

# Supporting Information

for

## **Pd/C-catalyzed aerobic oxidative esterification of alcohols and aldehydes: a highly efficient microwave-assisted green protocol**

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### **Copies of GC–MS chromatograms, MS, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra**

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## General experimental methods

All commercially available reagents and solvents were used without further purification. Pd sources from Sigma-Aldrich included 10 wt % Pd/C (Cat. No.75990) and Pd(OAc)<sub>2</sub> and PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> from Merck.

NMR spectra were recorded with a Bruker 300 Avance (300 MHz and 75 MHz for <sup>1</sup>H and <sup>13</sup>C, respectively) at 25 °C. Chemical shifts were calibrated to the residual proton and carbon resonances of the solvent; CDCl<sub>3</sub> (δ<sub>H</sub> = 7.26, δ<sub>C</sub> = 77.16).

GC–MS analyses were performed in a GC Agilent 6850 (Agilent Technologies, Palo Alto, CA, USA) that was fitted with a mass detector Agilent Network 5973, using a 30 m long capillary column HP 5-MS (5% phenyl methyl siloxane, i.d. 0.25 mm, film thickness 0.25 μm). GC conditions were: injection split 1:20, injector temperature 250 °C, detector temperature 280 °C. Carrier gas: helium (1.2 mL/min), temperature program: from 50 °C (held for 3 min) to 80 °C at 3 °C/min and to 300 °C at 10 °C/min.

Reactions were carried out in a professional MW reactor SynthWave (MLS GmbH, Milestone S.r.l.). This device enables high power density (1.5 kW/L) and inert barosphere and the possibility of simultaneously carrying out multiple reactions with gas insertion.

The sonochemical device was developed in collaboration with Danacamerini sas (Torino, Italy).

### General procedure for Pd/C catalyst regeneration

Pd/C filtered after a first cycle of oxidative esterification of benzylalcohol was regenerated. 50 mg were dispersed in toluene (1.45 mL). The reaction was carried out under magnetic stirring in a MW reactor SynthWave. The reactor was loaded with H<sub>2</sub> (6 bar) pressure followed by addition of N<sub>2</sub> up to 20 bar total pressure. The reaction was left at 60 °C for 8 hours (average power 150 W). The mixture was then filtered and the catalyst was recovered (0.042g).

### General procedure for the synthesis of Pd(OAc)<sub>2</sub>/C catalyst

Activated charcoal (0.200 g) was dispersed in MeOH (5 mL) and a solution of Pd(OAc)<sub>2</sub> (0.01 g) in MeOH (2.5 mL) was added to dropwise.

The reaction mixture was stirred at rt for 4 hours until the reaction mixture was colorless. The mixture was then filtered and washed with MeOH. The supported catalyst Pd(OAc)<sub>2</sub>/C was recovered (0.21 g).

**Table S1:** Oxidative esterification of benzylalcohol in the presence of methanol over different bases.

Entry	Pd catalyst	Base	Conversion [%]	<b>1</b> <sup>a</sup> [%]	<b>2</b> <sup>a</sup> [%]
1 <sup>a</sup>	Pd(OAc) <sub>2</sub>	K <sub>2</sub> CO <sub>3</sub>	100	–	100
2 <sup>a</sup>	Pd(OAc) <sub>2</sub>	Na <sub>2</sub> CO <sub>3</sub>	100	17	66
3 <sup>a</sup>	Pd(OAc) <sub>2</sub>	Li <sub>2</sub> CO <sub>3</sub>	95	59	36
4	Pd(OAc) <sub>2</sub>	Cs <sub>2</sub> CO <sub>3</sub>	94	73	21
5	Pd(OAc) <sub>2</sub>	<i>t</i> -BuOK	100	43	57
6	Pd(OAc) <sub>2</sub>	NEt <sub>3</sub>	42	30	7
7 <sup>b</sup>	Pd(OAc) <sub>2</sub>	KOMe	91	34	55
8 <sup>b</sup>	10% Pd/C	K <sub>2</sub> CO <sub>3</sub>	100	–	100
9 <sup>b,c</sup>	10% Pd/C	Na <sub>2</sub> CO <sub>3</sub>	100	–	100
10 <sup>b</sup>	10% Pd/C	Li <sub>2</sub> CO <sub>3</sub>	32	25	7
11 <sup>b</sup>	10% Pd/C	Cs <sub>2</sub> CO <sub>3</sub>	3	3	–
12 <sup>b</sup>	10% Pd/C	<i>t</i> -BuOK	12	10	3
13 <sup>b</sup>	10% Pd/C	NEt <sub>3</sub>	0	–	–
14 <sup>b</sup>	10% Pd/C	KOMe	87	44	43
15 <sup>d</sup>	5%Pd(OAc) <sub>2</sub> /C	K <sub>2</sub> CO <sub>3</sub>	100	11	89
16 <sup>d</sup>	5%Pd(OAc) <sub>2</sub> /C	Na <sub>2</sub> CO <sub>3</sub>	94	57	37
17 <sup>b</sup>	5%Pd(OAc) <sub>2</sub> /C	Li <sub>2</sub> CO <sub>3</sub>	96	56	38
18 <sup>b</sup>	5%Pd(OAc) <sub>2</sub> /C	Cs <sub>2</sub> CO <sub>3</sub>	20	18	2
19 <sup>b</sup>	5%Pd(OAc) <sub>2</sub> /C	<i>t</i> -BuOK	85	45	40
20 <sup>b</sup>	5%Pd(OAc) <sub>2</sub> /C	NEt <sub>3</sub>	0	–	–
21 <sup>b</sup>	5%Pd(OAc) <sub>2</sub> /C	KOMe	100	2	98

Reaction conditions: 1 mL MeOH, 0.1 g benzylalcohol, 2 equiv base, catalyst (5% Pd/mole of substrate), 2.5 bar O<sub>2</sub>/17.5 bar N<sub>2</sub>, MW reactor SynthWave, T = 90 °C (unless otherwise noted), 1 h.

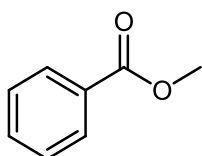
<sup>a</sup> Determined by GC–MS.

<sup>b</sup> The mixture of the base in MeOH was sonicated in an US bath for 10 s prior to the addition of the catalyst and the substrate.

<sup>c</sup> T = 100 °C.

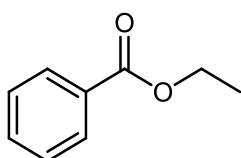
<sup>d</sup> 10% Pd/mole of substrate was added.

## Compound characterization



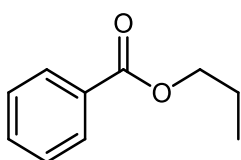
**Methyl benzoate (2a)**

General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with benzaldehyde (0.1 g, 0.942 mmol) to yield the desired ester as a colorless oil (0.126 g, 98%). When synthesized starting from benzylalcohol (0.1 g, 0.925 mmol), the reaction was performed at 100 °C for 1 h to yield the desired ester as a colorless oil (0.124 g, 98%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>8</sub>H<sub>8</sub>O<sub>2</sub> 136.05, found 136, 105, 77, 51; *t<sub>R</sub>* = 14.470 min.



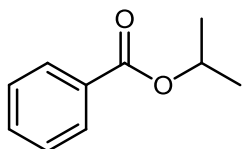
**Ethyl benzoate (2b)**

General procedure for aerobic alcohol esterification was performed in EtOH at 120 °C for 1.5 h with benzylalcohol (0.1 g, 0.925 mmol) in EtOH to yield the desired ester as a colorless oil (0.136 g, 98%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>9</sub>H<sub>10</sub>O<sub>2</sub> 150.07, found 150, 122, 105, 77, 51; *t<sub>R</sub>* = 16.538 min.



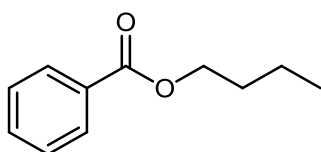
**Propyl benzoate (2c)**

General procedure for aerobic alcohol esterification was performed in *n*-PrOH at 120 °C for 1.5 h with benzylalcohol (0.1 g, 0.925 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired ester as a colorless oil (0.120 g, 79%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>2</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>12</sub>O<sub>2</sub> 164.08, found 164, 123, 105, 77, 51; *t<sub>R</sub>* = 18.562 min.



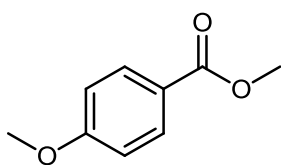
### Isopropyl benzoate (2d)

General procedure for aerobic alcohol esterification was performed in *i*PrOH at 120 °C for 1.5 h with benzylalcohol (0.1 g, 0.925 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired ester as a colorless oil (0.119 g, 78%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>3</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>12</sub>O<sub>2</sub> 164.08, found 164, 123, 105, 77, 51, 44, 32, 28, 18; *t<sub>R</sub>* = 17.335 min.



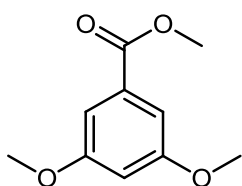
### Butyl benzoate (2e)

General procedure for aerobic alcohol esterification was performed in *n*-BuOH at 120 °C for 1.5 h with benzylalcohol (0.1 g, 0.925 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired ester as a colorless oil (0.126 g, 76%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>4</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>14</sub>O<sub>2</sub> 178.10, found 178, 123, 105, 77, 51; *t<sub>R</sub>* = 20.244 min.



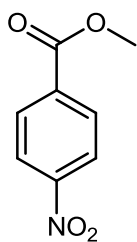
### Methyl 4-methoxybenzoate (3)

General procedure for aerobic aldehyde esterification was performed at 90 °C for 2 h with 4-methoxybenzaldehyde (0.1 g, 0.734 mmol) to yield the desired ester as a colorless oil (0.120 g, 98%). When synthesized starting from 4-methoxybenzylalcohol (0.1 g, 0.724 mmol), the reaction was performed at 120 °C for 1.5 h to yield the desired ester as a colorless oil (0.124 g, 98%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>9</sub>H<sub>10</sub>O<sub>3</sub> 166.06, found 166, 135, 107, 91, 77; *t<sub>R</sub>* = 20.270 min.



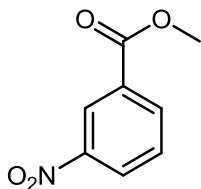
### Methyl 3,5-dimethoxybenzoate (4)

General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 3,5-dimethoxybenzaldehyde (0.1 g, 0.602 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.0354 g, 50%). When synthesized starting from 4-methoxybenzylalcohol (0.1 g, 0.724 mmol), the reaction was performed at 120 °C for 1.5 h and the same work-up purification was carried out. The desired ester was obtained as a colorless oil (0.0512 g, 36%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>5</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>12</sub>O<sub>4</sub> 196.07, found 196, 165, 138, 122, 107, 77, 63; *t<sub>R</sub>* = 22.995 min.



