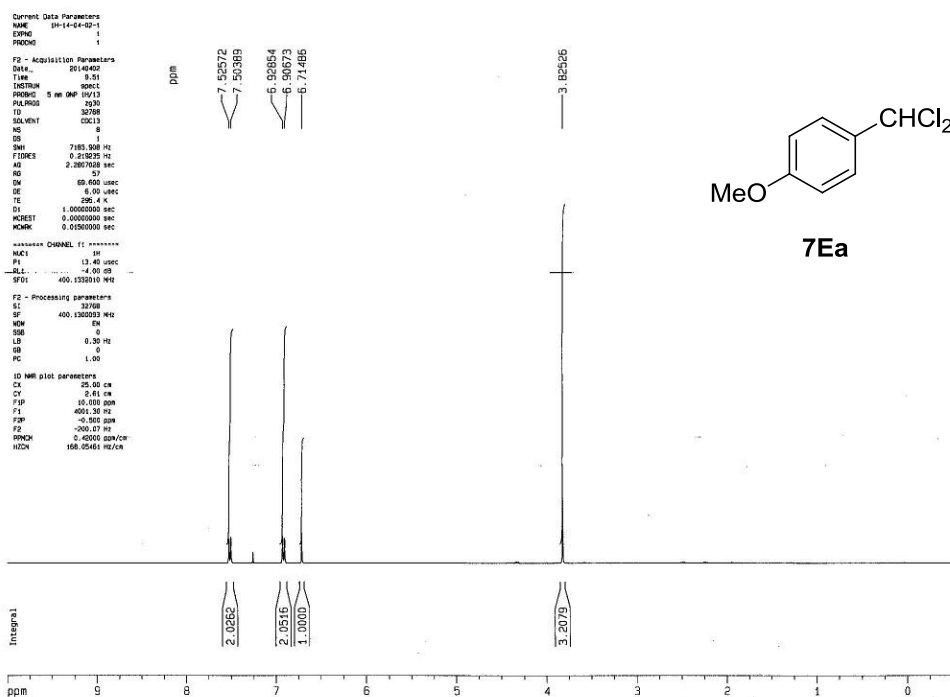
ppm

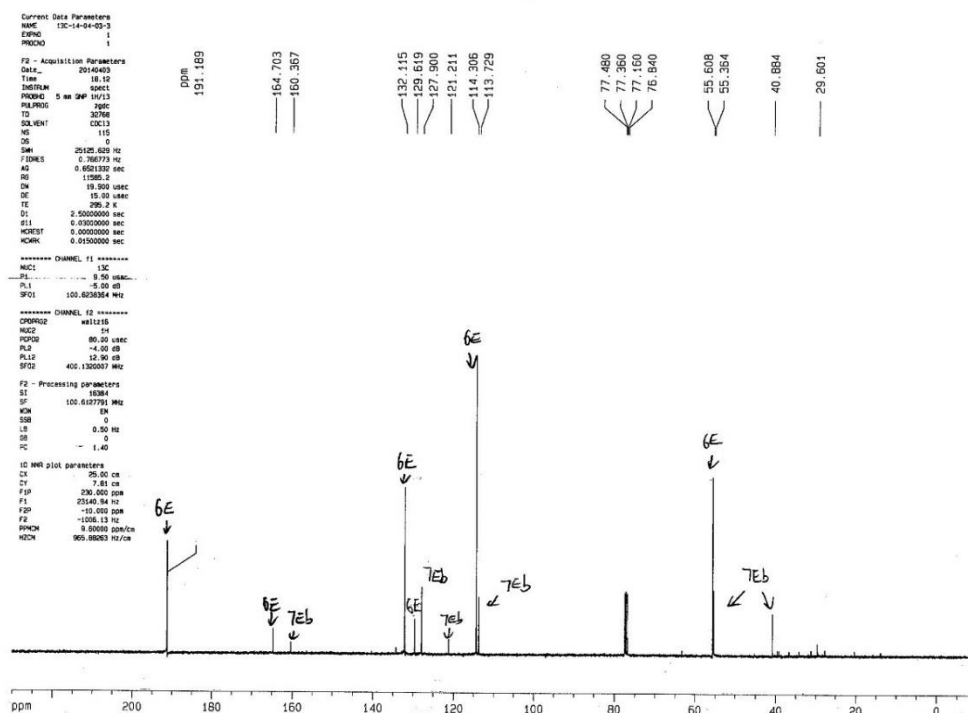
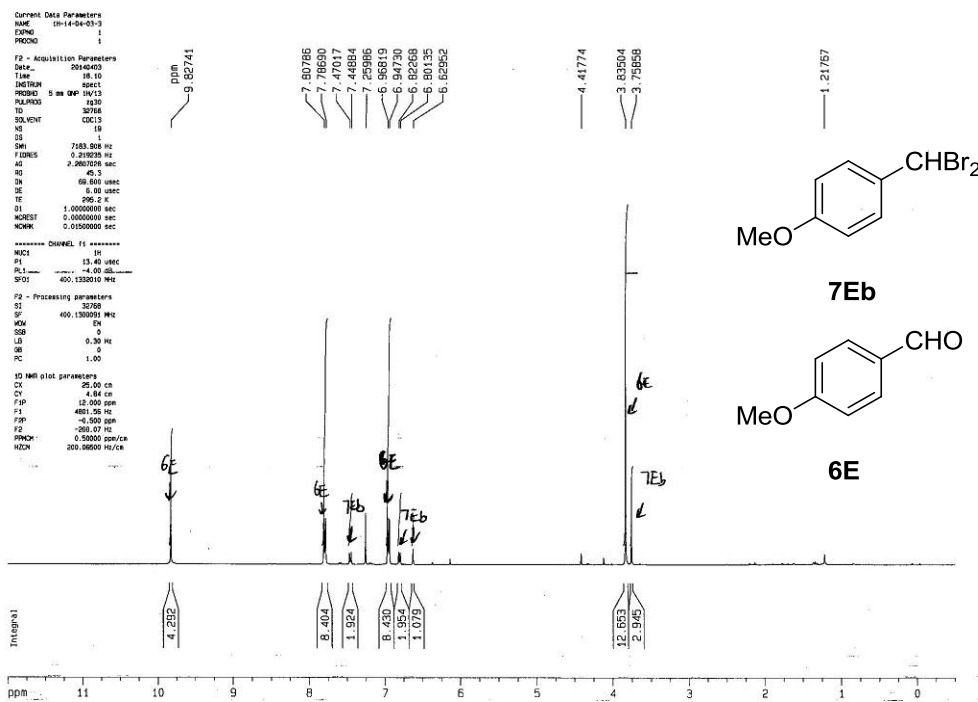
147.574
141.955
132.802
131.352
128.420
123.876

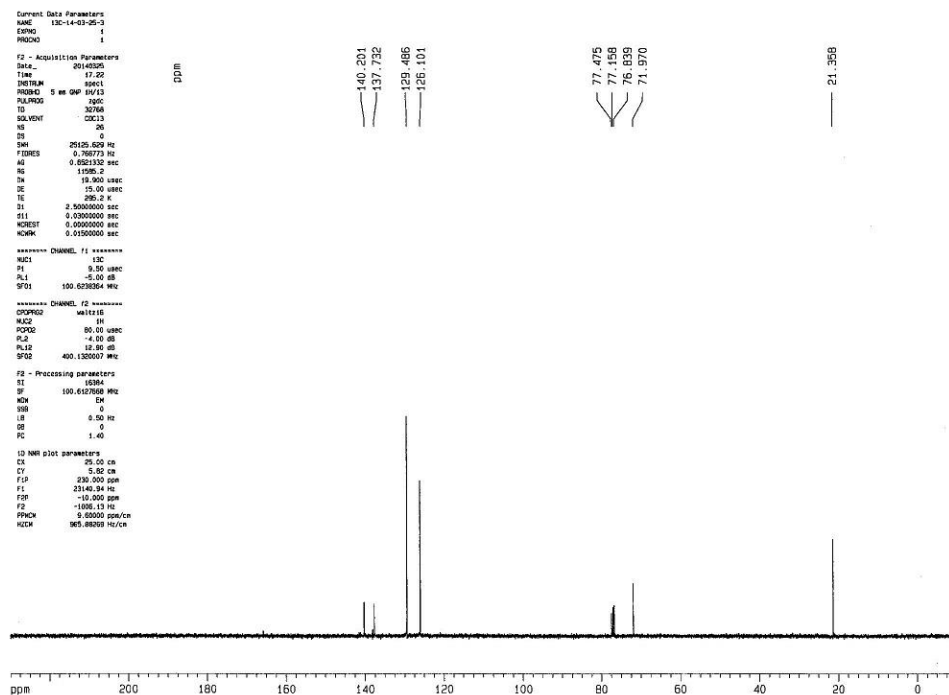
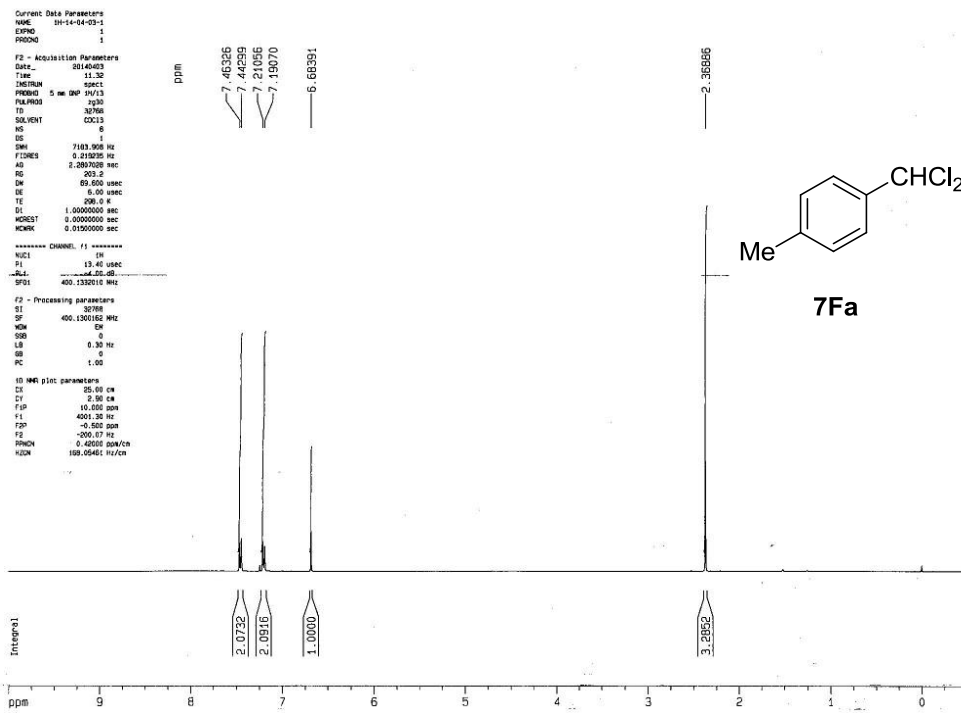
77.476
77.353
77.158
76.840

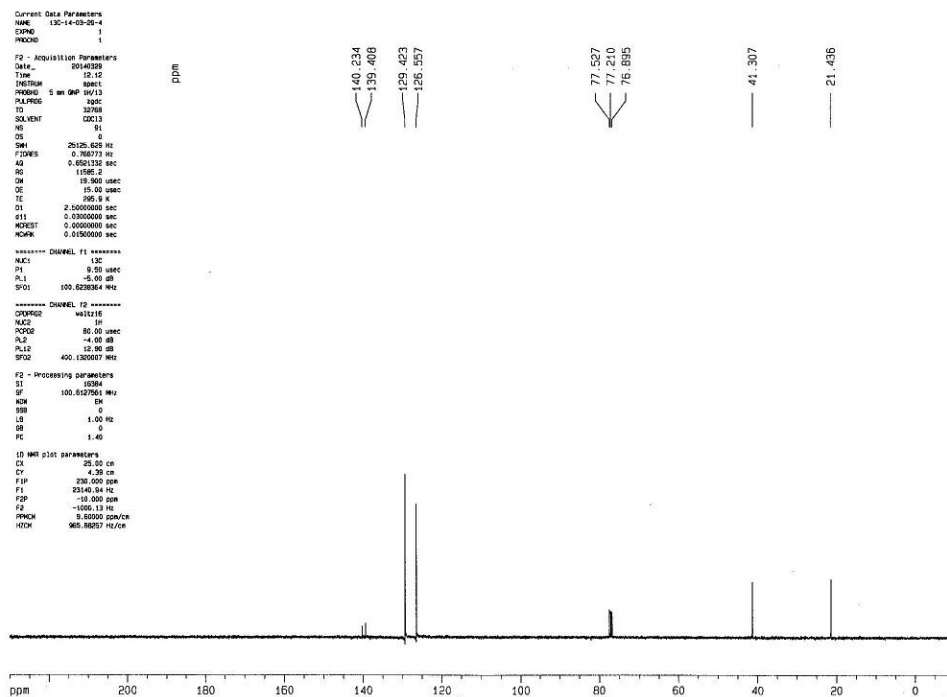
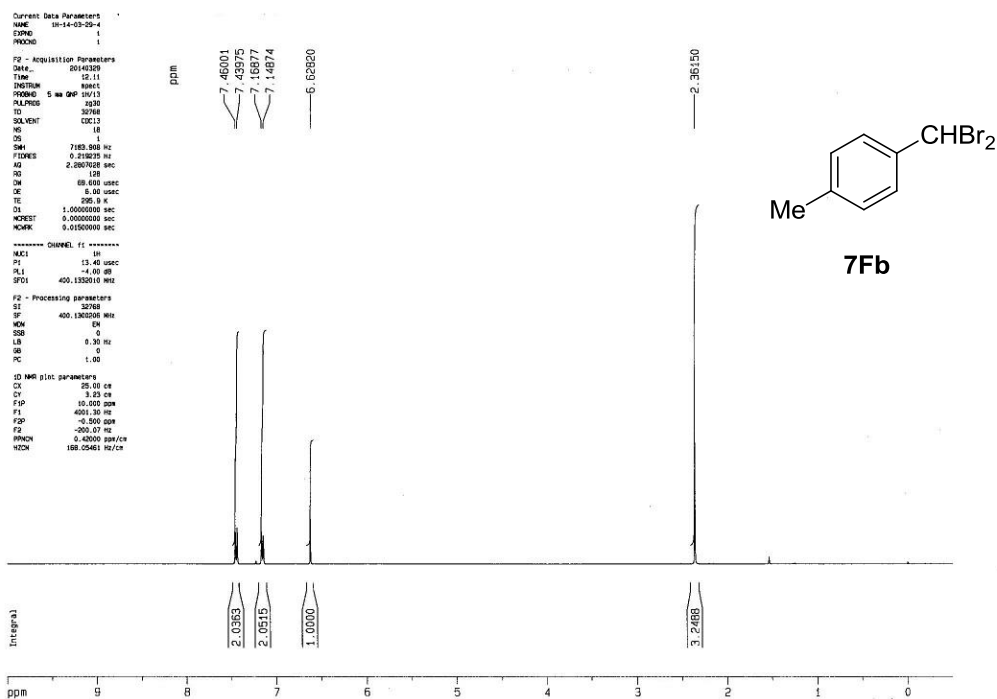
36.947

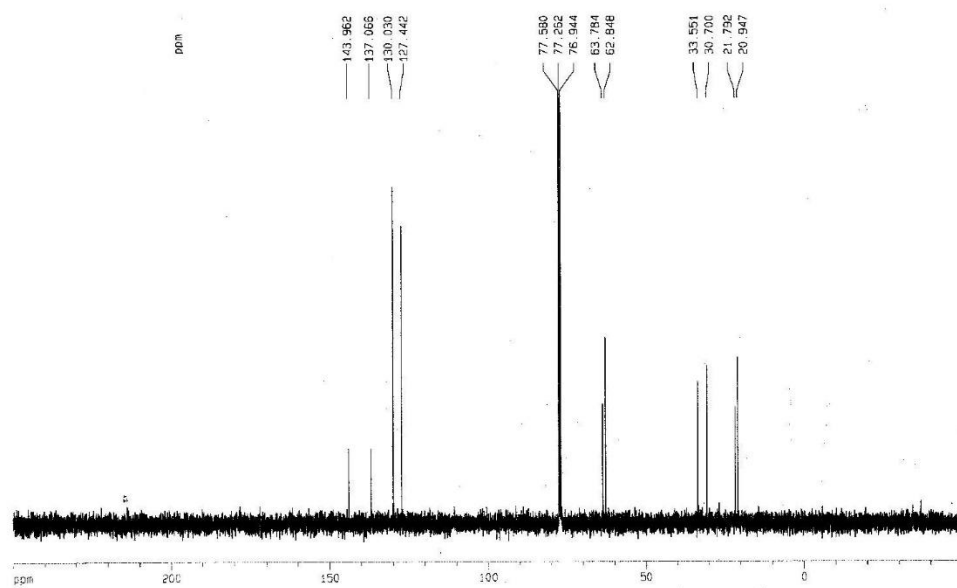
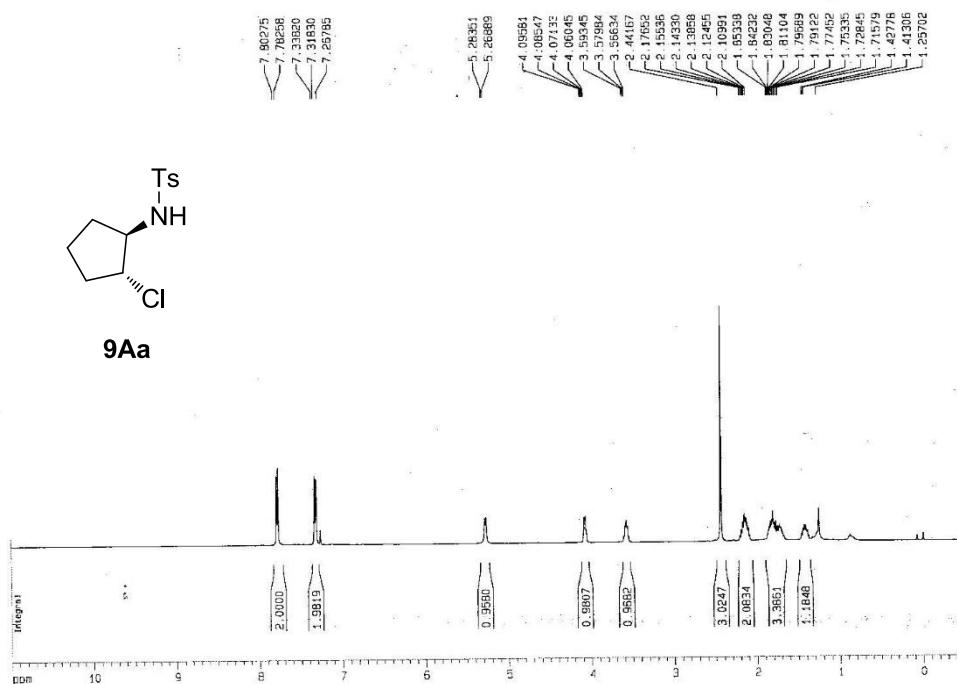
ppm 200 150 100 50 0

