

Supporting information

for

Synthesis of ergostane-type brassinosteroids with modifications in ring A

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General information, experimental details, characterization data and copies of ¹H and ¹³C NMR spectra

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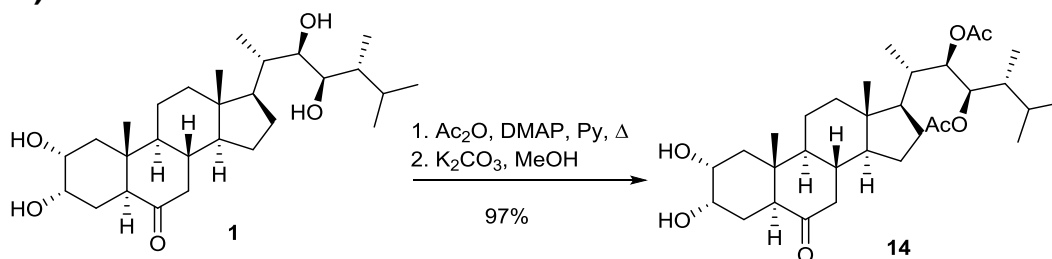
Experimental section

General remarks

All reactions that required anhydrous conditions were carried out under a positive argon flow with appropriately dried glassware, reagents and solvents. Commercially available reagents were used as received. Petroleum ether (PE) used had a boiling range of 60–90 °C. Reactions were monitored by TLC on silica gel GF₂₅₄ plates (Merck). Column chromatography was performed using silica gel (200–300 mesh). One and two-dimensional nuclear magnetic resonance (NMR) spectra were obtained in CDCl₃ on a Bruker AVANCE 500 spectrometer. Chemical shift values are given in δ (ppm) relative to the residual solvent peaks: δ_{H} 7.26 and δ_{C} 77.0. Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), ddd (doublet of doublet of doublets). High resolution mass spectra were recorded on a LTQ Orbitrap mass spectrometer coupled to an Accela HPLC System (HPLC column: Hypersyl GOLD, 50 mm \times 1 mm, 1.9 μm).

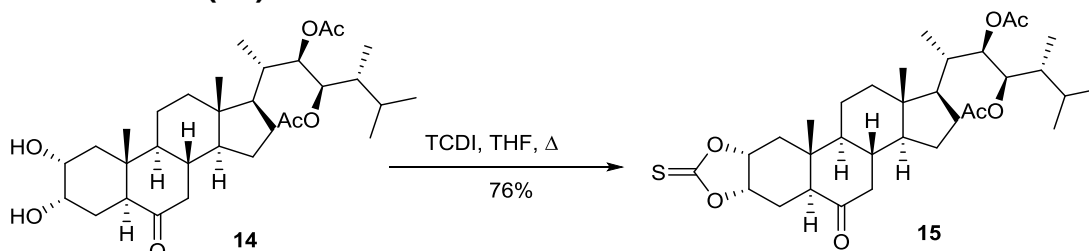
Procedures and spectroscopic analytical data

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-2 α ,3 α -dihydroxy-24-methyl-5 α -cholestan-6-one (**14**)



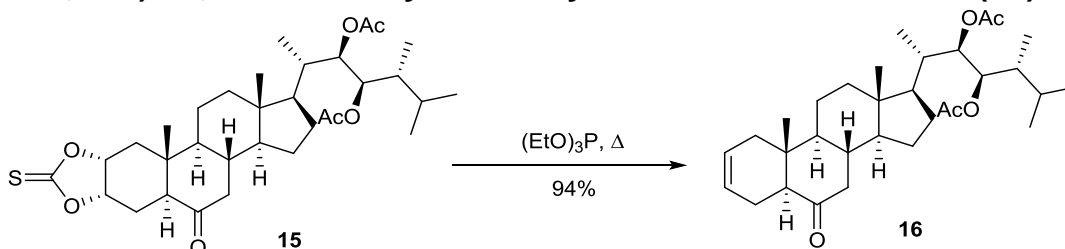
To a solution of epicastasterone (**1**, 406 mg, 0.874 mmol) in pyridine (2 mL), Ac₂O (495 μL , 5.24 mmol) and DMAP (8.5 mg, 0.070 mmol) were added. The mixture was kept at 60 °C for 16 h, then the solvent was evaporated in vacuo, the residue was dissolved in CHCl₃ (10 mL) and washed with saturated NaHCO₃ (3 \times 3 mL). The organic layers were dried over Na₂SO₄ and evaporated. The crude tetraacetate (609 mg) was dissolved in MeOH (8.6 mL), and then a solution of K₂CO₃ (483 mg, 3.49 mmol) in water (3 mL) was added. The mixture was stirred at 20 °C for 1 h, diluted with water (10 mL), and extracted with CHCl₃ (3 \times 20 mL). The combined organic extracts were dried over Na₂SO₄ and evaporated. The residue was chromatographed on SiO₂ (CHCl₃/MeOH 30:1 \rightarrow 24:1) to give diacetate **14** (467 mg, 97%) as white crystals. Mp 104–107 °C (hexane-EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 5.22 (dd, J = 7.4, 0.8 Hz, 1H, 22-H), 5.05 (dd, J = 7.3, 4.9 Hz, 1H, 23-H), 4.03 (d, J = 2.2 Hz, 1H, 3H), 3.74 (ddd, J = 11.3, 4.3, 3.2 Hz, 1H, 2-H), 2.65 (dd, J = 12.5, 2.6 Hz, 1H, 5-H), 2.27 (dd, J = 13.1, 4.5 Hz, 1H), 2.04 (s, 3H, OAc), 2.02 (s, 3H, OAc), 0.98 (d, J = 6.7 Hz, 3H), 0.91 (d, J = 6.9 Hz, 3H), 0.84 (d, J = 6.8 Hz, 3H), 0.81 (d, J = 7.1 Hz, 3H), 0.73 (s, 3H, 19-H), 0.65 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 212.0, 170.6, 170.6, 74.6, 68.3, 68.2, 56.4, 53.6, 52.9, 50.7, 46.6, 42.8, 42.5, 40.1, 39.3, 38.7, 37.7, 37.6, 27.8, 26.8, 26.3, 23.8, 22.4, 21.1, 20.9, 20.9, 17.1, 13.5, 13.4, 11.7, 10.8. HRMS (ESI): calcd for C₃₂H₅₂NaO₇⁺ [M+Na]⁺ 571.3605, found 571.3608.

(22*R*,23*R*,24*R*)-2α,3α-*O*-Thionocarbonyl-22,23-diacetoxy-24-methyl-5α-cholestan-6-one (15)



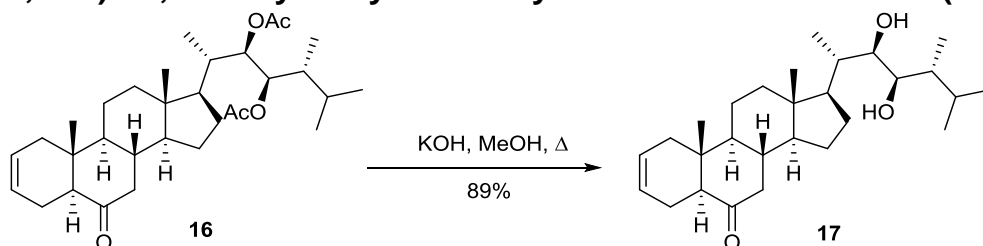
A mixture of diacetate **14** (959 mg, 1.75 mmol), 1,1'-thiocarbonyldiimidazole (934 mg, 5.24 mmol), DMAP (32 mg, 0.262 mmol), and dry THF (20 mL) was heated at 65 °C for 24 h. The solvent was evaporated and the residue was chromatographed on SiO₂ (PE/EtOAc 7:1→1:1) to give cyclic thionocarbonate **15** (783 mg, 76%) as white crystals. Mp 244–248°C (hexane-EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 5.21 (d, *J* = 7.1 Hz, 1H, 22-H), 5.03 (dd, *J* = 6.9, 5.2 Hz, 1H, 23-H), 4.97 – 4.93 (m, 1H, 2- or 3-H), 4.89 (dt, *J* = 10.0, 6.7 Hz, 1H, 3- or 2-H), 2.51 (dd, *J* = 12.7, 3.7 Hz, 1H, 5-H), 2.03 (s, 3H, OAc), 2.01 (s, 3H, OAc), 0.97 (d, *J* = 6.7 Hz, 3H), 0.90 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.7 Hz, 3H), 0.79 (d, *J* = 7.0 Hz, 3H), 0.68 (s, 3H, 19-H), 0.64 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 209.1, 191.6, 170.5, 79.8, 78.2, 74.4, 56.2, 52.8, 52.5, 50.5, 46.4, 42.6, 41.1, 38.9, 38.7, 38.5, 37.6, 37.3, 27.7, 26.8, 23.8, 22.4, 21.5, 21.0, 20.9, 20.8, 17.0, 13.3, 12.8, 11.5, 10.7. HRMS (ESI): calc for C₃₃H₅₀NaO₇S⁺ [M+Na]⁺ 613.3169, found 613.3171.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-24-methyl-5α-cholest-2-en-6-one (16)



