

Supporting Information

for

Efficient CO₂ capture by tertiary amine-functionalized ionic liquids through Li⁺-stabilized zwitterionic adduct formation

Zhen-Zhen Yang, Liang-Nian He*

Address: State Key Laboratory and Institute of Elemento-Organic Chemistry, Collaborative Innovation Center of Chemical Science and Engineering (Tianjin), Nankai University, Tianjin 300071, P. R. China.

Email: Liang-Nian He* - heln@nankai.edu.cn

*Corresponding author

General experimental methods, synthesis and characterization of the neutral ligands, lithium salts and the corresponding chelated ionic liquids

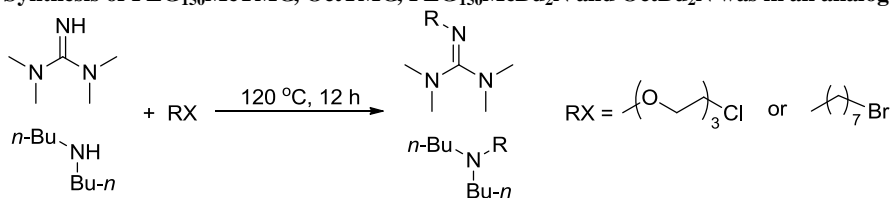
Table of contents

	page
1. Synthesis and characterization of the neutral ligands and lithium salts	S2
2. Coordination effect of different lithium salts with PEG ₁₅₀ MeTMG (Table S1)	S3
3. ¹ H NMR (CDCl ₃ , 400 MHz) spectrum of the neutral ligands and the corresponding chelate ionic liquids (Figure S1)	S4
4. ESI spectrum of PEG ₁₅₀ MeTMG, PEG ₁₅₀ MeBu ₂ N before and after coordinating with LiNTf ₂ (Figure S2)	S7
5. References	S9
6. NMR characterization and charts of all the neutral ligands and chelate ionic liquids	S9

1. Synthesis and characterization of the neutral ligands and lithium salts

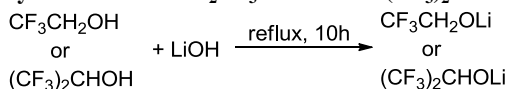
Synthesis of OctIm, PEG150MeIm and PEG150MeCl were performed in the same manner as described in the literature.¹

Synthesis of PEG₁₅₀MeTMG, OctTMG, PEG₁₅₀MeBu₂N and OctBu₂N was in an analogous manner as reported in the literature.²



In a dry flask, 1,1,3,3-tetramethylguanidine (TMG) or *n*-Bu₂NH (30 mmol) and xylene (1 mL) were mixed together. After being evacuated and purged with N₂ five times, the system was heated to 120 °C and maintained at this temperature for 2 h. Then PEG₁₅₀MeCl or 1-bromooctane (15 mmol) was added dropwise. After being stirred at 120 °C for 12 h under N₂ atmosphere, the resulting mixture was allowed to cool to room temperature. The formed salt precipitates were removed by filtration. The filtrate was distilled under reduced pressure and the product (PEG₁₅₀MeTMG, OctTMG, PEG₁₅₀MeBu₂N and OctBu₂N) obtained as colorless liquid.

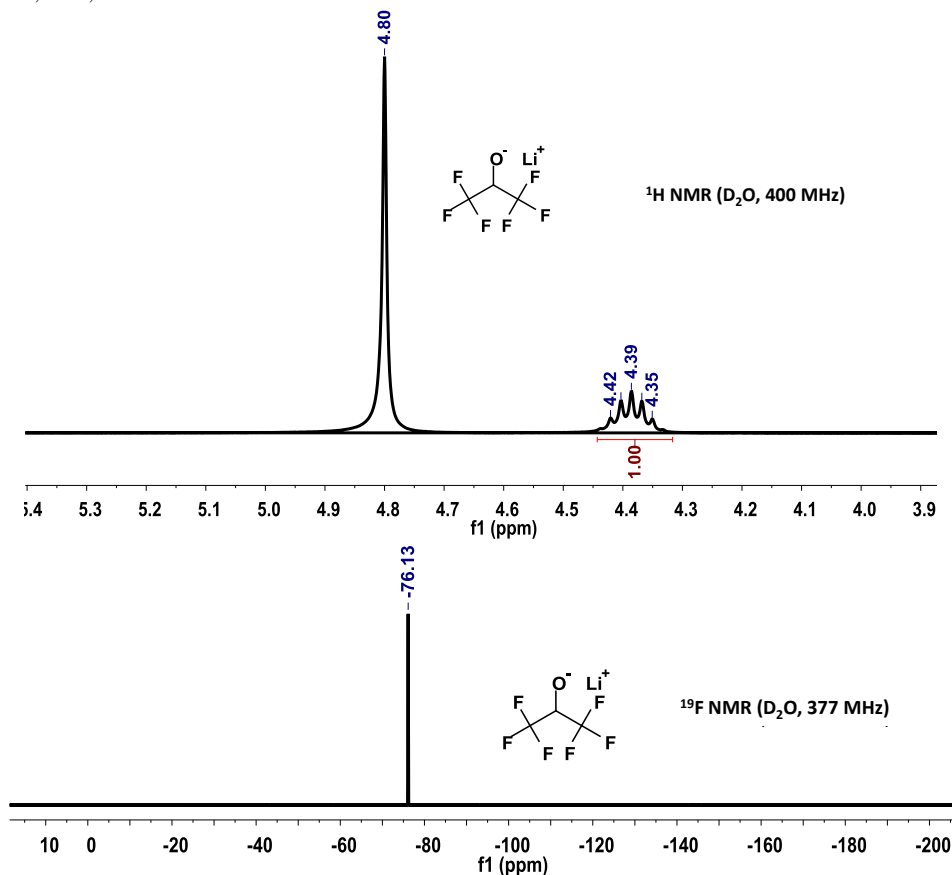
Synthesis of LiOCH₂CF₃ and LiOCH(CF₃)₂

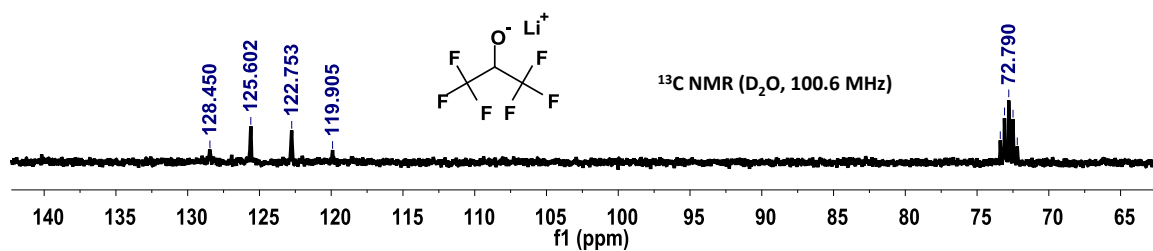


LiOH (10 mmol) and HOCH₂CF₃ or HOCH(CF₃)₂ (30 mmol) were stirred at reflux for 10 h, and then excess fluoro-substituted alcohol was removed on a rotator evaporator under reduced pressure. The residue was dried in vacuum to give LiOCH₂CF₃ or LiOCH(CF₃)₂ as a white solid.

LiOCH(CF₃)₂

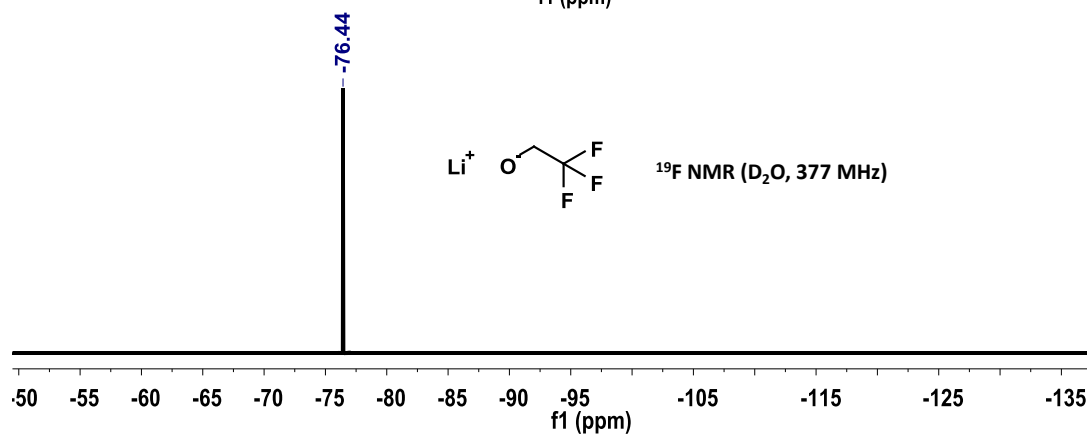
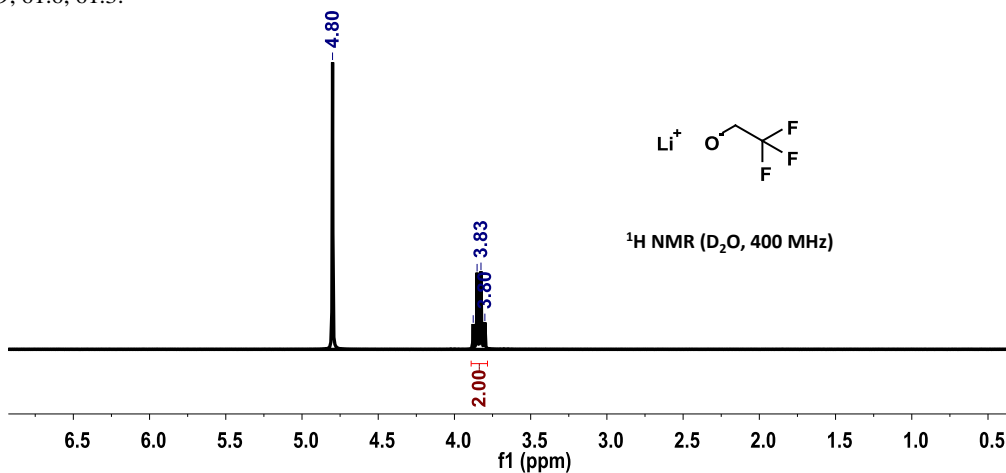
¹H NMR (D₂O, 400 MHz) δ 4.35-4.42 (m, 1H); ¹⁹F NMR (D₂O, 377 MHz) δ -76.13; ¹³C NMR (D₂O, 100.6 MHz) 128.4, 125.6, 122.8, 119.9, 73.4, 73.1, 72.8, 72.5, 72.2.





LiOCH₂CF₃

¹H NMR (D₂O, 400 MHz) δ 3.80-3.88 (m, 2H); ¹⁹F NMR (D₂O, 377 MHz) δ -76.44; ¹³C NMR (D₂O, 100.6 MHz) δ 131.0, 128.2, 125.4, 122.6, 62.2, 61.9, 61.6, 61.3.



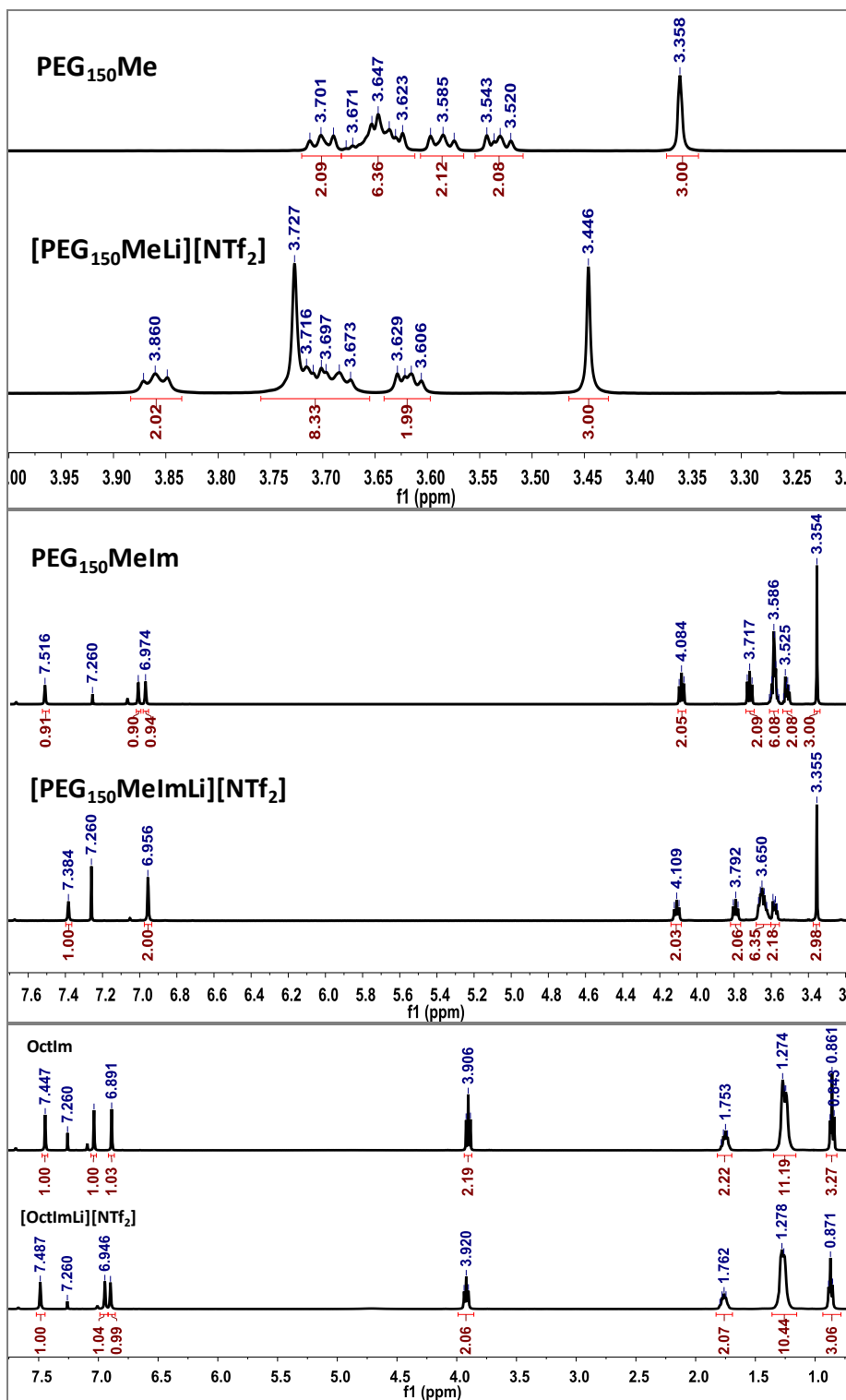
2. Coordination effect of different lithium salts with PEG₁₅₀MeTMG