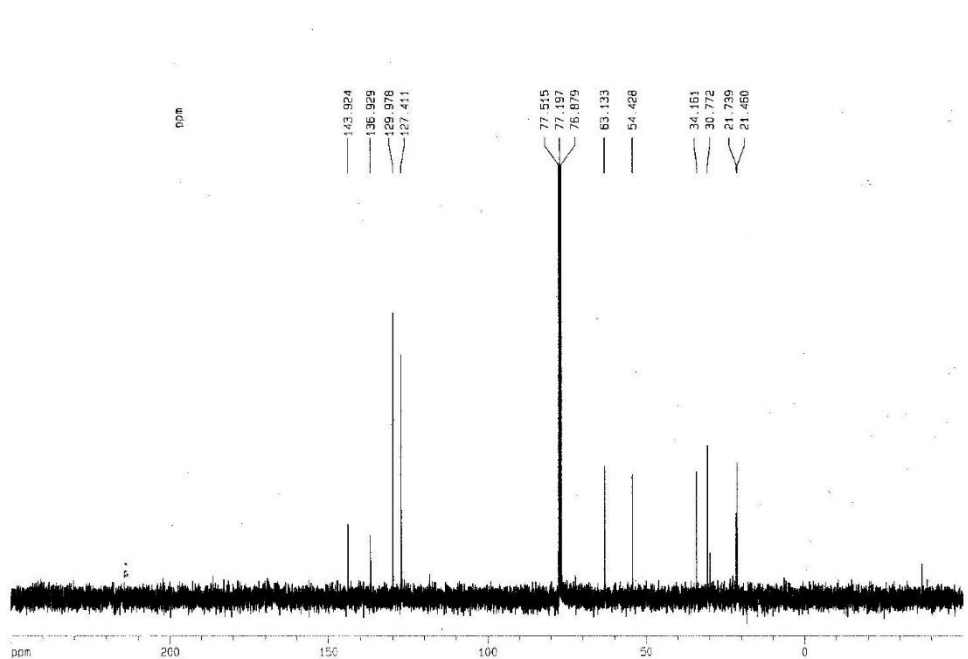
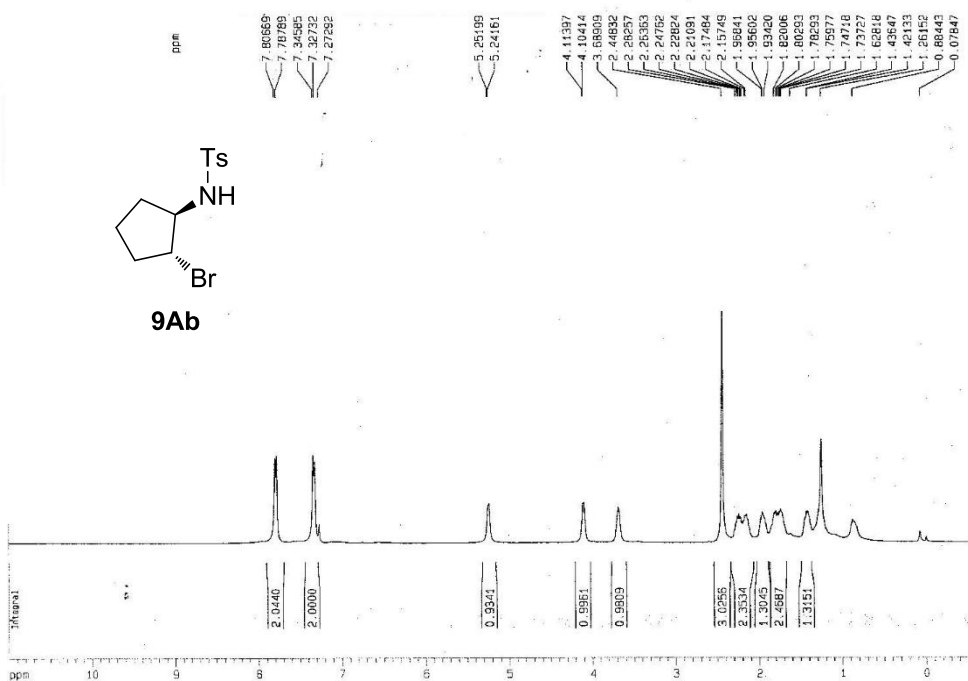


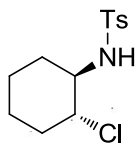




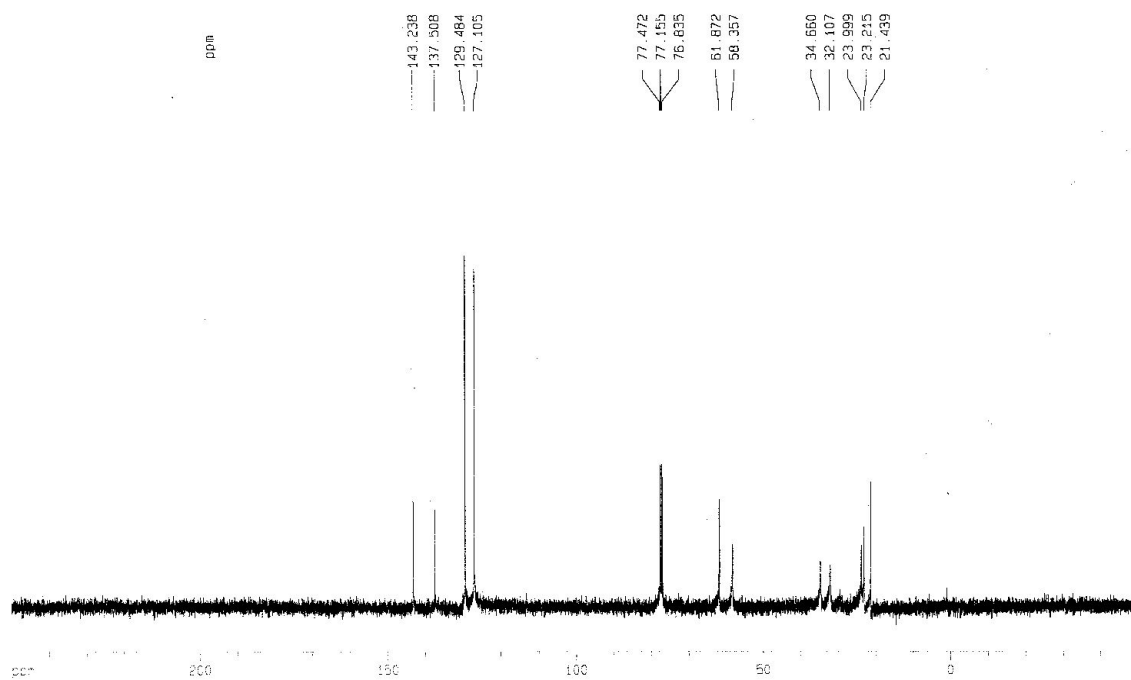
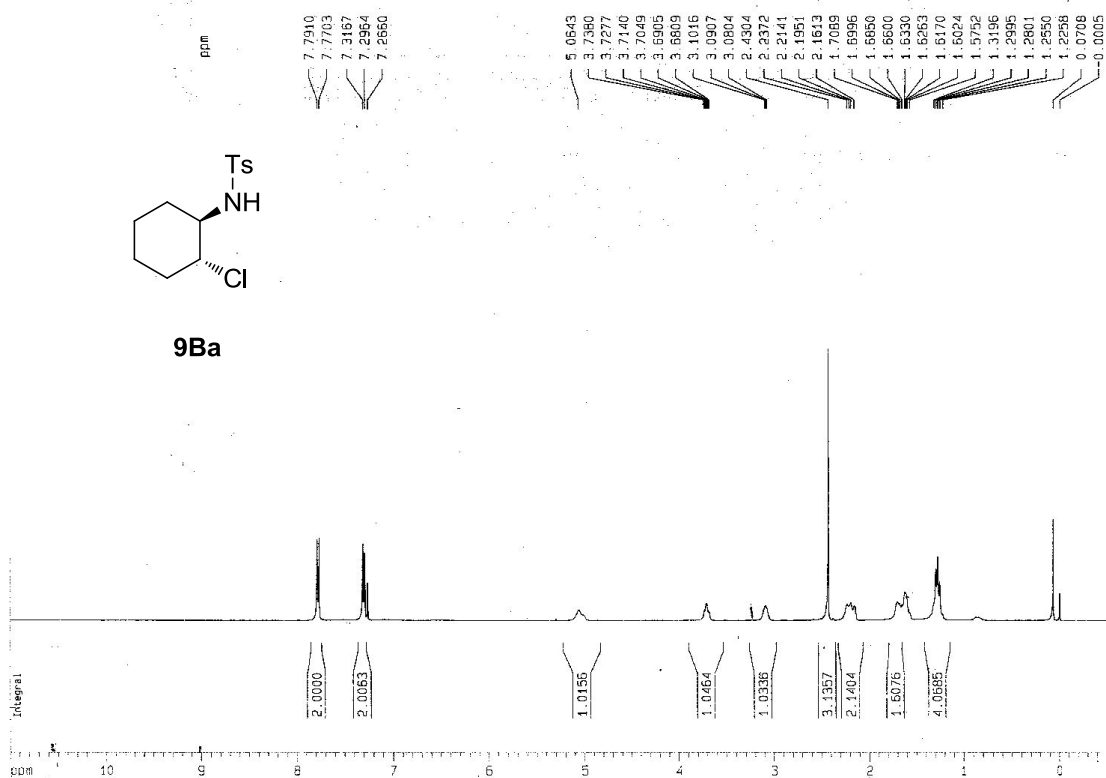


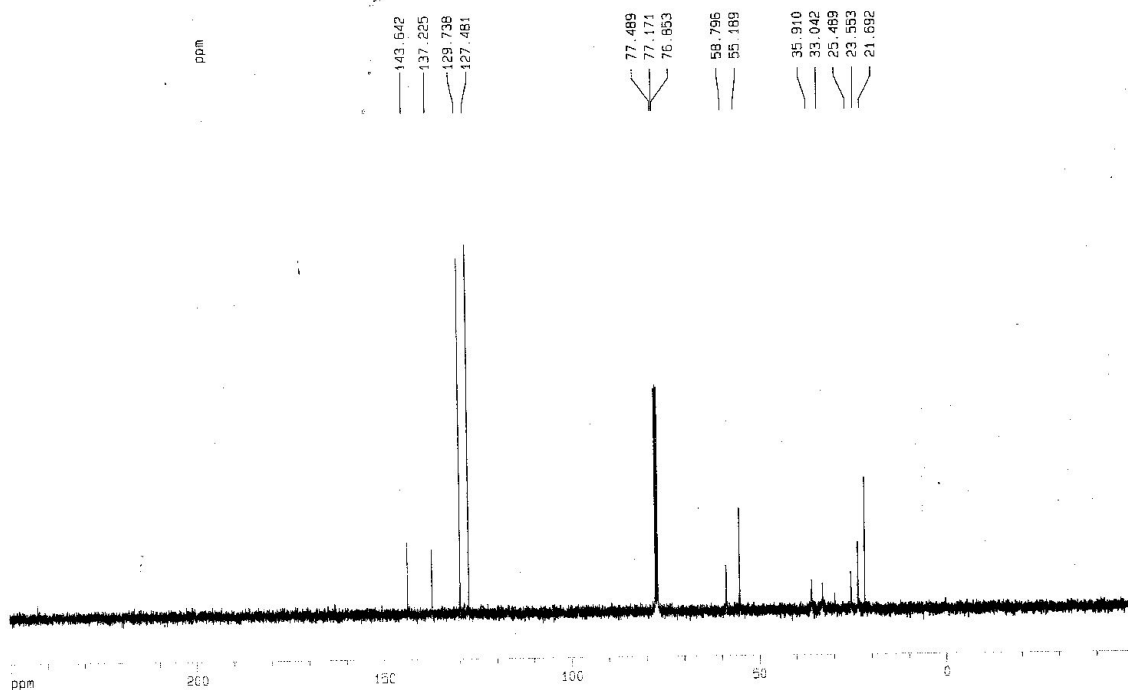
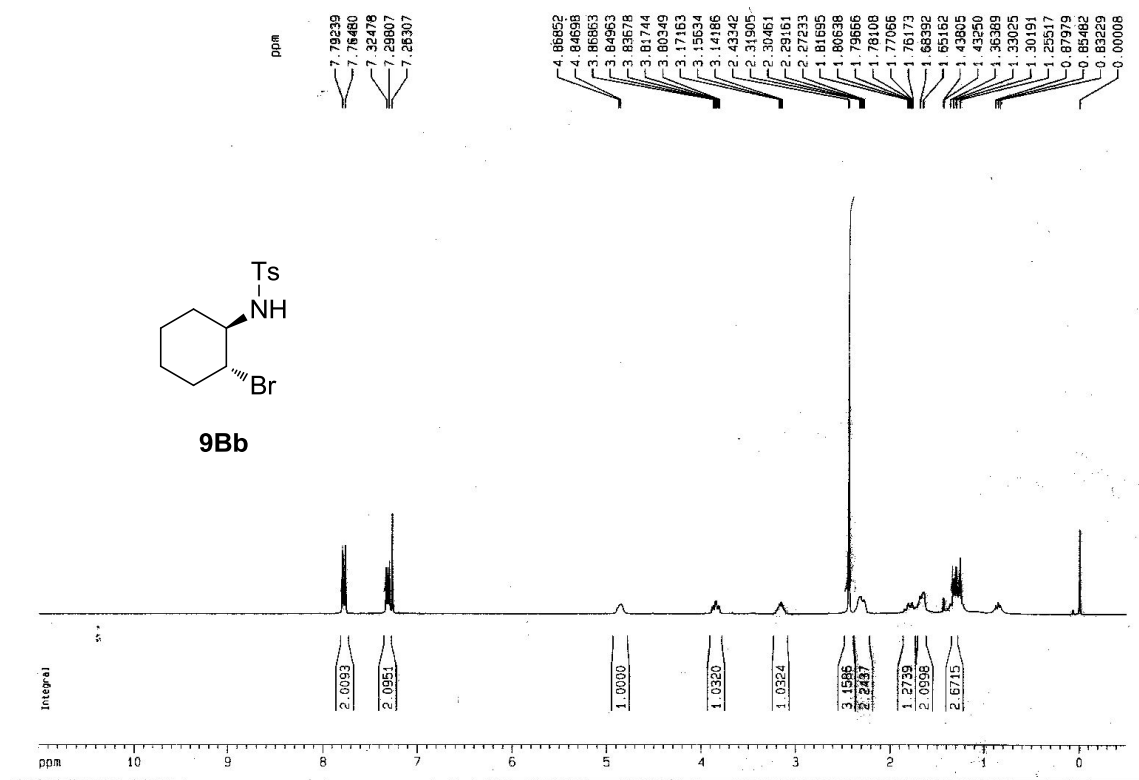