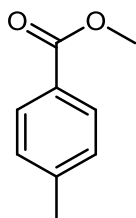
**Methyl 4-nitrobenzoate (5)**

General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 4-nitrobenzaldehyde (0.1 g, 0.662 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.113 g, 94%). When synthesized starting from 4-nitrobenzylalcohol (0.1 g, 0.653 mmol), the reaction was performed at 90 °C for 2 h and the same work-up purification was carried out. The desired ester was obtained as a colorless oil (0.0522 g, 44%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>5</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>12</sub>O<sub>4</sub> 196.07, found 196, 165, 138, 122, 107, 77, 63; *t<sub>R</sub>* = 22.995 min. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>6</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>8</sub>H<sub>7</sub>NO<sub>4</sub> 181.04, found 181, 164, 150, 104, 92, 76, 50; *t<sub>R</sub>* = 21.497 min.



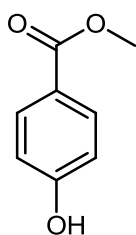
#### Methyl 3-nitrobenzoate (6)

General procedure for aerobic aldehyde esterification was performed at 90 °C for 2 h with 3-nitrobenzaldehyde (0.1 g, 0.662 mmol) to yield the desired ester as a pale yellow powder (0.118 g, 98%). When synthesized starting from 3-nitrobenzylalcohol (0.1 g, 0.653 mmol), the reaction was performed at 100 °C for 2 h. The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a pale yellow powder (0.0686 g, 58%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>7</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>8</sub>H<sub>7</sub>NO<sub>4</sub> 181.04, found 181, 150, 135, 119, 104, 92, 76, 50; *t<sub>R</sub>* = 21.497 min.



#### Methyl 4-methylbenzoate (7)

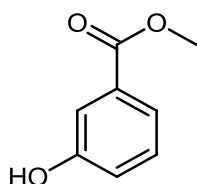
General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 4-methylbenzaldehyde (0.1 g, 0.832 mmol) to yield the desired ester as a colorless oil (0.123 g, 98%). When synthesized starting from 4-methylbenzylalcohol (0.1 g, 0.819 mmol), the reaction was performed at 120 °C for 1.5 h to yield the desired ester as a colorless oil (0.119 g, 97%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>9</sub>H<sub>10</sub>O<sub>2</sub> 150.07, found 150, 119, 91, 65; *t<sub>R</sub>* = 17.458 min.



#### Methyl 4-hydroxybenzoate (8)

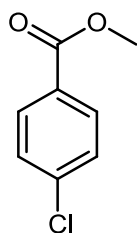
General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 4-hydroxybenzaldehyde (0.1 g, 0.819 mmol) to yield the desired ester as a white powder (0.120 g, 96%). When synthesized starting from 4-hydroxybenzylalcohol (0.1 g, 0.806 mmol), the reaction was performed at 120 °C for 1.5 h. The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.118 g, 96%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported

values in literature<sup>5</sup>. GC–MS ( $M^+$  electron impact),  $m/z$  calcd for  $C_8H_8O_3$  152.05, found 152, 121, 93, 65, 39;  $t_R$  = 21.024 min.



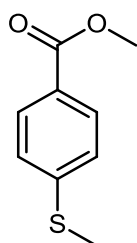
**Methyl 3-hydroxybenzoate (9)**

General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 3-hydroxybenzaldehyde (0.1 g, 0.819 mmol) to yield the desired ester as a white powder (0.122 g, 98%). When synthesized starting from 3-hydroxybenzylalcohol (0.1 g, 0.806 mmol), the reaction was performed at 120 °C for 1 h. The dried reaction mixture was diluted with  $CH_2Cl_2$ , washed with brine and finally dried (anhydrous  $Na_2SO_4$ ). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.110 g, 89%).  $^1H$  NMR (300 MHz,  $CDCl_3$ ) and  $^{13}C$  NMR (75 MHz,  $CDCl_3$ ) were in accordance with the reported values in literature<sup>2</sup>. GC–MS ( $M^+$  electron impact),  $m/z$  calcd for  $C_8H_8O_3$  152.05, found 152, 121, 93, 65, 28, 18;  $t_R$  = 21.383 min.



**Methyl 4-chlorobenzoate (10)**

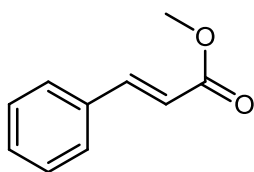
General procedure for aerobic aldehyde esterification was performed at 100 °C for 1 h with 4-chlorobenzaldehyde (0.1 g, 0.711 mmol). The dried reaction mixture was diluted with  $CH_2Cl_2$ , washed with brine and finally dried (anhydrous  $Na_2SO_4$ ). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a colorless oil (0.0157 g, 13%). When synthesized starting from 4-chlorobenzylalcohol (0.1 g, 0.701 mmol), the reaction was performed at 100 °C for 1 h and the same work-up purification was carried out. The desired ester was obtained as a colorless oil (0.00953 g, 8%).  $^1H$  NMR (300 MHz,  $CDCl_3$ ) and  $^{13}C$  NMR (75 MHz,  $CDCl_3$ ) were in accordance with the reported values in literature<sup>1</sup>. GC–MS ( $M^+$  electron impact),  $m/z$  calcd for  $C_8H_7ClO_2$  170.01, found 170, 139, 111, 75;  $t_R$  = 18.264 min.



**Methyl 4-(methylthio)benzoate (11)**

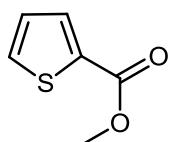


General procedure for aerobic aldehyde esterification was performed at 120 °C for 1.5 h with 4-(methylthio)benzaldehyde (0.1 g, 0.657 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.0754 g, 63%). When synthesized starting from 4-(methylthio)benzylalcohol (0.1 g, 0.648 mmol), the reaction was performed at 120 °C for 1.5 h and the same work-up purification was carried out. The desired ester was obtained as a white powder (0.100 g, 85%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>9</sub>H<sub>10</sub>O<sub>2</sub>S 182.04, found 182, 151, 123, 108; *t<sub>R</sub>* = 22.583 min.



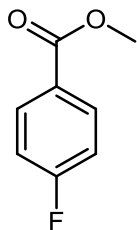
#### Methyl 3-phenylprop-2-enoate (12)

General procedure for aerobic aldehyde esterification was performed at 120 °C for 1.5 h with 3-phenylprop-2-enal (0.1 g, 0.757 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a white powder (0.0858 g, 70%). When synthesized starting from 3-phenylprop-2-enol (0.1 g, 0.745 mmol), the reaction was performed at 120 °C for 1.5 h and the same work-up procedure was carried out. The desired ester was obtained as a white powder (0.0422 g, 35%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>10</sub>H<sub>10</sub>O<sub>2</sub> 162.07, found 162, 131, 103, 77, 51; *t<sub>R</sub>* = 20.323 min.



#### Methyl thiophene-2-carboxylate (13)

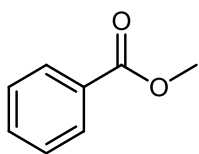
General procedure for aerobic aldehyde esterification was performed at 120 °C for 1.5 h with thiophene-2-carbaldehyde (0.1 g, 0.892 mmol). The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a colorless oil (0.0925 g, 73%). When synthesized starting from thiophene-2-ylmethanol (0.1 g, 0.876 mmol), the reaction was performed at 120 °C for 1.5 h and the same work-up procedure was carried out. The desired ester was obtained as a colorless oil (0.0933 g, 75%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>1</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>6</sub>H<sub>6</sub>O<sub>2</sub>S 142.01, found 142, 111; *t<sub>R</sub>* = 14.829 min.



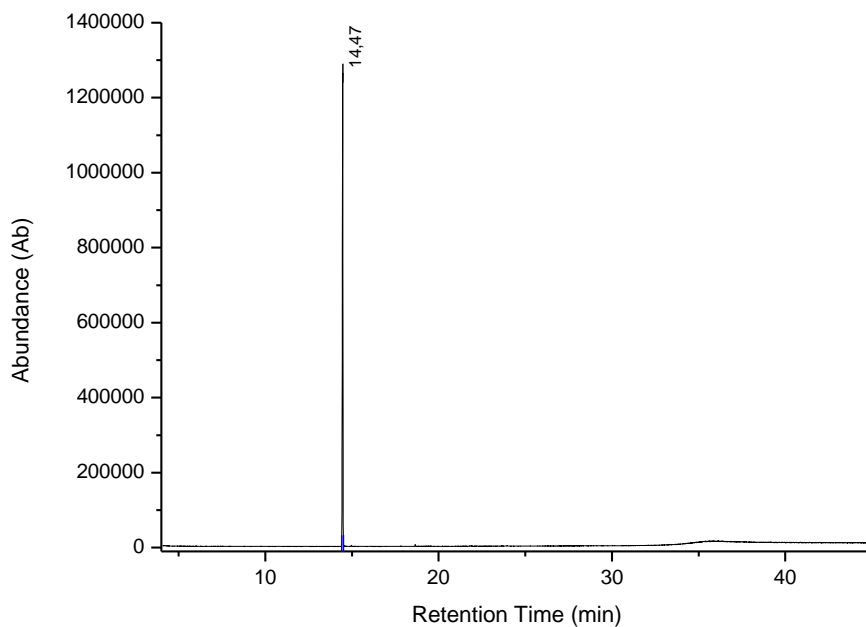
**Methyl 4-fluorobenzoate (14)**

General procedure for aerobic aldehyde esterification was performed at 120 °C for 1.5 h with 4-fluorobenzaldehyde (0.1 g, 0.806 mmol) to yield the desired ester as a colorless oil (0.121 g, 97%). When synthesized starting from 4-fluorobenzylalcohol (0.1 g, 0.793 mmol), the reaction was performed at 120 °C for 1.5 h. The dried reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine and finally dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>). The crude residue was purified by column chromatography (PE/EtOAc) to yield the desired methyl ester as a colorless oil (0.0355 g, 29%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) were in accordance with the reported values in literature<sup>8</sup>. GC–MS (M<sup>+</sup> electron impact), *m/z* calcd for C<sub>8</sub>H<sub>7</sub>FO<sub>2</sub> 154.04, found 154, 123, 95, 75; *t<sub>R</sub>* = 13.734 min.