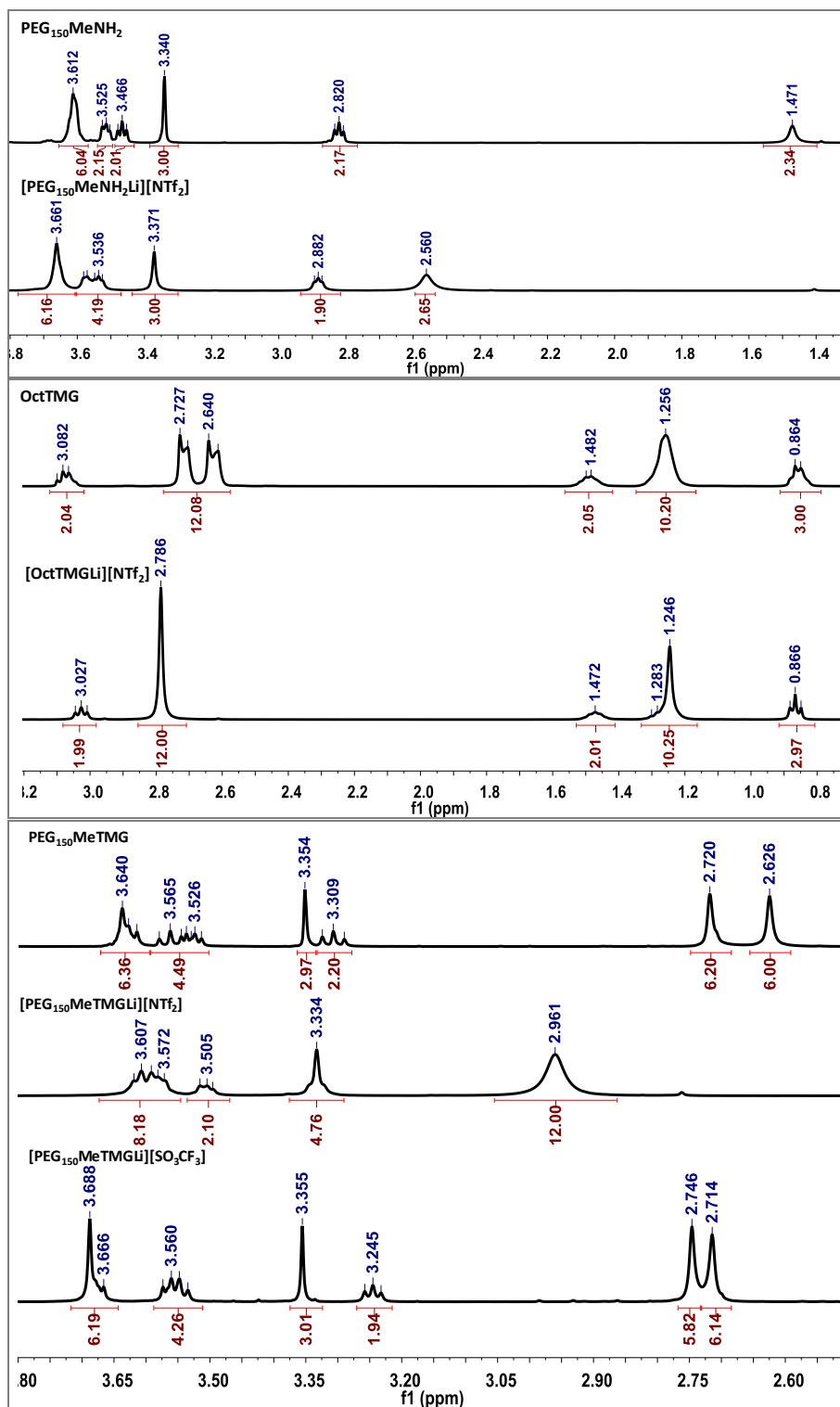
Table S1 Coordination effect of different lithium salts with PEG₁₅₀MeTMG^a

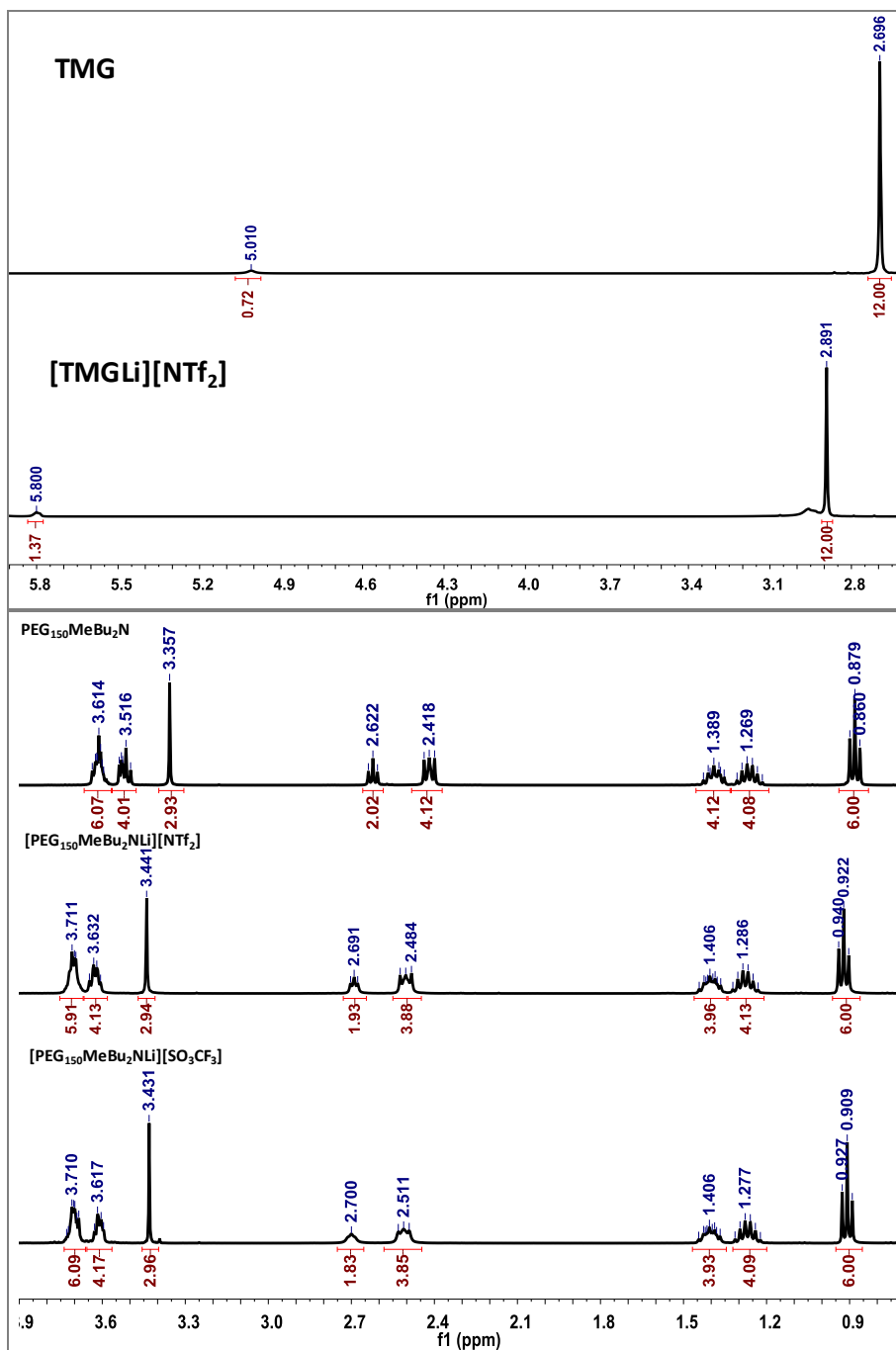
Entry	Lithium salt	Coordination
1	LiOAc	No
2	LiCl	No
3	LiClO ₄	No
4	Li ₂ WO ₄	No
5	LiOH	No
6	LiOCH ₂ CF ₃	No
7	LiOCH(CF ₃) ₂	No
8	LiSO ₃ CF ₃	Yes
9	LiNTf ₂	Yes

^a The reaction was conducted by mixing PEG₁₅₀MeTMG with the lithium salt in 1:1 molar ratio at room temperature. Homogeneous phases formed through multisite coordination interaction within 1 h. Otherwise, two separate phases were retained after stirring for 24 h.

3. ^1H NMR (CDCl_3 , 400 MHz) spectrum of the neutral ligands and the corresponding chelate ionic liquids







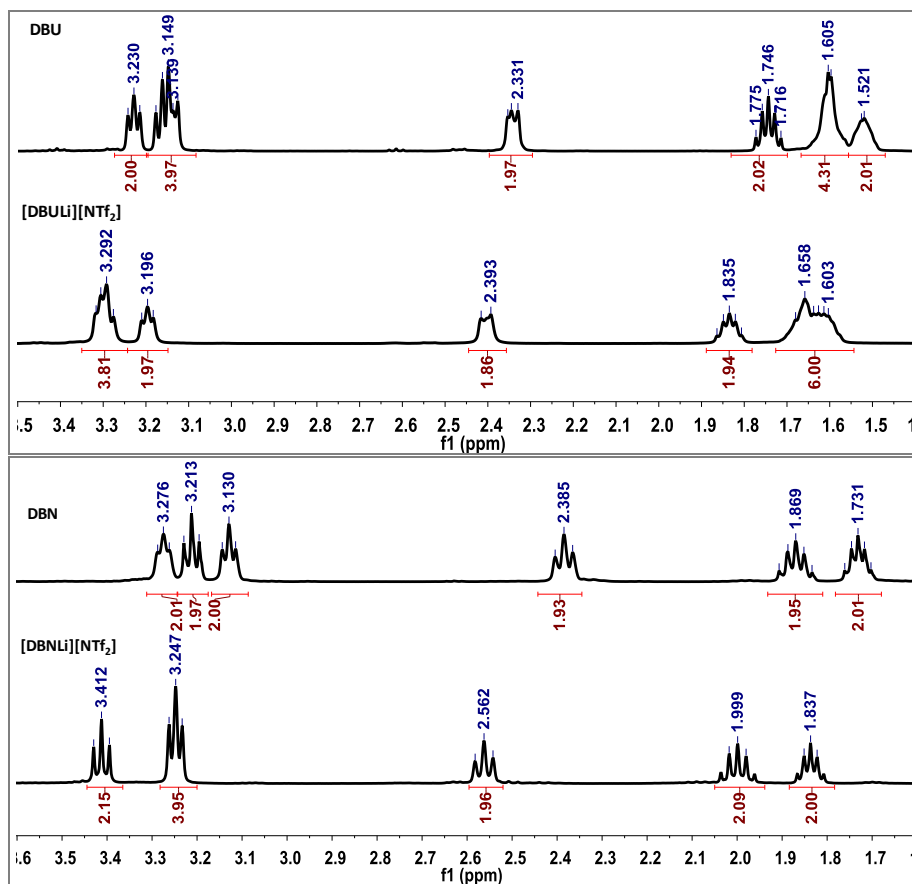
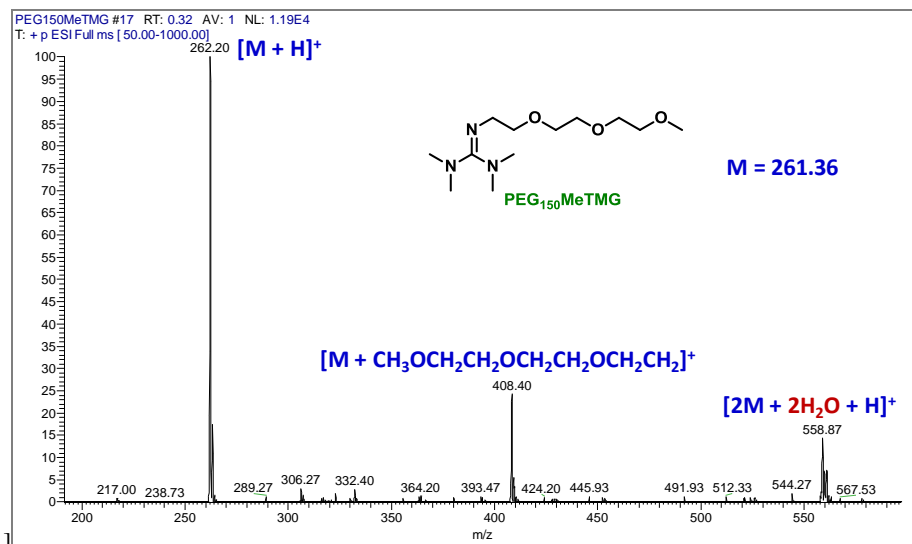


Figure S1 ¹H NMR (CDCl₃, 400 MHz) spectrum of the neutral ligands and the corresponding chelate ionic liquids after coordinated with lithium salts (LiNTf₂ and LiSO₃CF₃).

4. ESI spectrum of PEG₁₅₀MeTMG, PEG₁₅₀MeBu₂N before and after coordinating with LiNTf₂



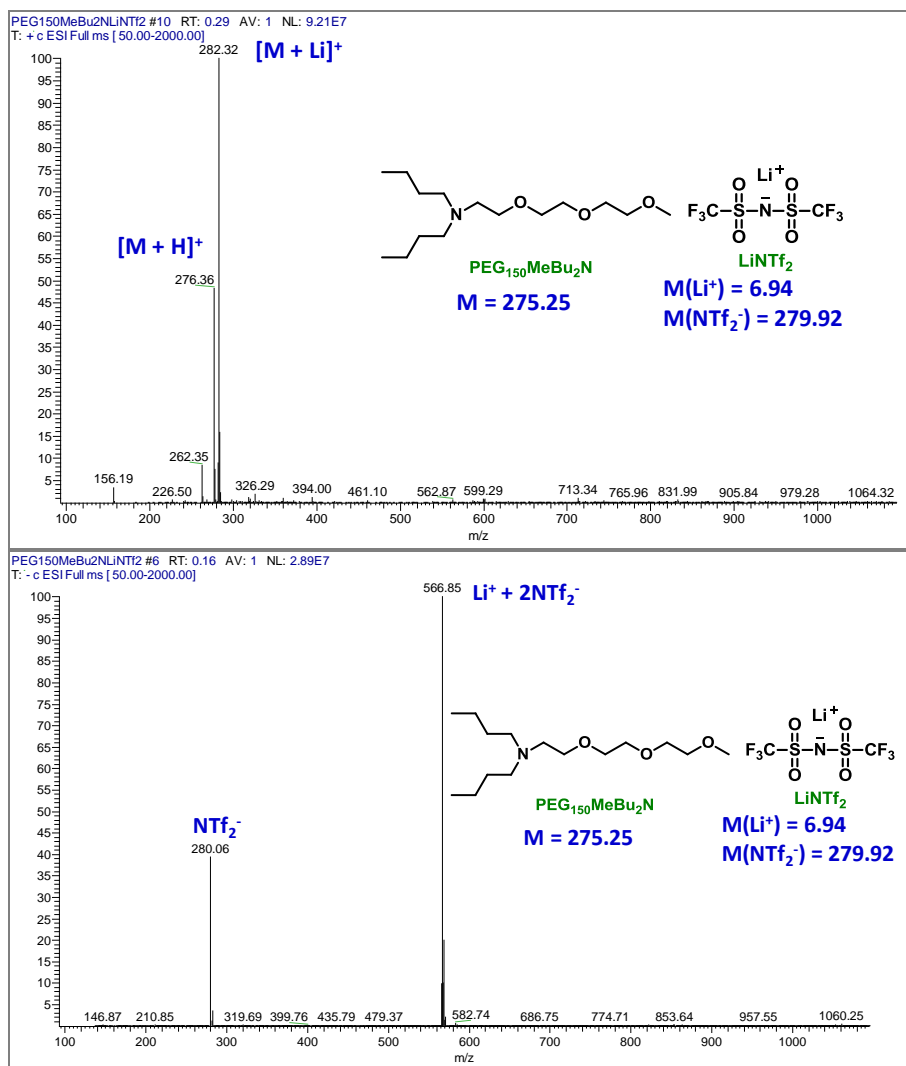


Figure S2 ESI spectrum of PEG₁₅₀MeTMG, PEG₁₅₀MeBu₂N before and after coordinating with LiNTf₂.