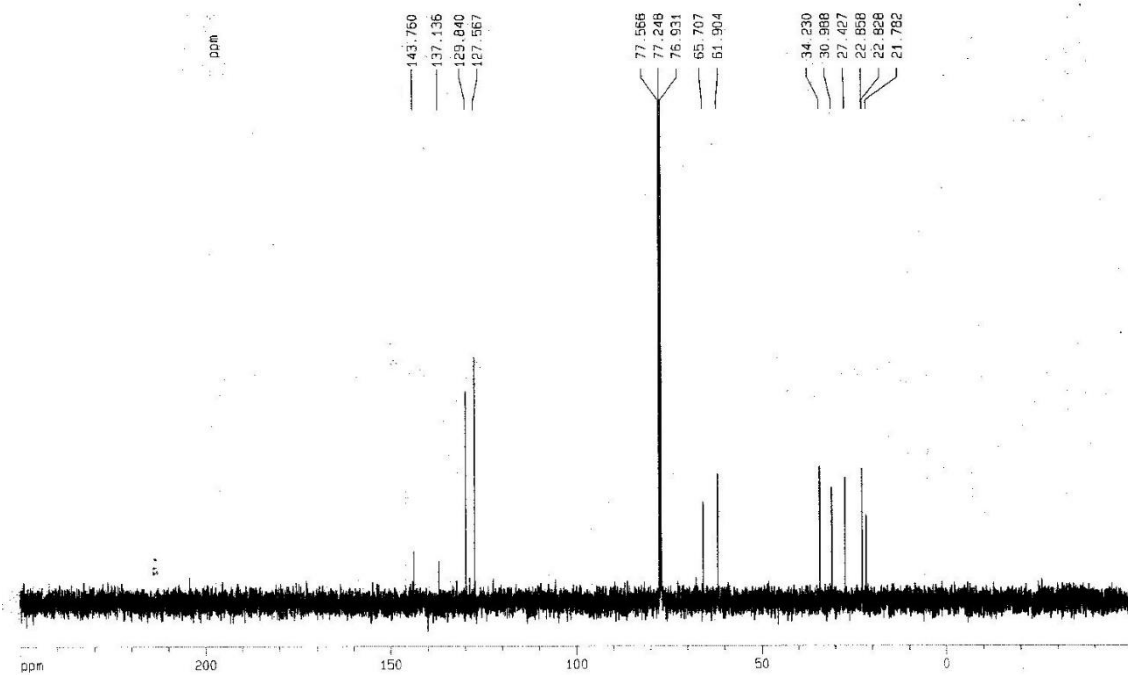
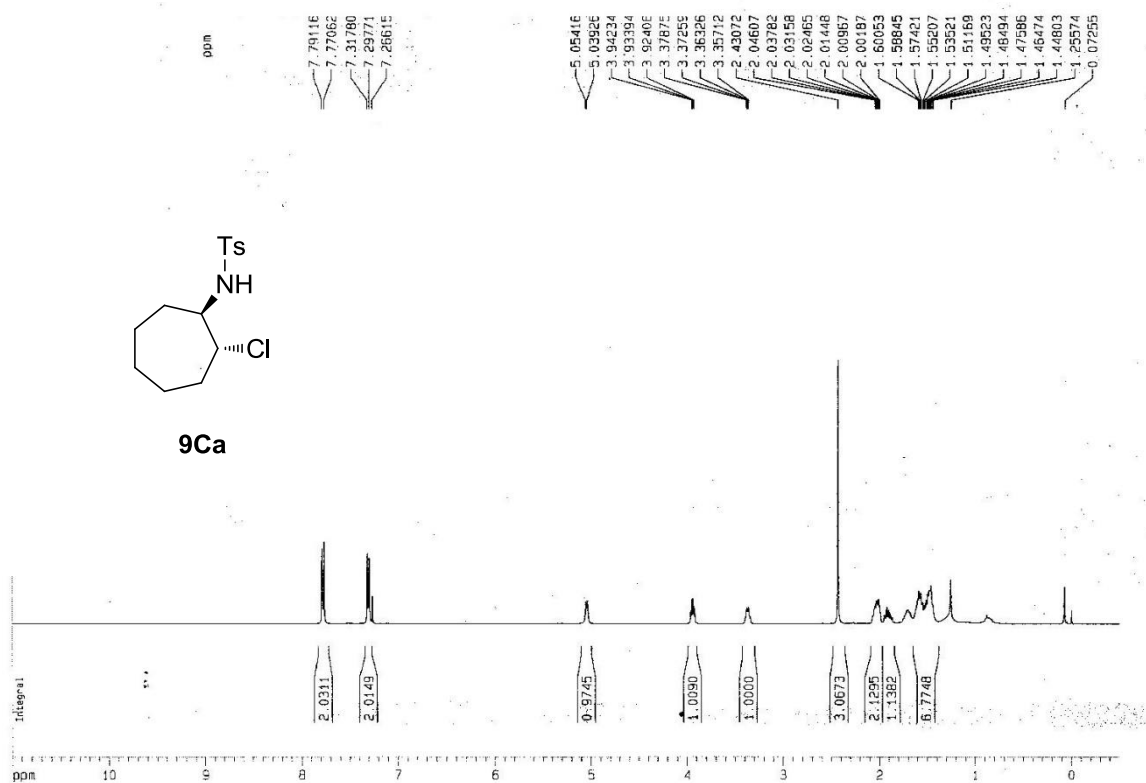


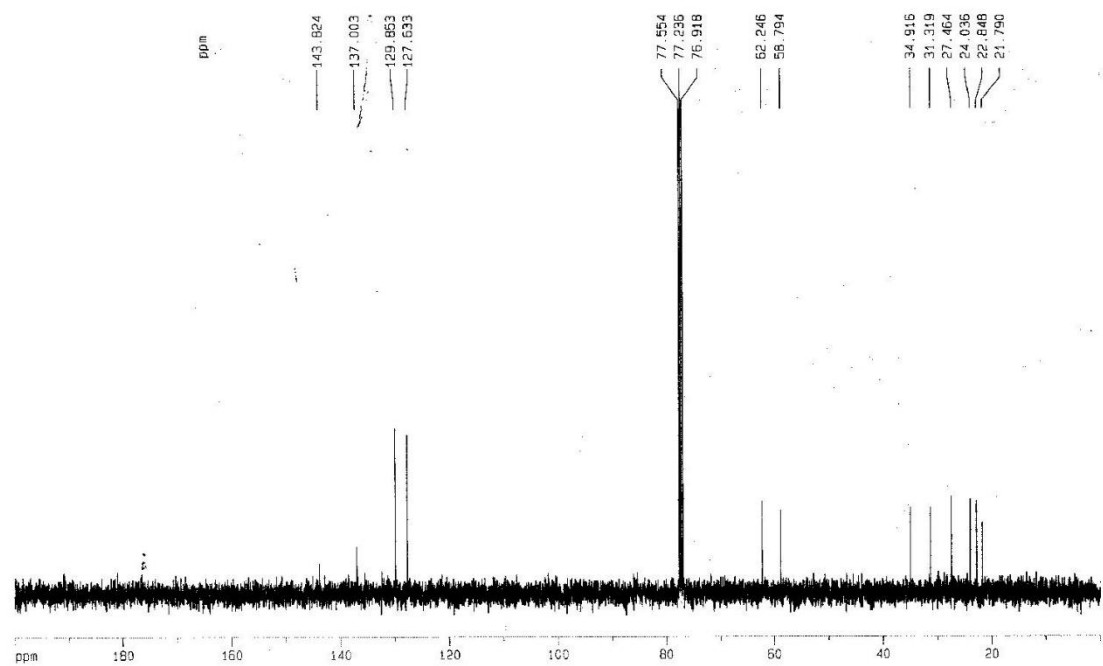
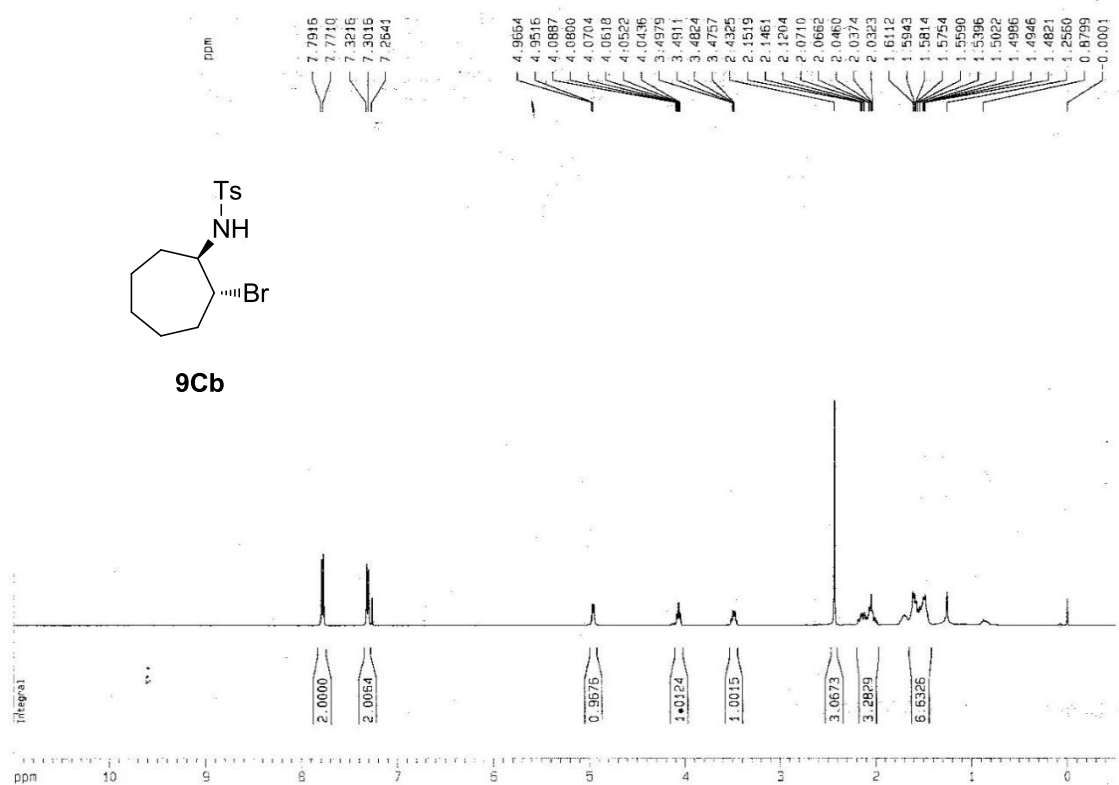


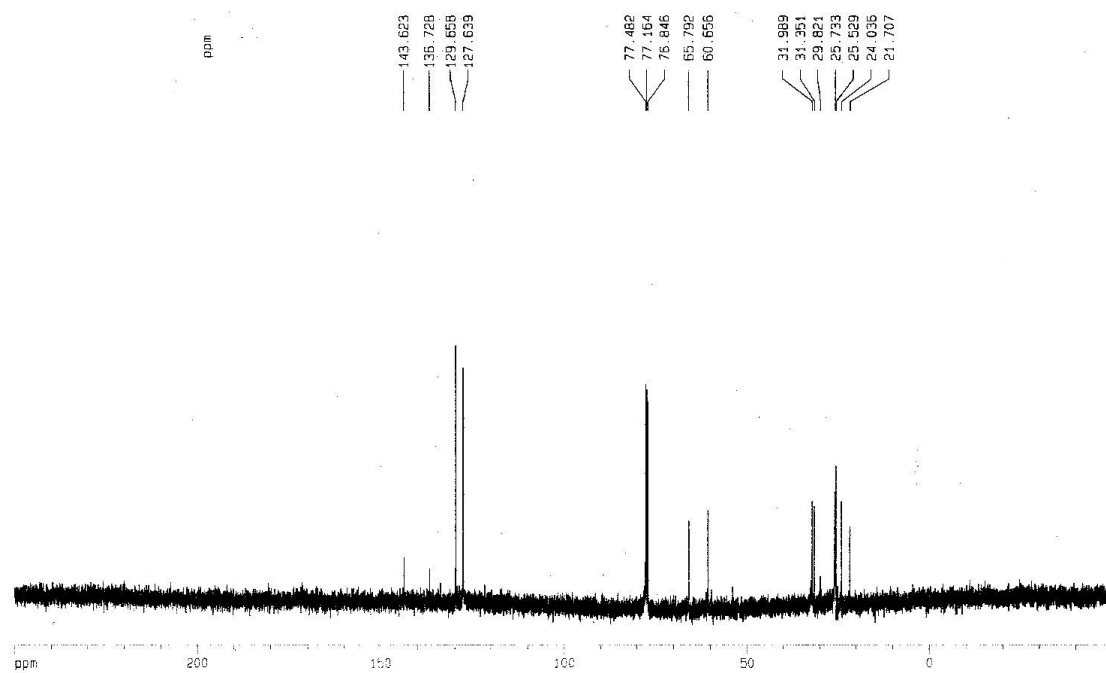
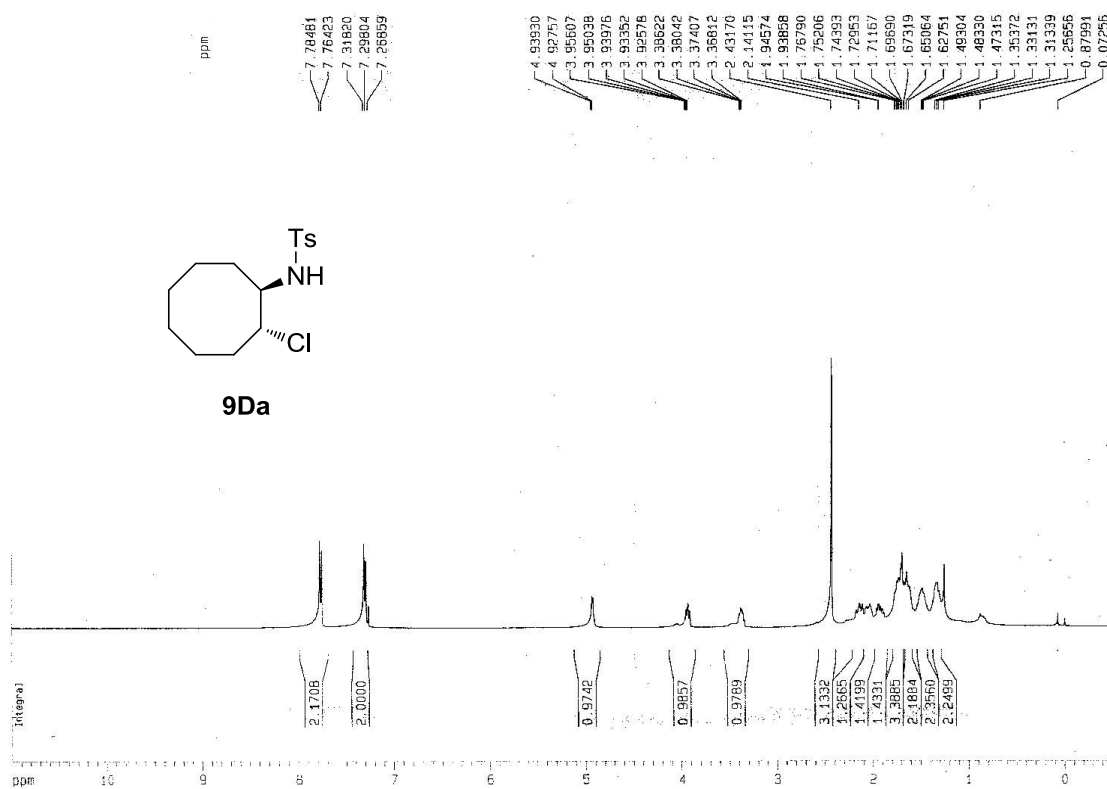
9Ba

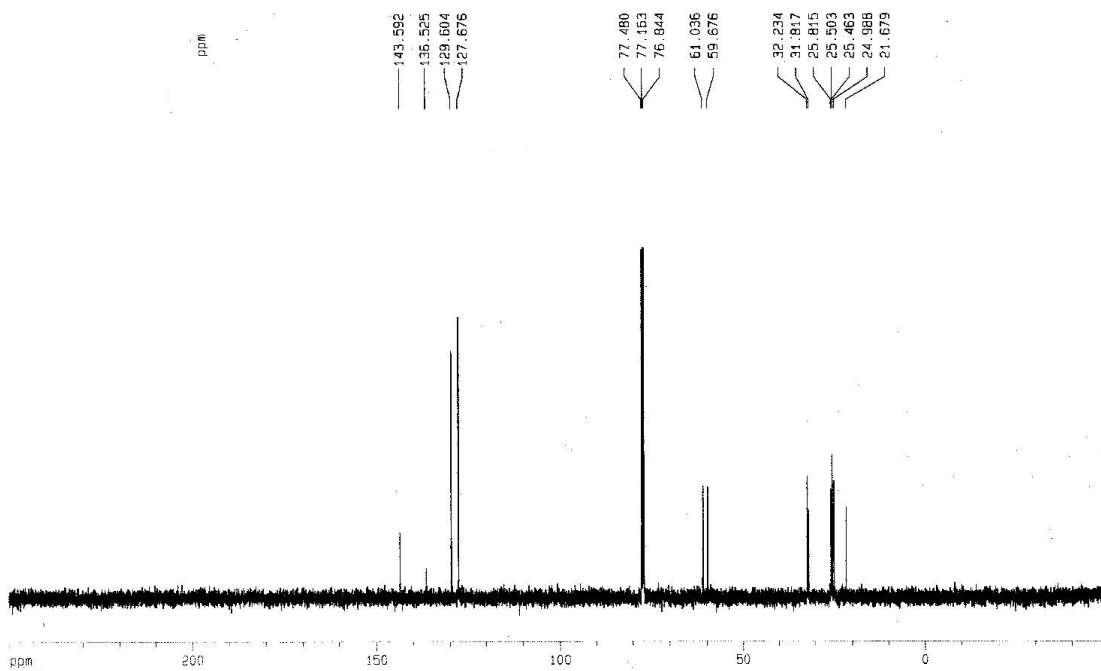
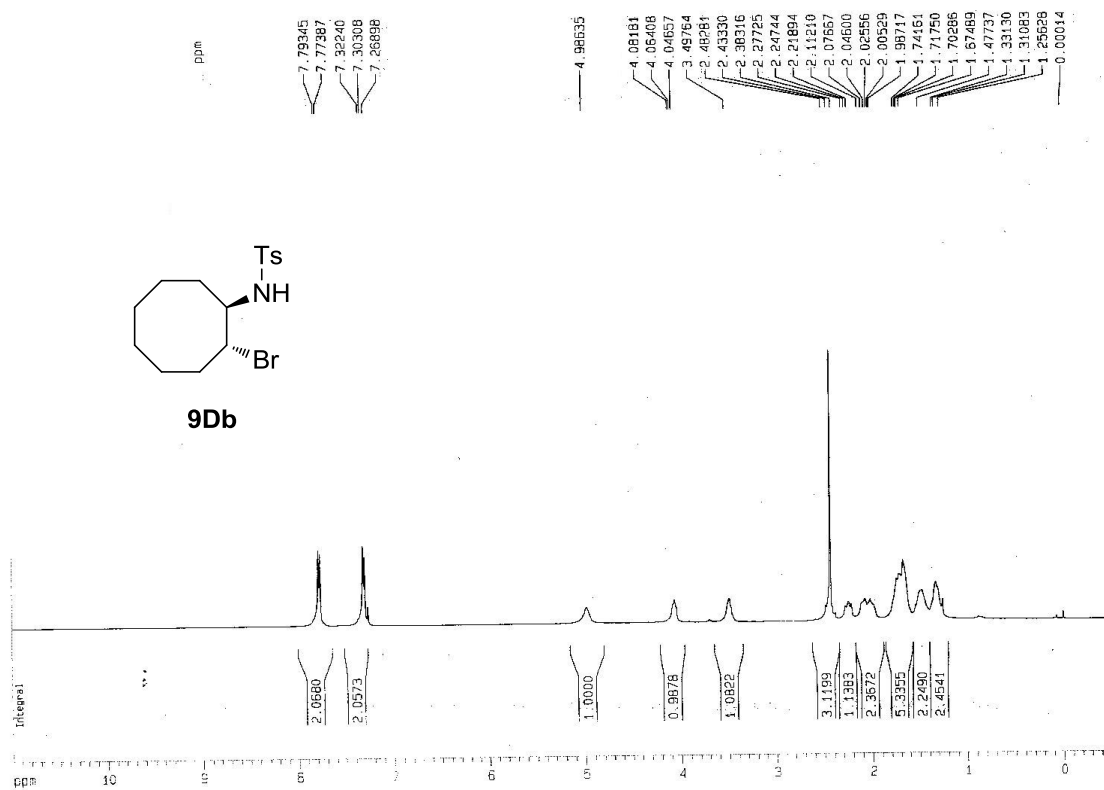