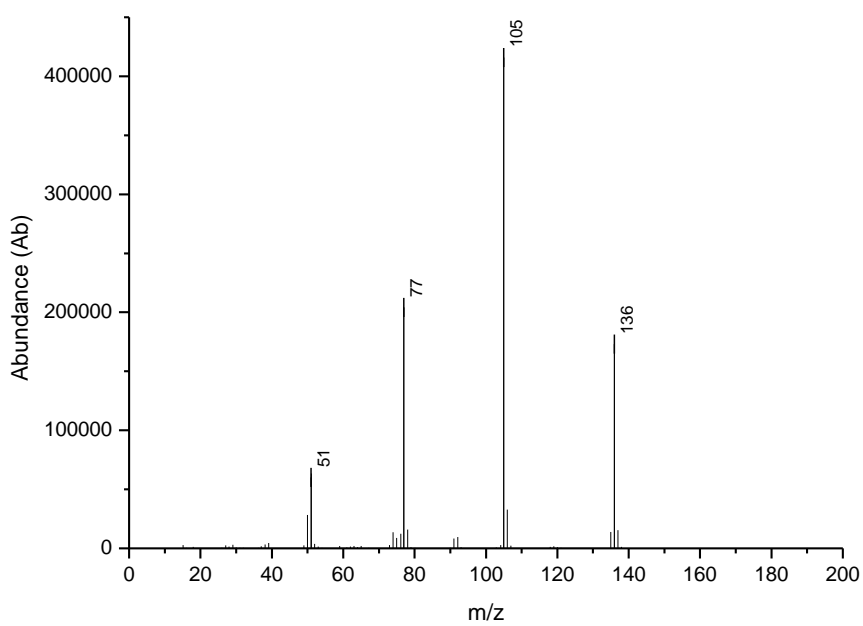
# GC-MS analysis chromatograms of crude reactions

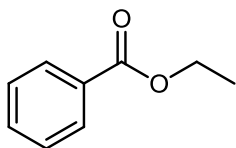


**Methyl benzoate (2a)**

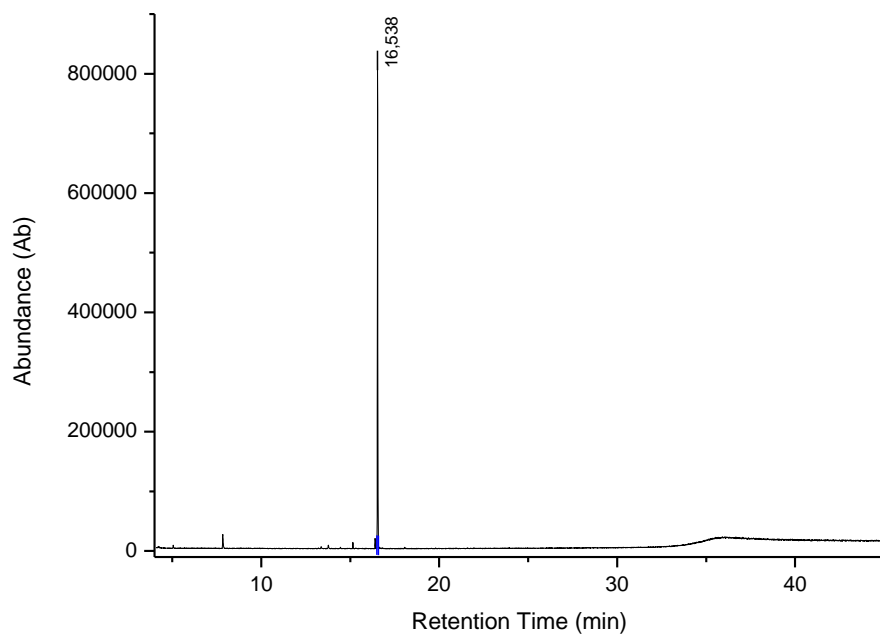


Peak	$t_R$	Area	% of Total
methyl benzoate	14.470	33436230	100

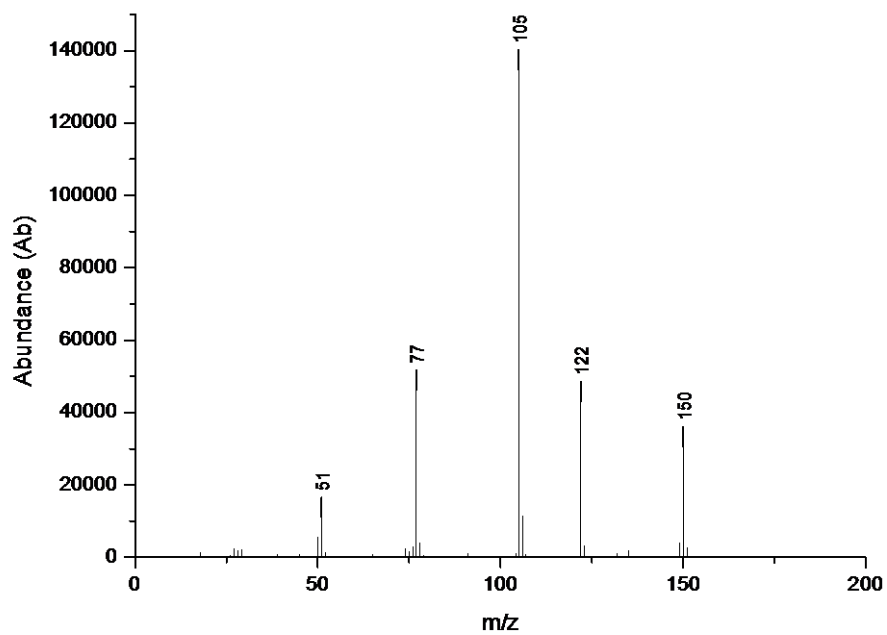


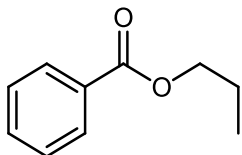


Ethyl benzoate (2b)

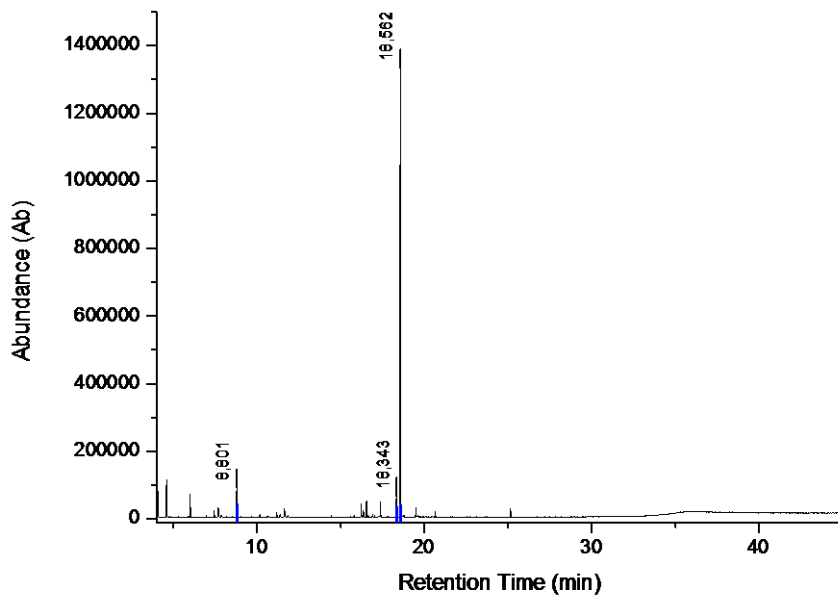


Peak	$t_R$	Area	% of Total
ethyl benzoate	16.538	14908543	100

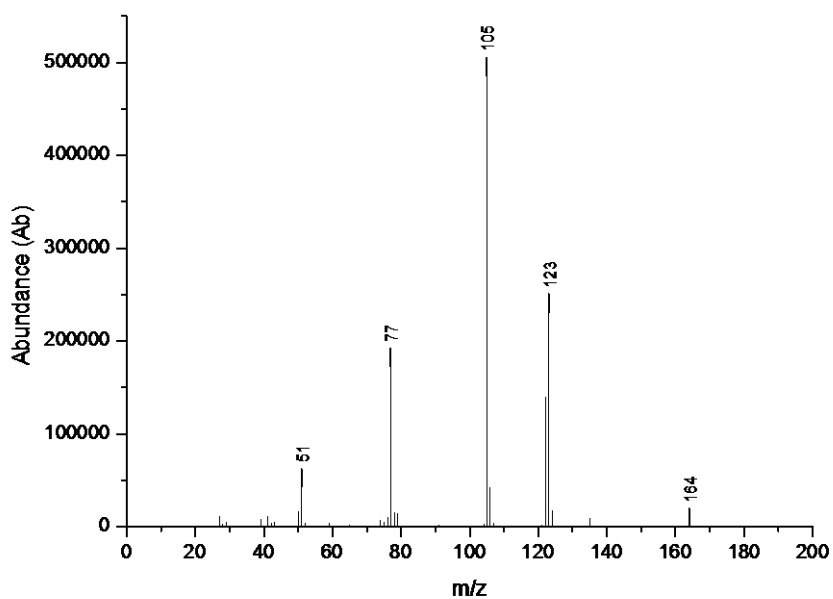


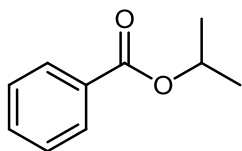


Propyl benzoate (2c)

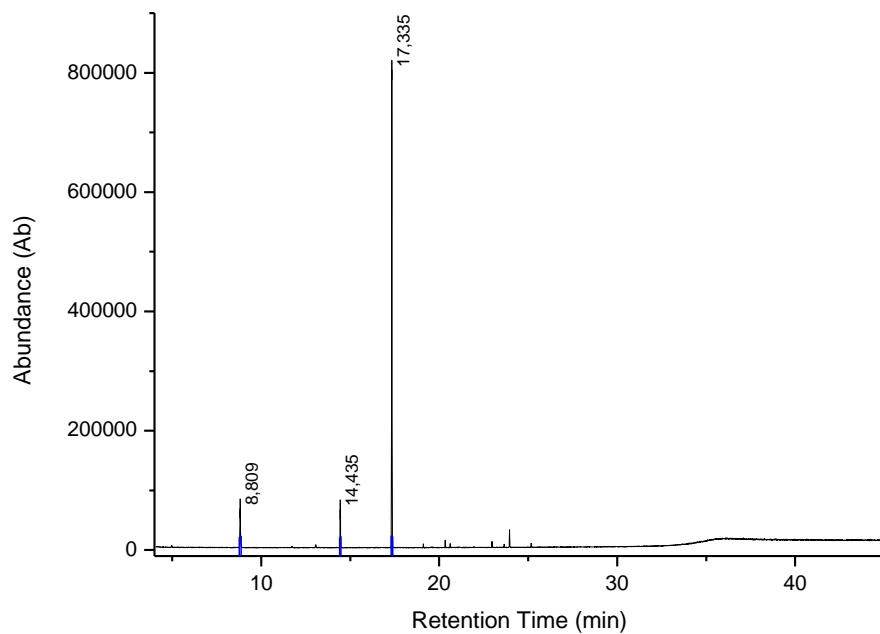


Peak	$t_R$	Area	% of Total
benzaldehyde	8.801	3072038	12
propanoic acid phenylmethyl ester	18.351	1398640	5
propyl benzoate	18.562	21511949	83