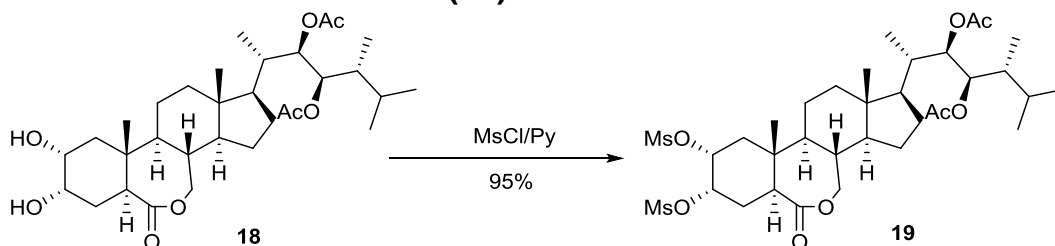
A mixture of thionocarbonate **15** (55 mg, 0.093 mmol) and (EtO)₃P (1 mL) was heated at 150 °C for 6 h. After cooling to 20 °C, the mixture was transferred to a column filled with SiO₂. The column was eluted with a gradient of PE/EtOAc 8:1→5:1 to give olefin **16** (45 mg, 94%) as white crystals. Mp 185–189°C (hexane). ¹H NMR (500 MHz, CDCl₃) δ 5.66 (dd, *J* = 10.0, 2.4 Hz, 1H, 2-H), 5.54 (dd, *J* = 9.5, 1.5 Hz, 1H, 3-H), 5.23 (d, *J* = 7.3 Hz, 1H, 22-H), 5.05 (dd, *J* = 7.2, 4.9 Hz, 1H, 23-H), 2.03 (s, 3H, OAc), 2.01 (s, 3H, OAc), 0.99 (d, *J* = 6.7 Hz, 3H), 0.91 (d, *J* = 6.9 Hz, 3H), 0.84 (d, *J* = 6.7 Hz, 3H), 0.81 (d, *J* = 7.0 Hz, 3H), 0.69 (s, 3H, 19-H), 0.66 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 211.8, 170.52, 170.51, 124.9, 124.4, 74.6, 56.5, 53.8, 53.3, 52.9, 46.8, 42.7, 39.9, 39.4, 39.3, 38.7, 37.7, 37.6, 27.8, 26.8, 23.8, 22.4, 21.7, 21.0, 20.9, 20.8, 17.1, 13.5, 13.4, 11.6, 10.8. HRMS (ESI): calc for C₃₂H₅₀NaO₅⁺ [M+Na]⁺ 537.3550, found 537.3551.

(22*R*,23*R*,24*R*)-22,23-Dihydroxy-24-methyl-5 α -cholest-2-en-6-one (17)



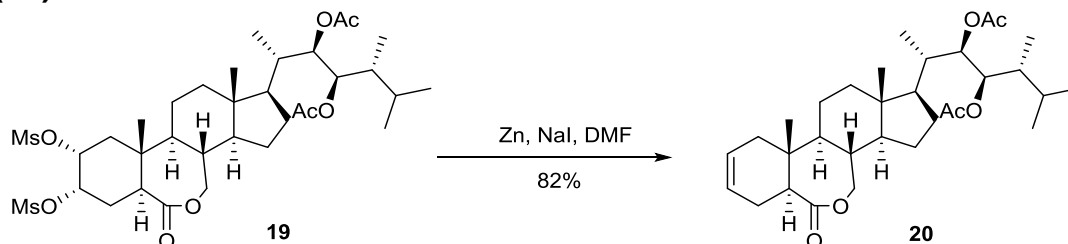
The diacetate **16** (44 mg, 0.085 mmol) was dissolved in a 5% KOH in MeOH (1 mL) and the solution was heated under heated at 65 °C for 1 h. The mixture was then cooled to 20 °C, acidified with 2 N HCl to pH 3 and extracted with CHCl₃ (3 × 5 mL). The combined organic extracts were dried over Na₂SO₄ and evaporated. The residue was chromatographed on SiO₂ (PE/EtOAc 5:1→1:1) to give diol **17** (33 mg, 89%) as white crystals. Mp 154–159°C (hexane-EtOAc). Lit. mp 166–168°C [1], 136–138°C [2]. ¹H NMR (500 MHz, CDCl₃) δ 5.67 (ddd, J = 9.7, 4.6, 1.9 Hz, 1H, 2-H), 5.56 (dd, J = 9.5, 1.9 Hz, 1H, 3-H), 3.69 (d, J = 3.5 Hz, 1H, 22-H), 3.40 (t, J = 5.2 Hz, 1H, 23-H), 0.97 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.9 Hz, 3H), 0.86 (d, J = 6.8 Hz, 3H), 0.84 (d, J = 7.0 Hz, 3H), 0.70 (s, 3H, 19-H), 0.68 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 212.1, 124.9, 124.5, 76.3, 72.6, 56.6, 53.8, 53.3, 52.6, 46.9, 42.7, 41.4, 40.2, 40.0, 39.4, 39.3, 37.7, 27.7, 27.0, 23.9, 22.1, 21.7, 21.1, 17.3, 13.5, 12.4, 11.7, 10.8. HRMS (APCI): calc for C₂₈H₄₃O⁺ [M-2H₂O+H]⁺ 395.3308, found 395.3314.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-2 α ,3 α -di(methylsulfonyloxy)-24-methyl-7 α -homo-7-oxa-5 α -cholestan-6-one (19)



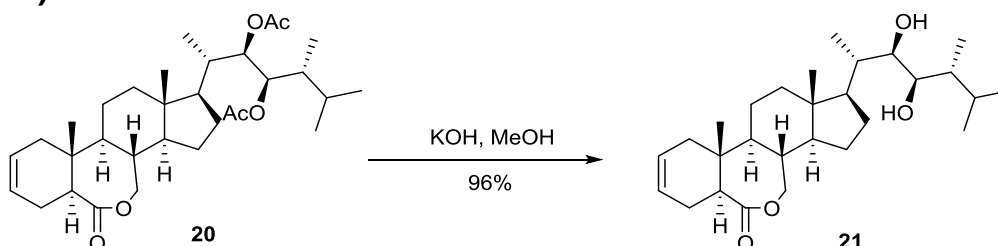
A mixture of diacetate **18** (302 mg, 0.535 mmol, prepared according to [3]), MsCl (0.206 mL, 2.68 mmol) and pyridine (1.5 mL) was kept at 20 °C overnight. The reaction mixture was diluted with water (3 mL) and extracted with CHCl₃ (3 × 3 mL). The combined extracts were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified on SiO₂ (CHCl₃/MeOH 40:1→36:1) to give dimesylate **19** (365 mg, 95%) as a white foam. ¹H NMR (500 MHz, CDCl₃) δ 5.20 (d, J = 6.9 Hz, 1H, 22-H), 5.10 (br.s, 1H, 3-H), 5.04 (dd, J = 7.1, 5.0 Hz, 1H, 23-H), 4.73 (ddd, J = 12.7, 4.3, 2.4 Hz, 1H, 2-H), 4.10 (d, J = 12.0 Hz, 1H, 7-H), 4.04 (dd, J = 12.4, 9.3 Hz, 1H, 7-H), 3.09 (s, 3H, OMs), 3.08 (s, 3H, OMs), 2.03 (s, 3H, OAc), 2.01 (s, 3H, OAc), 0.96 (s, 3H, 19-H), 0.96 (d, J = 6.3 Hz, 3H), 0.91 (d, J = 6.9 Hz, 3H), 0.83 (d, J = 6.7 Hz, 3H), 0.80 (d, J = 7.0 Hz, 3H), 0.68 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 174.2, 170.5, 74.9, 74.4, 70.4, 57.6, 52.8, 51.0, 42.4, 41.2, 39.7, 39.2, 39.0, 38.8, 38.7, 38.7, 38.5, 37.6, 30.4, 29.6, 27.7, 26.8, 24.6, 22.4, 22.2, 20.9, 20.8, 17.0, 15.5, 13.3, 11.4, 10.7. HRMS (ESI): calc for C₃₄H₅₆NaO₁₂S₂⁺ [M+Na]⁺ 743.3105, found 743.3124.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-24-methyl-7*a*-homo-7-oxa-5*α*-cholest-2-en-6-one (20)



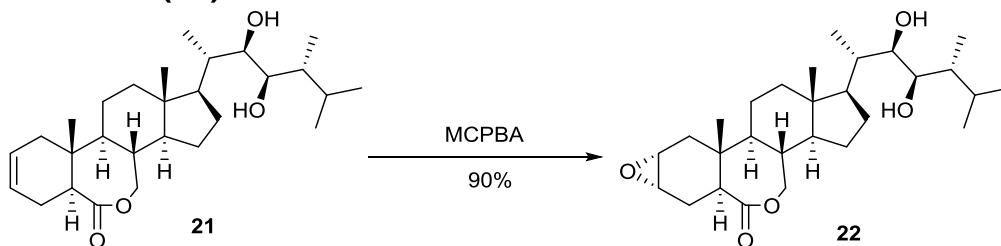
A mixture of dimesylate **19** (390 mg, 0.541 mmol), NaI (2.027 g, 13.5 mmol), Zn dust (884 mg, 13.5 mmol) and DMF (5.4 mL) was stirred at 150 °C for 2 h. After cooling to 20 °C, the precipitate was separated by filtration. The filtrate was diluted with water (50 mL) and extracted with EtOAc (2 × 40 mL). The combined organic layers were washed with saturated Na₂S₂O₃ (2 × 40 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was chromatographed on SiO₂ (PE/EtOAc 4:1→1:1) to give olefin **20** (235 mg, 82%) as white crystals. Mp 184–186 °C (PE-EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 5.67 (m, 1H, 3-H), 5.56 (m, 1H, 2-H), 5.22 (m, *J* = 7.2 Hz, 1H, 22-H), 5.06 (dd, *J* = 7.0, 5.0 Hz, 1H, 23-H), 4.10 (d, *J* = 12.4 Hz, 1H, 7-H), 4.02 (dd, *J* = 12.0, 9.1 Hz, 1H, 7-H), 2.91 (dd, *J* = 10.2, 5.5 Hz, 1H, 5-H), 2.85 (m, 1H), 2.03 (s, 3H, OAc), 2.03 (s, 3H, OAc), 0.99 (d, *J* = 6.7 Hz, 3H), 0.93 (d, *J* = 6.8 Hz, 3H), 0.89 (s, 3H, 19-H), 0.86 (d, *J* = 6.7 Hz, 3H), 0.82 (d, *J* = 7.0 Hz, 3H), 0.70 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 176.4, 170.6 (x2), 123.6, 123.3, 74.6, 70.3, 59.1, 52.9, 51.4, 44.2, 42.5, 41.3, 39.8, 39.6, 38.7, 37.7, 35.7, 29.7, 27.8, 26.9, 25.9, 24.7, 22.5, 20.9, 20.9, 17.2, 16.4, 13.3, 11.4, 10.8. HRMS (ESI): calc for C₃₂H₅₀NaO₆⁺ [M+Na]⁺ 553.3500, found 553.3506.