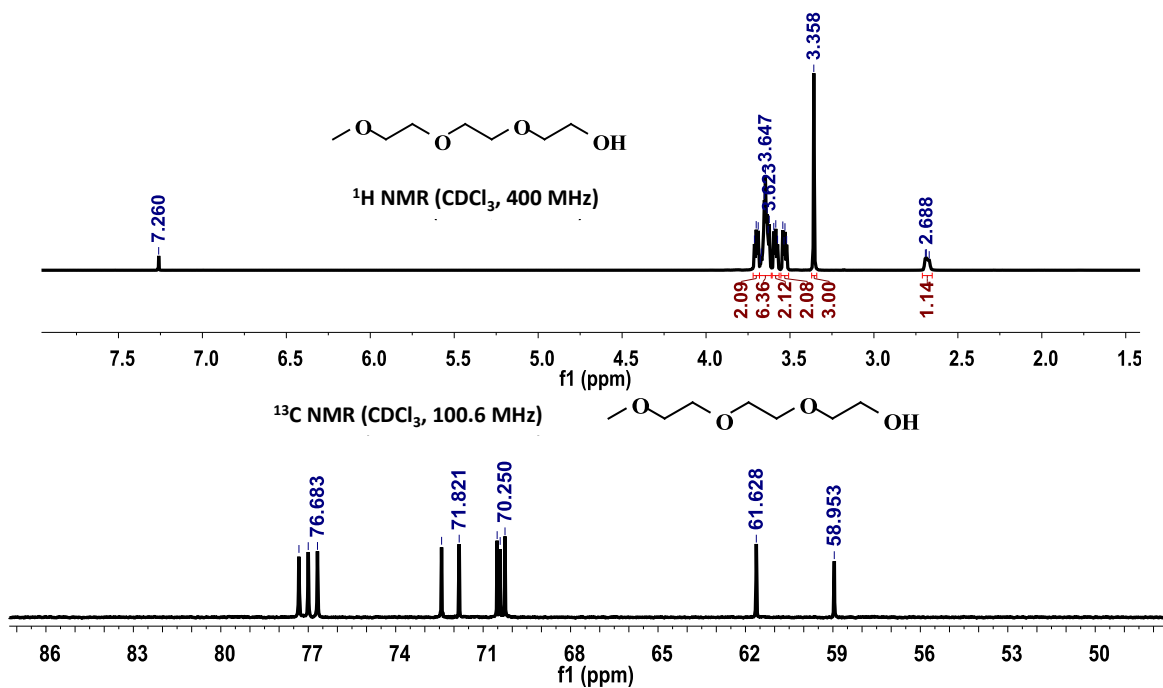
5. References

1. Z.-Z. Yang, L.-N. He, Y.-N. Zhao and B. Yu, *Environ. Sci. Technol.*, **2013**, 47, 1598-1605.
2. H. Yang, Z. Ma, Y. Qing, G. Xie, J. Gao, L. zhang, J. Gao and L. du, *Appl. Catal. A-Gen.*, **2010**, 382, 312-321.

6. NMR characterization and data on all neutral ligands and chelate ionic liquids

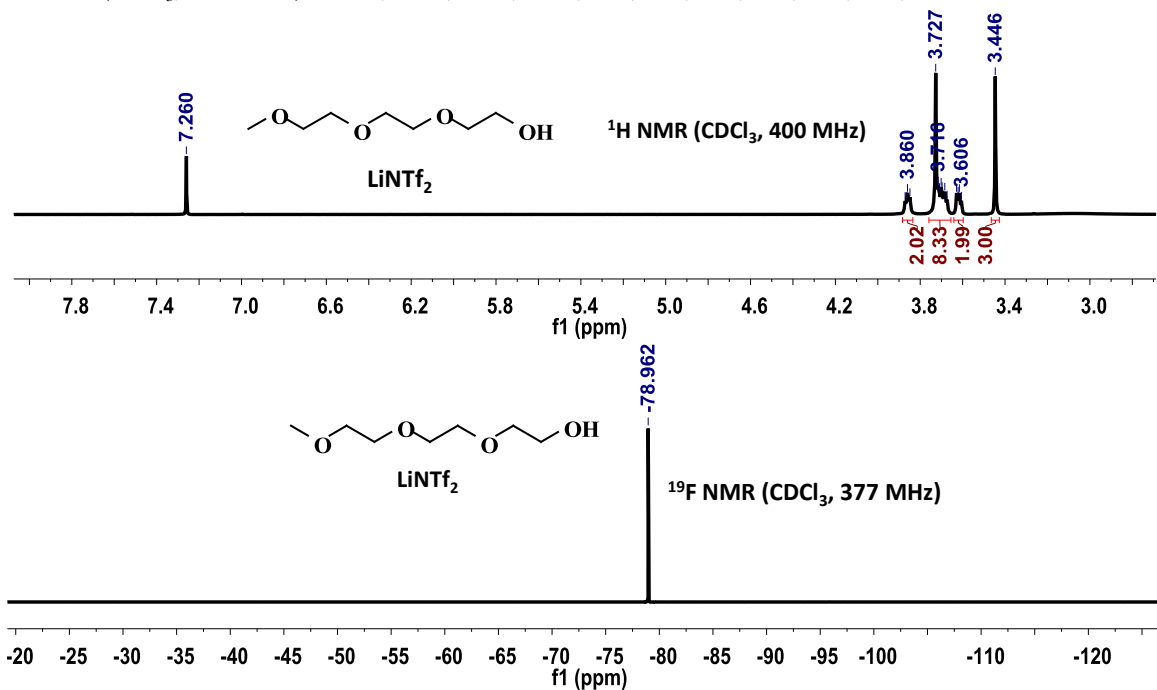
PEG₁₅₀Me

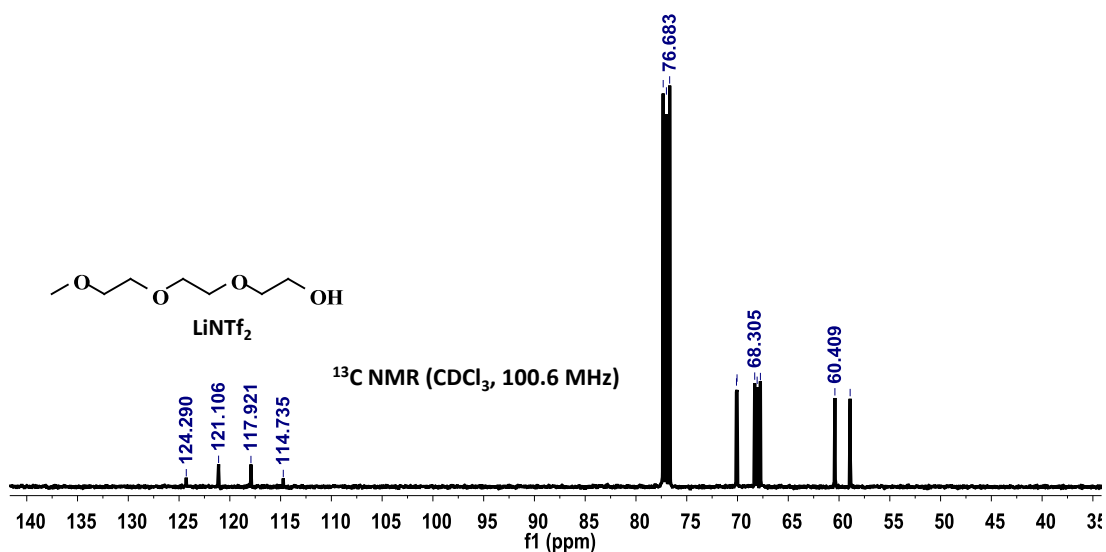
¹H NMR (CDCl₃, 400 MHz) δ 3.69-3.71 (m, 2H), 3.62-3.67 (m, 6H), 3.57-3.60 (m, 2H), 3.52-3.54 (m, 2H), 3.36 (s, 3H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 72.4, 71.8, 70.5, 70.4, 70.3, 61.6, 59.0.



PEG₁₅₀Me/LiNTf₂

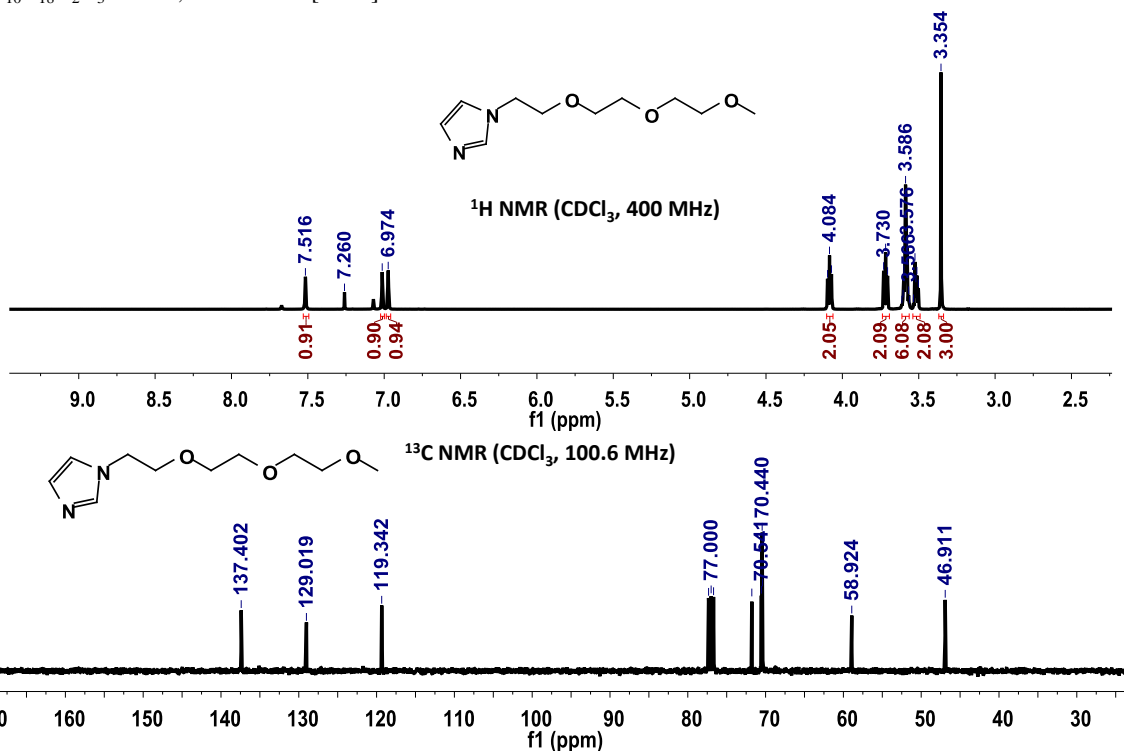
¹H NMR (CDCl₃, 400 MHz) δ 3.86 (t, ³J = 4.4 Hz, 2H), 3.67-3.73 (m, 8H), 3.61-3.63 (m, 2H), 3.45 (s, 3H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -79.0; ¹³C NMR (CDCl₃, 100.6 MHz) δ 124.3, 121.1, 117.9, 114.7, 70.1, 70.0, 68.3, 68.1, 67.7, 60.4, 58.9.





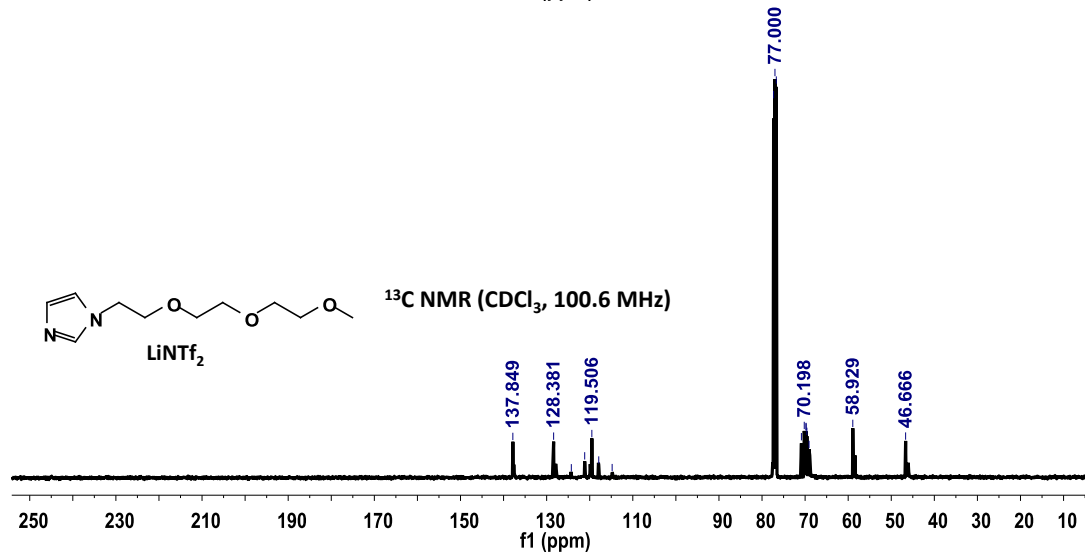
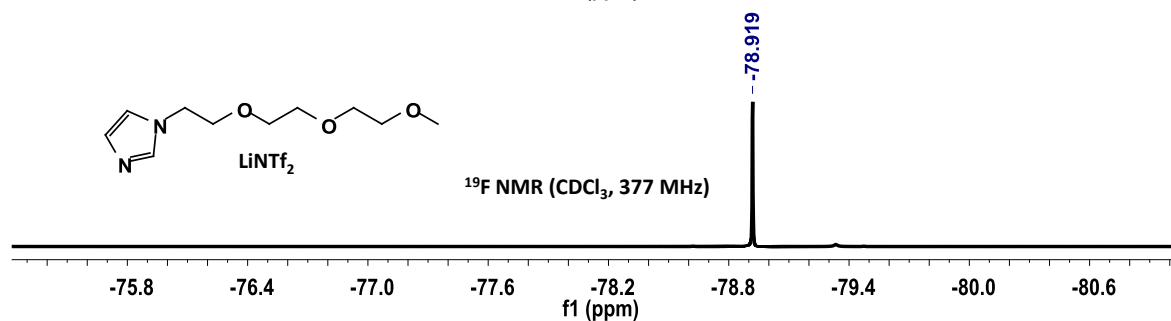
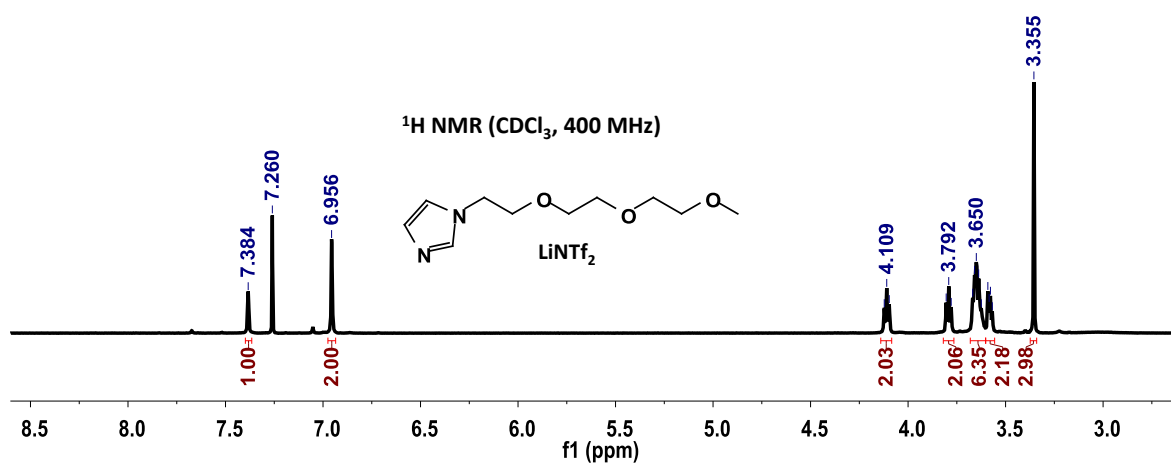
PEG₁₅₀MeIm

¹H NMR (CDCl₃, 400 MHz) δ 7.52 (s, 1H), 7.01 (s, 1H), 6.97 (s, 1H), 4.08 (t, ³J = 5.2 Hz, 2H), 3.72 (t, ³J = 5.2 Hz, 2H), 3.56-3.61 (m, 6H), 3.50-3.53 (m, 2H), 3.35 (s, 3H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 137.4, 129.0, 119.3, 71.8, 70.5, 70.4, 70.3, 58.9, 46.9. ESI-MS calcd for C₁₀H₁₈N₂O₃ 214.13, found 215.3 [M+H]⁺.