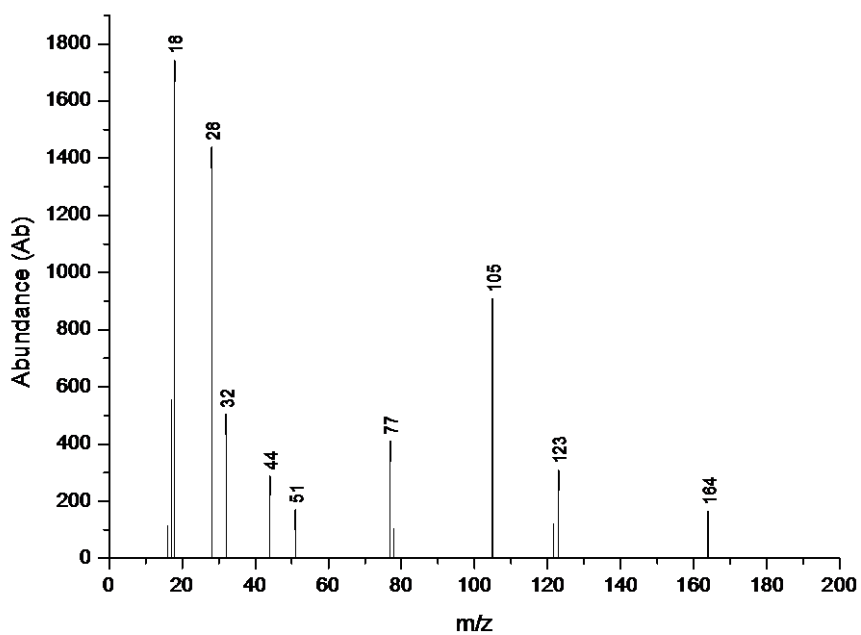


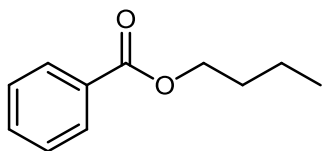


Isopropyl benzoate (2d)

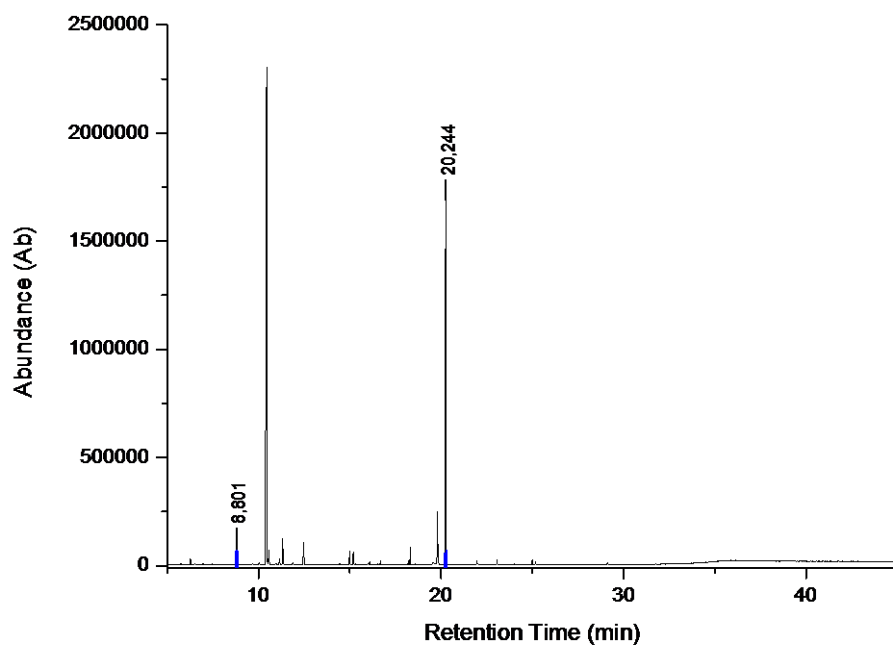


Peak	$t_R$	Area	% of Total
benzaldehyde	8.773	1781488	11
benzoate	14.416	1526945	9
isopropyl benzoate	17.335	12994139	80

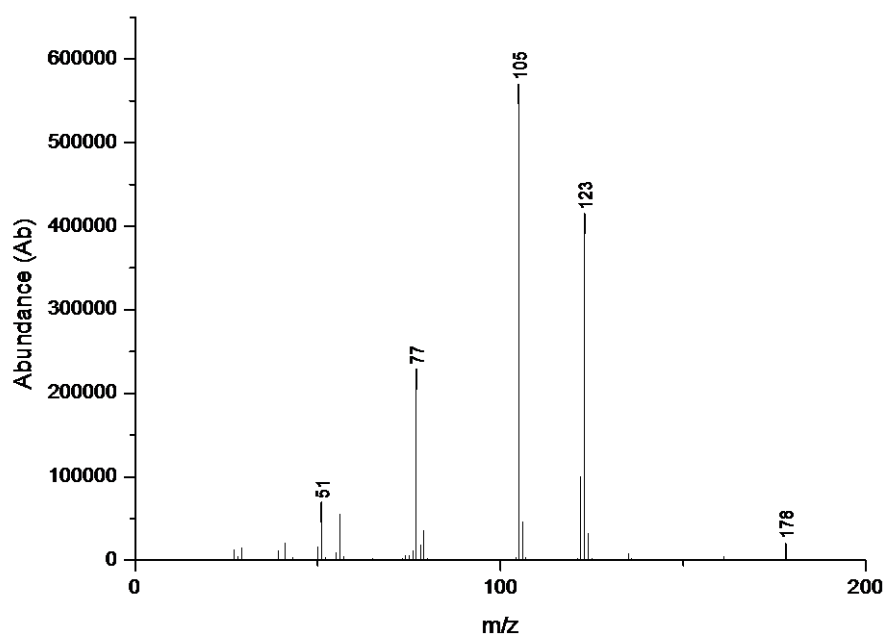


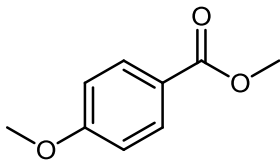


Butyl benzoate (2e)

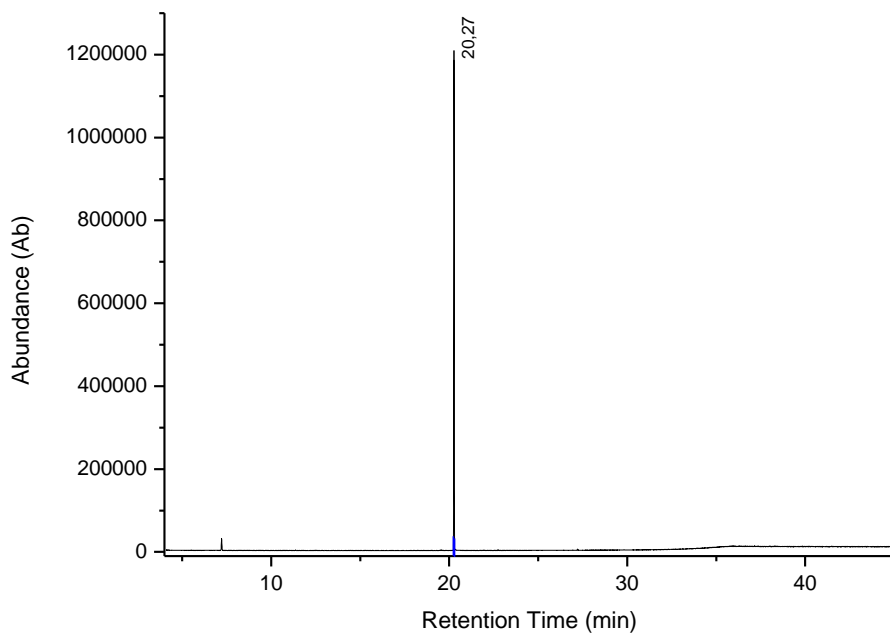


Peak	$t_R$	Area	% of Total
benzaldehyde	8.801	3368861	9
butyl butyrate	10.448		
butyl benzoate	20.244	28810928	78

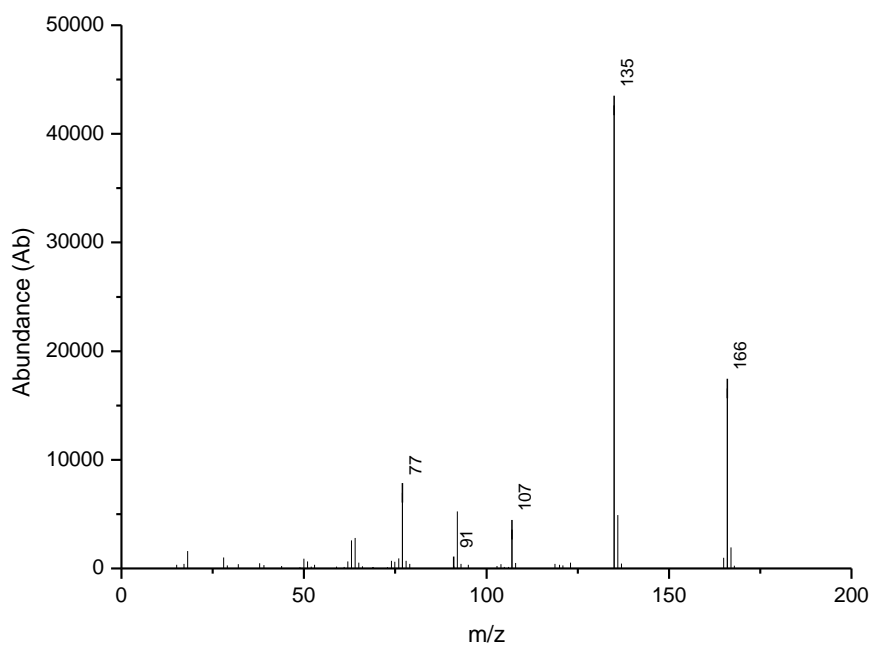




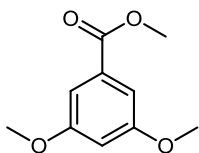
Methyl 4-methoxybenzoate (3)