(22*R*,23*R*,24*R*)-22,23-Dihydroxy-24-methyl-7*a*-homo-7-oxa-5*α*-cholest-2-en-6-one (21)



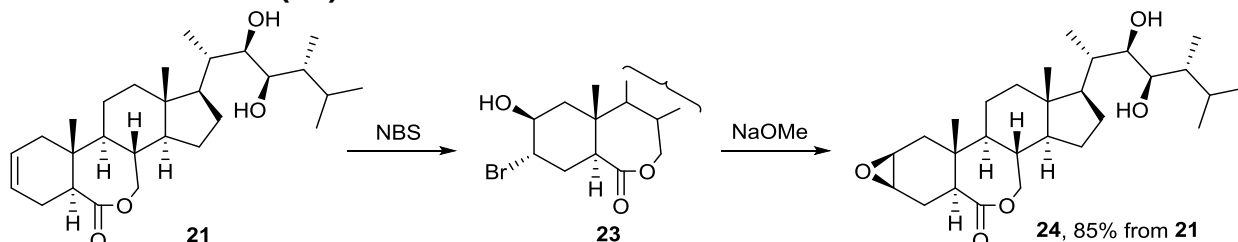
The title compound **21** (177 mg) was prepared as white crystals in 96% yield starting from diacetate **20** as described above for the preparation of **17**. Mp 177–179 °C (hexane-EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 5.71 – 5.64 (m, 1H, 3-H), 5.59 – 5.52 (m, 1H, 2-H), 4.11 (d, *J* = 12.2 Hz, 1H, 7-H), 4.03 (dd, *J* = 12.3, 8.7 Hz, 1H, 7-H), 3.67 (d, *J* = 3.6 Hz, 1H, 22-H), 3.39 (t, *J* = 5.2 Hz, 1H, 23-H), 2.92 (dd, *J* = 10.3, 5.7 Hz, 1H, 5-H), 2.88 – 2.77 (m, 1H), 0.95 (d, *J* = 6.7 Hz, 3H), 0.91 (d, *J* = 6.9 Hz, 3H), 0.88 (s, 3H, 19-H), 0.86 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 7.0 Hz, 3H), 0.70 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 176.5, 123.5, 123.4, 76.2, 72.5, 70.4, 59.0, 52.6, 51.3, 44.2, 42.4, 41.4, 41.2, 40.2, 39.8, 39.6, 35.7, 27.6, 27.0, 25.9, 24.7, 22.5, 22.1, 17.3, 16.4, 12.3, 11.6, 10.8. HRMS (ESI): calc for C₂₈H₄₆NaO₄⁺ [M+Na]⁺ 469.3288, found 469.3291.

(22*R*,23*R*,24*R*)-22,23-Dihydroxy-2 α ,3 α -epoxy-24-methyl-7 α -homo-7-oxa-5 α -cholestan-6-one (22**)**



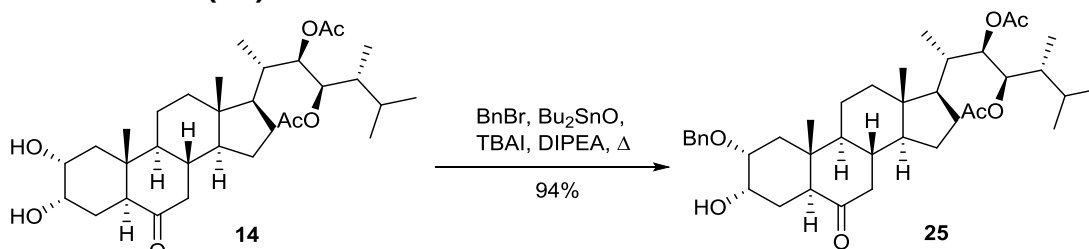
A solution of olefin **21** (56 mg, 0.125 mmol) and MCPBA (65 mg, 0.375 mmol) in CH_2Cl_2 (2 mL) was stirred at 20 °C for 1 h. The reaction mixture was washed with saturated NaHCO_3 (2 \times 2 mL), brine (2 \times 2 mL), dried over Na_2SO_4 and the solvent was evaporated under reduced pressure. The residue was purified on SiO_2 ($\text{CHCl}_3/\text{MeOH}$ 100:1) to give epoxide **22** (52 mg, 90%) as white crystals. Mp 198–200 °C (hexane-EtOAc). ^1H and ^{13}C NMR data are given in Table 1. HRMS (ESI): calc for $\text{C}_{28}\text{H}_{47}\text{O}_5^+$ $[\text{M}+\text{H}]^+$ 463.3418, found 469.3412.

(22*R*,23*R*,24*R*)-22,23-Dihydroxy-2 β ,3 β -epoxy-24-methyl-7 α -homo-7-oxa-5 α -cholestan-6-one (24**)**



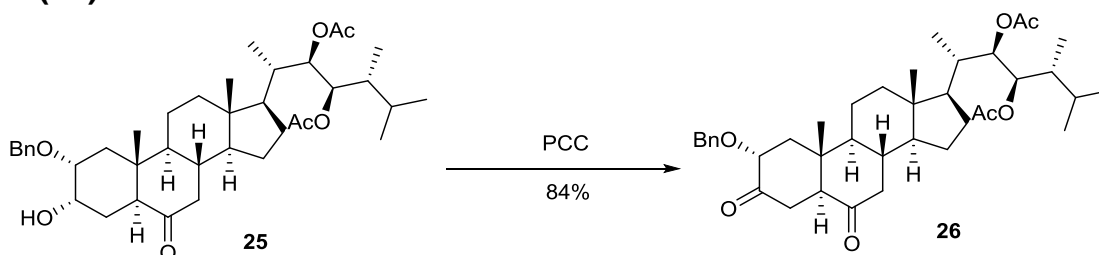
NBS (45 mg, 0.255 mmol) and water (1 mL) were added to a stirred solution of olefin **21** (57 mg, 0.128 mmol) in dimethoxyethane (6 mL). The reaction mixture was stirred at 20 °C for 1.5 h, then treated with saturated $\text{Na}_2\text{S}_2\text{O}_3$ (1 mL), diluted with water (3 mL) and extracted with EtOAc (3 \times 5 mL). The combined organic extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The crude bromohydrine **23** (75 mg) was dissolved in MeOH (12 mL) and treated with a 0.12 M MeONa in MeOH (2.4 mL, 0.281 mmol). The mixture was stirred at 20 °C for 15 min, then treated with saturated NH_4Cl (2 mL), diluted with water (2 mL) and extracted with EtOAc (2 \times 10 mL). The combined organic layers were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was chromatographed on SiO_2 ($\text{CHCl}_3/\text{MeOH}$ 69:1 \rightarrow 52:1) to give epoxide **24** (50 mg, 85%) as white crystals. Mp 200–203 °C (hexane-EtOAc). ^1H and ^{13}C NMR data are given in Table 1. HRMS (ESI): calc for $\text{C}_{28}\text{H}_{46}\text{NaO}_5^+$ $[\text{M}+\text{Na}]^+$ 485.3237, found 485.3234.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-3 α -hydroxy-2 α -benzyloxy-24-methyl-5 α -cholestan-6-one (25**)**



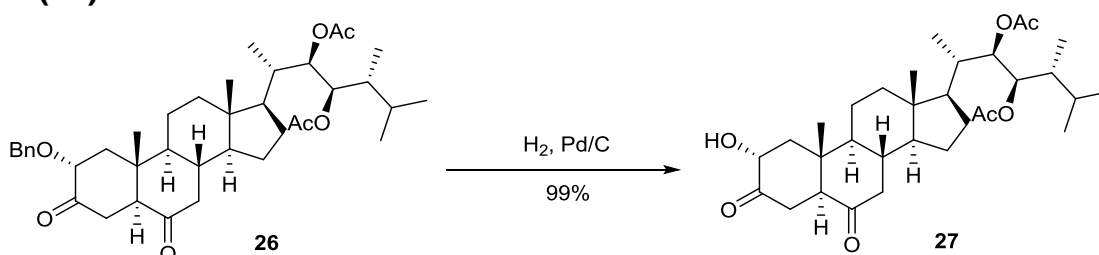
A mixture of diacetate **14** (148 mg, 0.270 mmol), Bu_2SnO (76.4 mg, 0.307 mmol), TBAI (38.5 mg, 0.104 mmol), DIPEA (118 μL , 0.675 mmol), BnBr (80.3 μL , 0.675 mmol) and dry toluene (3 mL) was stirred at 110 $^\circ\text{C}$ for 7 h. The reaction mixture was concentrated in vacuo and the residue was chromatographed on SiO_2 (PE/EtOAc 5:1 \rightarrow 1:3) to give benzyl ether **25** (162 mg, 94%) as a yellow oil. Its ^1H and ^{13}C NMR spectra were identical to those described by us previously [3].