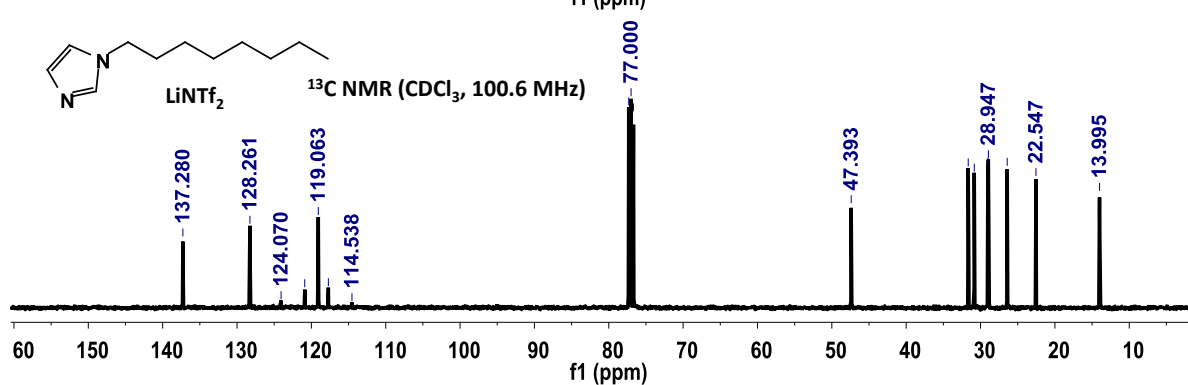
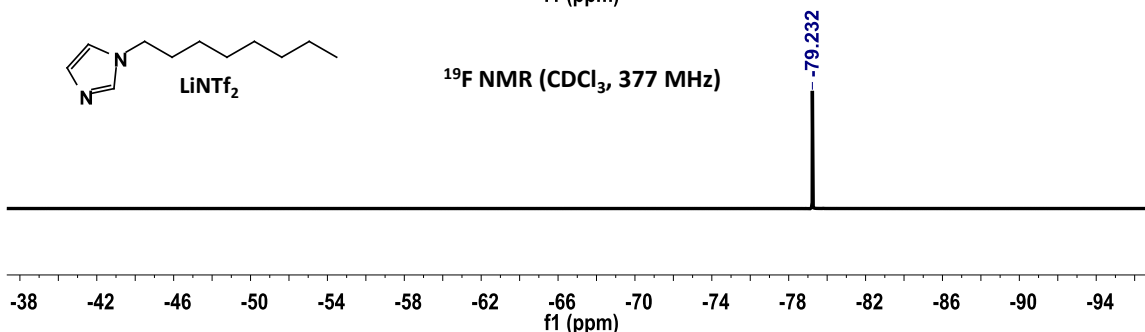
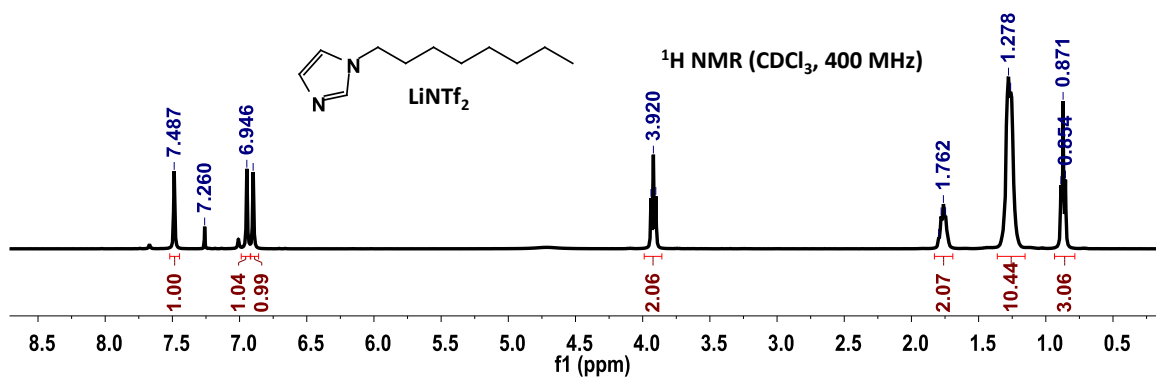
PEG₁₅₀MeIm/LiNTf₂

¹H NMR (CDCl₃, 400 MHz) δ 7.38 (s, 1H), 6.96 (s, 2H), 4.11 (t, ³J = 5.2 Hz, 2H), 3.79 (t, ³J = 5.2 Hz, 2H), 3.62-3.67 (m, 6H), 3.57-3.59 (m, 2H), 3.35 (s, 3H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -78.9; ¹³C NMR (CDCl₃, 100.6 MHz) δ 137.8, 128.4, 124.3, 121.2, 119.5, 117.9, 114.8, 70.9, 70.2, 69.8, 69.6, 69.1, 58.9, 46.7.



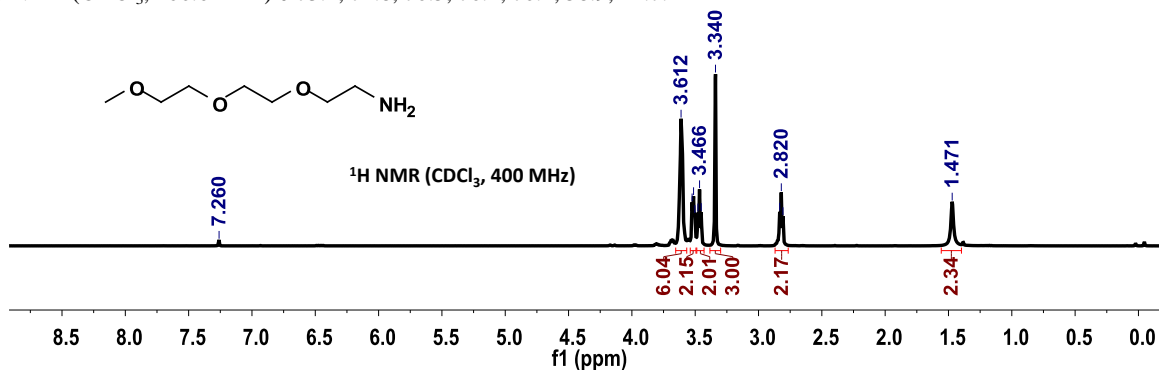
OctIm/LiNTf₂

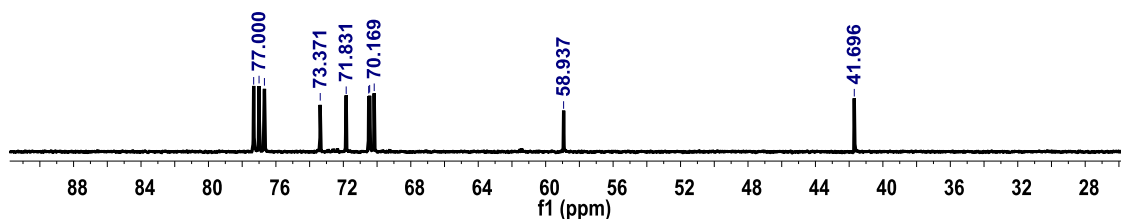
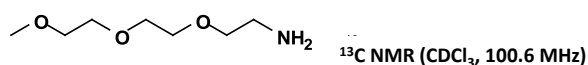
¹H NMR (CDCl₃, 400 MHz) δ 7.49 (s, 1H), 6.95 (s, 1H), 6.90 (s, 1H), 3.92 (t, ³J = 7.2 Hz, 2H), 1.75-1.80 (m, 2H), 1.26-1.28 (m, 10H), 0.87 (t, ³J = 6.4 Hz, 3H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -79.2; ¹³C NMR (CDCl₃, 100.6 MHz) δ 137.3, 128.3, 124.1, 120.9, 119.1, 117.7, 114.5, 47.4, 31.7, 30.8, 29.0, 28.9, 26.4, 22.5, 14.0.



PEG₁₅₀MeNH₂

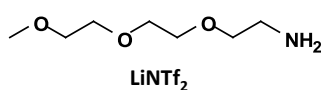
¹H NMR (CDCl₃, 400 MHz) δ 3.61 (s, 6H), 3.51 (t, ³J = 4 Hz, 2H), 3.47 (t, ³J = 4 Hz, 2H), 3.34 (s, 3H), 2.82 (t, ³J = 4 Hz, 2H), 1.47 (s, 2H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 73.4, 71.8, 70.5, 70.4, 70.2, 58.9, 41.7.



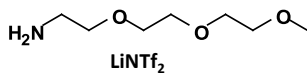
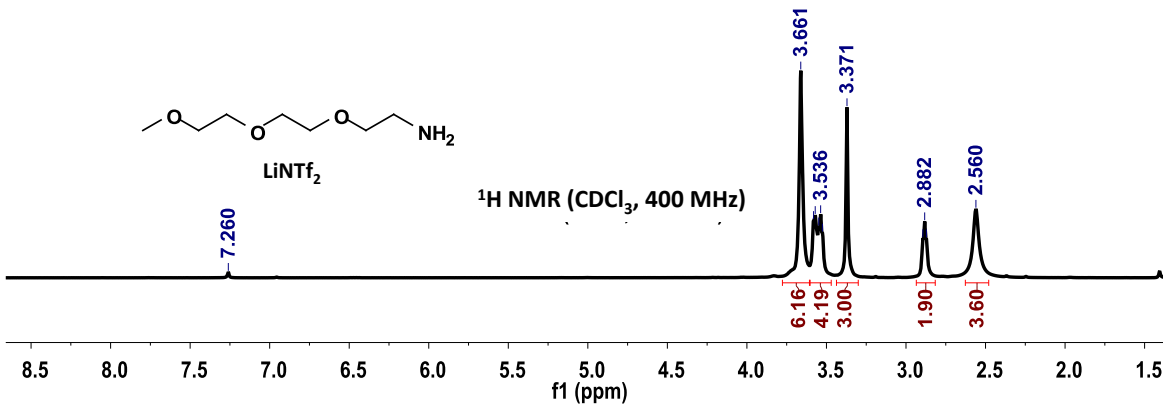


PEG₁₅₀MeNH₂/LiNTf₂

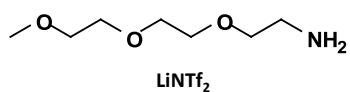
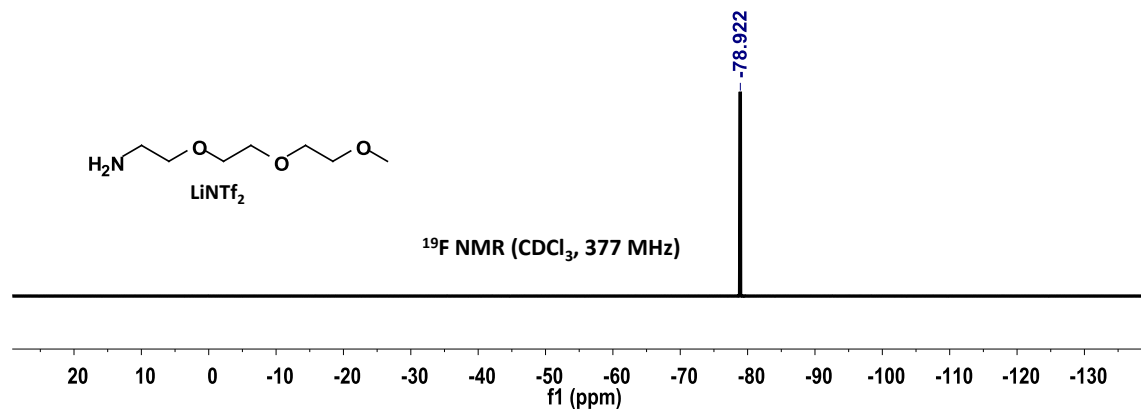
^1H NMR (CDCl_3 , 400 MHz) δ 3.66 (s, 6H), 3.53-3.58 (m, 4H), 3.37 (s, 3H), 2.88 (t, $^3J = 4$ Hz, 2H), 2.56 (s, 4H); ^{19}F NMR (CDCl_3 , 377 MHz) δ -78.9; ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 124.4, 121.2, 118.0, 114.8, 70.4, 70.3, 68.5, 68.3, 67.9, 58.7, 40.2.