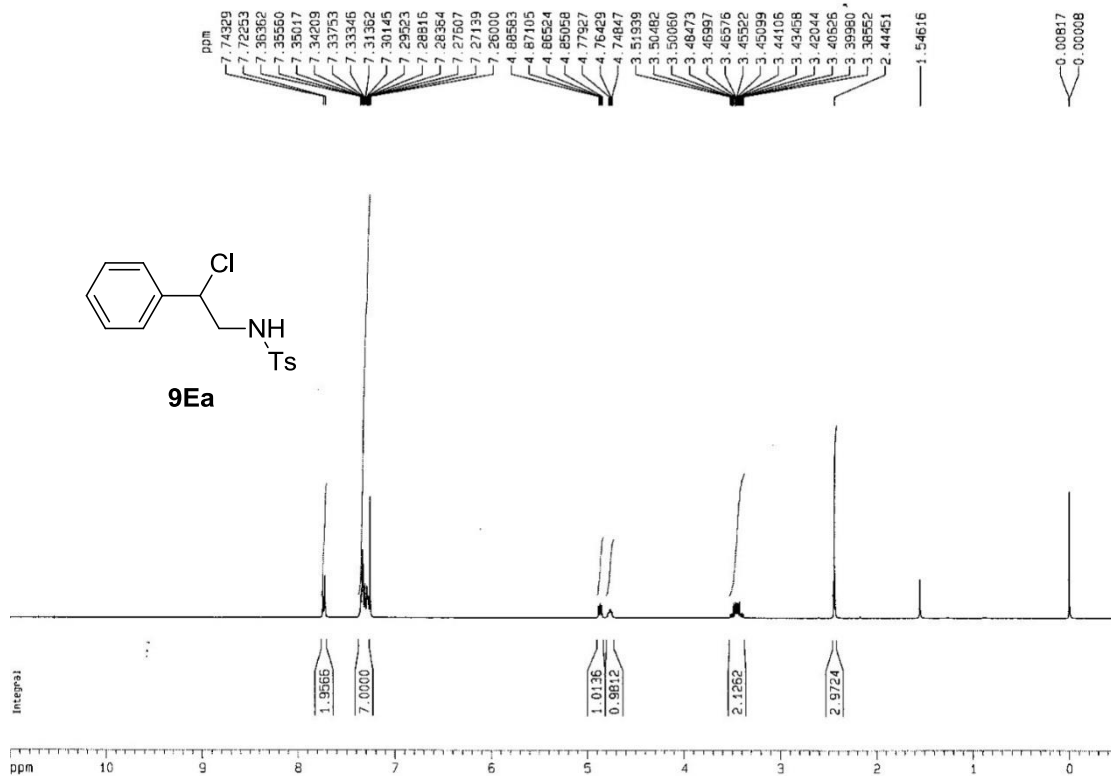












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 PROCNO 1

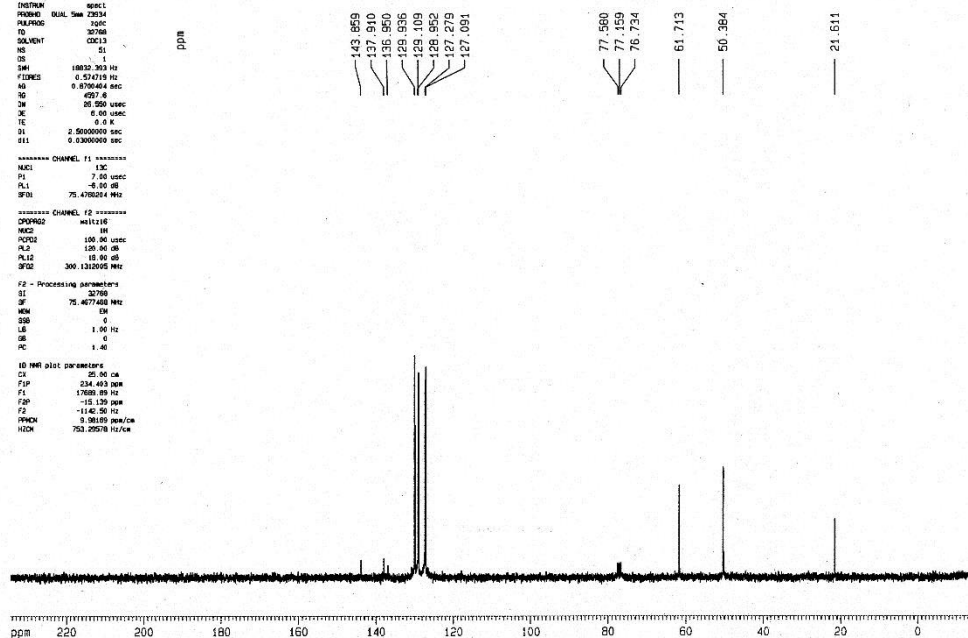
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 FIDRES 0.574719 Hz
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 RG 4997.6
 SN 25.550 uspc
 JC 6.00 uspc
 TC 0.0 K
 IN 2.50000000 sec
 SFI 0.03000000 sec

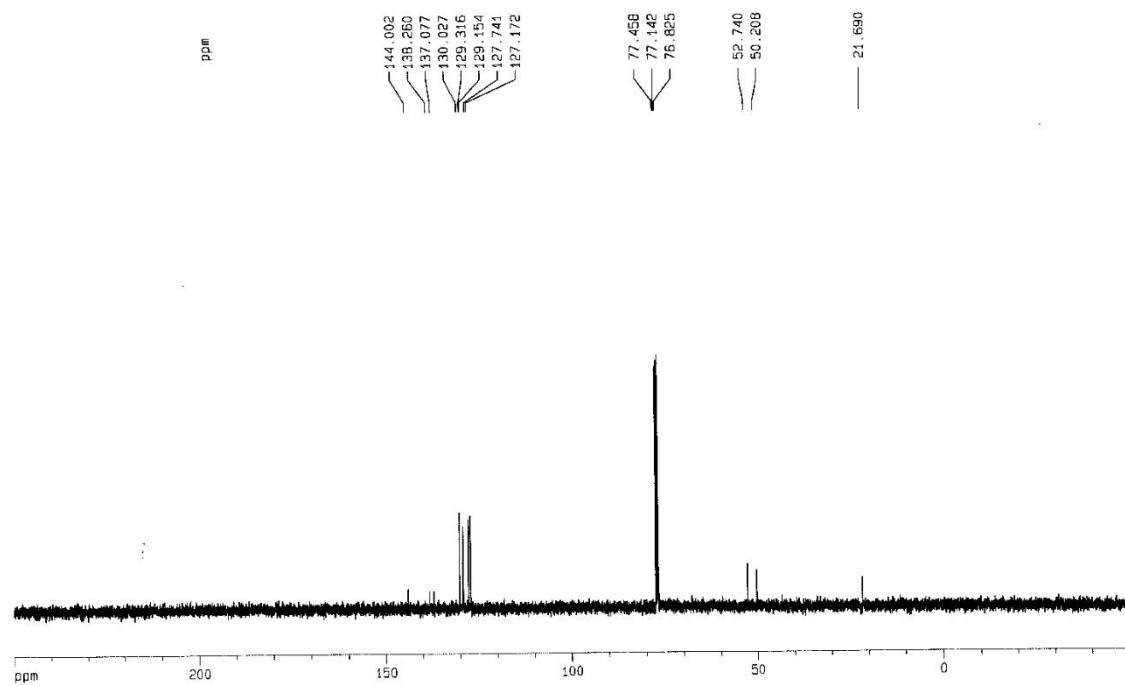
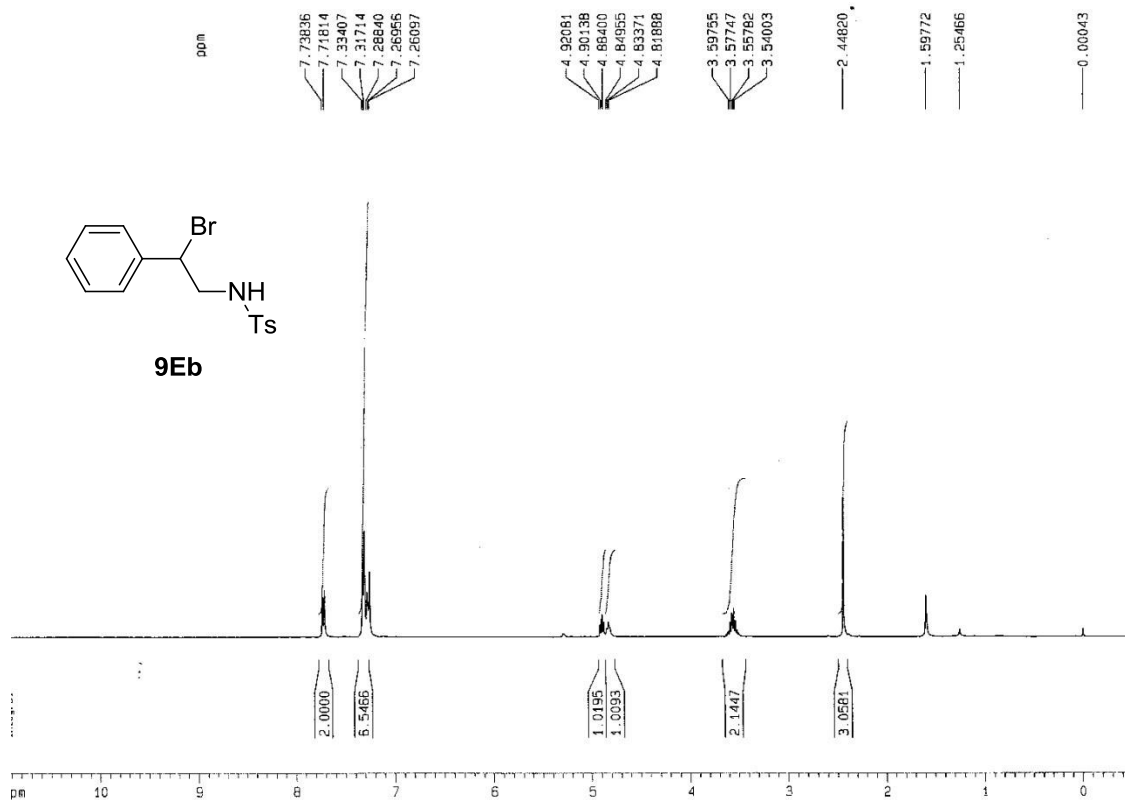
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 SFO1 75.4750000 MHz

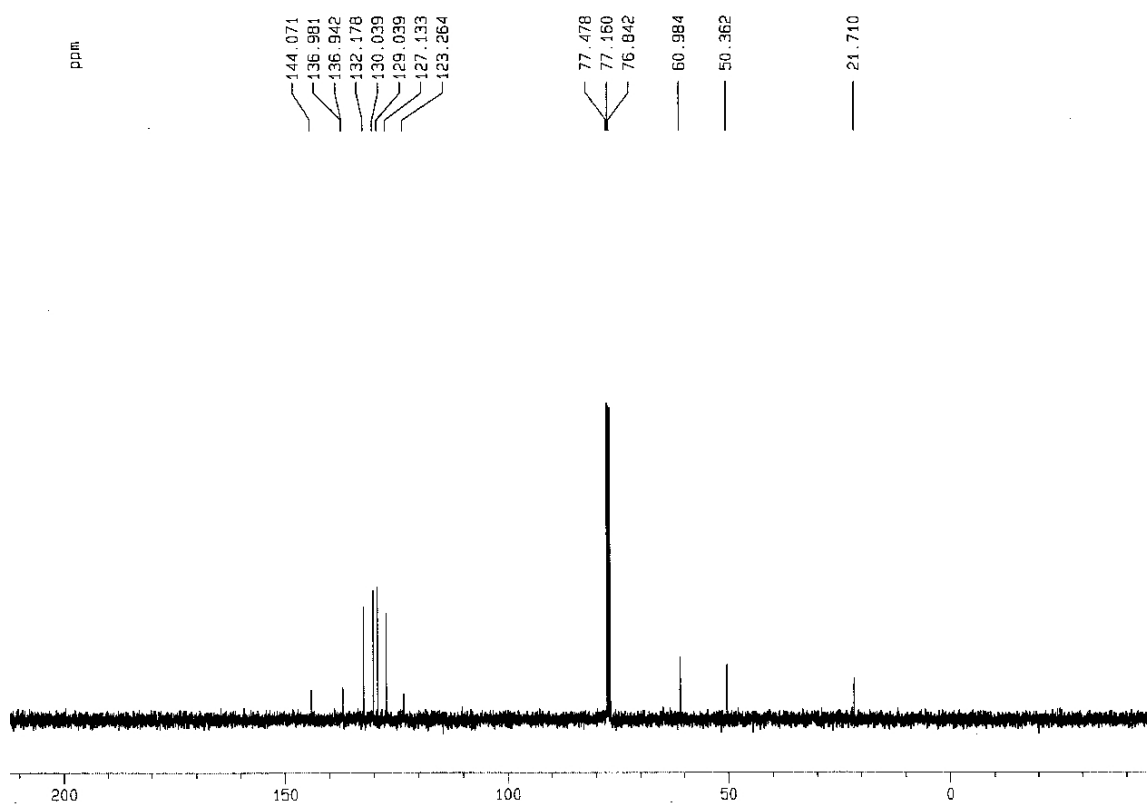
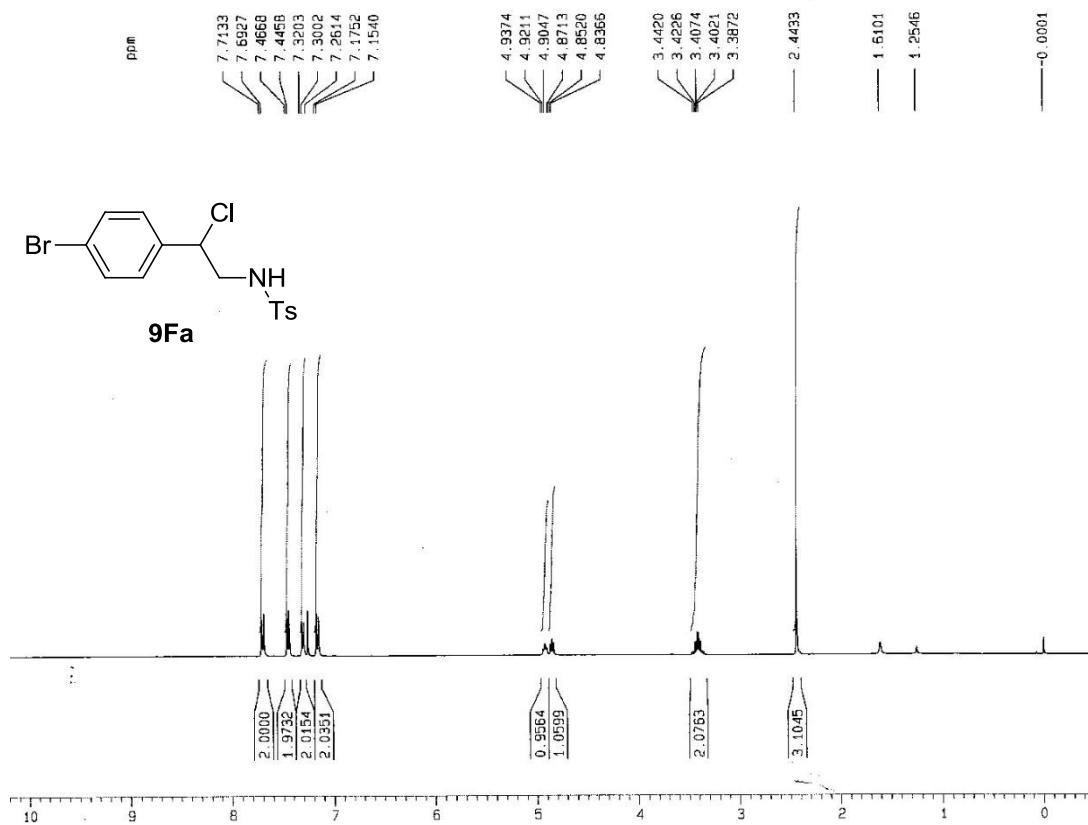
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 P2 100.00 uspc
 PL2 19.00 dB
 PL12 19.00 dB
 SFO2 300.1320000 MHz

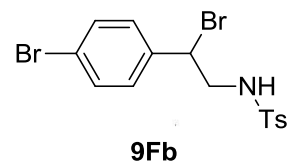
F2 - Processing parameters
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 MN 20
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
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 F1P 234.403 dB
 F1 17000.00 Hz
 F2P -15.120 dB
 F2 -1140.50 Hz
 FWHM 0.38100 dB/cw
 HZCX 753.29570 Hz/cw