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-2 α -benzyloxy-24-methyl-5 α -cholestan-3,6-dione (26**)**



PCC (432 mg, 2.00 mmol) was added to a stirred solution of **25** (160 mg, 0.250 mmol) in CH_2Cl_2 (2 mL). The mixture was stirred at 20 $^\circ\text{C}$ overnight and the reaction mixture was transferred to a column with SiO_2 . The column was eluted with PE/EtOAc 5:1 \rightarrow 1:1 to give diketone **26** (134 mg, 84%) as white crystals. Mp 180–184 $^\circ\text{C}$ (hexane-EtOAc). ^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.27 (m, 5H, - OCH_2Ph), 5.22 (d, J = 7.2 Hz, 1H, 22-H), 5.05 (dd, J = 7.2, 5.0 Hz, 1H, 23-H), 4.88 (d, J = 11.6 Hz, 1H, - OCH_2Ph), 4.46 (d, J = 11.6 Hz, 1H, - OCH_2Ph), 4.04 (dd, J = 12.2, 6.5 Hz, 1H, 2-H), 2.03 (s, 3H, OAc), 2.02 (s, 3H, OAc), 0.99 (d, J = 6.7 Hz, 3H), 0.96 (s, 3H, 19-H), 0.92 (d, J = 6.9 Hz, 3H), 0.85 (d, J = 6.8 Hz, 3H), 0.81 (d, J = 7.1 Hz, 3H), 0.67 (s, 3H, 18-H). ^{13}C NMR (125 MHz, CDCl_3) δ 208.9, 208.2, 170.5 (x2), 137.7, 128.4 (x2), 127.8 (x3), 78.4, 77.4, 74.5, 72.4, 58.4, 56.2, 53.4, 52.9, 46.2 (x2), 42.8, 42.6, 39.1, 38.7, 37.7, 37.6, 36.6, 27.8, 26.8, 23.8, 22.4, 21.7, 20.9, 20.8, 17.1, 14.0, 13.4, 11.7, 10.7. HRMS (ESI): calc for $\text{C}_{39}\text{H}_{56}\text{NaO}_7^+$ [M+Na] $^+$ 659.3918, found 659.3926.

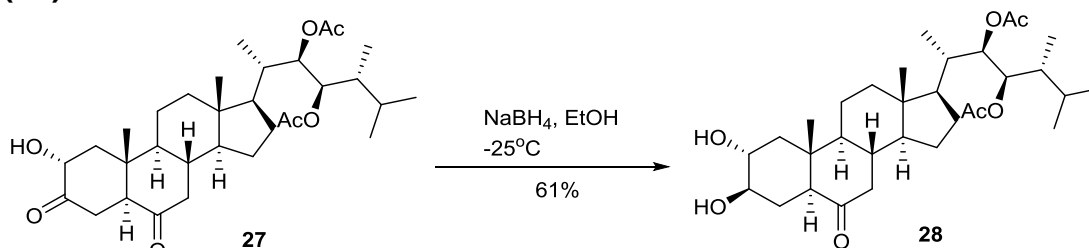
(22*R*,23*R*,24*R*)-22,23-Diacetoxy-2 α -hydroxy-24-methyl-5 α -cholestan-3,6-dione (27**)**



A solution of **26** (105 mg, 0.165 mmol) in THF (3.3 mL) was hydrogenated over Pd/C (10%, 17.5 mg) at 20 $^\circ\text{C}$ and atmospheric pressure. After 12 h, the mixture

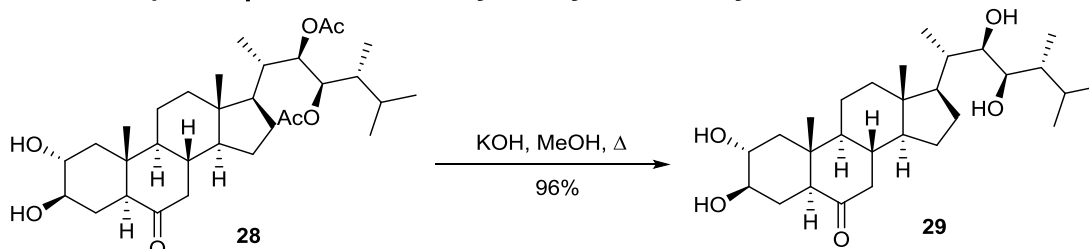
was filtered through Celite, the solvent removed and the crude material was purified by chromatography on SiO₂ (PE/EtOAc 4:1→1:4) to give compound **27** (89 mg, 99%) as a colorless oil. ¹H NMR (500 MHz, CDCl₃) δ 5.21 (d, *J* = 7.1 Hz, 1H, 22-H), 5.04 (dd, *J* = 7.1, 5.1 Hz, 1H, 23-H), 4.23 (dd, *J* = 11.8, 7.1 Hz, 1H, 2-H), 2.02 (s, 3H, OAc), 2.01 (s, 3H, OAc), 1.02 (s, 3H, 19-H), 0.98 (d, *J* = 6.7 Hz, 3H), 0.91 (d, *J* = 6.9 Hz, 3H), 0.84 (d, *J* = 6.8 Hz, 3H), 0.80 (d, *J* = 7.0 Hz, 3H), 0.67 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 210.9, 207.9, 170.6 (x2), 74.5, 71.9, 58.5, 56.2, 53.3, 52.9, 47.7, 46.2, 42.8, 42.6, 39.1, 38.7, 37.7, 37.4, 35.0, 27.8, 26.8, 23.8, 22.4, 21.7, 20.9, 20.8, 17.1, 13.8, 13.4, 11.6, 10.7.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-2α,3β-dihydroxy-24-methyl-5α-cholestan-6-one (28)



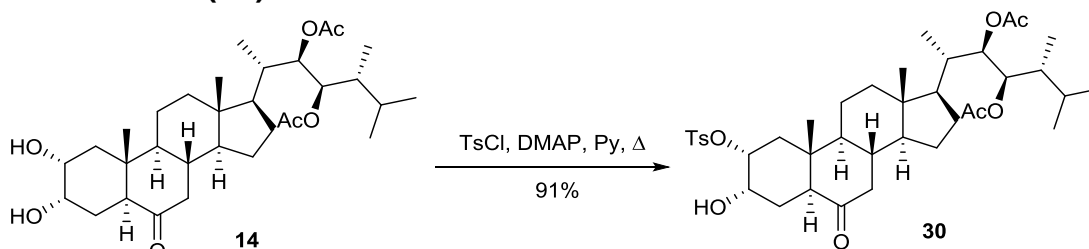
To a cooled (−25 °C) and stirred solution of diketone **27** (32 mg, 0.059 mmol) in absolute EtOH (19.5 mL), NaBH₄ (2.4 mg, 0.064 mmol) was added. Stirring was continued for 30 min at the same temperature, then water (10 mL) was added and the mixture was extracted with CHCl₃ (3 × 20 mL). The combined extracts were dried over Na₂SO₄ and concentrated in vacuo. The residue was chromatographed on SiO₂ (CHCl₃/MeOH 70:1→10:1) to give compound **28** (19.5 mg, 61%) as a white solid. ¹H NMR (500 MHz, CDCl₃) δ 5.23 (d, *J* = 7.1 Hz, 1H, 22-H), 5.06 (dd, *J* = 7.2, 5.0 Hz, 1H, 23-H), 3.66-3.54 (m, 1H, 2-H), 3.43-3.34 (m, 1H, 3-H), 2.04 (s, 3H, OAc), 2.02 (s, 3H, OAc), 0.99 (d, *J* = 6.7 Hz, 3H), 0.92 (d, *J* = 6.9 Hz, 4H), 0.85 (d, *J* = 6.8 Hz, 4H), 0.81 (d, *J* = 7.1 Hz, 4H), 0.79 (s, 3H, 19-H), 0.66 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 210.0, 170.6 (x2), 77.5, 75.7, 74.6, 72.0, 56.6, 56.4, 53.8, 52.9, 46.4, 44.2, 42.8, 39.3, 38.7, 37.7, 37.5, 29.7, 27.9, 27.8, 26.8, 23.8, 22.4, 21.6, 20.9, 20.9, 17.1, 14.3, 13.4, 11.7, 10.8.

(22*R*,23*R*,24*R*)-2α,3β,22,23-Tetrahydroxy-24-methyl-5α-cholestan-6-one (29)



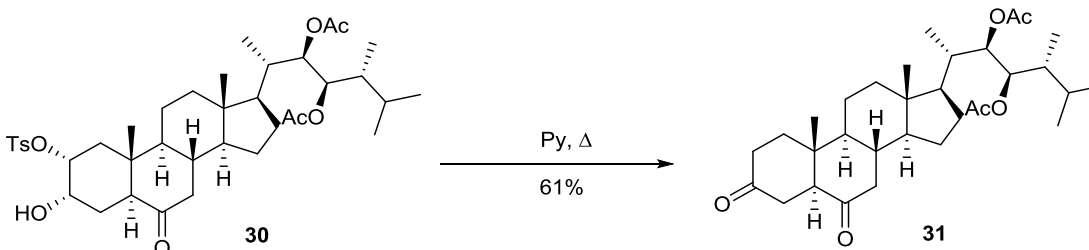
The title compound **29** (13 mg) was prepared as white crystals in 96% yield starting from diacetate **28** as described above for the preparation of **17**. Mp 210–213 °C (EtOAc). Lit. mp 209–212 °C [4], 213–215 °C [5]. ¹H NMR (500 MHz, C₅D₅N) δ 4.06 (dd, *J* = 14.2, 7.0 Hz, 2H, 3- and 22-H), 3.86 (ddd, *J* = 11.8, 8.9, 4.9 Hz, 1H, 2-H), 3.71 (dd, *J* = 5.9, 4.8 Hz, 1H, 23-H), 1.31 (d, *J* = 6.7 Hz, 3H), 1.07 (d, *J* = 6.8 Hz, 3H), 1.02 (d, *J* = 6.9 Hz, 3H), 0.98 (d, *J* = 6.9 Hz, 3H), 0.85 (s, 3H, 19-H), 0.70 (s, 3H, 18-H). ¹³C NMR (125 MHz, C₅D₅N) δ 210.5, 76.7, 76.4, 72.8, 72.7, 57.3, 57.1, 54.3, 53.7, 47.1, 46.3, 43.4, 43.2, 42.6, 41.7, 40.3, 38.1, 29.8, 28.7, 27.6, 24.5, 22.9, 22.3, 17.9, 14.6, 13.7, 12.4, 11.8. HRMS (ESI): calc for C₂₈H₄₈NaO₅⁺ [*M*+*H*]⁺ 487.3394, found 487.3392.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-3 α -hydroxy-2 α -tosyloxy-24-methyl-5 α -cholestan-6-one (30**)**