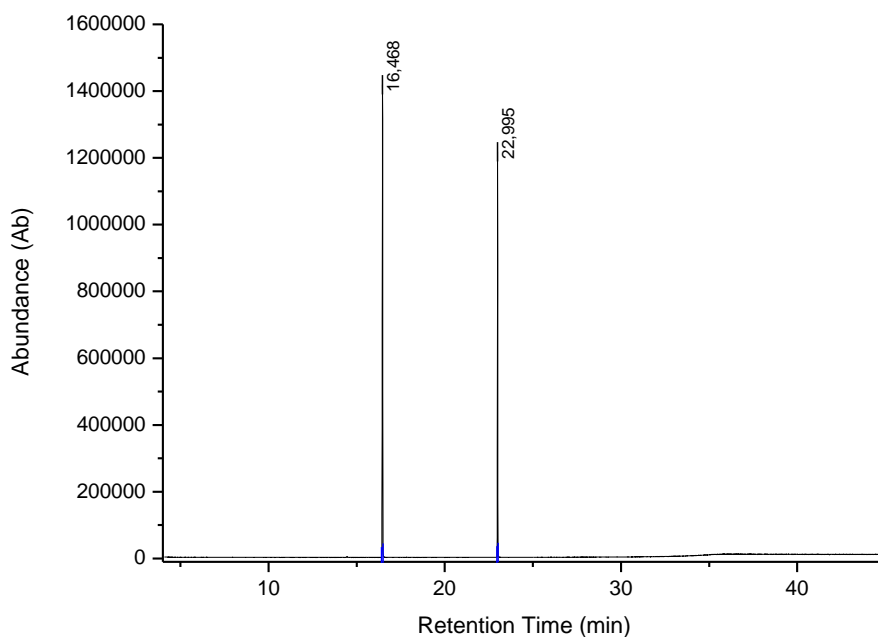
Peak	$t_R$	Area	% of Total
methyl 3-methoxybenzoate	20.270	18340786	100



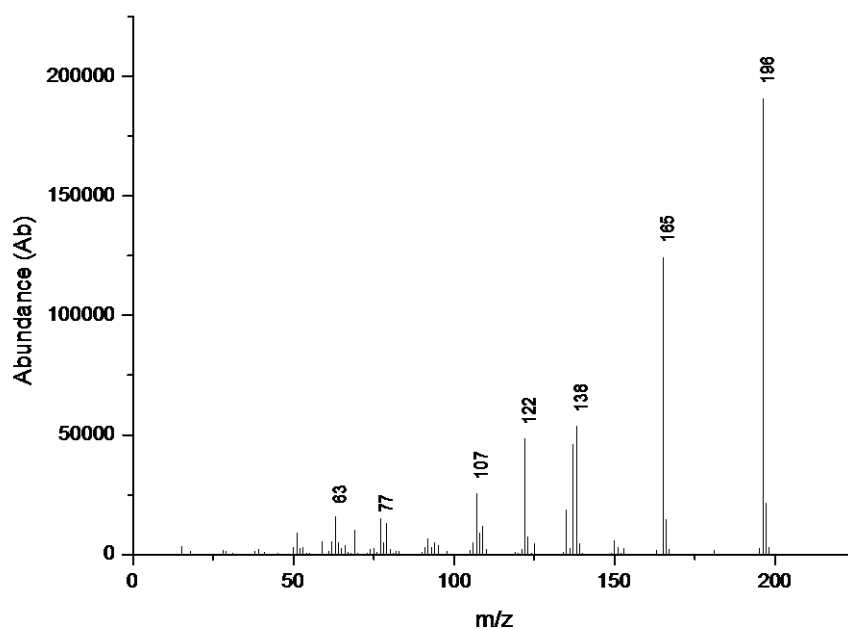


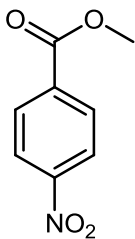


### Methyl 3,5-dimethoxybenzoate (4)

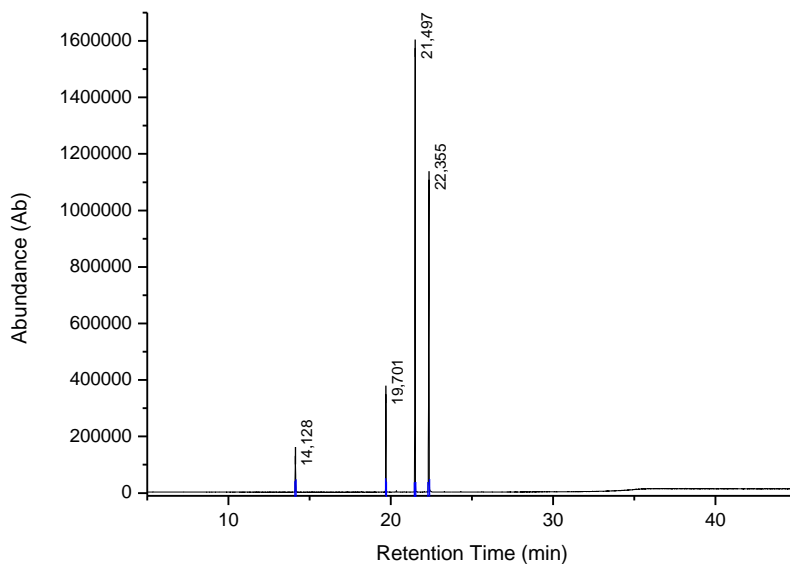


Peak	$t_R$	Area	% of Total
1,3-dimethoxybenzene	16.468	29952311	62
methyl 3,5-dimethoxybenzoate	22.995	18173718	38

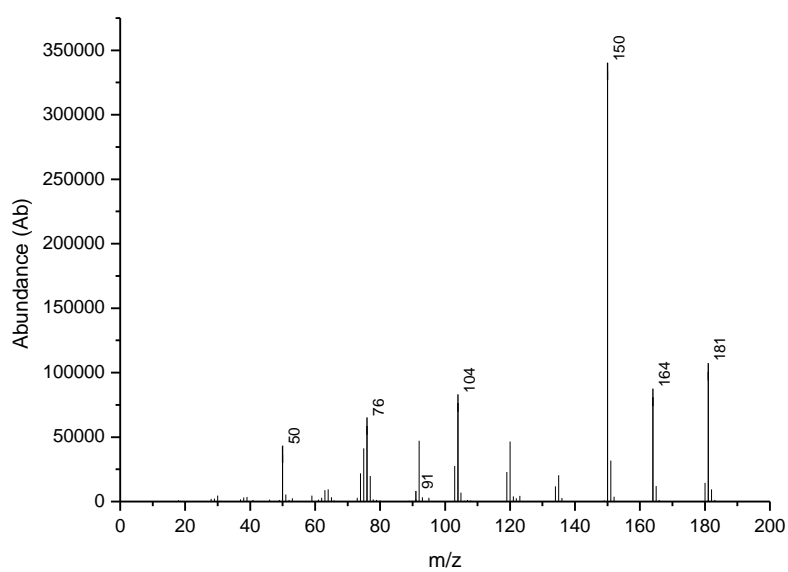


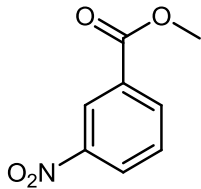


**Methyl 4-nitrobenzoate (5)**

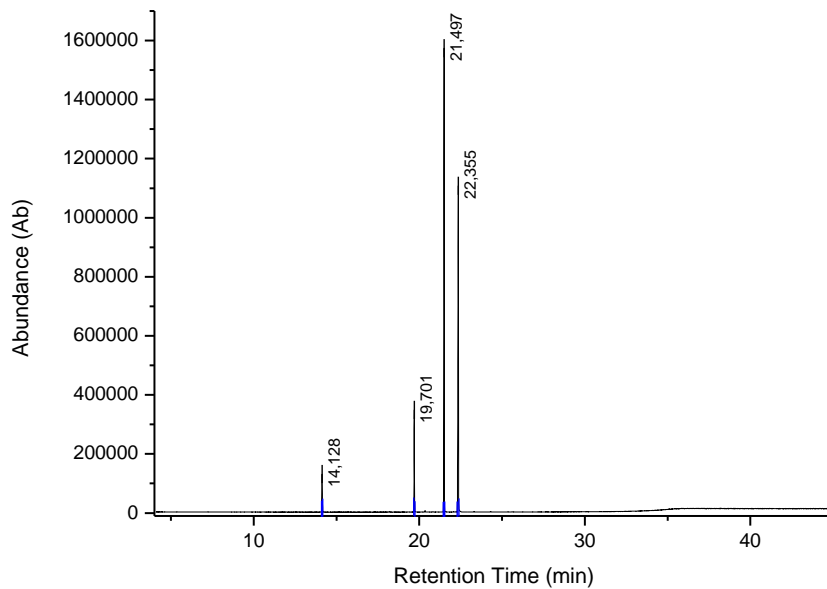


Peak	$t_R$	Area	% of Total
nitrobenzene	14.128	3172715	5
4-nitrobenzaldehyde	19.701	5144260	8
methyl 4-nitrobenzoate	21.497	29652299	46
4-nitrobenzylalcohol	22.355	26565151	41