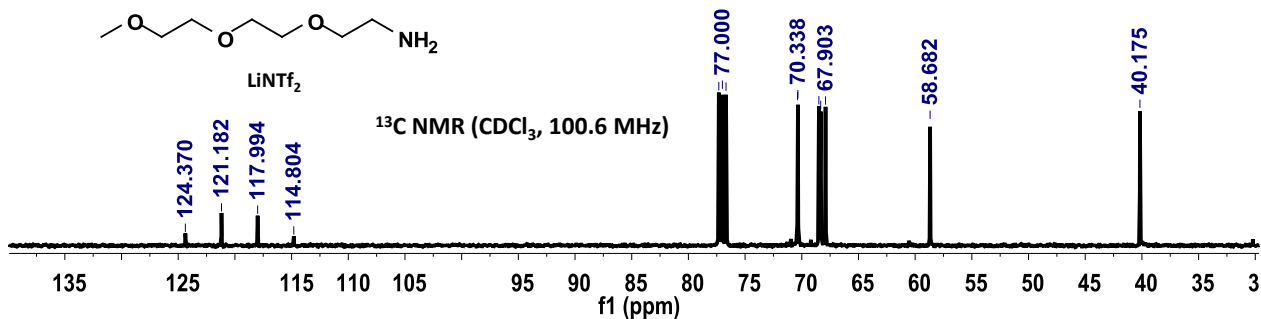
^1H NMR (CDCl_3 , 400 MHz)



^{19}F NMR (CDCl_3 , 377 MHz)

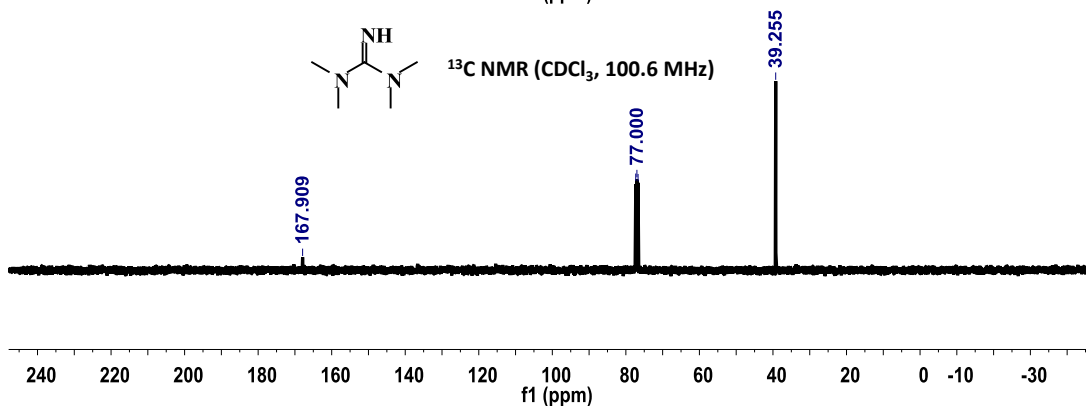
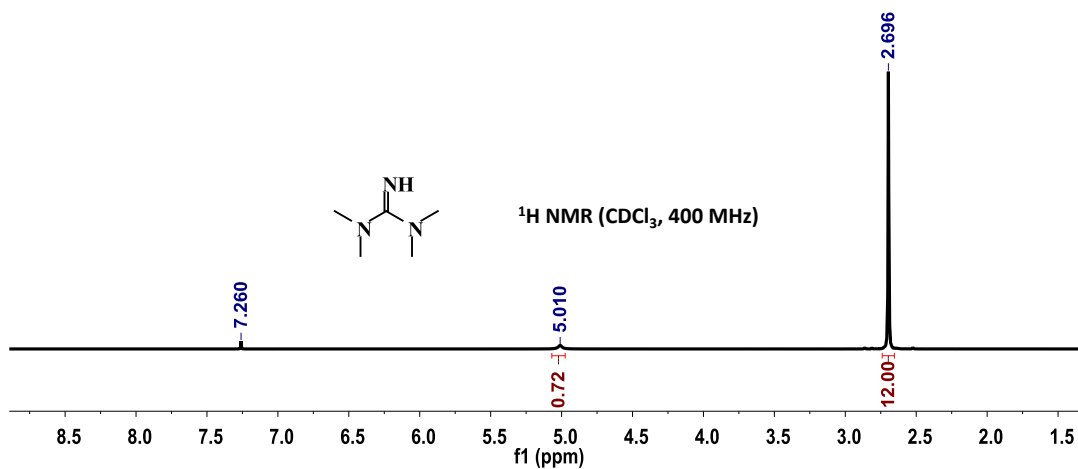


^{13}C NMR (CDCl_3 , 100.6 MHz)



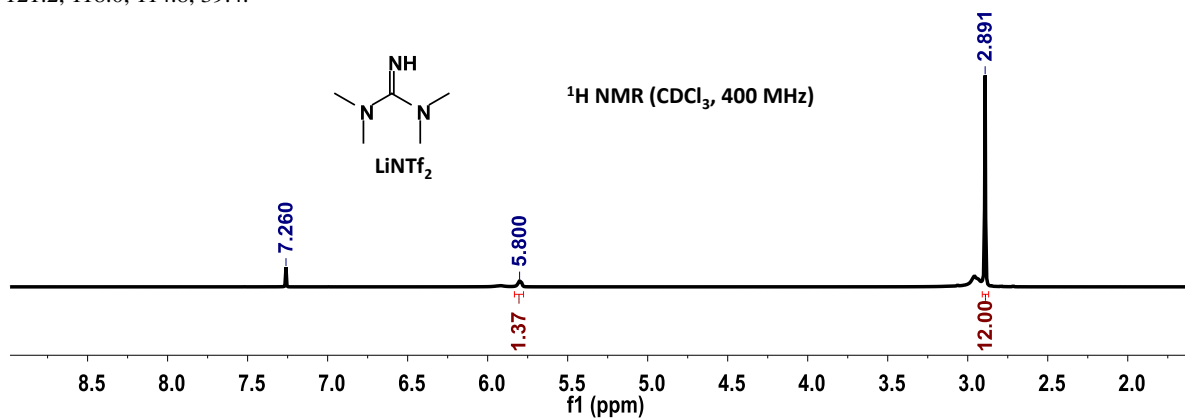
TMG

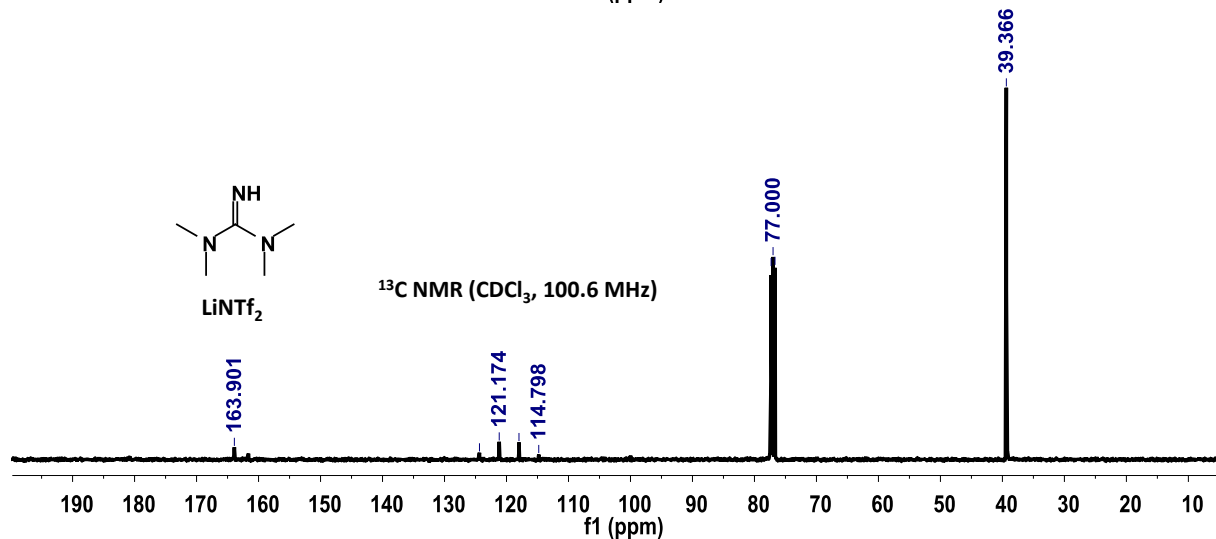
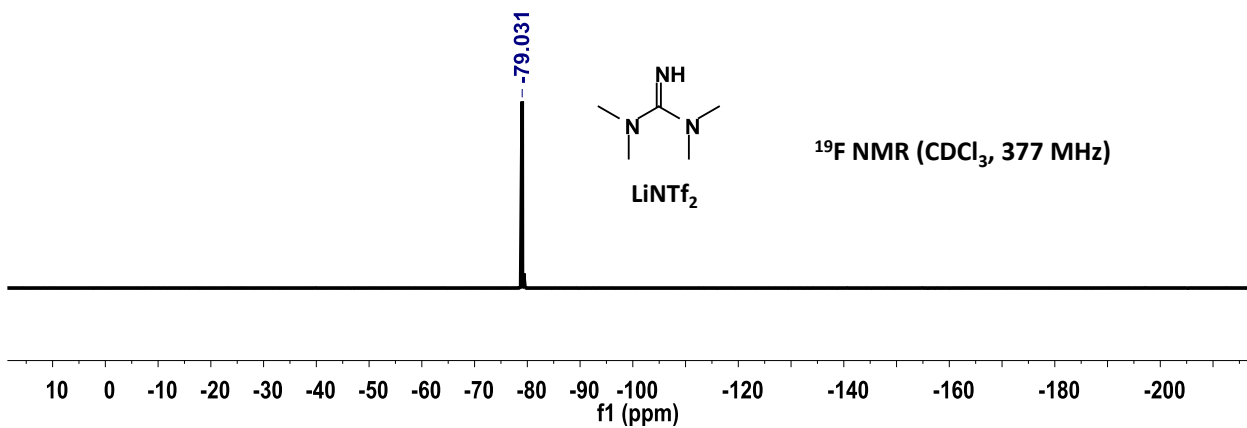
^1H NMR (CDCl_3 , 400 MHz) δ 5.01 (s, 1H), 2.70 (s, 12H); ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 167.9, 39.3.



TMG/LiNTf₂

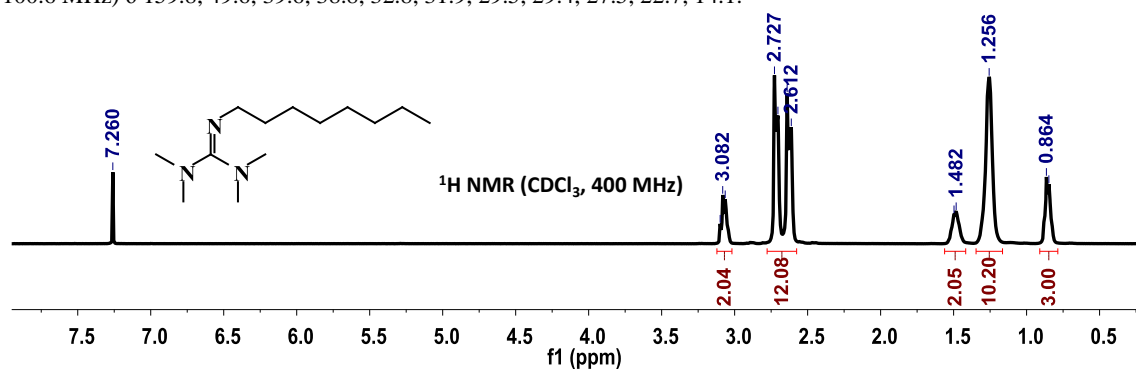
¹H NMR (CDCl₃, 400 MHz) δ 5.80 (s, 1H), 2.89 (s, 12H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -79.0; ¹³C NMR (CDCl₃, 100.6 MHz) δ 163.9, 124.4, 121.2, 118.0, 114.8, 39.4.

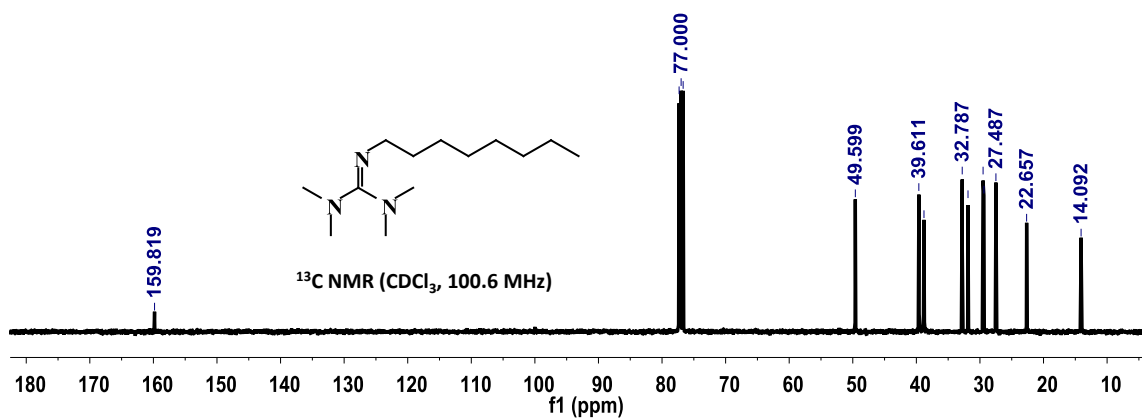




OctTMG

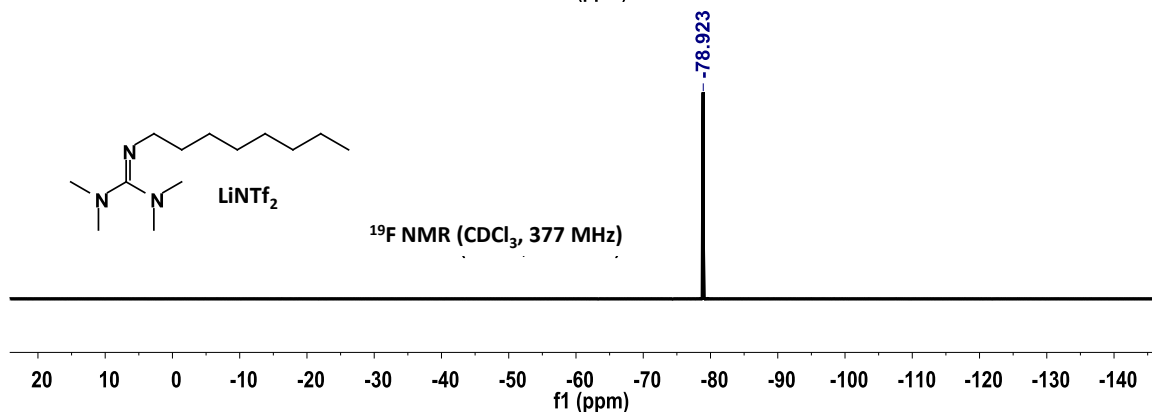
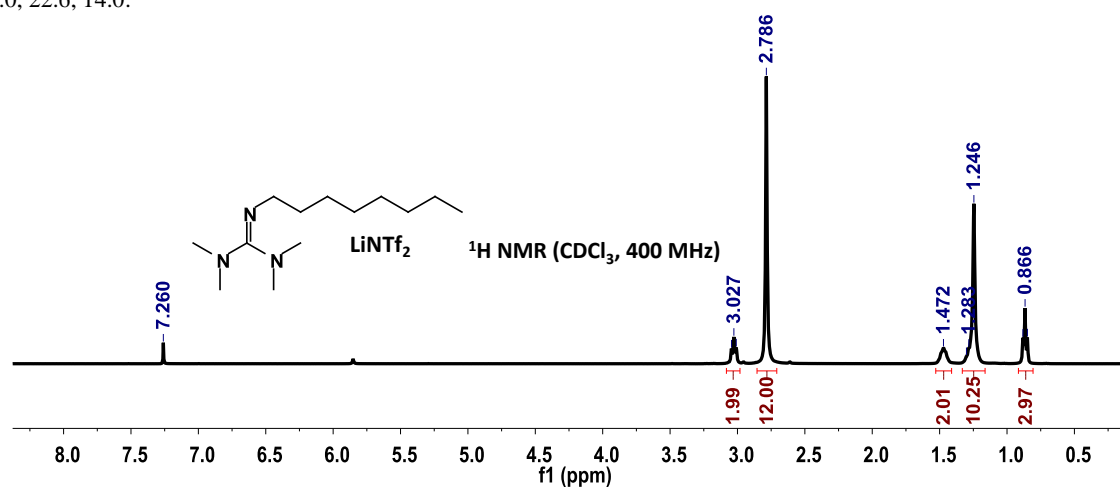
¹H NMR (CDCl₃, 400 MHz) δ 3.07-3.10 (m, 2H), 2.61-2.73 (m, 12H), 1.48-1.50 (m, 2H), 1.26 (m, 10H), 0.85-0.86 (m, 3H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 159.8, 49.6, 39.6, 38.8, 32.8, 31.9, 29.5, 29.4, 27.5, 22.7, 14.1.