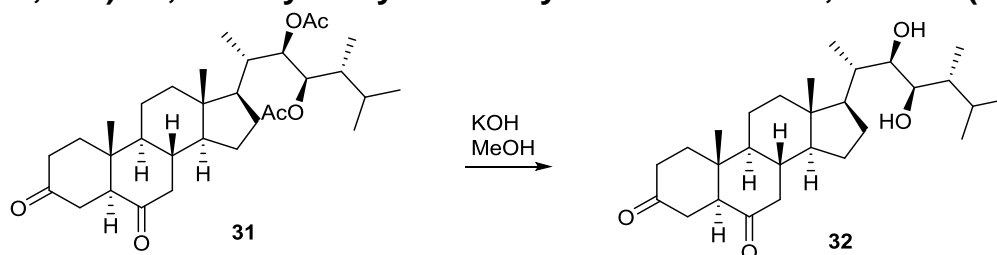
A mixture of diacetate **14** (206 mg, 0.375 mmol), pyridine (2 mL), TsCl (214 mg, 1.126 mmol) and DMAP (9 mg, 0.075 mmol) was kept overnight at 30 °C. Then water (10 mL) was added and the mixture was extracted with EtOAc (3 × 10 mL). The combined organic extracts were washed with water (2 × 10 mL), dried over Na₂SO₄ and evaporated. The residue was chromatographed on SiO₂ (PE/EtOAc 3:1→1:3) to give tosylate **30** (240 mg, 91%) as a colorless oil. ¹H NMR (500 MHz, CDCl₃) δ 7.78 (d, *J* = 8.2 Hz, 2H, OTs), 7.35 (d, *J* = 8.2 Hz, 2H, OTs), 5.22 (d, *J* = 7.2 Hz, 1H, 22-H), 5.05 (dd, *J* = 7.2, 5.0 Hz, 1H, 23-H), 4.54 (d, *J* = 11.7 Hz, 1H, 2-H), 4.07 (d, *J* = 2.2 Hz, 1H, 3-H), 2.65 (dd, *J* = 12.5, 2.8 Hz, 1H, 5-H), 2.45 (s, 3H, OTs), 2.25 (dd, *J* = 13.2, 4.5 Hz, 1H), 2.03 (s, 3H, OAc), 2.02 (s, 3H, OAc), 0.97 (d, *J* = 6.7 Hz, 3H), 0.91 (d, *J* = 6.9 Hz, 3H), 0.84 (d, *J* = 6.8 Hz, 3H), 0.80 (d, *J* = 7.0 Hz, 3H), 0.68 (s, 3H, 19-H), 0.63 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 210.8, 170.5, 145.1, 133.9, 129.9, 127.6, 80.7, 74.5, 66.7, 56.3, 53.4, 52.9, 50.1, 46.4, 42.7, 42.7, 39.2, 38.8, 37.7, 37.5, 37.3, 27.8, 26.8, 26.1, 23.8, 22.4, 21.6, 21.1, 20.9, 20.8, 17.1, 13.4, 13.3, 11.6, 10.8. HRMS (ESI): calc for C₃₉H₅₈NaO₉S⁺ [*M*+Na]⁺ 725.3694, found 725.3694.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-24-methyl-5 α -cholestan-3,6-dione (31**)**



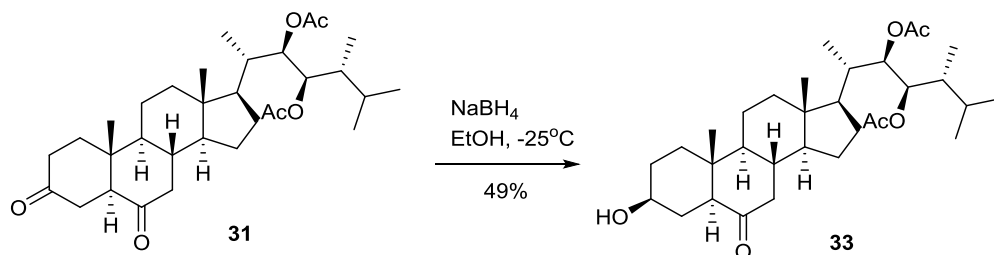
A solution of tosylate **30** (240 mg, 0.342 mmol) in pyridine (3 mL) was heated at 115 °C for 6 h. The solvent was removed in vacuo and the residue was purified by column chromatography on SiO₂ (PE/EtOAc 7:1→1:1) to give less polar diketone **31** (110 mg, 61%) as a colorless oil and starting tosylate **30** (17 mg, 7%). ¹H NMR (500 MHz, CDCl₃) δ 5.24 (dd, *J* = 7.3, 0.9 Hz, 1H, 22-H), 5.06 (dd, *J* = 7.3, 5.0 Hz, 1H, 23-H), 2.04 (s, 3H, OAc), 2.03 (s, 3H, OAc), 1.00 (d, *J* = 6.8 Hz, 3H), 0.95 (s, 3H, 19-H), 0.93 (d, *J* = 6.9 Hz, 3H), 0.85 (d, *J* = 6.8 Hz, 3H), 0.82 (d, *J* = 7.1 Hz, 3H), 0.69 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 211.1, 208.8, 170.5, 74.6, 57.5, 56.4, 53.4, 53.0, 46.5, 42.9, 41.2, 39.3, 38.8, 38.1, 38.0, 37.7, 37.3, 36.9, 27.9, 26.9, 23.9, 22.4, 21.6, 20.9, 20.8, 17.1, 13.4, 12.6, 11.7, 10.8. HRMS (ESI): calc for C₃₂H₅₀NaO₆⁺ [*M*+Na]⁺ 553.3500, found 553.3485.

(22*R*,23*R*,24*R*)-22,23-Dihydroxy-24-methyl-5 α -cholestan-3,6-dione (32**)**



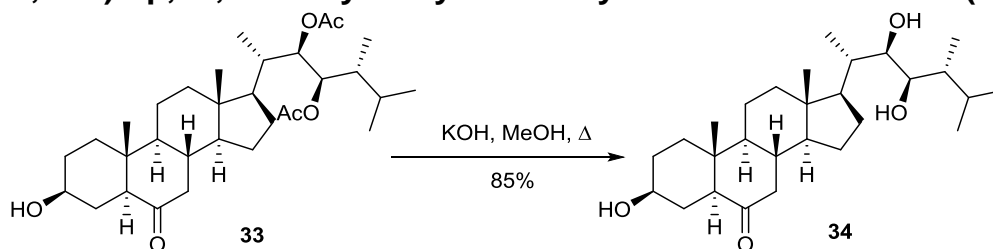
The diacetate **31** (32 mg 0.0603 mmol) was dissolved in a 5% KOH in MeOH (1 mL) and the solution was kept under argon at 20 °C for 5 h. The mixture was then acidified with 2 N HCl to pH 3 and extracted with CHCl₃ (3 × 3 mL). The combined organic extracts were dried over Na₂SO₄ and evaporated. The residue was chromatographed on SiO₂ (PE/EtOAc 3:1) to give diol **32** (14 mg, 52%) as an amorphous white powder. ¹H NMR (500 MHz, CDCl₃) δ 3.70 (dd, J = 4.4, 1.3 Hz, 1H, 22-H), 3.43 – 3.39 (m, 1H, 23-H), 0.98 (d, J = 6.7 Hz, 3H), 0.96 (s, 3H, 19-H), 0.92 (d, J = 6.9 Hz, 3H), 0.87 (d, J = 6.8 Hz, 3H), 0.85 (d, J = 7.0 Hz, 3H), 0.70 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 211.2, 209.0, 76.4, 72.6, 57.5, 56.4, 53.4, 52.6, 46.5, 42.9, 41.4, 41.2, 40.2, 39.3, 38.1, 37.3, 37.0, 27.7, 27.0, 23.9, 22.1, 21.7, 17.3, 12.6, 12.4, 11.9, 10.8. HRMS (ESI): calc for C₂₈H₄₆NaO₄⁺ [M+Na]⁺ 469.3288, found 469.3289.