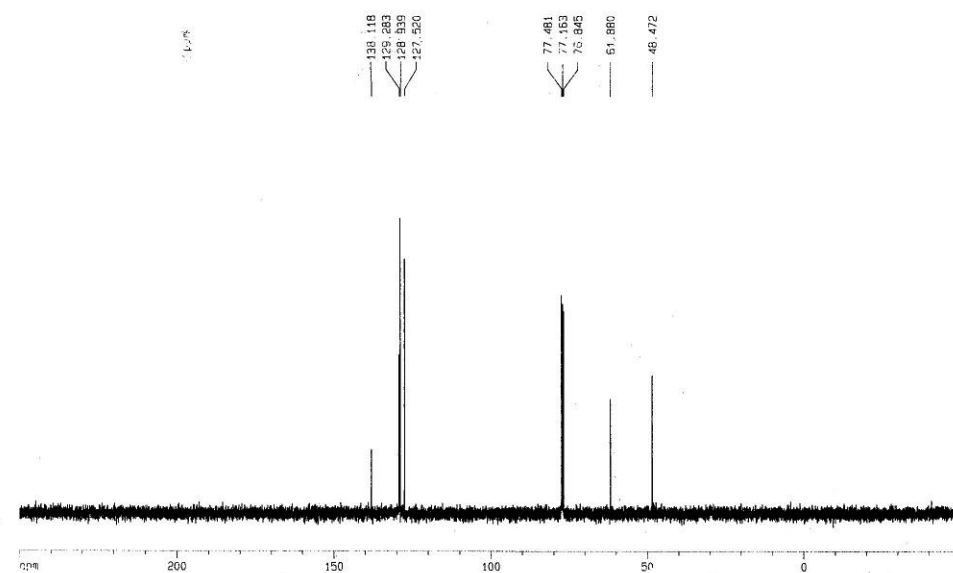
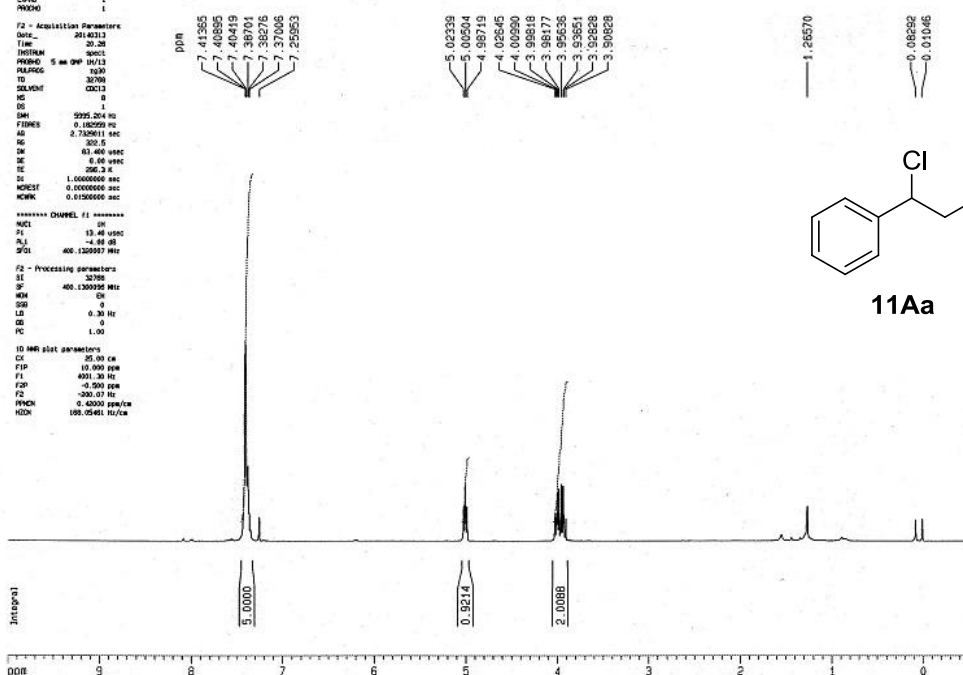


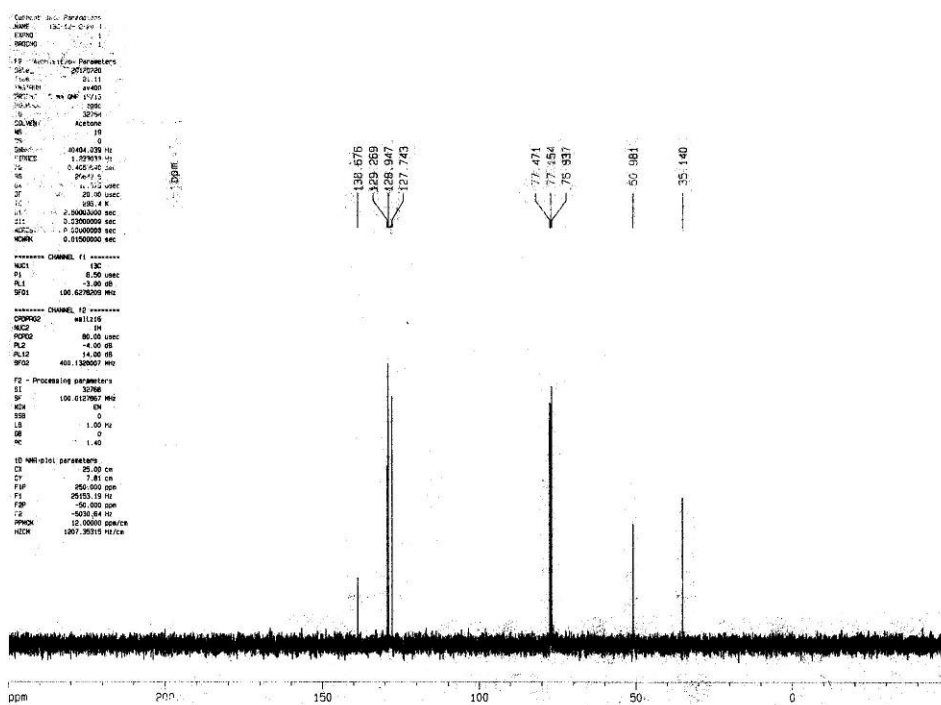
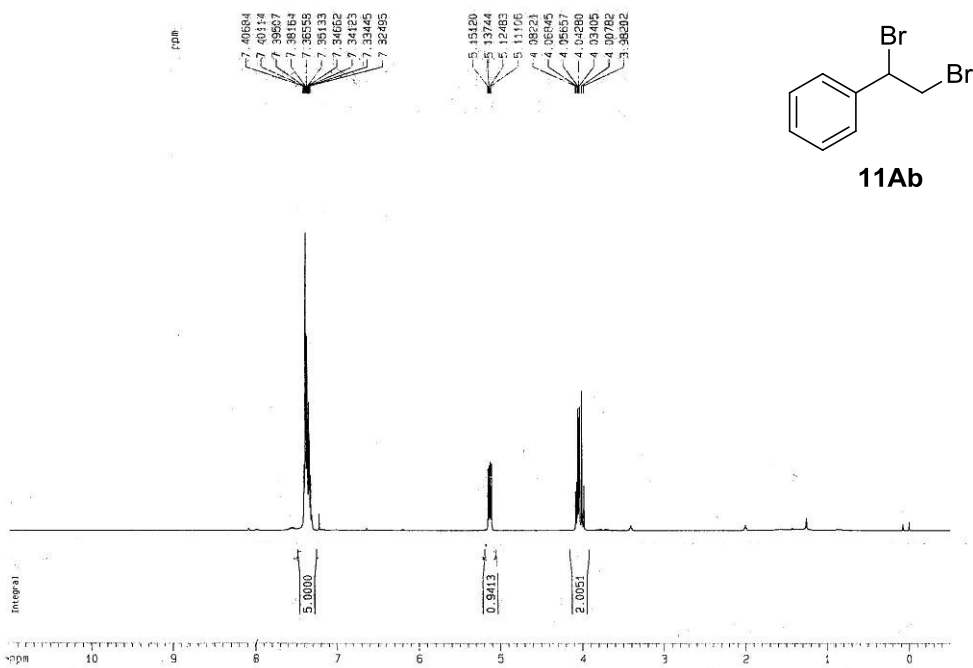


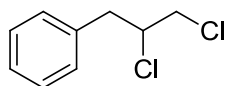




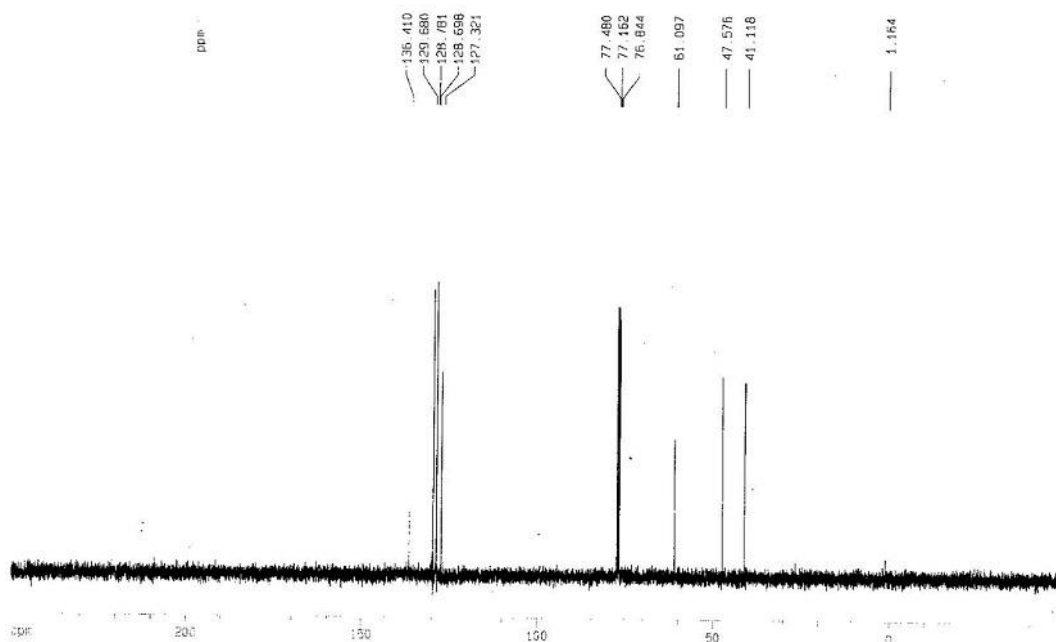
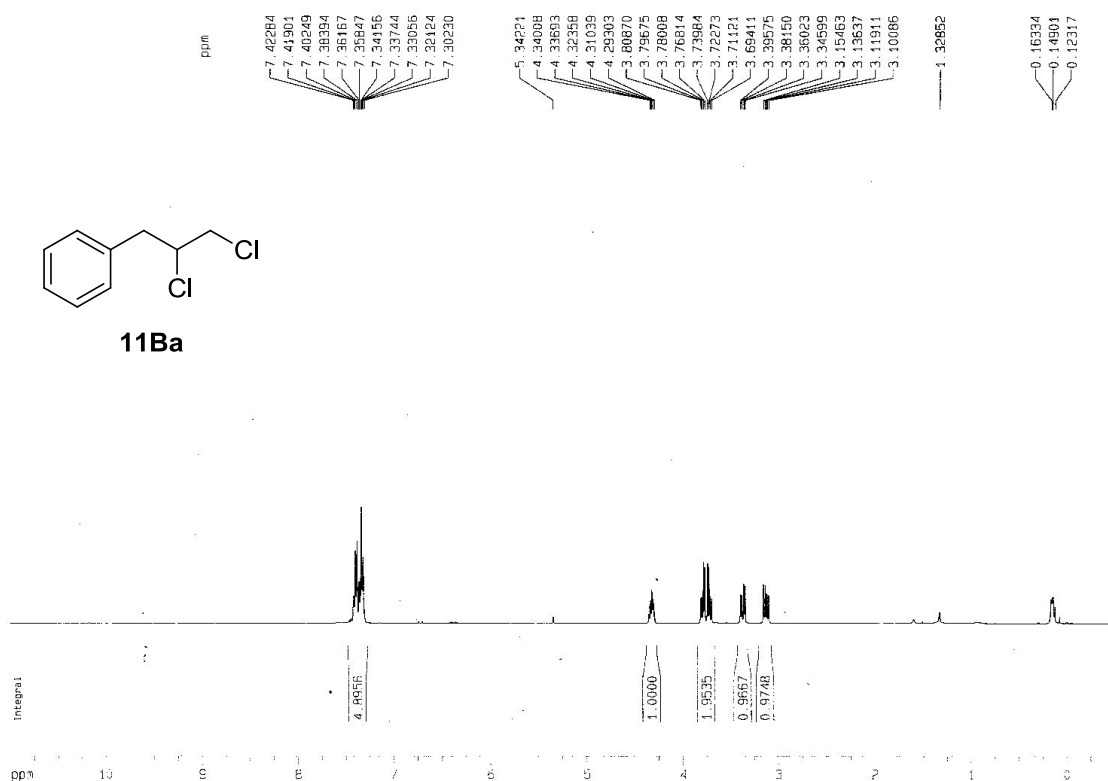
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 PROCNO 1
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 PULPROG zgpg30
 TO 32768
 SOLVENT CDCl3
 NS 0
 DS 4
 SWH 2295.204 Hz
 FIDRES 0.180259 Hz
 AQ 2.732611 sec
 RG 320.5
 DE 63.400 usec
 SE 0.80 usec
 DI 1.0000000 sec
 NUC1 13C
 P1 13.40 usec
 PL1 -1.80 dB
 SFO1 400.1300937 MHz
 F2 - Processing parameters
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 SF 400.1300937 MHz
 NH 655
 SH 0
 LB 0.30 Hz
 GB 0
 PC 1.00
 IO NMR plot parameters
 CX 20.00 cm
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 FI 4001.30 Hz
 FZ0 -0.500 ppm
 FZ -390.07 Hz
 PPM0 0.0000 ppm/cm
 HZ0 100.0541 Hz/cm

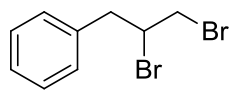




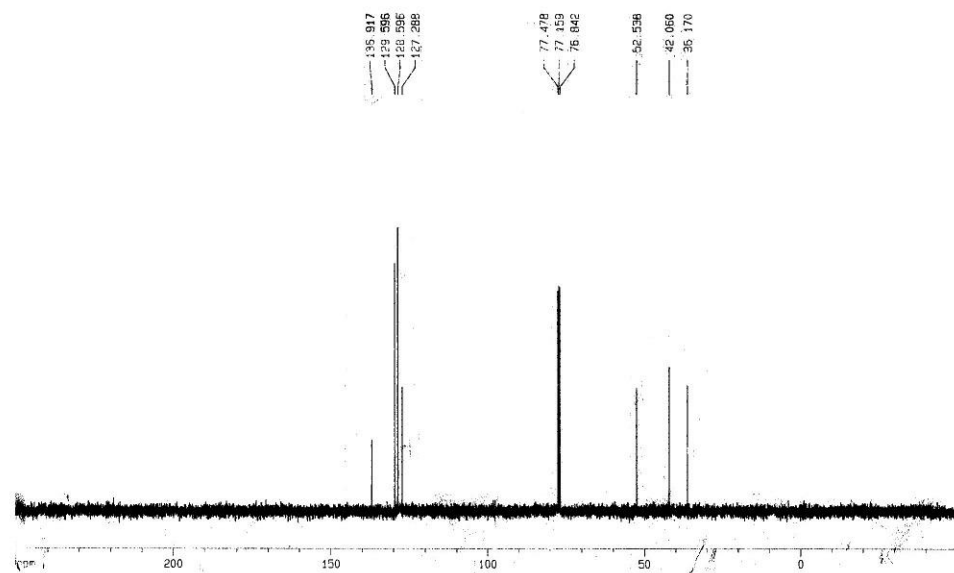
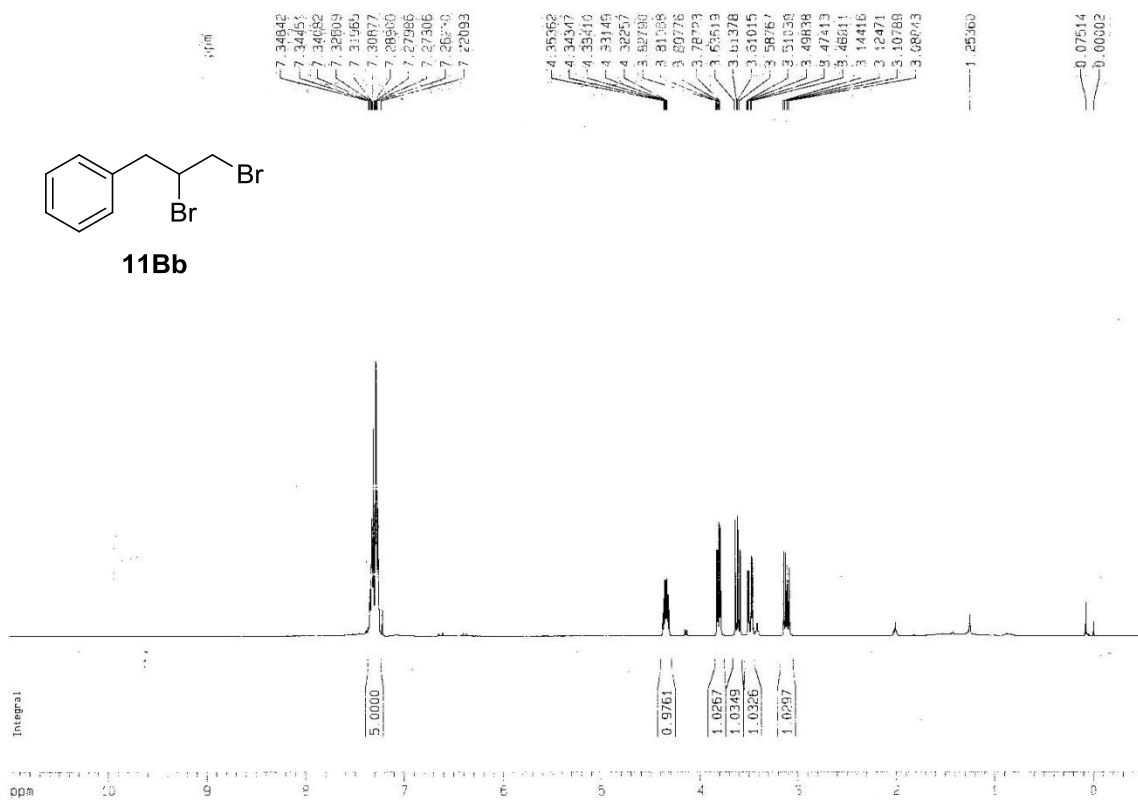