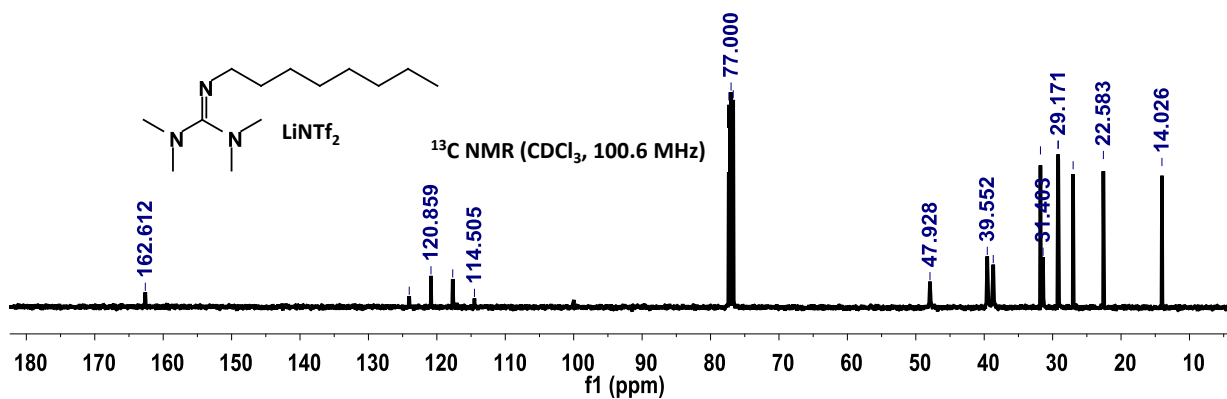




OctTMG/LiNTf₂

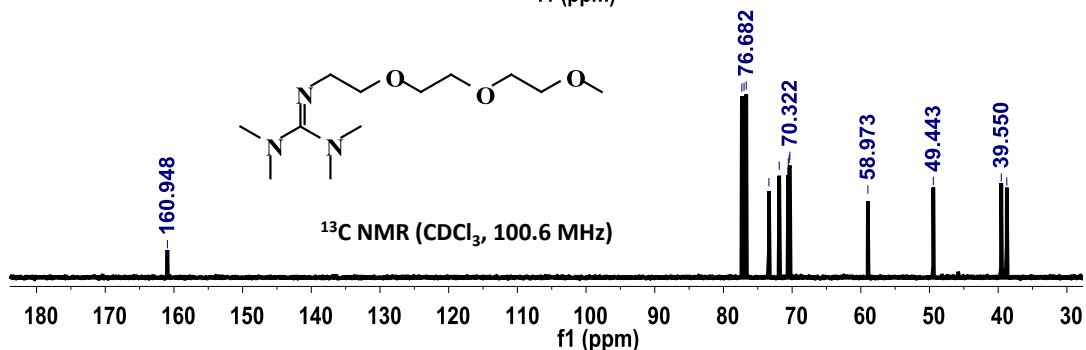
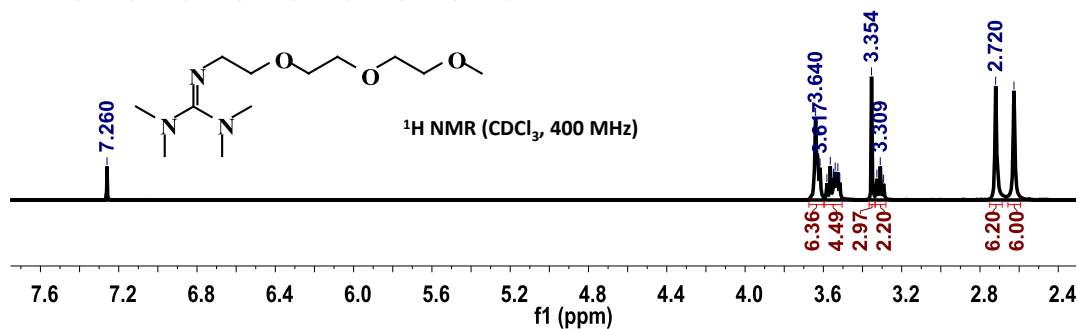
¹H NMR (CDCl₃, 400 MHz) δ 3.03 (t, ³J = 6.8 Hz, 2H), 2.79 (s, 12H), 1.47 (s, 2H), 1.25-1.30 (m, 10H), 0.87 (t, ³J = 6.4 Hz, 3H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -78.9; ¹³C NMR (CDCl₃, 100.6 MHz) δ 162.6, 124.0, 120.9, 117.7, 114.5, 47.9, 39.6, 38.7, 31.8, 31.4, 29.22, 29.17, 27.0, 22.6, 14.0.





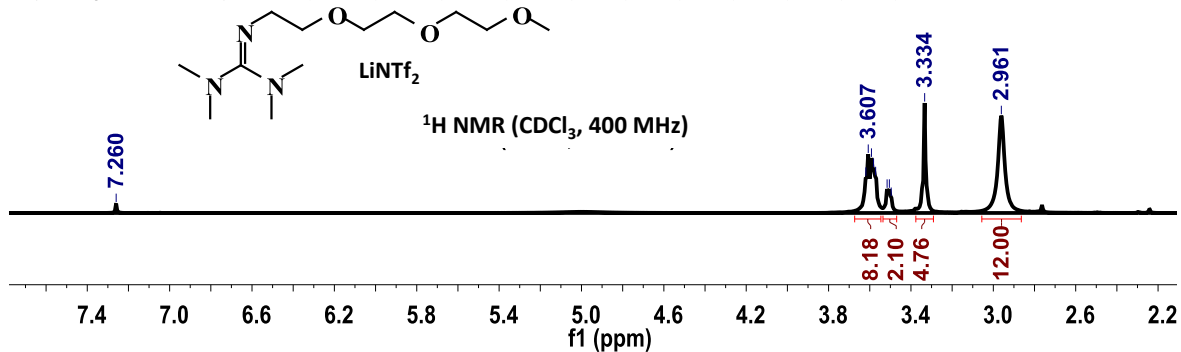
PEG₁₅₀MeTMG

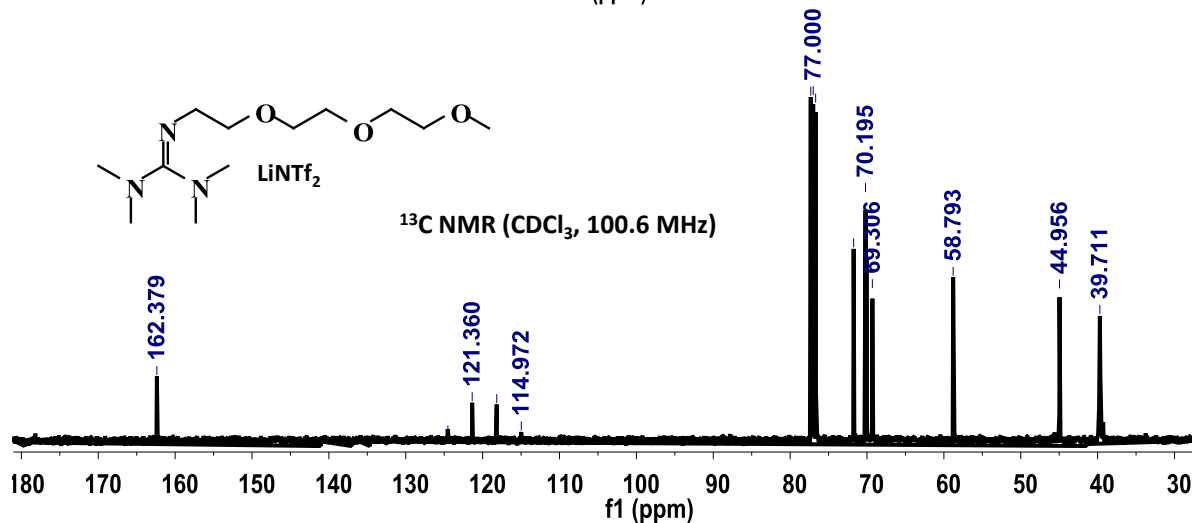
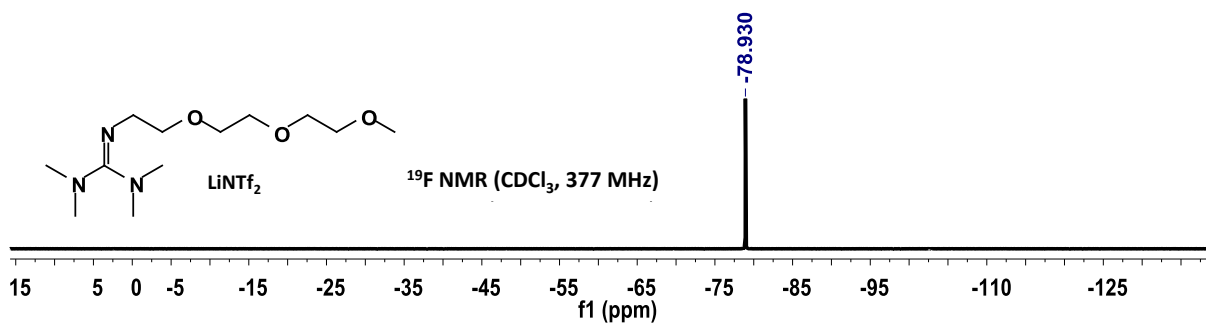
¹H NMR (CDCl₃, 400 MHz) δ 3.52-3.64 (m, 10H), 3.35 (s, 3H), 3.31 (t, ³J = 8 Hz, 2H), 2.72 (s, 6H), 2.63 (s, 6H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 160.9, 73.4, 71.9, 70.6, 70.4, 70.3, 59.0, 49.4, 39.6, 38.8.



PEG₁₅₀MeTMG/LiNTf₂

¹H NMR (CDCl₃, 400 MHz) δ 3.57-3.62 (m, 8H), 3.50-3.52 (m, 2H), 3.33 (s, 5H), 2.96 (s, 12H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -78.9; ¹³C NMR (CDCl₃, 100.6 MHz) δ 162.4, 124.5, 121.4, 118.2, 115.0, 71.7, 70.2, 70.1, 69.3, 58.8, 45.0, 39.7.

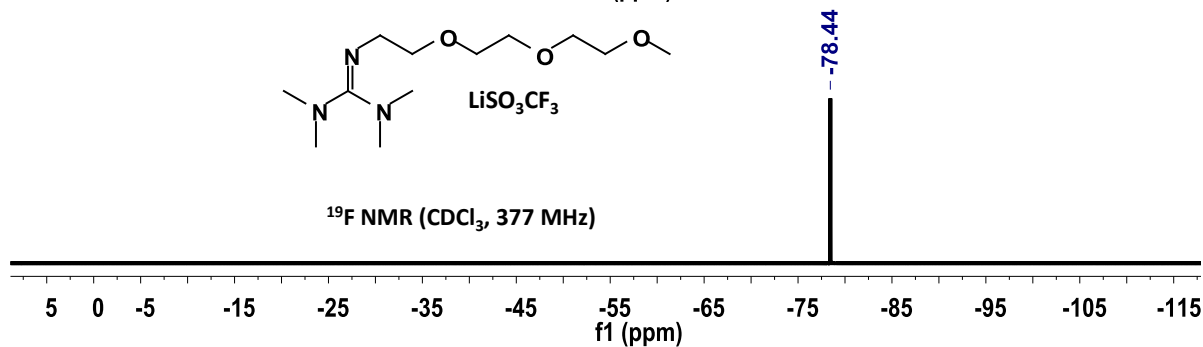
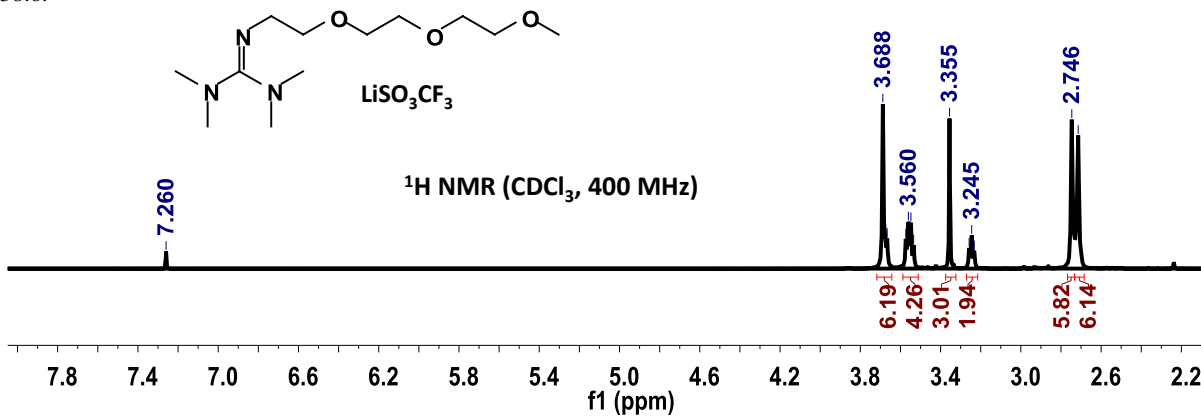


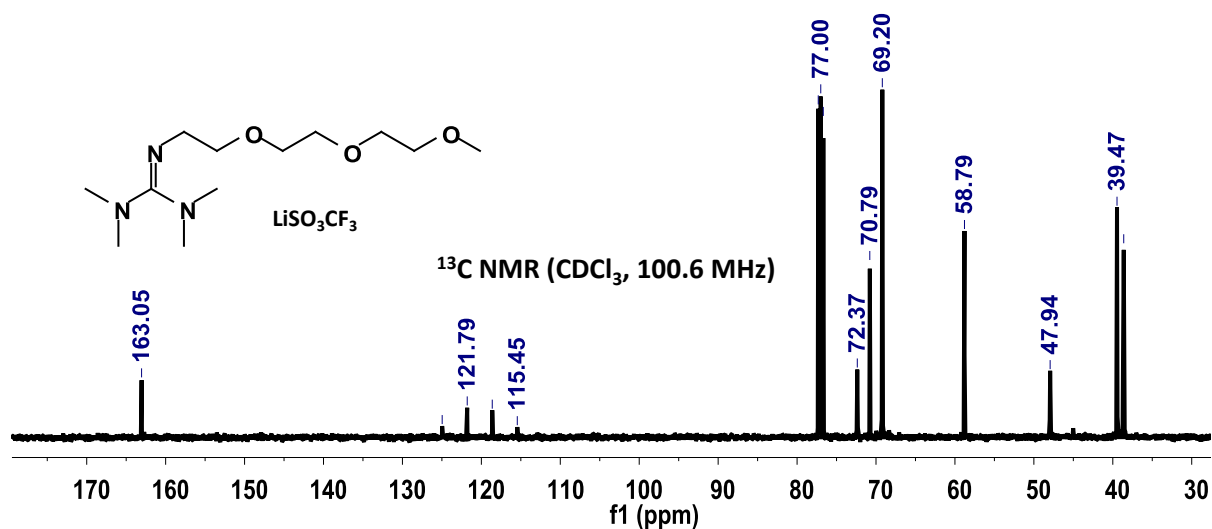


PEG₁₅₀MeTMG/LiSO₃CF₃

¹H NMR (CDCl₃, 400 MHz) δ 3.67-3.69 (m, 6H), 3.53-3.57 (m, 4H), 3.36 (s, 3H), 3.24 (t, ³J = 5.2 Hz, 2H), 2.75 (s, 6H), 2.71 (s, 6H);