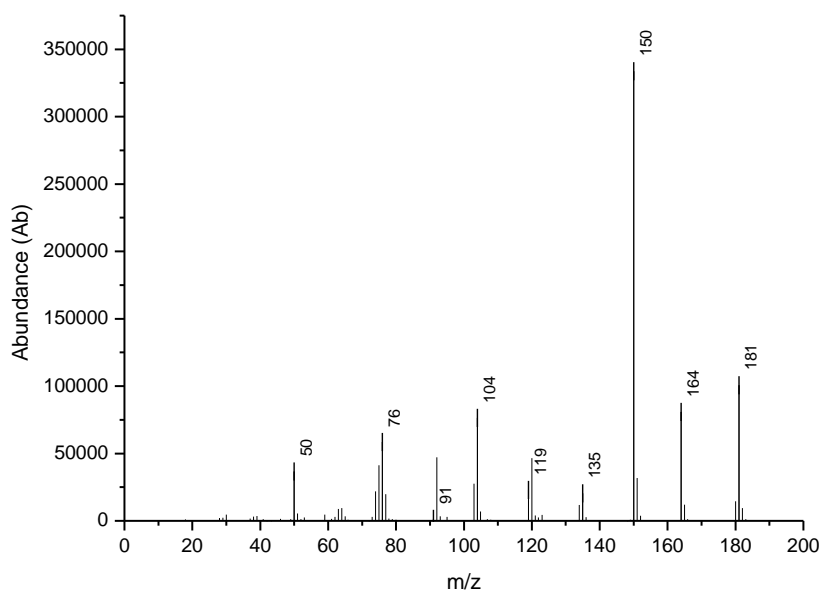


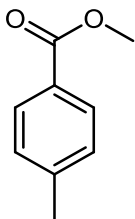


### Methyl 3-nitrobenzoate (6)

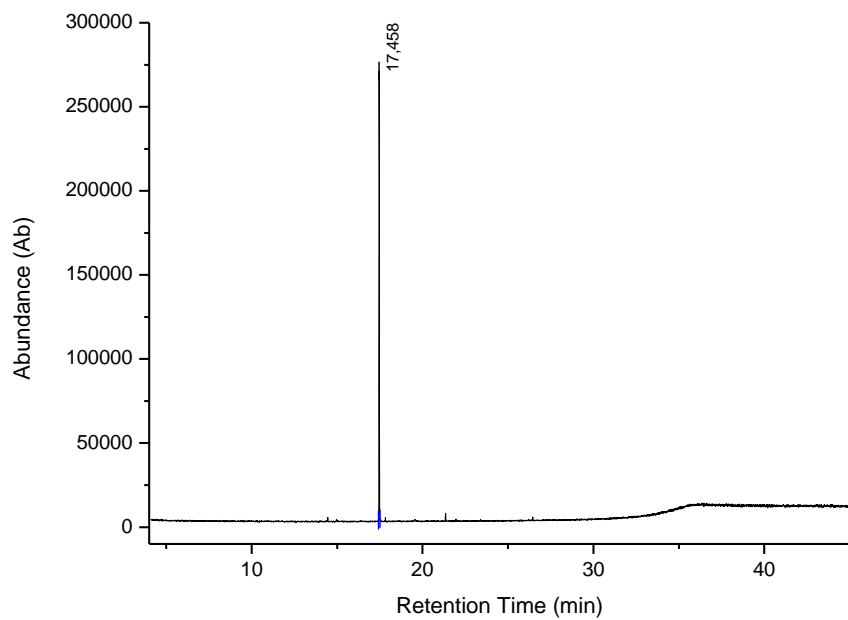


Peak	$t_R$	Area	% of Total
nitrobenzene	14.110	4064251	5
3-nitrobenzaldehyde	19.893	5098567	6
methyl 3-nitrobenzoate	21.751	49211596	60
3-nitrobenzylalcohol	22.206	23474229	29

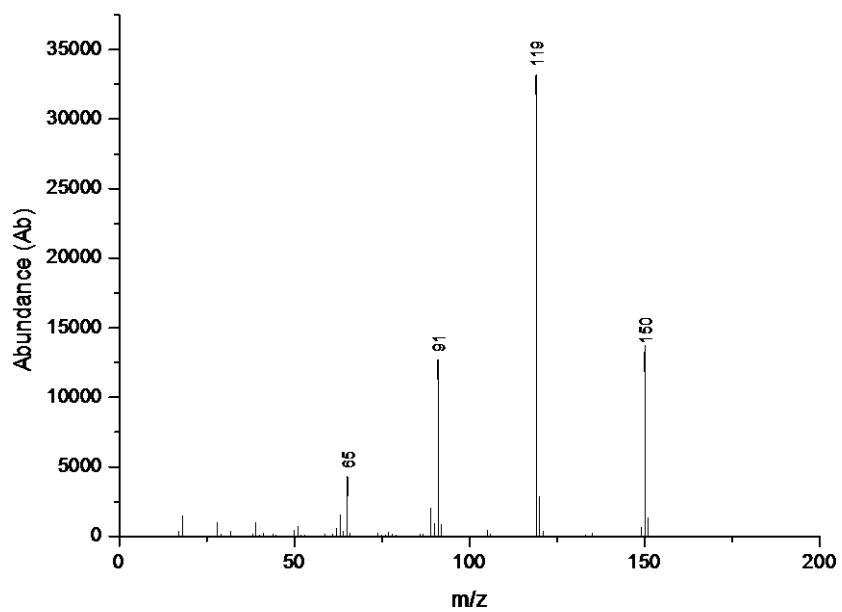


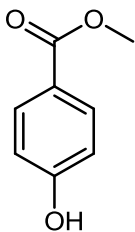


Methyl 4-methylbenzoate (7)

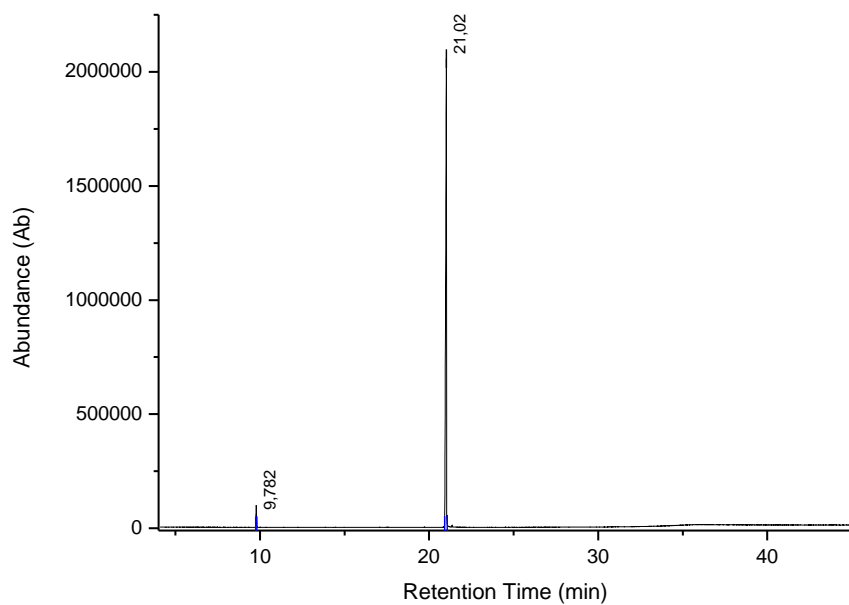


Peak	$t_R$	Area	% of Total
methyl 4-methylbenzoate	17.458	4331510	100

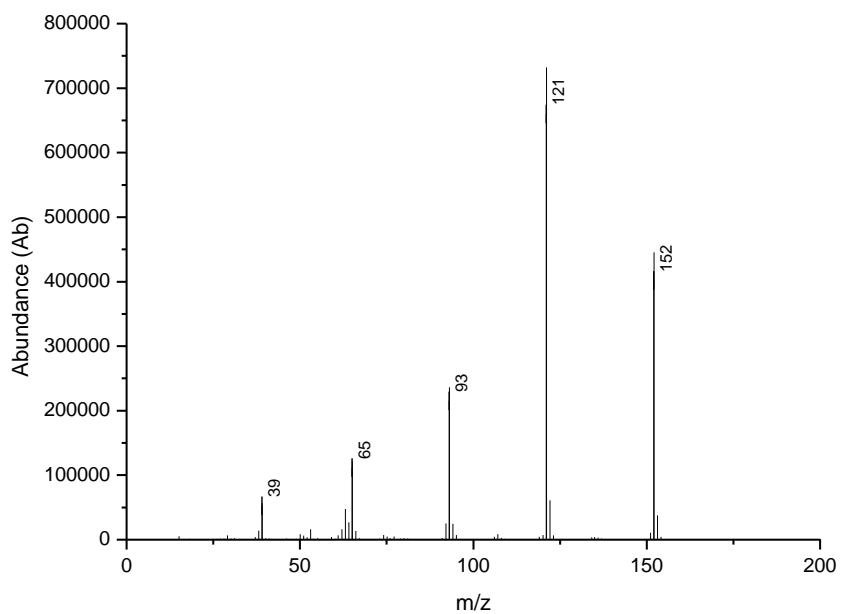


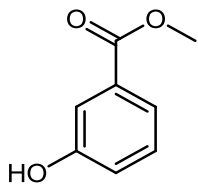


**Methyl 4-hydroxybenzoate (8)**

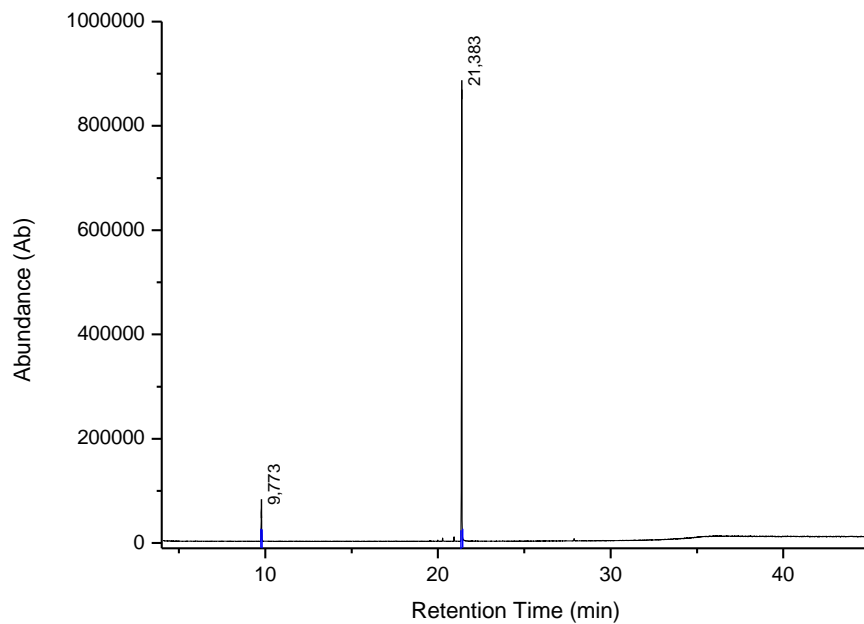


Peak	$t_R$	Area	% of Total
phenol	9.782	1559671	2
methyl 4-hydroxybenzoate	21.024	75112648	98