(22*R*,23*R*,24*R*)-22,23-Diacetoxy-3 β -hydroxy-24-methyl-5 α -cholestan-6-one (33**)**



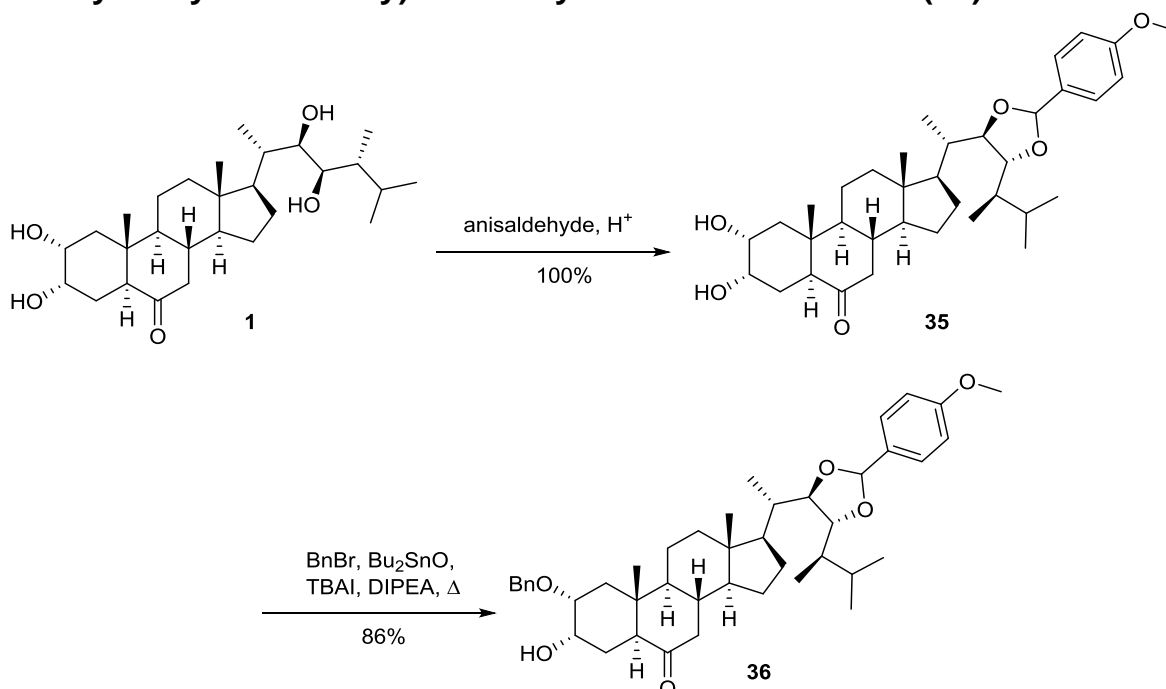
NaBH₄ (1 mg, 0.0456 mmol) was added to a cooled (–25 °C) solution of diketone **31** (22 mg, 0.0415 mmol) in absolute EtOH (13.5 mL) and the mixture was stirred at the same temperature for 30 min. Water (5 mL) was added and the mixture was extracted with CHCl₃ (3 × 10 mL). The combined extracts were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was chromatographed on SiO₂ (CHCl₃/MeOH 100:1→94:1) to give alcohol **33** (9 mg, 49%) as white crystals. Mp 221 – 224 °C (CHCl₃-MeOH). ¹H NMR (500 MHz, CDCl₃) δ 5.24 (dd, J = 7.4, 1.1 Hz, 1H, 22-H), 5.06 (dd, J = 7.3, 4.9 Hz, 1H, 23-H), 3.61 – 3.53 (m, 1H, 3-H), 2.04 (s, 3H, OAc), 2.03 (s, 3H, OAc), 0.99 (d, J = 6.8 Hz, 3H), 0.92 (d, J = 6.9 Hz, 3H), 0.85 (d, J = 6.8 Hz, 3H), 0.82 (d, J = 7.1 Hz, 3H), 0.75 (s, 3H, 19-H), 0.67 (s, 3H, 18-H). ¹³C NMR (125 MHz, CDCl₃) δ 210.6, 2x170.6, 77.5, 74.6, 70.6, 56.7, 56.6, 53.8, 53.0, 46.6, 42.9, 40.9, 39.4, 38.8, 37.9, 37.7, 36.7, 30.7, 30.0, 27.9, 26.9, 23.9, 22.5, 21.5, 20.9, 20.9, 17.2, 13.4, 13.1, 11.7, 10.8. HRMS (ESI): calc for C₃₂H₅₂NaO₆⁺ [M+Na]⁺ 555.3656. Найдено 555.3636.

(22*R*,23*R*,24*R*)-3 β ,22,23-Trihydroxy-24-methyl-5 α -cholestan-6-one (34)



The title compound **34** (10 mg) was prepared as white crystals in 85% yield starting from diacetate **33** as described above for the preparation of **17**. Mp 233–236 °C (CHCl₃:MeOH). Lit. mp 233–234 °C [6]. ¹H NMR (500 MHz, C₅D₅N) δ 4.06 (br.s, 1H, 22-H), 3.85 (m, 1H, 3-H), 3.72 (br.s, 1H, 23-H), 1.33 (d, J = 5.9 Hz, 5H), 1.07 (d, J = 6.2 Hz, 3H), 1.03 (d, J = 6.4 Hz, 4H), 0.98 (d, J = 6.7 Hz, 3H), 0.79 (s, 3H, 19-H), 0.73 (s, 3H, 18-H). ¹³C NMR (125 MHz, Pyr) δ 210.7, 76.7, 72.8, 70.4, 57.3, 57.2, 54.2, 53.8, 47.2, 43.4, 42.6, 41.7, 41.3, 40.4, 38.4, 37.4, 32.2, 31.6, 28.7, 27.7, 24.6, 22.9, 22.2, 17.9, 13.7, 13.6, 12.4, 11.8. HRMS (APCI): calc for C₂₈H₄₅O₂⁺ [M-2H₂O+H]⁺ 413.3413, found 413.3417.

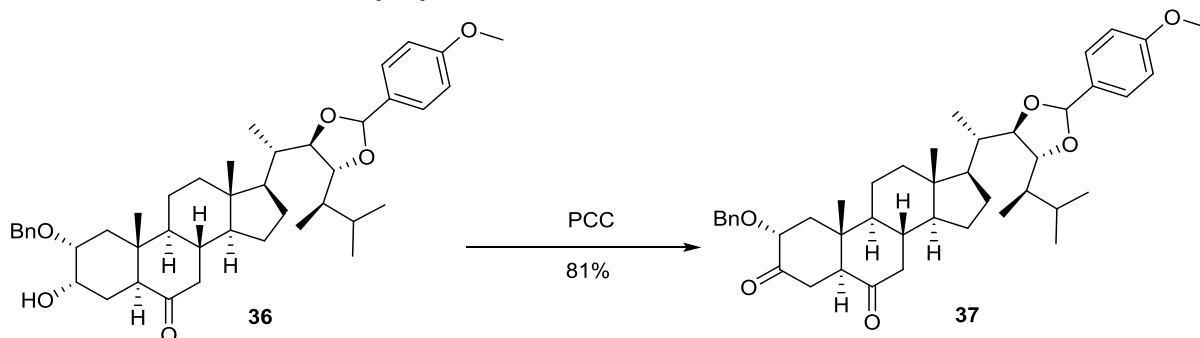
(22*R*,23*R*,24*R*)-2 α -Benzyloxy-3 α -hydroxy-22,23-(*p*-methoxybenzylidenedioxy)-24-methyl-5 α -cholestan-6-one (36)



TMSCl (28 μ L, 0.221 mmol) was added to MeOH (17 mL) and the solution was stirred at 20 °C for 10 min. Then, epicastasterone **1** (205 mg, 0.441 mmol) and anisaldehyde (161 μ L, 1.323 mmol) were added and the mixture was stirred for further 40 min. The mixture was then treated with saturated NaHCO₃ (10 mL) and extracted with CHCl₃ (3 \times 20 mL). The combined extracts were dried over Na₂SO₄ and concentrated in vacuo to give 257 mg of crude **35**. Its benzylation was performed according to the procedure described above for the preparation of **25**. The benzyl ether **36** (256 mg, 86%) was isolated as a yellow oil. ¹H NMR (500 MHz, CDCl₃) δ 7.44 (d, J = 8.6 Hz, 2H, arom.), 7.41 – 7.27 (m, 5H, arom.), 6.89 (dd, J = 8.4, 6.8 Hz, 2H, arom.), 4.59 (d, J = 11.7 Hz, 1H, -OCH₂Ph), 4.54 (d, J = 11.7 Hz, 1H, -OCH₂Ph), 4.18 (br.s, 1H, 22-H), 4.01 (m, 1H, 23-H), 3.799 and 3.796 (s, 3H, OMe), 3.70 (dd, J = 10.2, 5.3 Hz, 1H, 2-H), 3.53 – 3.46 (m, 1H, 3-H), 2.72 (d, J = 12.4 Hz, 1H), 1.17 – 0.62 (m, 18H). ¹³C NMR (125 MHz, CDCl₃)

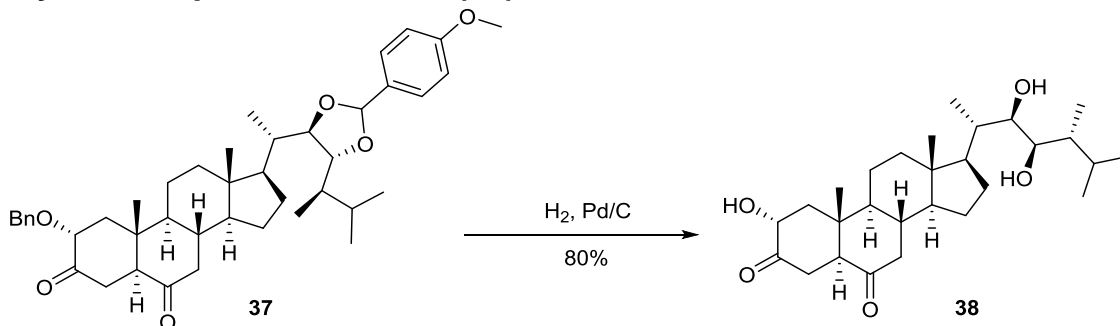
δ 212.1, 160.5, 138.0, 129.6, 128.5, 128.0, 127.8, 127.5, 113.7, 113.6, 104.1, 101.9, 83.4, 82.6, 80.9, 80.8, 75.6, 70.4, 65.4, 56.3, 55.3, 53.6, 53.1, 50.8, 46.6, 42.8, 42.4, 41.6, 40.0, 39.2, 39.0, 37.7, 37.4, 27.8, 27.7, 27.4, 25.4, 23.8, 21.2, 21.0, 20.9, 16.2, 15.7, 13.6, 13.2, 12.9, 11.7, 10.1, 8.8. HRMS (ESI): calc for $C_{43}H_{61}O_6^+$ $[M+H]^+$ 673.4463, found 673.4459.