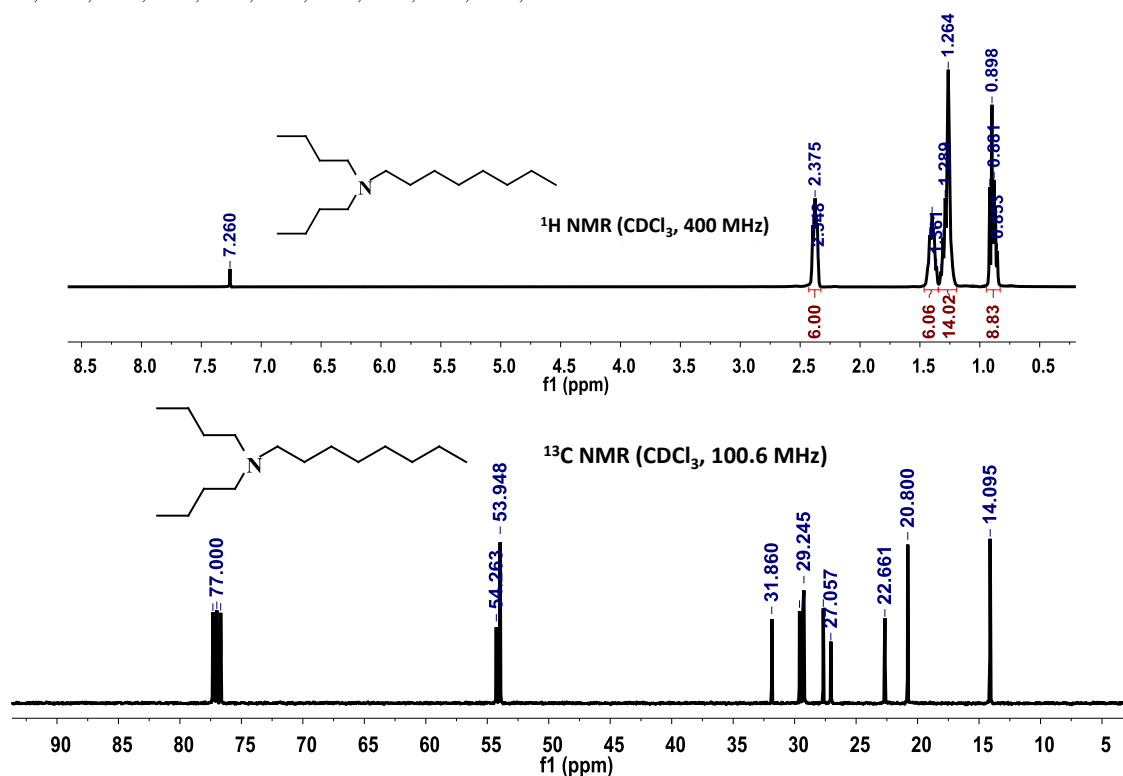
¹⁹F NMR (CDCl₃, 377 MHz) δ -78.4; ¹³C NMR (CDCl₃, 100.6 MHz) δ 163.1, 125.0, 121.8, 118.6, 115.5, 72.4, 70.8, 69.2, 58.8, 47.9, 39.5, 38.6.





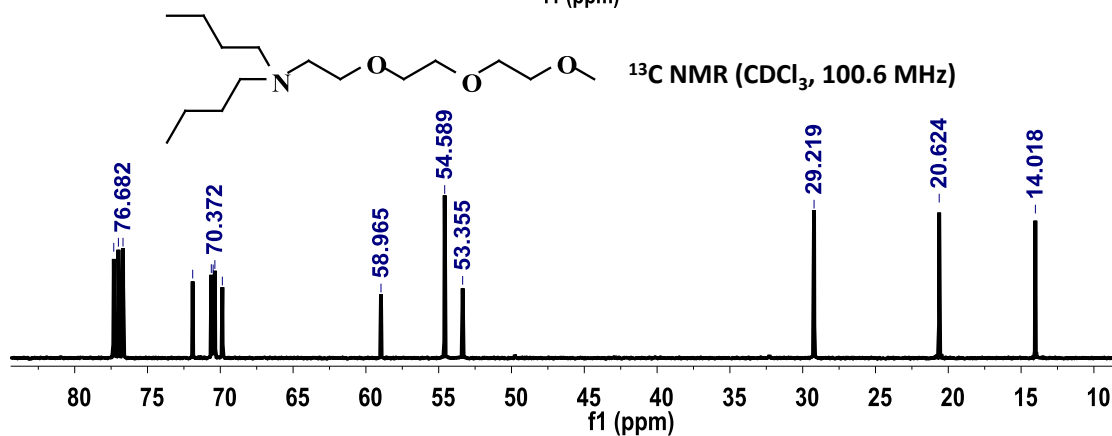
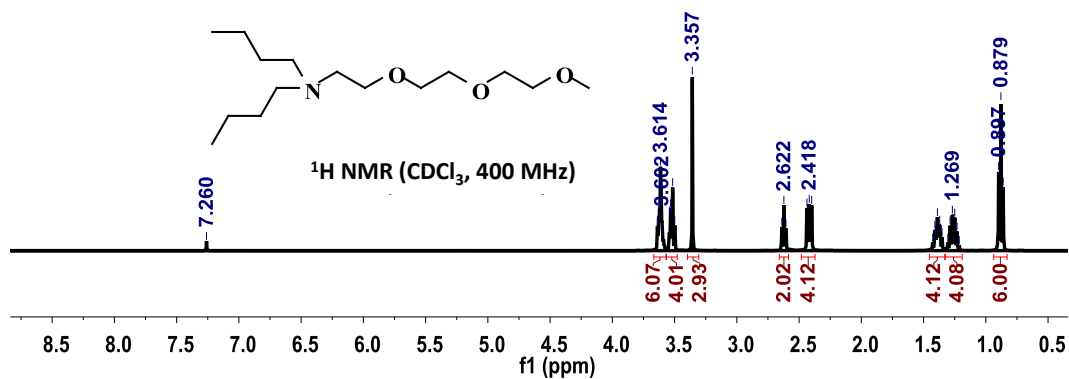
OctBu₂N

¹H NMR (CDCl₃, 400 MHz) δ 2.35-2.39 (m, 6H), 1.36-1.43 (m, 6H), 1.26-1.33 (m, 14H), 0.85-0.92 (m, 9H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 54.3, 53.9, 31.9, 29.6, 29.3, 29.2, 27.7, 27.1, 22.7, 20.8, 14.1.



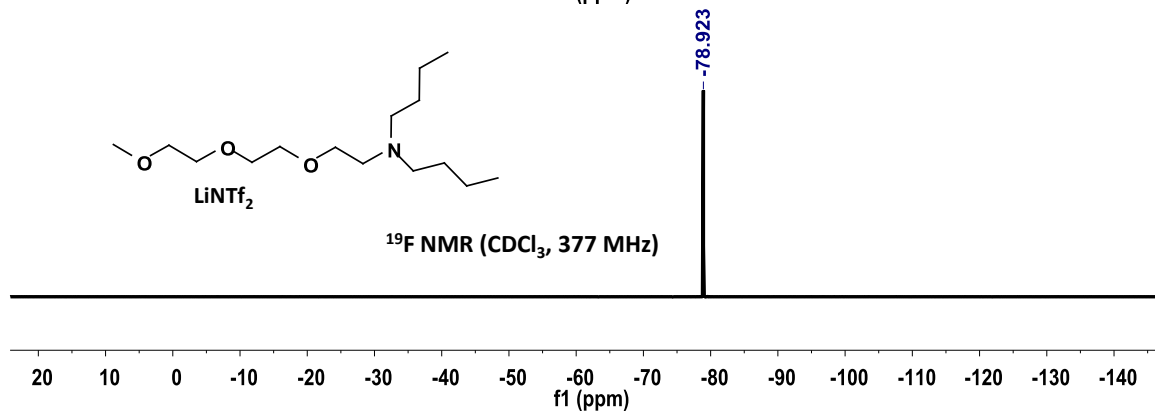
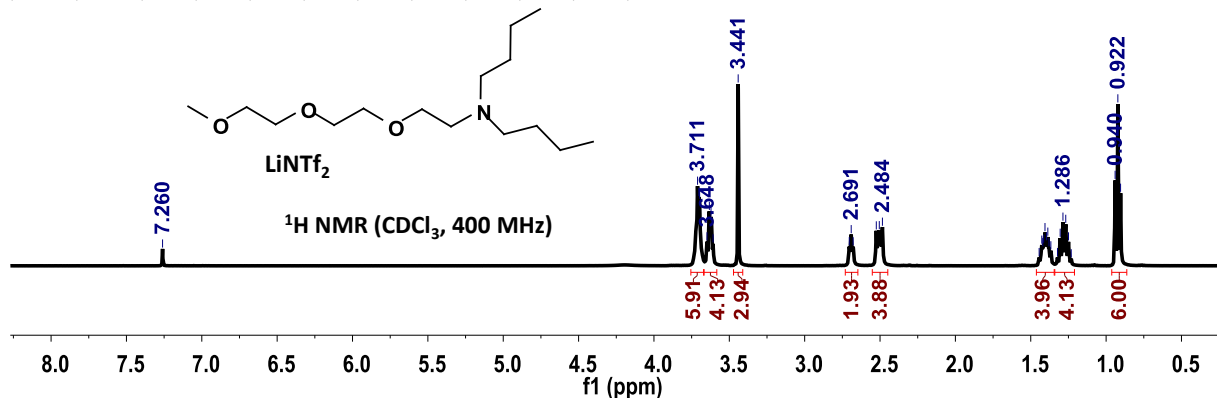
PEG₁₅₀MeBu₂N

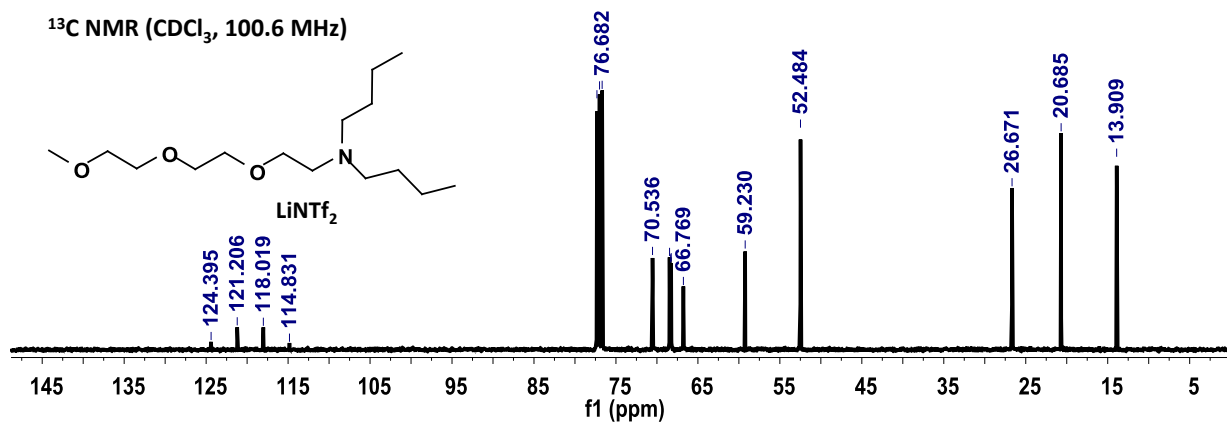
¹H NMR (CDCl₃, 400 MHz) δ 3.60-3.64 (m, 6H), 3.50-3.54 (m, 4H), 3.36 (s, 3H), 2.62 (t, ³J = 6.8 Hz, 2H), 2.42 (t, ³J = 7.6 Hz, 4H), 1.35-1.43 (m, 4H), 1.21-1.30 (m, 4H), 0.88 (t, ³J = 7.2 Hz, 6H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 71.9, 70.6, 70.5, 70.4, 69.9, 59.0, 54.6, 53.4, 29.2, 20.6, 14.0.



PEG₁₅₀MeBu₂N/LiNTf₂

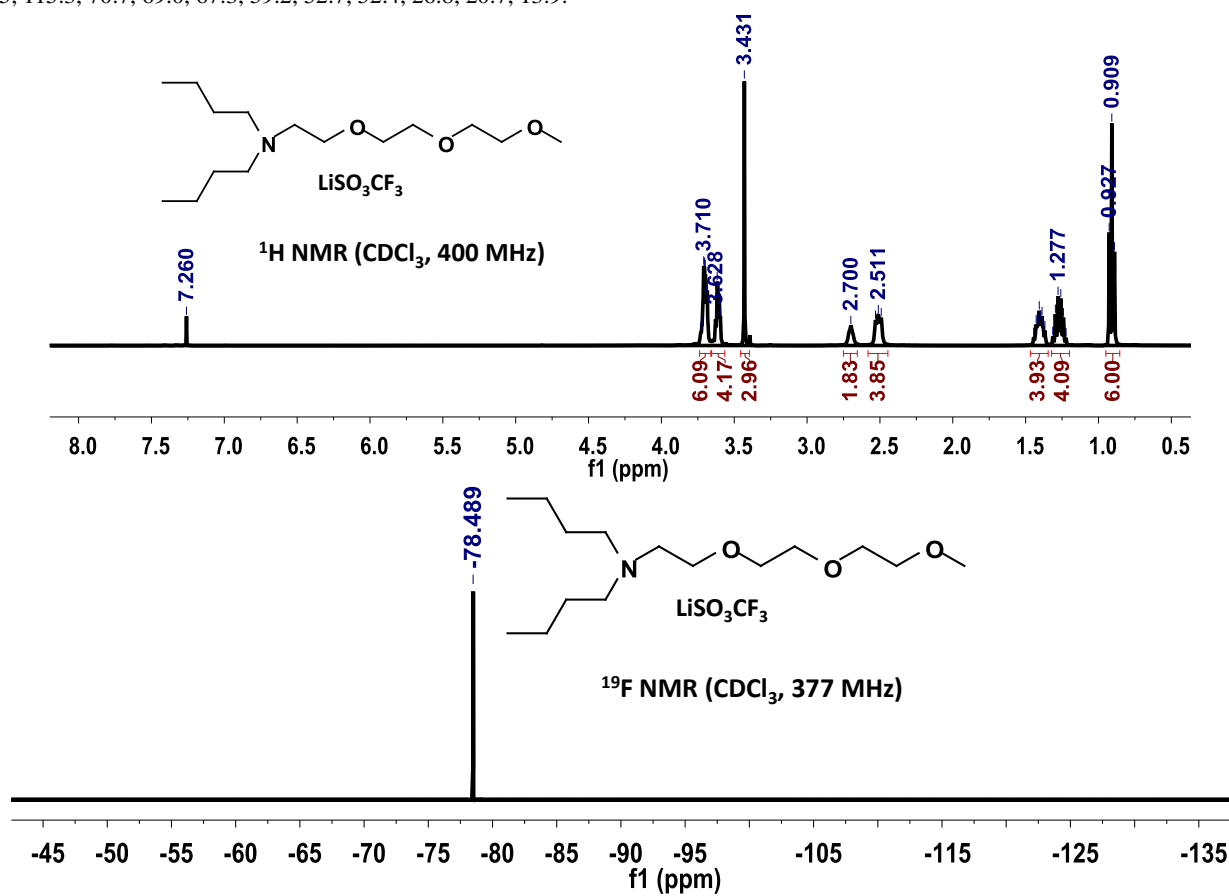
¹H NMR (CDCl₃, 400 MHz) δ 3.70-3.71 (m, 6H), 3.61-3.65 (m, 4H), 3.44 (s, 3H), 2.69 (t, ³J = 5.2 Hz, 2H), 2.51 (t, ³J = 8 Hz, 4H), 1.37-1.44 (m, 4H), 1.23-1.32 (m, 4H), 0.92 (t, ³J = 7.2 Hz, 6H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -78.9; ¹³C NMR (CDCl₃, 100.6 MHz) δ 124.4, 121.2, 118.0, 114.8, 70.5, 68.5, 68.3, 68.2, 66.8, 59.2, 52.5, 26.7, 20.7, 13.9.