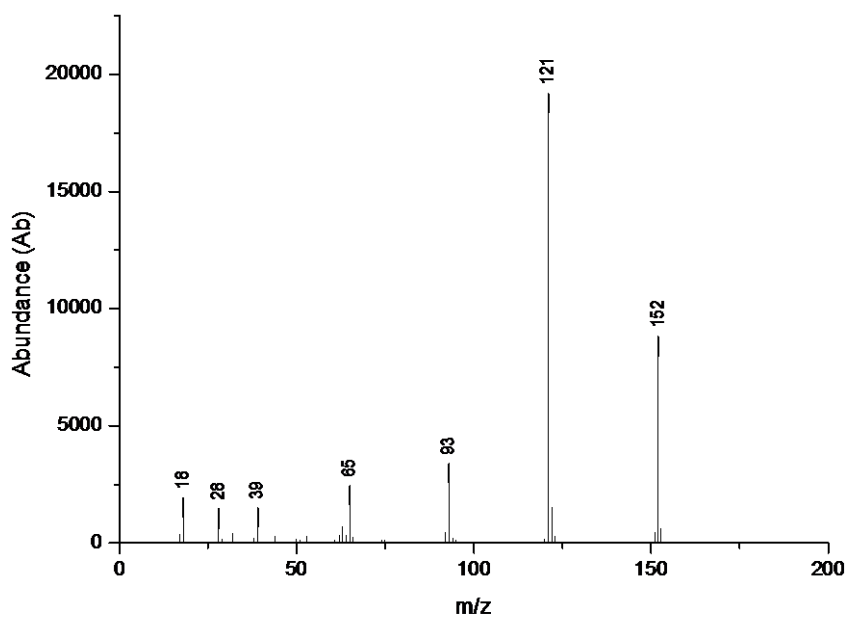


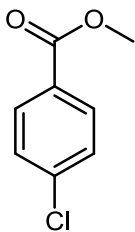


**Methyl 3-hydroxybenzoate (9)**

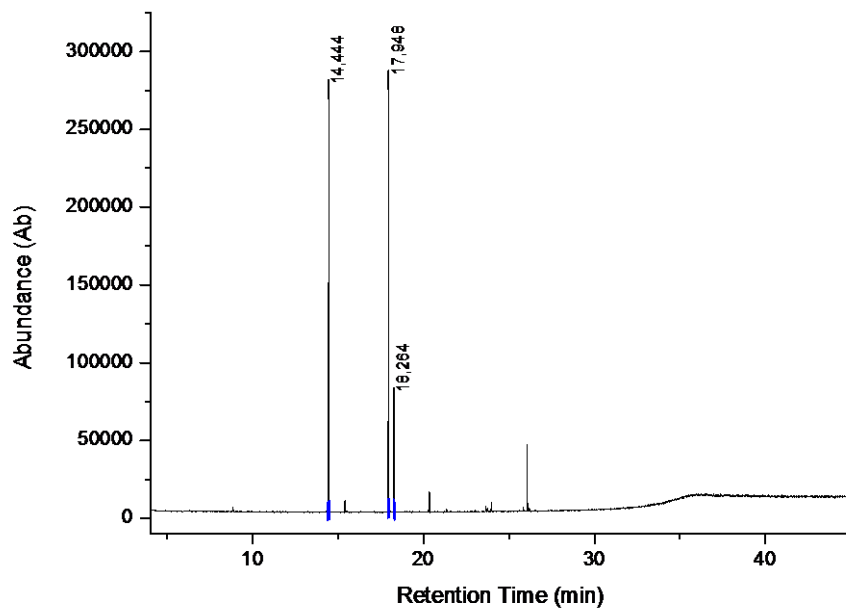


Peak	$t_R$	Area	% of Total
phenol	9.773	1671136	9
methyl 3-hydroxybenzoate	21.383	17862872	91

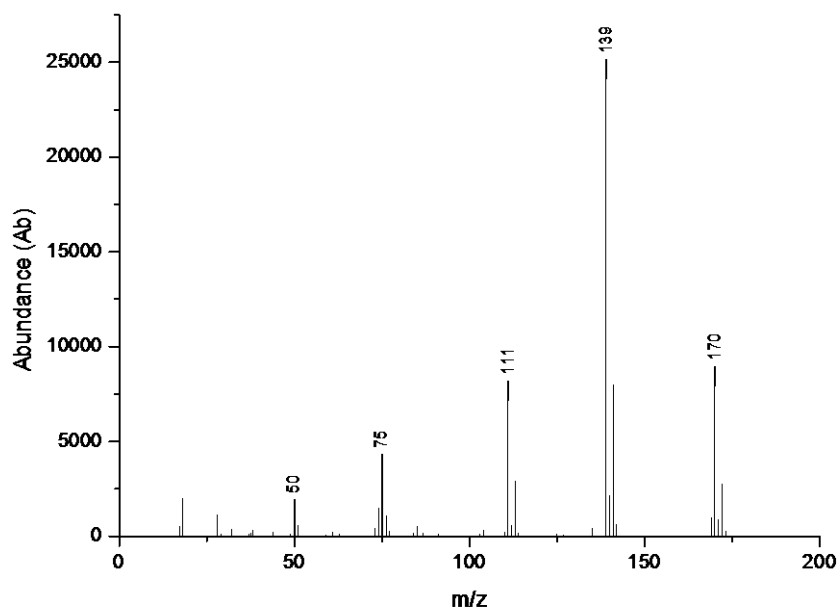


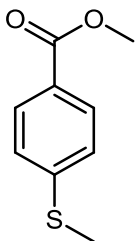


Methyl 4-chlorobenzoate (10)

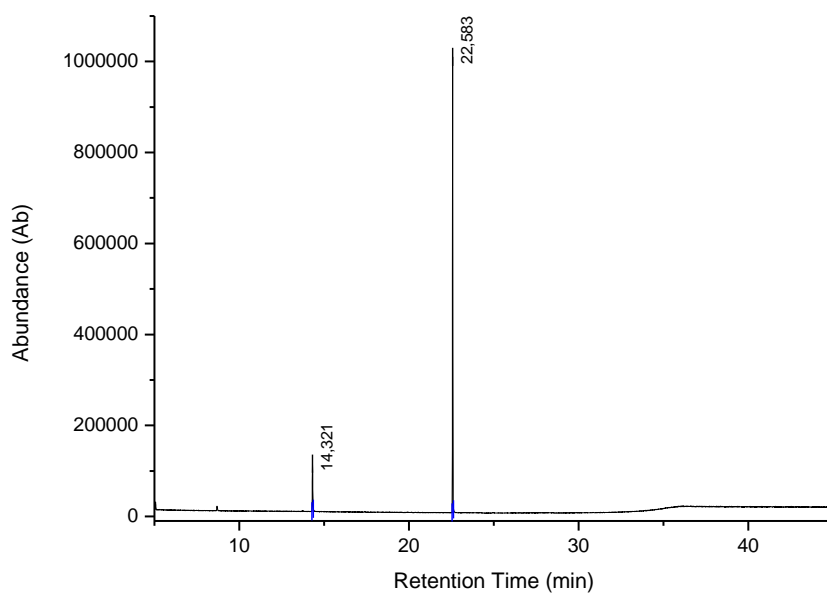


Peak	$t_R$	Area	% of Total
methyl benzoate	14.452	6239520	52
4-chlorobenzylalcohol	17.948	4638083	38
methyl 4-chlorobenzoate	18.264	1190777	10

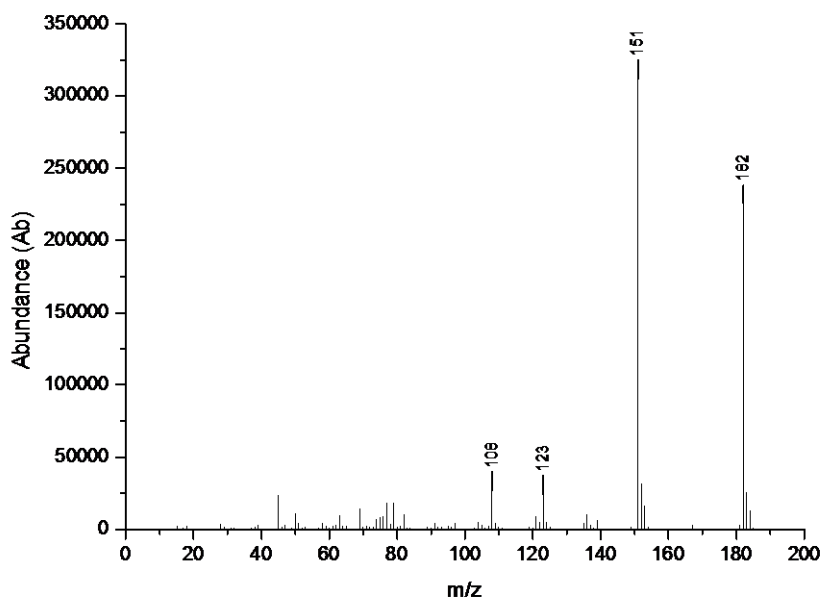




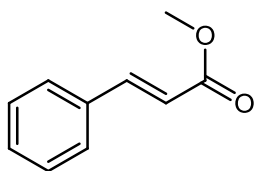
Methyl 4-(methylthio)benzoate (11)



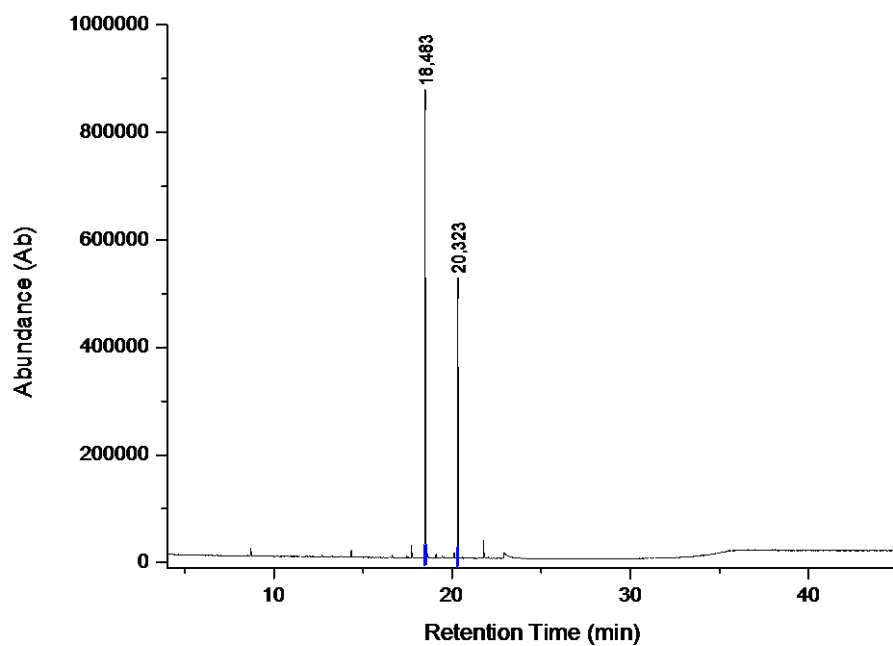
Peak	$t_R$	Area	% of Total
methyl benzoate	14.321	2561672	13
methyl 4-(methylthio)benzoate	22.583	16434629	87