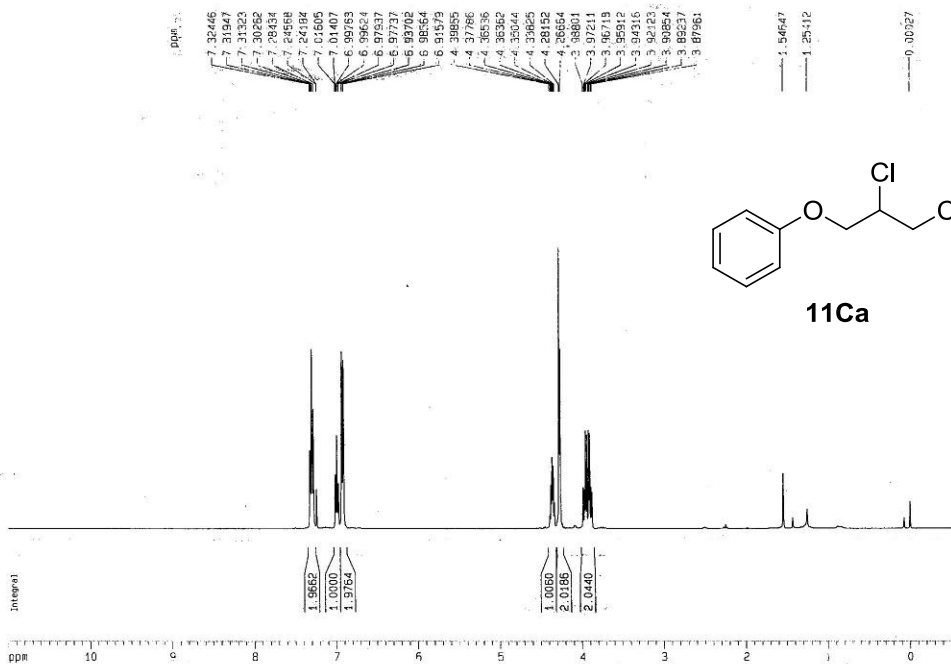
11Ba





11Bb





Current Data Parameters
NAME 13C-14-04-04-2
EXPNO 1
PROCNO 1

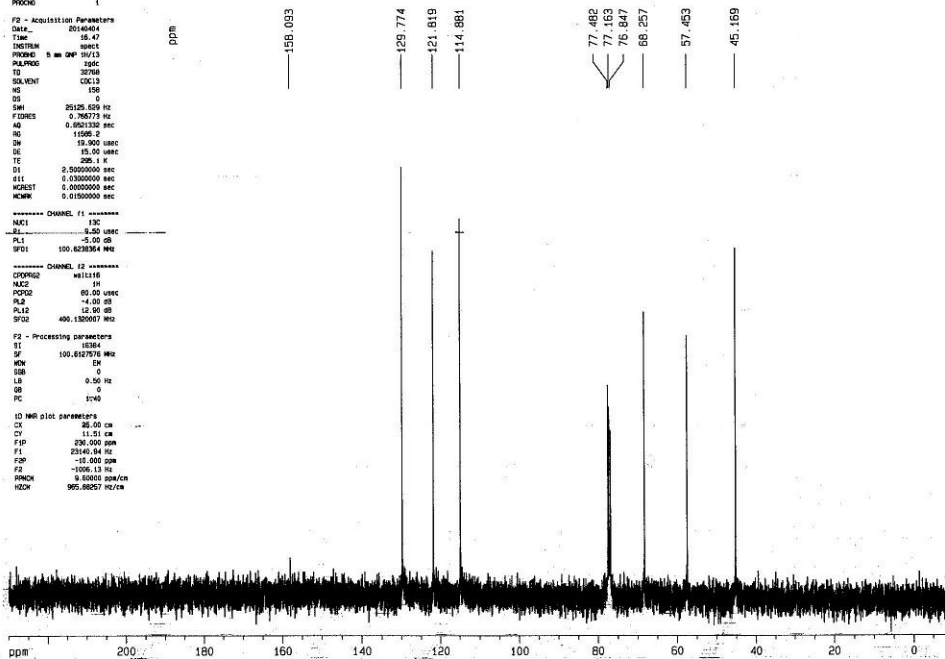
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TD 32768
SOLVENT CDCl3
NS 150
DS 4
SWH 25125.620 Hz
FIDRES 0.3601330 Hz
AQ 0.0011330 sec
RG 11500.2
SN 13.900 dB
DE 15.00 dB
TE 300.2 K
D1 2.50000000 sec
S11 0.03000000 sec
MCHST 0.00000000 sec
MCHW 0.01500000 sec

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NUC1 13C
P1 130
PL1 0.00 usec
SFO1 100.626364 MHz

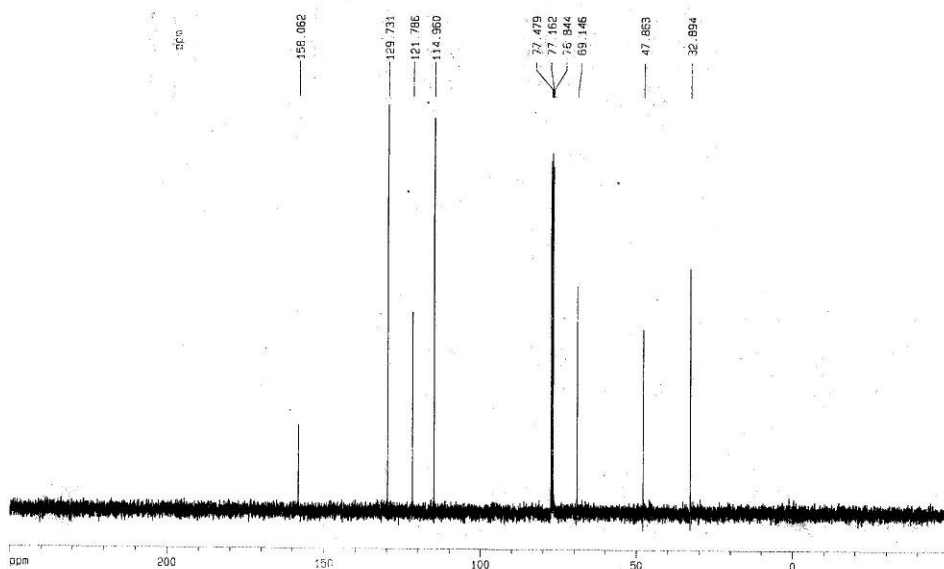
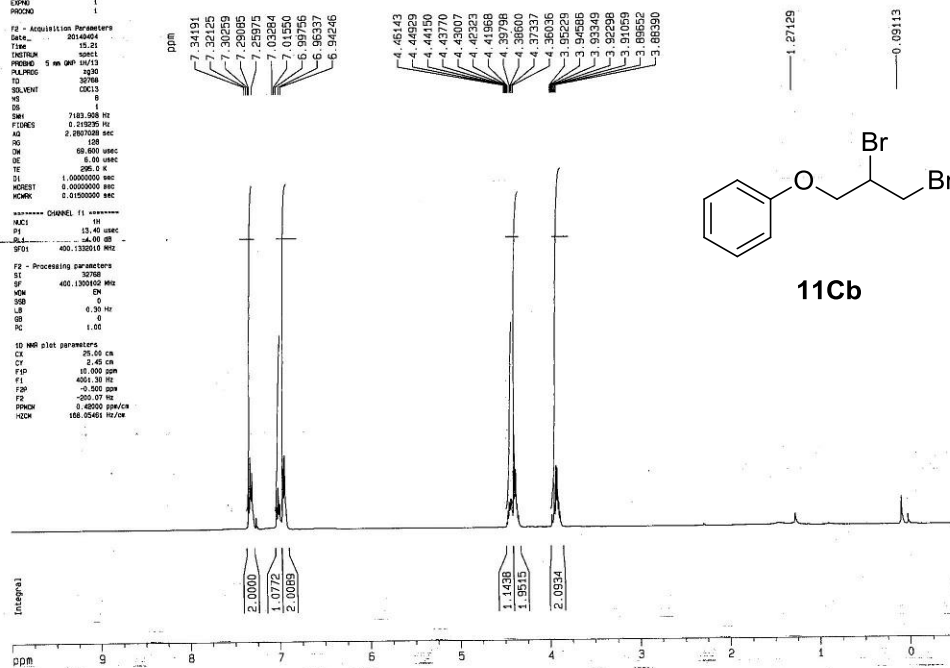
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PCPD2 60.00 usec
PL2 14.00 dB
PL12 12.00 dB
SFO2 400.150907 MHz

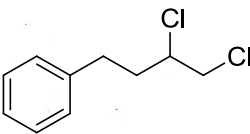
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GB 0
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1D NMR plot parameters
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F2 -1000.13 Hz
PRCK 6.000000 Hz/cm
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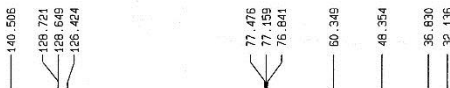


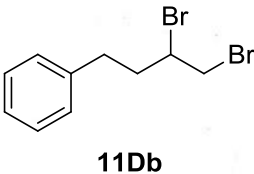
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 INSTRUM: spect
 PULPROG: zgpg30
 TO: 30768
 SOLVENT: CDCl3
 NS: 8
 DS: 4
 SWH: 7183.500 MHz
 FIDRES: 0.2190295 MHz
 AQ: 2.2801028 sec
 RG: 328
 DM: 69.600 usec
 DE: 8.80 usec
 TE: 296.2 K
 D1: 1.00000000 sec
 HOREST: 0.00000000 sec
 HOSHR: 0.01000000 sec
 ----- CHANNEL f1 -----
 NUC1: 1H
 P1: 13.40 usec
 PL1: -1.80 dB
 SFO1: 400.132610 MHz
 F2 - Processing parameters
 SI: 32768
 SF: 400.1306102 MHz
 KCM: 0
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 1.00
 1D NMR plot parameters
 CT: 32.00 cm
 CY: 2.40 cm
 FID: 15.000 ppm
 F1: 4001.30 Hz
 F2: -4.200 ppm
 F2: -200.07 Hz
 PPMH: 0.40000 ppm/cv
 NDC: 166.15461 Hz/cv



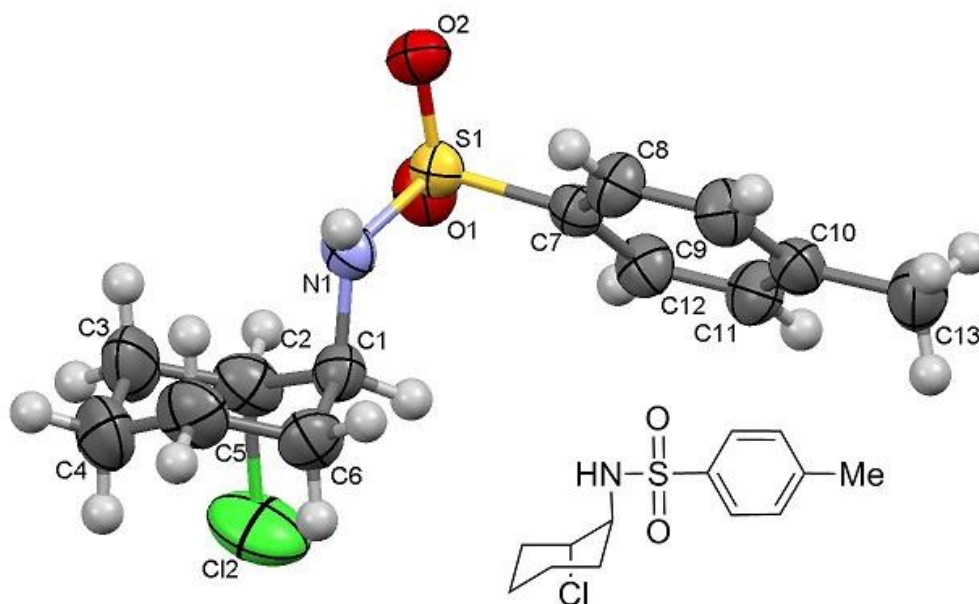


11Da

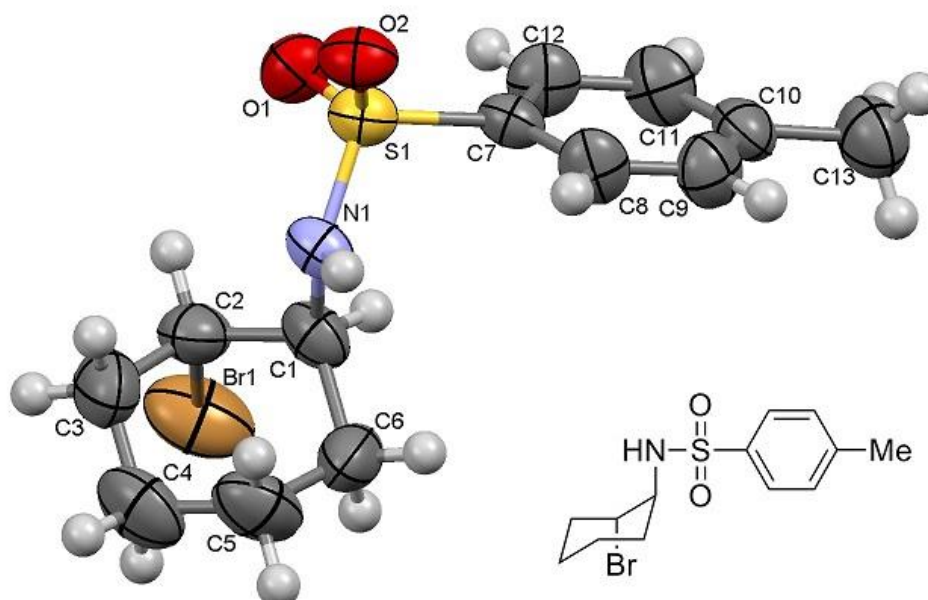




X-ray Structure Details



X-ray structure of *N*-(2-chlorocyclohexyl)-4-methylbenzenesulfonamide



X-ray structure of *N*-(2-bromocyclohexyl)-4-methylbenzenesulfonamide

Refinement

The structures were solved by direct methods, *SHELXS-97* (Sheldrick, 2008). All non-H atoms were refined anisotropically.

All of the C-bound H atoms were observable from difference Fourier map but were all placed at geometrical positions with C—H = 0.93, 0.96, 0.97 and 0.98 Å for phenyl, methyl, methylene and methine H-atoms. The N-bound H atom was located from difference Fourier map. All H-atoms were refined using riding model with $U_{\text{iso}}(\text{H}) = 1.2U_{\text{eq}}(\text{Carrier})$.

Highest peak is 0.44 at (0.5145, 0.9409, 0.6173) [0.96 Å from Cl1] Deepest hole is -0.48 at (0.2792, 0.0893, 0.3290) [0.70 Å from Cl1]

Computing details

Data collection: *APEX* (Bruker AXS Inc, 2007); cell refinement: *SAINT* v7.34A (Bruker AXS Inc, 2007); data reduction: *CrystalStructure* (Rigaku/MSC and Rigaku Corporation, 2006); program(s) used to solve structure: *SHELXS97* (Sheldrick, 2008); program(s) used to refine structure: *SHELXL97* (Sheldrick, 2008); molecular graphics: Mercury (Macrae *et al.*, 2008); software used to prepare material for publication: *SHELXL97* (Sheldrick, 2008).

References

- Bruker AXS Inc. (2007). *ApexII*, Madison, Wisconsin, USA.
- Macrae, C. F., Bruno, I. J., Chisholm, J. A., Edgington, P. R., McCabe, P., Pidcock, E., Rodriguez-Monge, L., Taylor, R., van de Streek, J. & Wood, P. A. (2008). *Mercury J. Appl. Cryst.* **41**, 466–470.
- Rigaku/MSC and Rigaku Corporation. (2006). *CrystalStructure*. Single Crystal Structure Analysis Software. Version 3.8.1. Rigaku/MSC, 9009 New Trails Drive, The Woodlands, TX, USA 77381-5209. Rigaku, 3-9-12 Akishima, Tokyo 196-8666, Japan.
- Sheldrick, G. M. (2008). *SADABS*, Göttingen University, Göttingen, Germany.
- Sheldrick, G. M. (2008). *SHELX* programs. (*SHELXL97*, *SHELXS97*.) *Acta Cryst.* **E68**, 112-122.

X-ray report of *N*-(2-chlorocyclohexyl)-4-methylbenzenesulfonamide

The compound *N*-(2-chlorocyclohexyl)-4-methylbenzenesulfonamide crystallized in a centrosymmetric triclinic primitive space group, *P* -1 (#2). There are two complex molecules in the unit cell. The cyclo-hexyl ring is in the chair form with the puckering amplitude (*Q*) of 0.535(4)Å, θ value of 176.3(4)° and ϕ value of 35(6)°. Both the chloride and the sulphonamide groups are at axial positions. The compound is racemic. The atoms C1 and C2 are of the same configuration.