(22*R*,23*R*,24*R*)-2 α -Benzyloxy-22,23-(*p*-methoxybenzylidenedioxy)-24-methyl-5 α -cholestan-3,6-dione (37**)**



A mixture of **36** (256 mg, 0.381 mmol), PCC (657 mg, 3.05 mmol) and CH_2Cl_2 (3 mL) was stirred at 20 °C overnight. The reaction mixture was filtered through a plug of SiO_2 (EtOAc) and concentrated in vacuo. The residue was purified by column chromatography on SiO_2 (PE/EtOAc 7:1→1:1) to give ketone **37** (208 mg, 81%) as white crystals. Mp 184–188 °C (hexane-EtOAc). 1H NMR (500 MHz, $CDCl_3$) δ 7.47 – 7.28 (m, 7H, arom.), 6.92 – 6.86 (m, 2H, arom.), 5.85 and 5.69 (s, 1H, $-CH-C_6H_4OMe$), 4.89 (d, $J = 11.6$ Hz, 1H, $-CH_2Ph$), 4.47 (d, $J = 11.6$ Hz, 1H, $-CH_2Ph$), 4.10 – 3.97 (m, 2H, 22- and 23-H), 3.796 and 3.792 (s, 3H, OMe), 3.70 (dd, $J = 10.3, 5.3$ Hz, 1H, 2-H), 1.12 (dd, $J = 12.6, 6.5$ Hz, 3H), 0.97 (s, 3H, 19-H), 0.95 – 0.71 (m, 9H), 0.70 (s, 3H, 18-H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 208.8, 208.3, 160.5, 160.2, 137.7, 131.1, 129.6, 128.5, 128.4 (x2), 128.0, 127.8 (x4), 113.70, 113.65, 104.2, 101.9, 83.3, 82.6, 80.9, 80.8, 78.4, 77.3, 77.0, 76.7, 72.3, 58.3, 56.1, 55.3, 53.3, 53.1, 46.3, 46.2, 42.9, 42.6, 41.6, 40.0, 39.0, 38.9, 37.7, 36.6, 27.8, 27.6, 27.4, 23.9, 21.8, 21.0, 20.9, 16.2, 15.7, 13.9, 13.2, 12.9, 11.7, 10.1, 8.9. HRMS (ESI): calc for $C_{43}H_{58}NaO_6^+$ $[M+Na]^+$ 693.4126, found 693.4132.

(22*R*,23*R*,24*R*)-2 α ,22,23-Trihydroxy-24-methyl-5 α -cholestan-3,6-dione / 3-dehydro-24-epicastasterone/ (38**)**



The title compound **38** (115 mg) was prepared as white crystals in 80% yield starting from **37** as described above for the preparation of **27**. Mp 190–193 °C (EtOAc). 1H NMR (500 MHz, $CDCl_3$) δ 4.24 (dd, $J = 11.9, 7.0$ Hz, 1H, 2-H), 3.67 (br.s, 1H, 22-H), 3.39 (br.s, 1H, 23-H), 1.03 (s, 3H, 19-H), 0.96 (d, $J = 6.3$ Hz, 3H), 0.90 (d, $J = 6.8$ Hz, 3H), 0.85 (d, $J = 6.7$ Hz, 3H), 0.83 (d, $J = 6.9$ Hz, 3H),

0.69 (s, 3H, 18-H). ^{13}C NMR (125 MHz, CDCl_3) δ 211.0, 208.1, 76.3, 72.5, 72.0, 58.5, 56.3, 53.3, 52.5, 47.7, 46.3, 42.8, 42.7, 41.4, 40.2, 39.2, 37.5, 35.0, 27.7, 27.0, 23.9, 22.1, 21.8, 17.2, 13.8, 12.4, 11.8, 10.8. HRMS (ESI): calc for $\text{C}_{28}\text{H}_{43}\text{O}_3^+$ $[\text{M}-2\text{H}_2\text{O}+\text{H}]^+$ 427.3207, found 427.3210.

Table 1 – ^1H and ^{13}C NMR spectroscopic data for epoxides **22** and **24**^{a,b,c}

Position	Compound			
	epoxide 22		epoxide 24	
	δ , H	δ , C	δ , H	δ , C
1	1.62 m; 2.02	39.8	1.97 m; 2.77 m	24.5
2	3.29 m	52.0	3.21 m	52.1
3	3.13 dd (4.1, 4.1)	50.4	3.22 m	49.7
4	2.12 ddd (16.0, 4.5, 2.6); 2.51 ddd (16.0, 11.6, 1.0)	25.2	1.60 m; 2.38 d (15.6)	38.7
5	3.08 dd (11.5, 4.7)	41.2	2.72 m	42.8
6	-	176.0	-	175.5
7	3.99 dd (12.3, 8.8); 4.05 dd (12.3, 1.2)	70.1	3.98 dd (12.5, 9.0); 4.08 (12.5)	70.3
8	1.44	40.0	1.65 m	39.3
9	1.15	59.4	1.11 m	59.9
10	-	33.8	-	36.5
11	1.44 m; 1.66 m	22.6 ^d	1.41 m; 1.72 m	22.3 ^d
12	1.22 m; 1.97 m	39.6	1.20 m; 1.98 m	39.5
13	-	42.5	-	42.3
14	1.14 m	51.3	1.12	51.2
15	1.65 m	24.6 ^d	1.21 m; 1.67 m	24.7 ^d
16	1.31 m; 1.97 m	27.6 ^d	1.31 m; 1.98 m	27.5 ^d
17	1.53 m	52.5	1.53 m	52.5
18	0.68 s	11.7	0.68 s	11.5
19	0.89 s	17.9	0.90 s	22.1
20	1.44 m	40.2	1.45 m	40.2
21	0.94 d (6.7)	12.3	0.94 d (6.6)	12.3
22	3.65 br.s	72.4	3.66 d (3.4)	72.5
23	3.37 br.s	76.2	3.38 dd (5.2, 5.2)	76.2
24	1.47 m	41.4	1.48 m	41.4
25	1.88 m	26.9	1.88 m	26.9
26	0.84 d (6.8) ^e	17.2 ^e	0.85 d (6.8) ^e	17.2
27	0.89 d (6.7) ^e	22.1 ^e	0.89 d (6.6) ^e	17.6
28	0.82 d (7.0)	10.8	0.82 d (7.0)	10.8

^a NMR chemical shifts (δ) are from spectra obtained in CDCl_3 solutions.

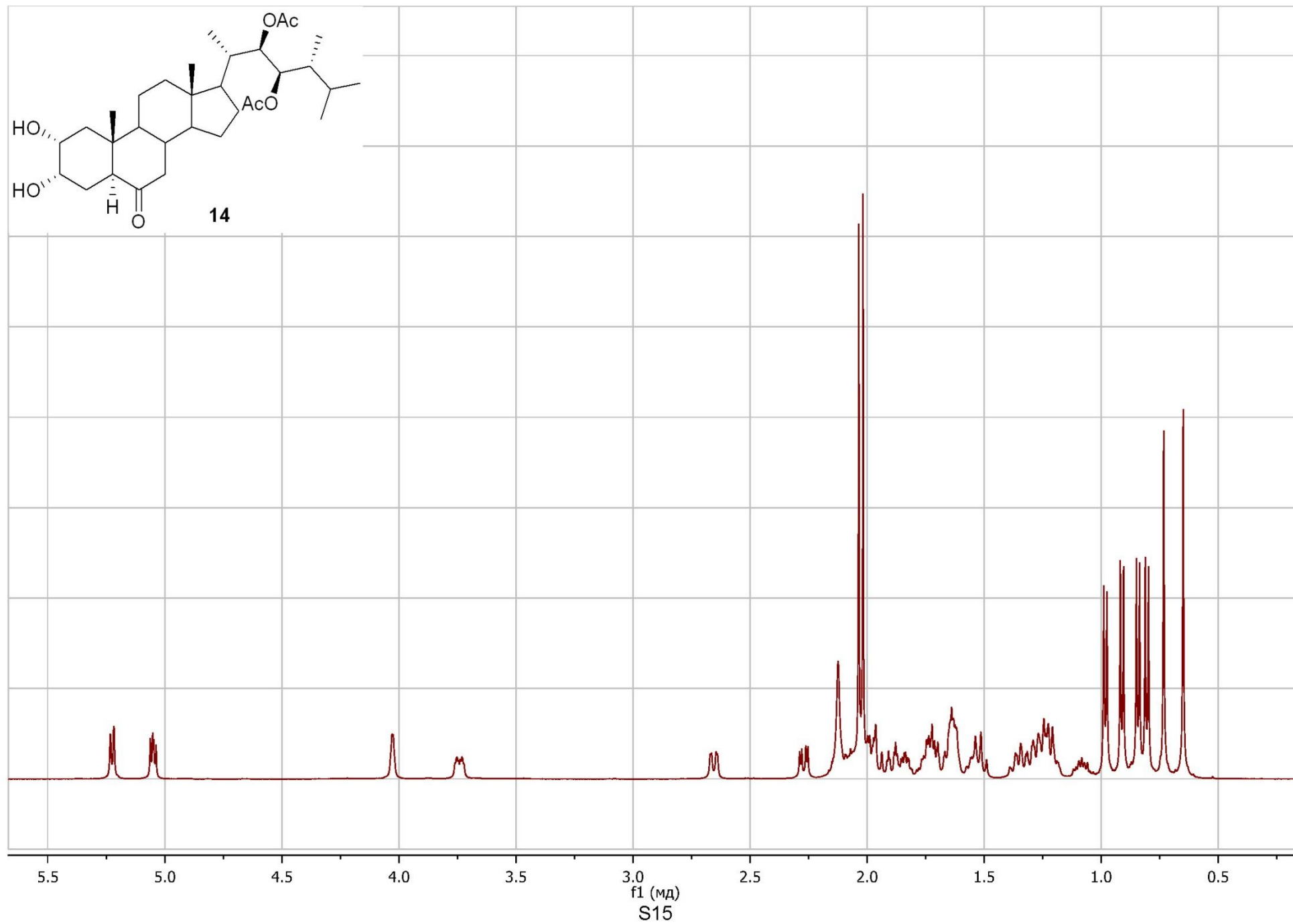
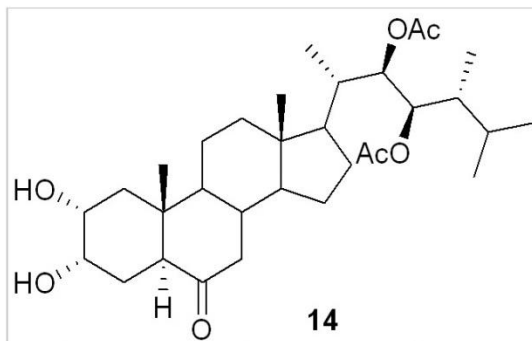
^b Assigned by DEPT, COSY, HSQC, and HMBC experiments.