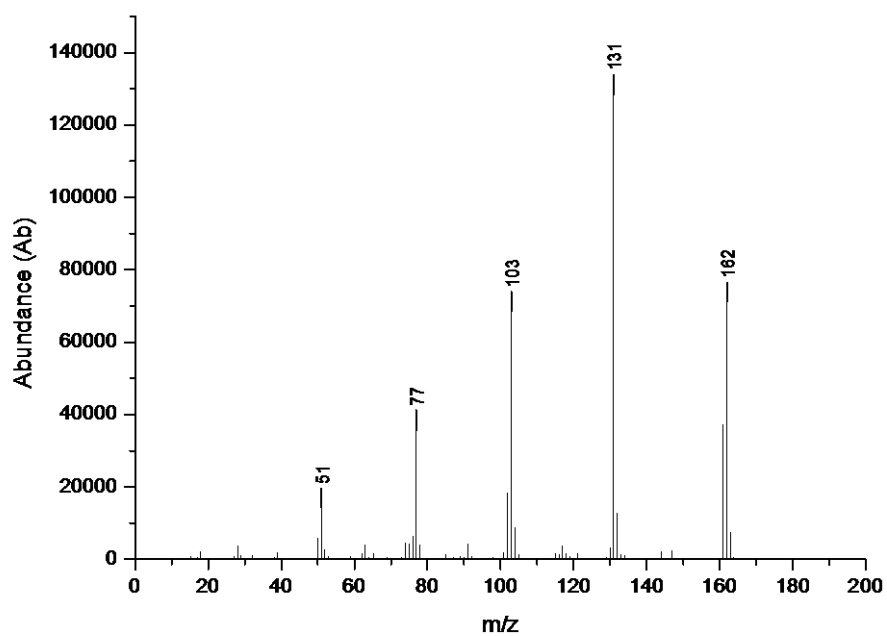


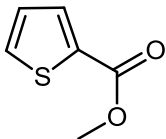


Methyl 3-phenylprop-2-enoate (12)

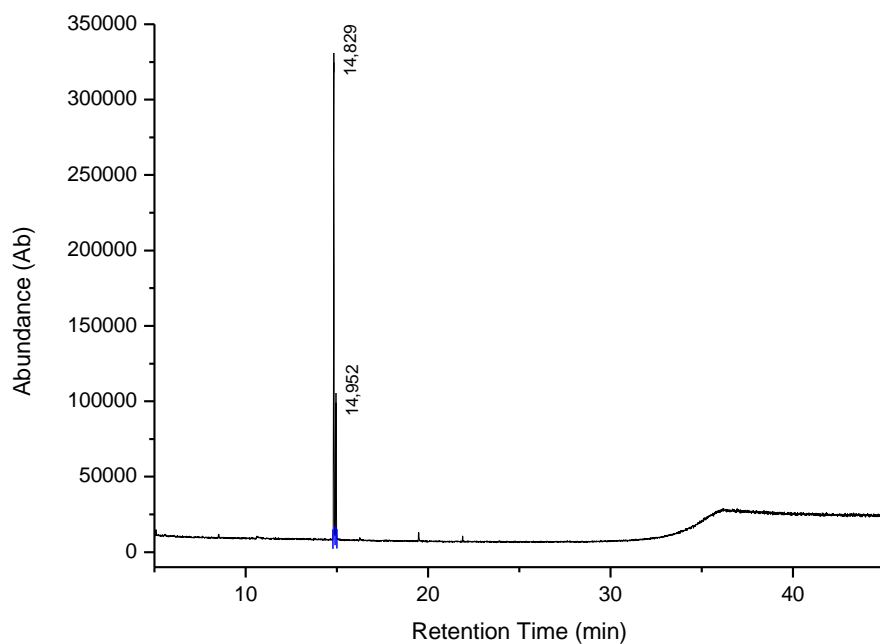


Peak	$t_R$	Area	% of Total
3-phenylprop-2-enol	18.492	14545844	63
methyl 3-phenylprop-2-enoate	20.323	7586659	37

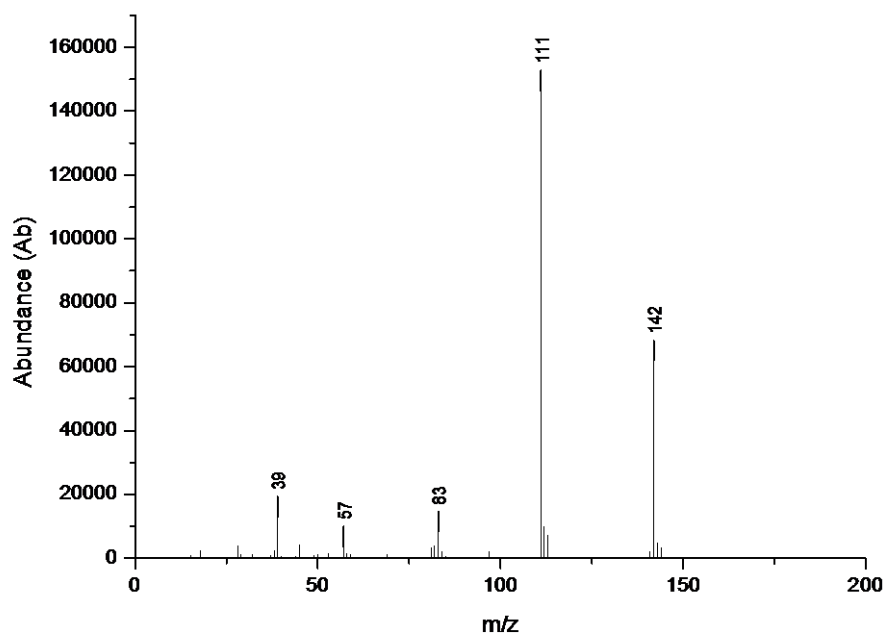


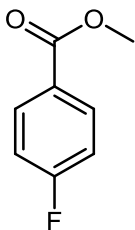


Methyl thiophene-2-carboxylate (13)

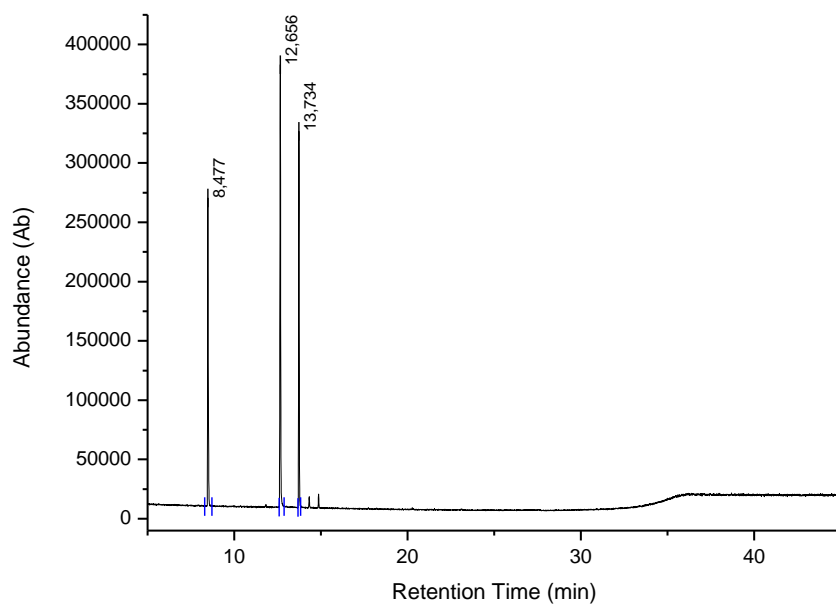


Peak	$t_R$	Area	% of Total
methyl thiophene-2-caboxylate	14.829	6922544	78
2-dimethoxymethyl thiopene	14.952	1977090	22

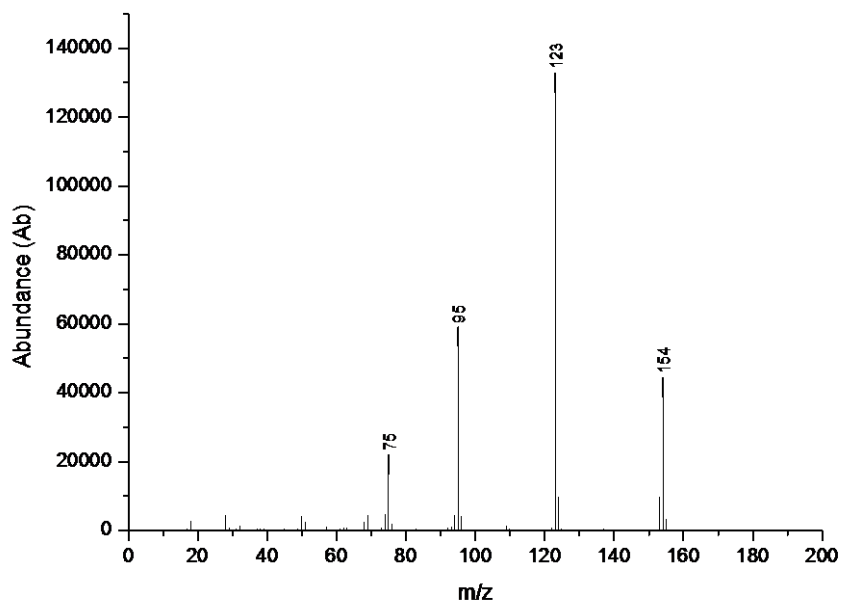




Methyl 4-fluorobenzoate (14)



Peak	$t_R$	Area	% of Total
4-fluorobenzaldehyde	8.485	6786469	26
4-fluorobenzylalcohol	12.665	11355713	43
methyl 4-fluorobenzoate	13.734	8194213	31



## References

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- [2] Spectral Database for Organic Compounds SDBS, SDBS #2319. Available at [http://sdb.sdb.aist.go.jp/sdb/cgi-bin/direct\\_frame\\_top.cgi](http://sdb.sdb.aist.go.jp/sdb/cgi-bin/direct_frame_top.cgi)
- [3] B. R. Kim, G. H. Sung, S.-G. Lee, *Tetrahedron*, **2013**, 69, 3234-3237.
- [4] C. Liu, J. Wang, L. Meng, Y. Deng, Y. Li, A. Lei, *Angew. Chem. Int. Ed.*, **2011**, 50, 5144-5148.
- [5] P. Li, J. Zhao, R. Lang, C. Xia, F. Li, *Tetrahedron Lett.*, **2014**, 55, 390-393.
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- [8] B. A. Tschaen, J. R. Schmink, G. A. Molander, *Org. Lett.*, **2013**, 15 (3), 500-503.