All the bond parameters are comparable to its bromide analogue. C—Cl is 1.818(3)Å, S=O is 1.4268(18)–1.4365(18)Å; S—N is 1.616(2)Å.

There are only very weak $\pi \cdots \pi$ inter-actions between the neighbouring tolyl rings. The complementary pair of inter-molecular N1—H1N $\cdots$ O2 H-bonding inter-actions link the molecules into dimers. There is no residual solvent accessible void volume found in the unit cell.

Experimental

A colourless block crystal of, C₁₃H₁₈ClNO₂S, having approximate dimensions of 0.38 mm x 0.56 mm x 0.56 mm was mounted in glass capillary. All measurements were made on a Bruker *Apex* CCD detector with graphite monochromated Mo—K α radiation.

Cell constants and an orientation matrix for data collection corresponded to a primitive monoclinic cell with dimensions: *a* = 6.9260(2)Å, *b* = 9.4804(3)Å, *c* = 11.6065(4)Å, *V* = 714.23(4)Å³ α = 73.524(2)°, β = 77.837(2)° γ = 87.759(2)°.

For *Z* = 2 and F.W. = 287.79, the calculated density is 1.338 g/cm³. Based on a statistical analysis of intensity distribution, and the successful solution and refinement of the structure, the space group was determined to be: *P* -1 (#2)

The data were collected at a temperature of 23(1)°C to a maximum 2θ value of 25.02°.

Of the 11110 reflections that were collected, 2480 reflections were unique. (*R*_{int} = 0.0275); equivalent reflections were merged.

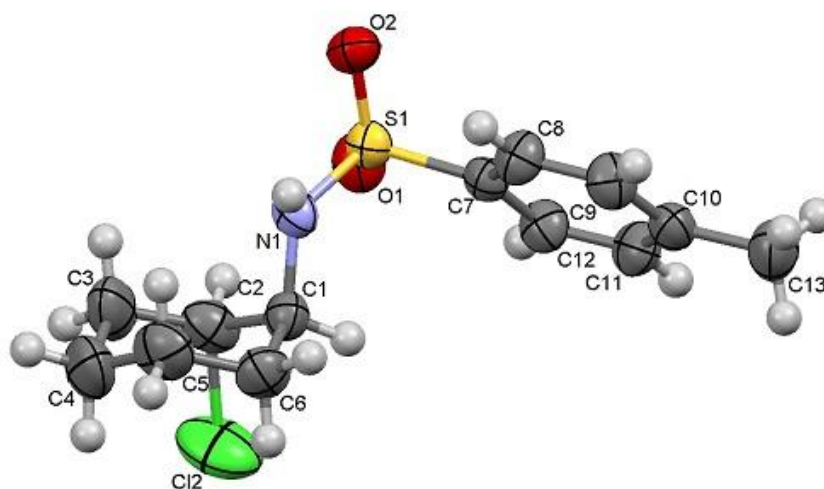


Figure 1 The compound shown at 50% probability thermal ellipsoids with the atom numbering scheme.

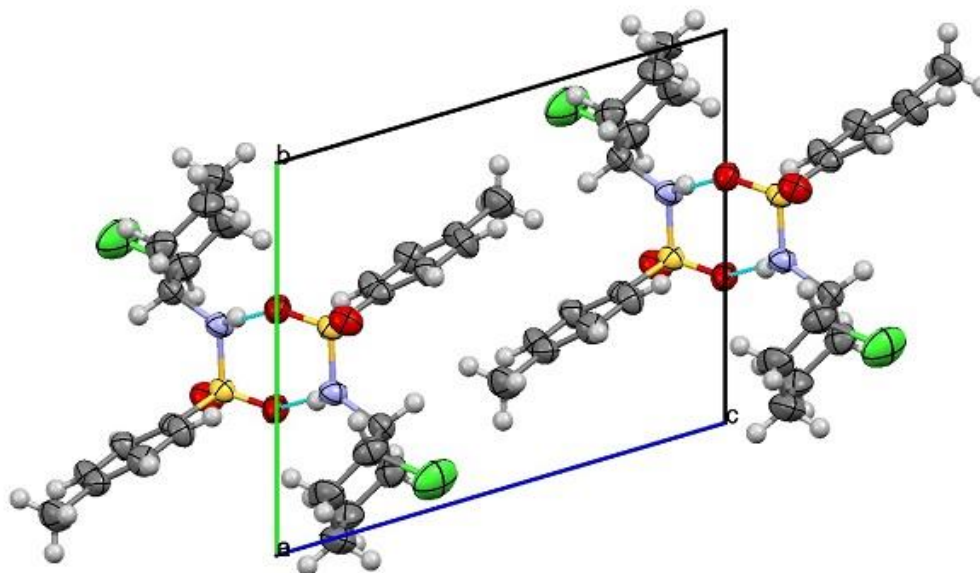


Figure 2 The packing diagram of the unit cell was projected along the *a* axis, shown at 50% probability thermal ellipsoids. The cyan dotted lines represent the complementary pair of intermolecular N1-H1N $\cdots$ O2 H-bonding interactions linking the molecules into dimers.

Crystal data

$C_{13}H_{18}ClNO_2S$	$Z = 2$
$M_r = 287.79$	$F(000) = 304$
Triclinic, $P\bar{1}$	$D_x = 1.338 \text{ Mg m}^{-3}$
Hall symbol: -P 1	Mo $K\alpha$ radiation, $\lambda = 0.71073 \text{ \AA}$
$a = 6.9260(2) \text{ \AA}$	Cell parameters from 11110 reflections
$b = 9.4804(3) \text{ \AA}$	$\theta = 1.9\text{--}25.0^\circ$
$c = 11.6065(4) \text{ \AA}$	$\mu = 0.41 \text{ mm}^{-1}$
$\alpha = 73.524(2)^\circ$	$T = 296 \text{ K}$
$\beta = 77.837(2)^\circ$	Block, colourless
$\gamma = 87.759(2)^\circ$	$0.56 \times 0.56 \times 0.38 \text{ mm}$
$V = 714.23(4) \text{ \AA}^3$	

Data collection

Bruker APEX CCD diffractometer	2480 independent reflections
Radiation source: fine-focus sealed tube	2119 reflections with $I > 2\sigma(I)$
graphite	$R_{\text{int}} = 0.028$
ω & ϕ scans	$\theta_{\text{max}} = 25.0^\circ$, $\theta_{\text{min}} = 1.9^\circ$
Absorption correction: multi-scan <i>SADABS</i> (Sheldrick, 2008)	$h = -8 \rightarrow 8$
$T_{\text{min}} = 0.804$, $T_{\text{max}} = 0.861$	$k = -11 \rightarrow 11$
11110 measured reflections	$l = -13 \rightarrow 13$

Refinement

Refinement on F^2	Primary atom site location: structure-invariant direct methods
Least-squares matrix: full	Secondary atom site location: difference Fourier map
$R[F^2 > 2\sigma(F^2)] = 0.045$	Hydrogen site location: inferred from neighbouring sites
$wR(F^2) = 0.127$	H-atom parameters constrained
$S = 1.06$	$w = 1/[\sigma^2(F_o^2) + (0.0607P)^2 + 0.4124P]$ where $P = (F_o^2 + 2F_c^2)/3$
2480 reflections	$(\Delta/\sigma)_{\text{max}} = 0.006$
164 parameters	$\Delta\rho_{\text{max}} = 0.44 \text{ e \AA}^{-3}$
0 restraints	$\Delta\rho_{\text{min}} = -0.48 \text{ e \AA}^{-3}$

Special details

Geometry. All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Refinement. Refinement of F^2 against ALL reflections. The weighted R-factor wR and goodness of fit S are based on F^2 , conventional R-factors R are based on F, with F set to zero for negative F^2 . The threshold expression of $F^2 > 2\sigma(F^2)$ is used only for calculating R-factors(gt) etc. and is not relevant to the choice of reflections for refinement. R-factors based on F^2 are statistically about twice as large as those based on F, and R-factors based on ALL data will be even larger.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters ($\text{\AA}^2$)

	x	y	z	$U_{\text{iso}}^*/U_{\text{eq}}$
Cl1	0.63511 (16)	0.92618 (12)	0.64754 (11)	0.1109 (4)
S1	0.28780 (9)	0.46056 (6)	0.87910 (6)	0.0428 (2)
O1	0.4977 (3)	0.4550 (2)	0.84511 (18)	0.0566 (5)
O2	0.1860 (3)	0.37354 (19)	0.99763 (16)	0.0521 (5)
N1	0.2296 (3)	0.6289 (2)	0.87325 (19)	0.0461 (5)
H1N	0.0941	0.6362	0.9106	0.055*
C1	0.3156 (4)	0.7518 (3)	0.7656 (2)	0.0487 (6)
H1	0.3512	0.7157	0.6929	0.058*
C2	0.5015 (4)	0.8056 (3)	0.7917 (3)	0.0621 (7)
H2	0.5832	0.7209	0.8187	0.075*
C3	0.4571 (6)	0.8854 (4)	0.8886 (3)	0.0798 (10)
H3A	0.5790	0.9275	0.8943	0.096*
H3B	0.4052	0.8154	0.9674	0.096*
C4	0.3092 (6)	1.0073 (4)	0.8630 (4)	0.0861 (11)
H4A	0.2784	1.0499	0.9313	0.103*
H4B	0.3672	1.0843	0.7899	0.103*

C5	0.1231 (5)	0.9493 (3)	0.8448 (3)	0.0741 (9)
H5A	0.0345	1.0299	0.8238	0.089*
H5B	0.0578	0.8802	0.9209	0.089*
C6	0.1654 (4)	0.8733 (3)	0.7437 (3)	0.0614 (7)
H6A	0.2151	0.9456	0.6659	0.074*
H6B	0.0432	0.8314	0.7383	0.074*
C7	0.1883 (3)	0.4103 (2)	0.7678 (2)	0.0409 (5)
C8	-0.0132 (4)	0.3866 (3)	0.7871 (2)	0.0513 (6)
H8	-0.0967	0.4013	0.8565	0.062*
C9	-0.0891 (4)	0.3411 (3)	0.7029 (3)	0.0563 (7)
H9	-0.2246	0.3248	0.7166	0.068*
C10	0.0314 (4)	0.3191 (3)	0.5979 (2)	0.0528 (7)
C11	0.2314 (4)	0.3457 (3)	0.5796 (3)	0.0591 (7)
H11	0.3144	0.3327	0.5093	0.071*
C12	0.3122 (4)	0.3913 (3)	0.6627 (2)	0.0512 (6)
H12	0.4475	0.4089	0.6484	0.061*
C13	-0.0555 (5)	0.2675 (4)	0.5079 (3)	0.0749 (9)
H13A	-0.0202	0.3364	0.4280	0.090*
H13B	-0.1967	0.2605	0.5339	0.090*
H13C	-0.0048	0.1727	0.5047	0.090*

Atomic displacement parameters ($\text{\AA}^2$)

	U^{11}	U^{22}	U^{33}	U^{12}	U^{13}	U^{23}
Cl1	0.0846 (7)	0.0878 (7)	0.1249 (9)	-0.0193 (5)	0.0215 (6)	-0.0025 (6)
S1	0.0387 (4)	0.0433 (3)	0.0469 (4)	0.0074 (2)	-0.0075 (3)	-0.0155 (3)
O1	0.0388 (10)	0.0639 (12)	0.0721 (13)	0.0100 (8)	-0.0116 (9)	-0.0283 (10)
O2	0.0547 (11)	0.0511 (10)	0.0458 (10)	0.0072 (8)	-0.0090 (8)	-0.0082 (8)
N1	0.0435 (12)	0.0427 (11)	0.0515 (12)	0.0032 (9)	-0.0023 (9)	-0.0182 (9)
C1	0.0539 (15)	0.0502 (14)	0.0452 (14)	0.0029 (12)	-0.0098 (12)	-0.0192 (11)
C2	0.0540 (17)	0.0529 (16)	0.077 (2)	0.0019 (13)	-0.0178 (14)	-0.0117 (14)
C3	0.101 (3)	0.067 (2)	0.089 (2)	-0.0015 (18)	-0.051 (2)	-0.0276 (18)
C4	0.128 (3)	0.0575 (18)	0.086 (2)	0.009 (2)	-0.032 (2)	-0.0351 (18)

C5	0.086 (2)	0.0538 (17)	0.081 (2)	0.0253 (16)	-0.0157 (18)	-0.0216 (16)
C6	0.0635 (18)	0.0592 (17)	0.0641 (18)	0.0080 (14)	-0.0281 (14)	-0.0116 (14)
C7	0.0411 (13)	0.0354 (12)	0.0458 (13)	0.0062 (10)	-0.0072 (10)	-0.0129 (10)
C8	0.0422 (14)	0.0623 (16)	0.0510 (15)	0.0070 (12)	-0.0053 (11)	-0.0222 (13)
C9	0.0444 (15)	0.0625 (17)	0.0648 (17)	0.0019 (12)	-0.0129 (13)	-0.0211 (14)
C10	0.0658 (18)	0.0428 (13)	0.0532 (15)	0.0041 (12)	-0.0183 (13)	-0.0148 (12)
C11	0.0670 (19)	0.0608 (17)	0.0505 (16)	0.0057 (14)	-0.0015 (13)	-0.0254 (13)
C12	0.0441 (15)	0.0547 (15)	0.0543 (16)	0.0035 (12)	-0.0015 (12)	-0.0209 (12)
C13	0.094 (2)	0.070 (2)	0.074 (2)	0.0020 (18)	-0.0320 (19)	-0.0303 (17)

Geometric parameters (Å, °)

Cl1—C2	1.818 (3)	C5—H5A	0.9700
S1—O1	1.4268 (18)	C5—H5B	0.9700
S1—O2	1.4365 (18)	C6—H6A	0.9700
S1—N1	1.616 (2)	C6—H6B	0.9700
S1—C7	1.767 (2)	C7—C8	1.383 (3)
N1—C1	1.480 (3)	C7—C12	1.388 (3)
N1—H1N	0.9561	C8—C9	1.374 (4)
C1—C2	1.520 (4)	C8—H8	0.9300
C1—C6	1.525 (4)	C9—C10	1.389 (4)
C1—H1	0.9800	C9—H9	0.9300
C2—C3	1.501 (5)	C10—C11	1.379 (4)
C2—H2	0.9800	C10—C13	1.506 (4)
C3—C4	1.521 (5)	C11—C12	1.383 (4)
C3—H3A	0.9700	C11—H11	0.9300
C3—H3B	0.9700	C12—H12	0.9300
C4—C5	1.499 (5)	C13—H13A	0.9600
C4—H4A	0.9700	C13—H13B	0.9600
C4—H4B	0.9700	C13—H13C	0.9600
C5—C6	1.516 (4)		
O1—S1—O2	119.78 (11)	C6—C5—H5A	109.4

O1—S1—N1	108.11 (11)	C4—C5—H5B	109.4
O2—S1—N1	105.61 (11)	C6—C5—H5B	109.4
O1—S1—C7	107.50 (11)	H5A—C5—H5B	108.0
O2—S1—C7	107.29 (11)	C5—C6—C1	112.5 (2)
N1—S1—C7	108.09 (11)	C5—C6—H6A	109.1
C1—N1—S1	121.06 (16)	C1—C6—H6A	109.1
C1—N1—H1N	116.9	C5—C6—H6B	109.1
S1—N1—H1N	111.7	C1—C6—H6B	109.1
N1—C1—C2	107.4 (2)	H6A—C6—H6B	107.8
N1—C1—C6	110.0 (2)	C8—C7—C12	120.1 (2)
C2—C1—C6	111.9 (2)	C8—C7—S1	119.70 (19)
N1—C1—H1	109.2	C12—C7—S1	120.18 (19)
C2—C1—H1	109.2	C9—C8—C7	119.5 (2)
C6—C1—H1	109.2	C9—C8—H8	120.2
C3—C2—C1	112.6 (3)	C7—C8—H8	120.2
C3—C2—Cl1	110.5 (2)	C8—C9—C10	121.7 (3)
C1—C2—Cl1	107.0 (2)	C8—C9—H9	119.2
C3—C2—H2	108.9	C10—C9—H9	119.2
C1—C2—H2	108.9	C11—C10—C9	117.8 (2)
Cl1—C2—H2	108.9	C11—C10—C13	121.7 (3)
C2—C3—C4	113.0 (3)	C9—C10—C13	120.6 (3)
C2—C3—H3A	109.0	C10—C11—C12	121.9 (2)
C4—C3—H3A	109.0	C10—C11—H11	119.1
C2—C3—H3B	109.0	C12—C11—H11	119.1
C4—C3—H3B	109.0	C11—C12—C7	119.0 (2)
H3A—C3—H3B	107.8	C11—C12—H12	120.5
C5—C4—C3	110.9 (3)	C7—C12—H12	120.5
C5—C4—H4A	109.5	C10—C13—H13A	109.5
C3—C4—H4A	109.5	C10—C13—H13B	109.5
C5—C4—H4B	109.5	H13A—C13—H13B	109.5
C3—C4—H4B	109.5	C10—C13—H13C	109.5
H4A—C4—H4B	108.0	H13A—C13—H13C	109.5

C4—C5—C6	111.4 (3)	H13B—C13—H13C	109.5
C4—C5—H5A	109.4		

Hydrogen-bond geometry (Å, °)

<i>D</i> —H $\cdots$ <i>A</i>	<i>D</i> —H	H $\cdots$ <i>A</i>	<i>D</i> $\cdots$ <i>A</i>	<i>D</i> —H $\cdots$ <i>A</i>
N1—H1N $\cdots$ O2 ⁱ	0.96	2.00	2.949 (3)	172.9
C2—H2 $\cdots$ O1	0.98	2.53	3.204 (3)	126
C12—H12 $\cdots$ O1	0.93	2.54	2.903 (3)	104

Symmetry code: (i) -*x*, -*y*+1, -*z*+2.