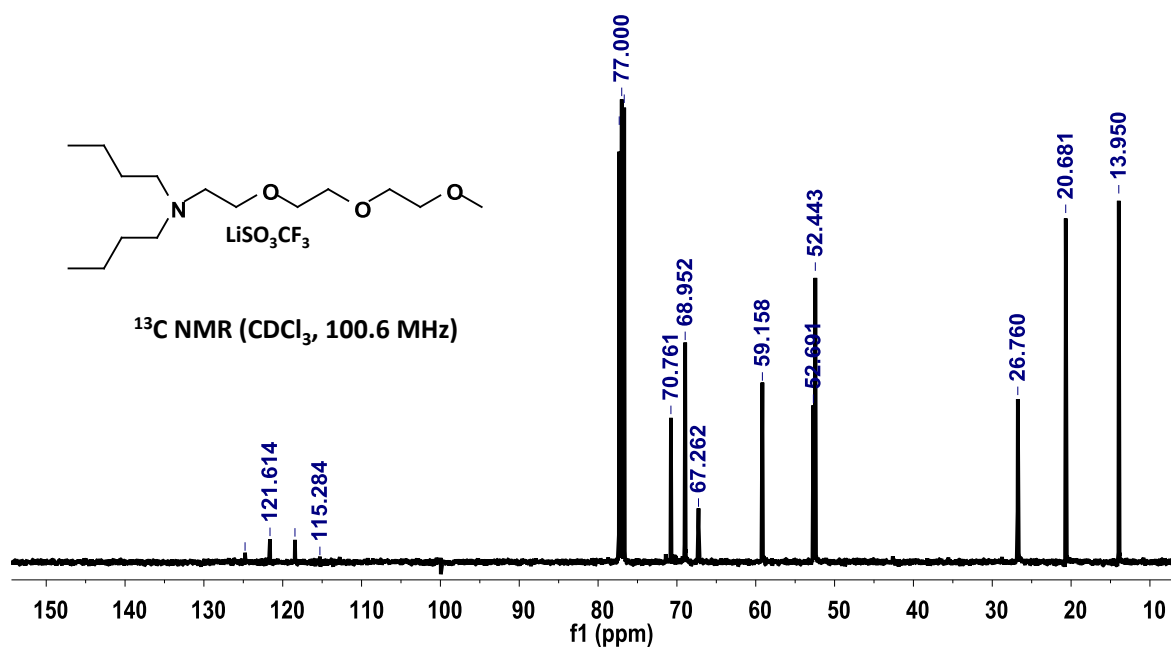




PEG₁₅₀MeBu₂N/LiSO₃CF₃

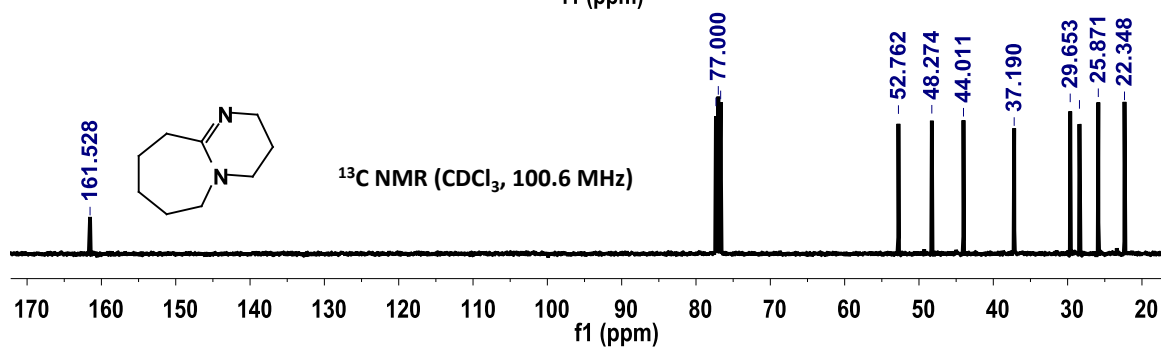
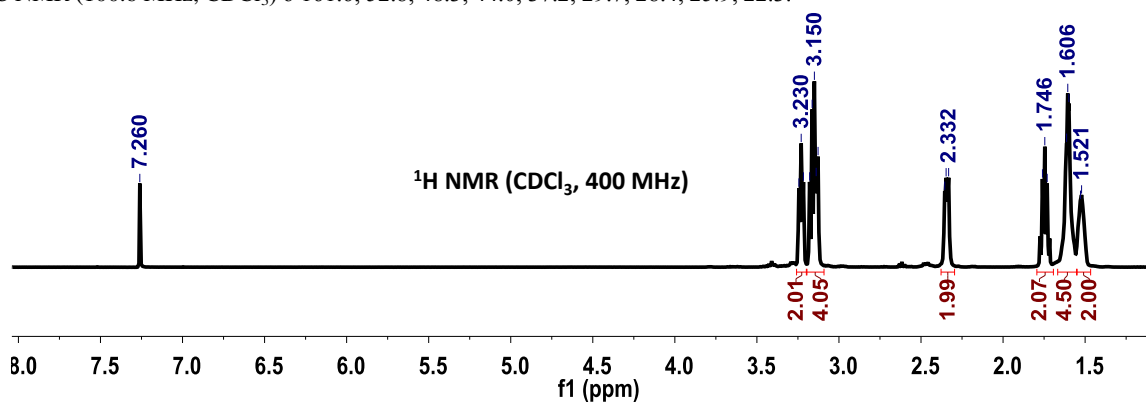
¹H NMR (CDCl₃, 400 MHz) δ 3.68-3.73 (m, 6H), 3.59-3.63 (m, 4H), 3.43 (s, 3H), 2.70 (s, 2H), 2.51 (t, ³J = 7.6 Hz, 4H), 1.37-1.44 (m, 4H), 1.22-1.31 (m, 4H), 0.91 (t, ³J = 7.2 Hz, 6H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -78.5; ¹³C NMR (CDCl₃, 100.6 MHz) δ 124.8, 121.6, 118.5, 115.3, 70.7, 69.0, 67.3, 59.2, 52.7, 52.4, 26.8, 20.7, 13.9.





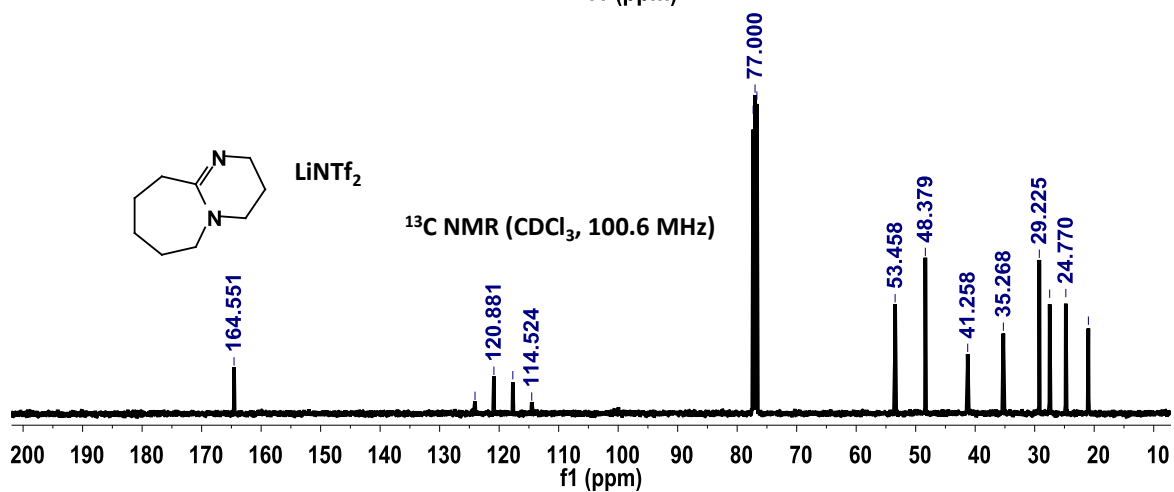
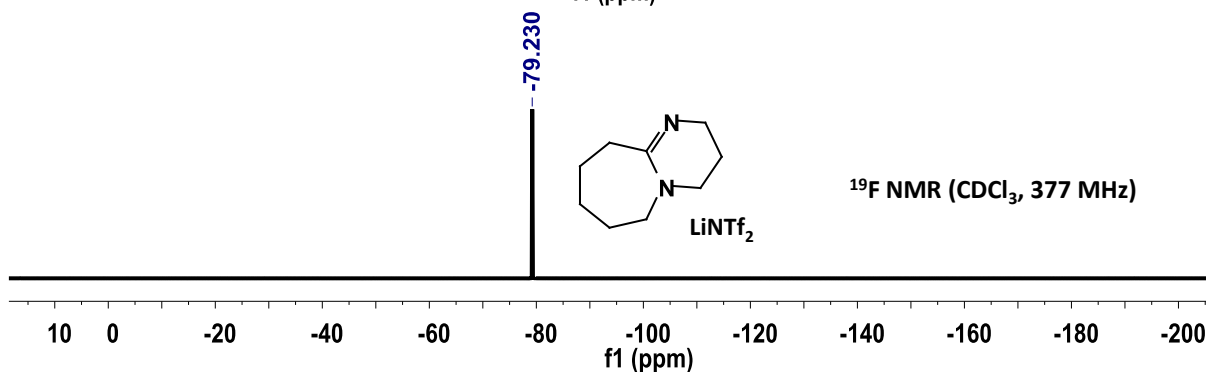
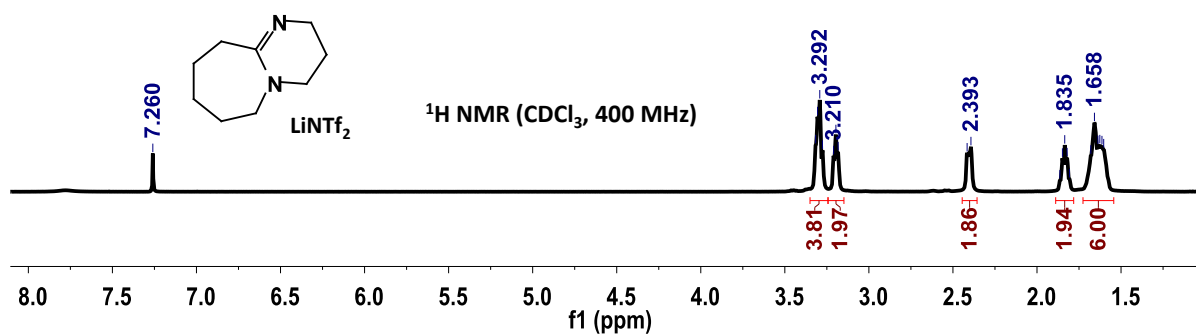
DBU (1,8-diazabicyclo[5.4.0]undec-7-ene):

¹H NMR (400 MHz, CDCl₃) δ 3.23 (t, ³J = 5.6 Hz, 2 H), 3.13-3.18 (m, 4 H), 2.33-2.35 (m, 2 H), 1.72-1.77 (m, 2 H), 1.60 (s, 4 H), 1.52 (s, 2 H); ¹³C NMR (100.6 MHz, CDCl₃) δ 161.6, 52.8, 48.3, 44.0, 37.2, 29.7, 28.4, 25.9, 22.3.



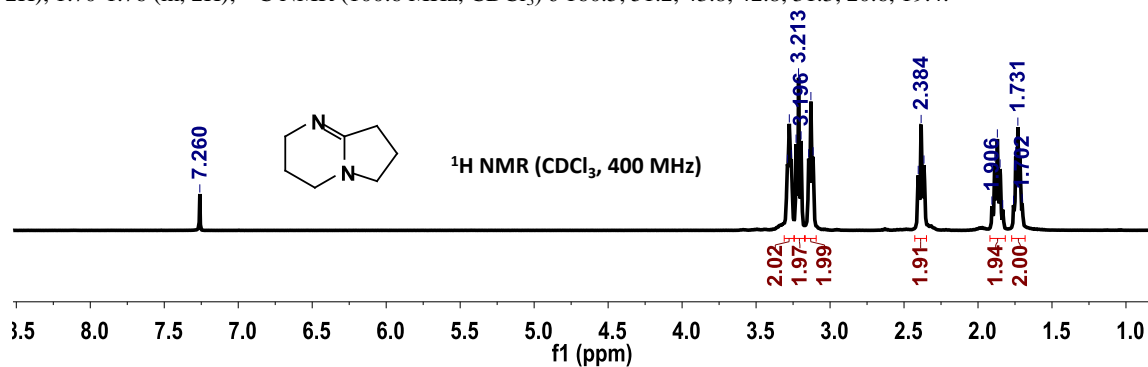
DBU/LiNTf₂

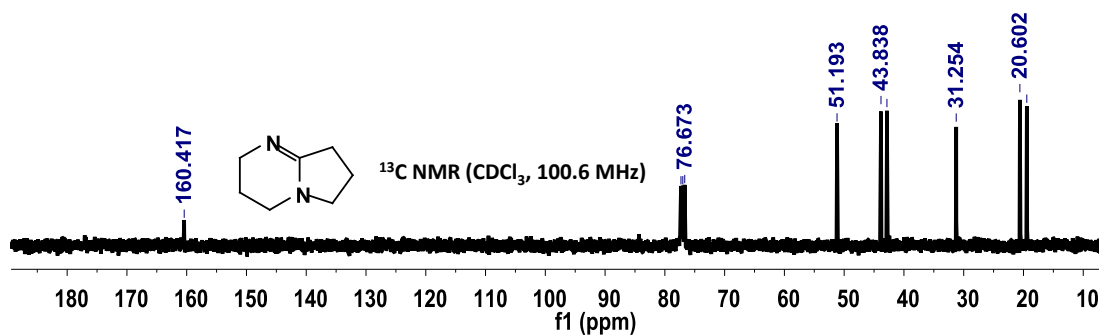
¹H NMR (CDCl₃, 400 MHz) δ 3.28-3.32 (m, 4H), 3.20 (t, 3J = 5.6 Hz, 2H), 2.39-2.42 (m, 2H), 1.81-1.86 (m, 2H), 1.60-1.68 (m, 6H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -79.2; ¹³C NMR (CDCl₃, 100.6 MHz) δ 164.6, 124.1, 120.9, 117.7, 114.5, 53.5, 48.4, 41.3, 35.3, 29.2, 27.5, 24.8, 21.0.



DBN (1,5-diazabicyclo[4.3.0]non-5-ene):

¹H NMR (400 MHz, CDCl₃) δ 3.28 (t, ³J = 4.8 Hz, 2H), 3.21 (t, ³J = 6.8 Hz, 2H), 3.13 (t, ³J = 6 Hz, 2H), 2.38 (t, ³J = 7.6 Hz, 2H), 1.83-1.91 (m, 2H), 1.70-1.76 (m, 2H); ¹³C NMR (100.6 MHz, CDCl₃) δ 160.5, 51.2, 43.8, 42.8, 31.3, 20.6, 19.4.





DBN/LiNTf₂

¹H NMR (400 MHz, CDCl₃) δ 3.41 (t, 3J = 7.2 Hz, 2H), 3.25 (t, 3J = 6 Hz, 4H), 2.56 (t, 3J = 8 Hz, 2H), 1.96-2.04 (m, 2H), 1.81-1.87 (m, 2H); ¹⁹F NMR (CDCl₃, 377 MHz) δ -79.2; ¹³C NMR (100.6 MHz, CDCl₃) δ 163.2, 124.1, 121.0, 117.8, 114.6, 52.3, 42.8, 41.2, 30.6, 19.7, 19.1.