X-ray report of *N*-(2-bromocyclohexyl)-4-methylbenzenesulfonamide

The compound *N*-(2-bromocyclohexyl)-4-methylbenzenesulfonamide crystallized in a centrosymmetric monoclinic primitive space group, *P* 2₁/c (# 14). There are a total of four complex molecules in the unit cell. The cyclo-hexyl ring is in the chair form with the puckering amplitude (*Q*) of 0.529(9)Å, θ value of 176.1(10)° and ϕ value of 59(12)°. Both the bromide and the sulphonamide groups are at axial positions. The compound is racemic. The atoms C1 and C2 are of the same configuration.

All the bond parameters are comparable within normal ranges. C—Br is 1.985(7)Å, S=O is 1.429(5)–1.441(4)Å; S—N is 1.615(5)Å.

There are weak $\pi\cdots\pi$ inter-actions between the neighbouring tolyl rings. Inter-molecular H-bonding inter-actions present. There is no residual solvent accessible void volume found in the unit cell.

Experimental

A colourless plate crystal of, C₁₃H₁₈BrNO₂S, having approximate dimensions of 0.14 mm x 0.26 mm x 0.38 mm was mounted in glass capillary. All measurements were made on a Bruker Apex CCD detector with graphite monochromated Mo—K α radiation.

Cell constants and an orientation matrix for data collection corresponded to a primitive monoclinic cell with dimensions: *a* = 10.8559(4)Å, *b* = 13.6017(5)Å, *c* = 10.5811(4)Å, *V* = 1482.46(10)Å³ β = 108.406(2)°.

For $Z = 4$ and F.W. = 332.25, the calculated density is 1.489 g/cm^3 . Based on a statistical analysis of intensity distribution, and the successful solution and refinement of the structure, the space group was determined to be: $P 2_1/c$ (#14)

The data were collected at a temperature of $23(1)^\circ\text{C}$ to a maximum 2θ value of 66.1° .

Of the 17462 reflections that were collected, 2616 reflections were unique. ($R_{\text{int}} = 0.0738$); equivalent reflections were merged.

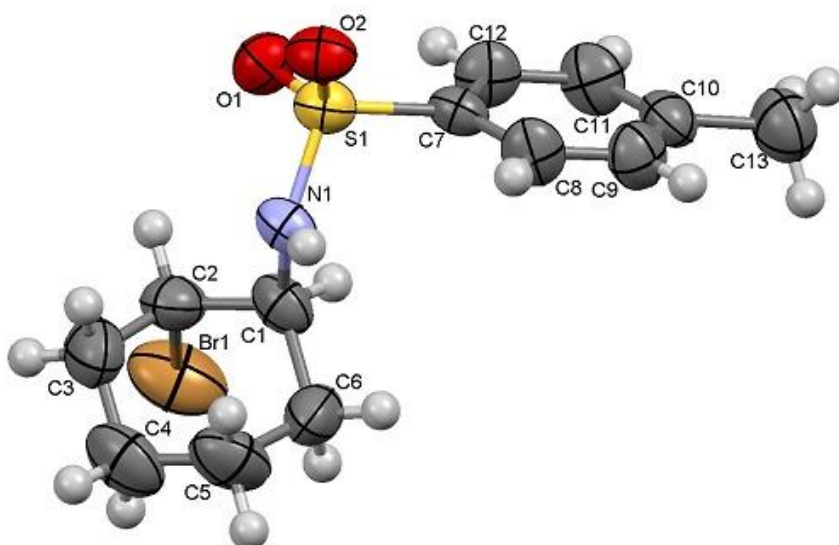


Figure 3 The ORTEP plot of the compound was shown at 50% probability thermal ellipsoids with the atom numbering scheme.

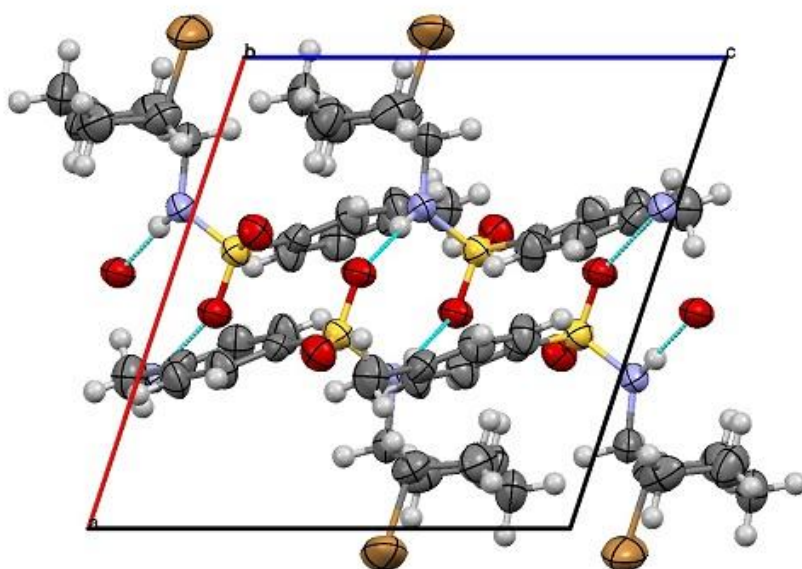


Figure 4 The packing diagram of the unit cell was shown at 50% probability thermal ellipsoids. The cyan dotted line represent the complementary pair of intermolecular N1-H1N...O2 H-bonding interactions linking the molecules into dimers.

Crystal data

$C_{13}H_{18}BrNO_2S$	$F(000) = 680$
$M_r = 332.25$	$D_x = 1.489 \text{ Mg m}^{-3}$
Monoclinic, $P2_1/c$	Mo $K\alpha$ radiation, $\lambda = 0.71073 \text{ \AA}$
Hall symbol: -P 2ybc	Cell parameters from 17462 reflections
$a = 10.8559(4) \text{ \AA}$	$\theta = 1.5\text{--}25.0^\circ$
$b = 13.6017(5) \text{ \AA}$	$\mu = 2.91 \text{ mm}^{-1}$
$c = 10.5811(4) \text{ \AA}$	$T = 296 \text{ K}$
$\beta = 108.406(2)^\circ$	Plate, colourless
$V = 1482.46(10) \text{ \AA}^3$	$0.38 \times 0.26 \times 0.14 \text{ mm}$
$Z = 4$	

Data collection

Bruker APEX CCD diffractometer	2616 independent reflections
Radiation source: fine-focus sealed tube	1601 reflections with $I > 2\sigma(I)$
graphite	$R_{\text{int}} = 0.074$
ω & ϕ scans	$\theta_{\text{max}} = 25.0^\circ$, $q_{\text{min}} = 2.0^\circ$
Absorption correction: multi-scan <i>SADABS</i> (Sheldrick, 2008)	$h = -11 \rightarrow 12$
$T_{\text{min}} = 0.405$, $T_{\text{max}} = 0.686$	$k = -16 \rightarrow 16$
17462 measured reflections	$l = -12 \rightarrow 12$

Refinement

Refinement on F^2	Primary atom site location: structure-invariant direct methods
Least-squares matrix: full	Secondary atom site location: difference Fourier map
$R[F^2 > 2\sigma(F^2)] = 0.068$	Hydrogen site location: inferred from neighbouring sites
$wR(F^2) = 0.201$	H-atom parameters constrained
$S = 1.04$	$w = 1/[\sigma^2(F_o^2) + (0.0872P)^2 + 3.5474P]$ where $P = (F_o^2 + 2F_c^2)/3$
2616 reflections	$(\Delta/\sigma)_{\text{max}} = 0.019$
164 parameters	$\Delta\rho_{\text{max}} = 1.55 \text{ e } \text{\AA}^{-3}$
0 restraints	$\Delta\rho_{\text{min}} = -0.81 \text{ e } \text{\AA}^{-3}$

Special details

Geometry. All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Refinement. Refinement of F^2 against ALL reflections. The weighted R-factor wR and goodness of fit S are based on F^2 , conventional R-factors R are based on F , with F set to zero for negative F^2 . The threshold expression of $F^2 > 2\sigma(F^2)$ is used only for calculating R-factors(gt) etc. and is not relevant to the choice of reflections for refinement. R-factors based on F^2 are statistically about twice as large as those based on F , and R-factors based on ALL data will be even larger.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters ($\text{\AA}^2$)

	x	y	z	$U_{\text{iso}}^*/U_{\text{eq}}$
Br1	1.05259 (9)	0.31013 (9)	0.63054 (10)	0.1096 (5)
S1	0.58845 (16)	0.39459 (12)	0.38391 (16)	0.0520 (5)
O1	0.6280 (5)	0.2988 (3)	0.3570 (5)	0.0646 (12)
O2	0.4575 (4)	0.4086 (3)	0.3858 (4)	0.0632 (12)
N1	0.6808 (5)	0.4284 (4)	0.5293 (5)	0.0537 (13)
H1N	0.6400	0.4783	0.5605	0.064*
C1	0.8245 (7)	0.4251 (5)	0.5627 (6)	0.0612 (18)
H1	0.8472	0.4297	0.4803	0.073*
C2	0.8705 (7)	0.3288 (5)	0.6279 (7)	0.0683 (19)
H2	0.8172	0.2766	0.5735	0.082*
C3	0.8607 (9)	0.3220 (6)	0.7644 (8)	0.084 (2)
H3A	0.9024	0.2618	0.8056	0.101*
H3B	0.7698	0.3180	0.7584	0.101*
C4	0.9212 (9)	0.4070 (8)	0.8520 (7)	0.094 (3)
H4A	0.9050	0.4007	0.9367	0.113*
H4B	1.0144	0.4059	0.8691	0.113*
C5	0.8689 (9)	0.5011 (7)	0.7901 (8)	0.091 (3)
H5A	0.9133	0.5546	0.8471	0.109*
H5B	0.7774	0.5051	0.7816	0.109*
C6	0.8855 (8)	0.5124 (6)	0.6535 (8)	0.081 (2)
H6A	0.9772	0.5163	0.6630	0.097*
H6B	0.8447	0.5730	0.6129	0.097*
C7	0.6156 (6)	0.4763 (5)	0.2655 (5)	0.0485 (15)
C8	0.5881 (7)	0.5746 (5)	0.2724 (7)	0.0647 (19)
H8	0.5562	0.5977	0.3387	0.078*
C9	0.6087 (7)	0.6380 (5)	0.1799 (7)	0.0677 (19)
H9	0.5903	0.7044	0.1843	0.081*
C10	0.6559 (7)	0.6054 (6)	0.0807 (6)	0.0620 (18)
C11	0.6824 (7)	0.5081 (6)	0.0772 (7)	0.071 (2)
H11	0.7148	0.4853	0.0111	0.085*

C12	0.6632 (7)	0.4419 (5)	0.1674 (7)	0.0630 (18)
H12	0.6819	0.3755	0.1624	0.076*
C13	0.6785 (9)	0.6785 (6)	-0.0179 (8)	0.089 (3)
H13A	0.7094	0.6444	-0.0814	0.106*
H13B	0.7419	0.7260	0.0288	0.106*
H13C	0.5985	0.7113	-0.0635	0.106*

Atomic displacement parameters (Å²)

	U^{11}	U^{22}	U^{33}	U^{12}	U^{13}	U^{23}
Br1	0.0643 (6)	0.1545 (11)	0.1021 (8)	0.0326 (6)	0.0149 (5)	-0.0239 (6)
S1	0.0471 (9)	0.0586 (10)	0.0518 (9)	0.0023 (7)	0.0177 (7)	-0.0078 (8)
O1	0.069 (3)	0.057 (3)	0.071 (3)	0.001 (2)	0.028 (2)	-0.007 (2)
O2	0.046 (3)	0.079 (3)	0.065 (3)	-0.002 (2)	0.019 (2)	-0.013 (2)
N1	0.046 (3)	0.074 (4)	0.043 (3)	0.008 (3)	0.015 (2)	-0.002 (2)
C1	0.061 (4)	0.081 (5)	0.043 (4)	0.006 (4)	0.018 (3)	0.003 (3)
C2	0.058 (4)	0.072 (5)	0.065 (5)	-0.007 (4)	0.007 (3)	-0.006 (4)
C3	0.083 (6)	0.089 (6)	0.080 (6)	-0.011 (5)	0.024 (4)	0.021 (5)
C4	0.095 (7)	0.136 (9)	0.052 (5)	0.010 (6)	0.025 (4)	0.000 (5)
C5	0.099 (7)	0.101 (7)	0.063 (5)	0.008 (5)	0.010 (5)	-0.023 (5)
C6	0.073 (5)	0.063 (5)	0.091 (6)	-0.013 (4)	0.005 (4)	0.015 (4)
C7	0.042 (3)	0.062 (4)	0.039 (3)	0.002 (3)	0.010 (3)	-0.006 (3)
C8	0.085 (5)	0.059 (5)	0.058 (4)	0.017 (4)	0.033 (4)	-0.008 (3)
C9	0.079 (5)	0.056 (4)	0.069 (5)	0.014 (4)	0.025 (4)	0.003 (4)
C10	0.055 (4)	0.076 (5)	0.048 (4)	0.002 (4)	0.007 (3)	0.008 (4)
C11	0.081 (5)	0.084 (6)	0.060 (4)	0.010 (4)	0.040 (4)	-0.001 (4)
C12	0.073 (5)	0.058 (4)	0.067 (4)	0.012 (3)	0.034 (4)	-0.010 (4)
C13	0.099 (7)	0.097 (6)	0.071 (5)	0.001 (5)	0.029 (5)	0.013 (5)

Geometric parameters (Å, °)

Br1—C2	1.985 (7)	C5—H5A	0.9700
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S1—O1	1.429 (5)	C5—H5B	0.9700
S1—O2	1.441 (4)	C6—H6A	0.9700
S1—N1	1.615 (5)	C6—H6B	0.9700
S1—C7	1.768 (6)	C7—C8	1.377 (9)
N1—C1	1.488 (8)	C7—C12	1.380 (8)
N1—H1N	0.9264	C8—C9	1.375 (9)
C1—C2	1.491 (10)	C8—H8	0.9300
C1—C6	1.539 (10)	C9—C10	1.378 (10)
C1—H1	0.9800	C9—H9	0.9300
C2—C3	1.485 (11)	C10—C11	1.357 (10)
C2—H2	0.9800	C10—C13	1.517 (10)
C3—C4	1.497 (12)	C11—C12	1.375 (10)
C3—H3A	0.9700	C11—H11	0.9300
C3—H3B	0.9700	C12—H12	0.9300
C4—C5	1.468 (12)	C13—H13A	0.9600
C4—H4A	0.9700	C13—H13B	0.9600
C4—H4B	0.9700	C13—H13C	0.9600
C5—C6	1.520 (11)		
O1—S1—O2	118.9 (3)	C6—C5—H5A	109.3
O1—S1—N1	108.3 (3)	C4—C5—H5B	109.3
O2—S1—N1	105.5 (3)	C6—C5—H5B	109.3
O1—S1—C7	107.7 (3)	H5A—C5—H5B	108.0
O2—S1—C7	108.2 (3)	C5—C6—C1	111.1 (6)
N1—S1—C7	107.7 (3)	C5—C6—H6A	109.4
C1—N1—S1	120.4 (4)	C1—C6—H6A	109.4
C1—N1—H1N	121.4	C5—C6—H6B	109.4
S1—N1—H1N	109.0	C1—C6—H6B	109.4
N1—C1—C2	108.0 (6)	H6A—C6—H6B	108.0
N1—C1—C6	109.7 (6)	C8—C7—C12	120.6 (6)
C2—C1—C6	111.9 (6)	C8—C7—S1	119.0 (5)
N1—C1—H1	109.1	C12—C7—S1	120.4 (5)

C2—C1—H1	109.1	C9—C8—C7	119.0 (6)
C6—C1—H1	109.1	C9—C8—H8	120.5
C3—C2—C1	112.9 (6)	C7—C8—H8	120.5
C3—C2—Br1	110.8 (5)	C8—C9—C10	121.6 (7)
C1—C2—Br1	107.8 (5)	C8—C9—H9	119.2
C3—C2—H2	108.4	C10—C9—H9	119.2
C1—C2—H2	108.4	C11—C10—C9	117.9 (7)
Br1—C2—H2	108.4	C11—C10—C13	122.6 (7)
C2—C3—C4	113.7 (7)	C9—C10—C13	119.5 (7)
C2—C3—H3A	108.8	C10—C11—C12	122.6 (6)
C4—C3—H3A	108.8	C10—C11—H11	118.7
C2—C3—H3B	108.8	C12—C11—H11	118.7
C4—C3—H3B	108.8	C11—C12—C7	118.4 (6)
H3A—C3—H3B	107.7	C11—C12—H12	120.8
C5—C4—C3	111.4 (7)	C7—C12—H12	120.8
C5—C4—H4A	109.3	C10—C13—H13A	109.5
C3—C4—H4A	109.3	C10—C13—H13B	109.5
C5—C4—H4B	109.3	H13A—C13—H13B	109.5
C3—C4—H4B	109.3	C10—C13—H13C	109.5
H4A—C4—H4B	108.0	H13A—C13—H13C	109.5
C4—C5—C6	111.5 (7)	H13B—C13—H13C	109.5
C4—C5—H5A	109.3		

Hydrogen-bond geometry (Å, °)

<i>D</i> —H $\cdots$ <i>A</i>	<i>D</i> —H	H $\cdots$ <i>A</i>	<i>D</i> $\cdots$ <i>A</i>	<i>D</i> —H $\cdots$ <i>A</i>
N1—H1N $\cdots$ O2 ⁱ	0.93	2.05	2.971 (7)	175.5
C2—H2 $\cdots$ O1	0.98	2.57	3.248(9)	127
C12—H12 $\cdots$ O1	0.93	2.54	2.906(8)	104

Symmetry code: (i) -x+1, -y+1, -z+1.