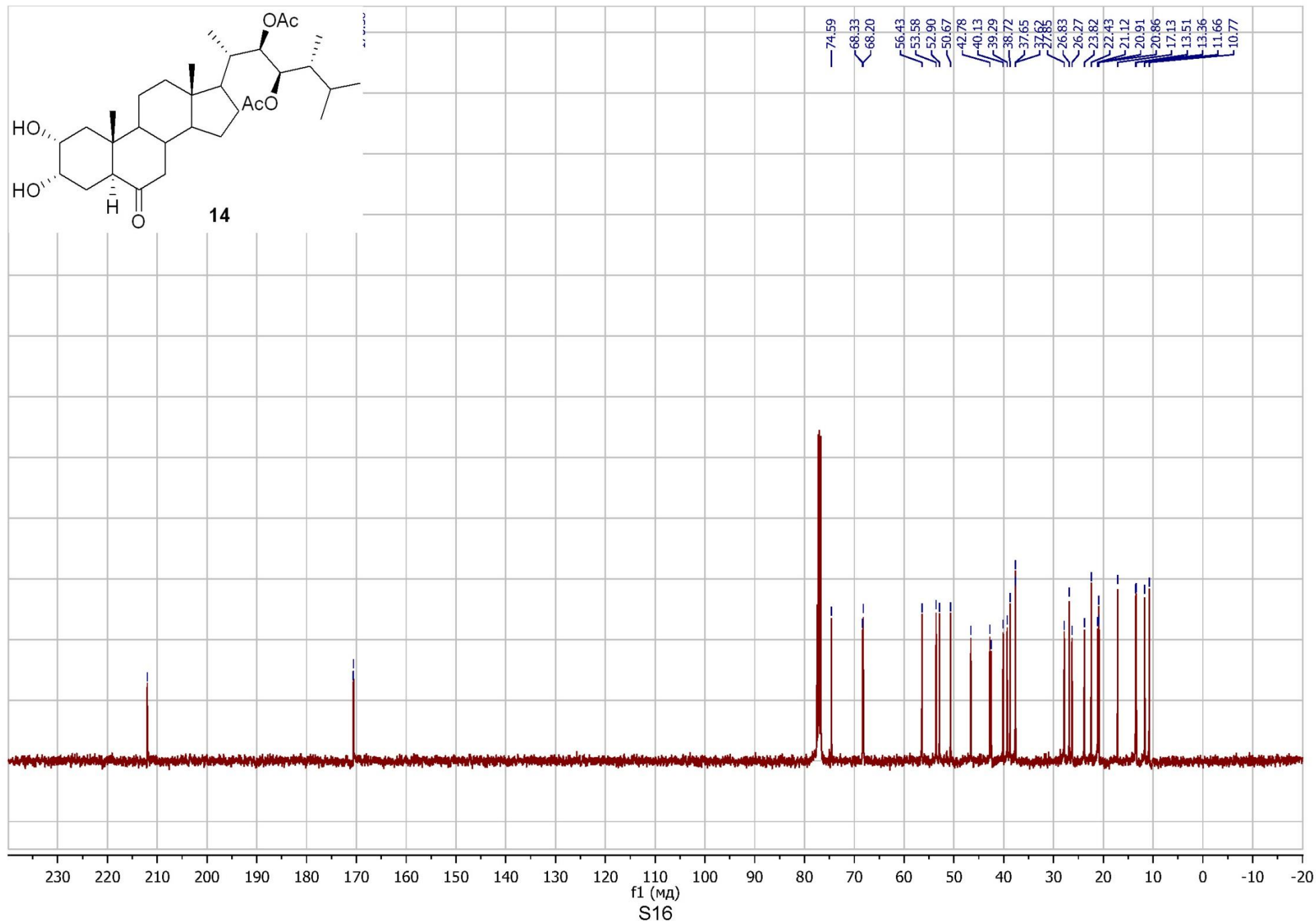
^c J values (in Hz) in parentheses.

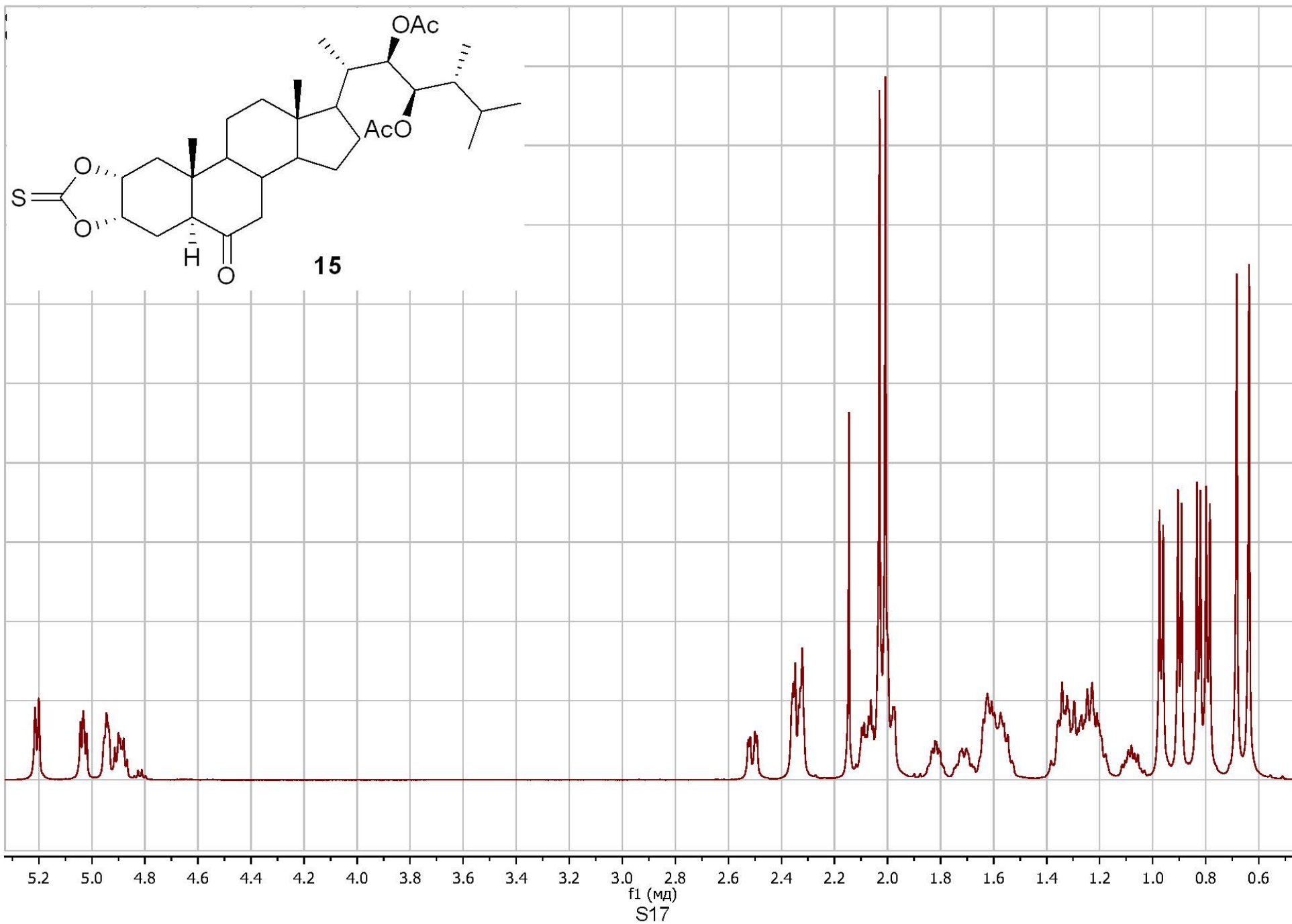
^{d,e} May be reversed.

References

1. Khripach, V. A.; Zhabinskii, V. N.; Gulyakevich, O. V.; Konstantinova, O. V.; Misharin, A. Y.; Mekhtiev, A. R.; Timofeev, V. P.; Tkachev, Y. V. *Russ. J. Bioorg. Chem.* **2010**, 36, 746-754. doi: 10.1134/S1068162010060117
2. Voigt, B.; Takatsuto, S.; Yokota, T.; Adam, G. *J. Chem. Soc., Perkin Trans. 1* **1995**, 1495-1498. doi: 10.1039/P19950001495
3. Zhylitskaya, H. A.; Litvinovskaya, R. P.; Zhabinskii, V. N.; Khripach, V. A. *Steroids* **2017**, 117, 2-10. doi: 10.1016/j.steroids.2016.06.006
4. Voigt, B.; Porzel, A.; Adam, G.; Golsch, D.; Adam, W.; Wagner, C.; Merzweiler, K. *Collect. Czech. Chem. Commun.* **2002**, 67, 91-102. doi: 10.1135/cccc20020091
5. Levinson, E. E.; Traven, V. F. *J Chem Res (S)* **1996**, 196-197
6. Voigt, B.; Schmidt, J.; Adam, G. *Tetrahedron* **1996**, 52, 1997-2004. doi: 10.1016/0040-4020(95)01042-4

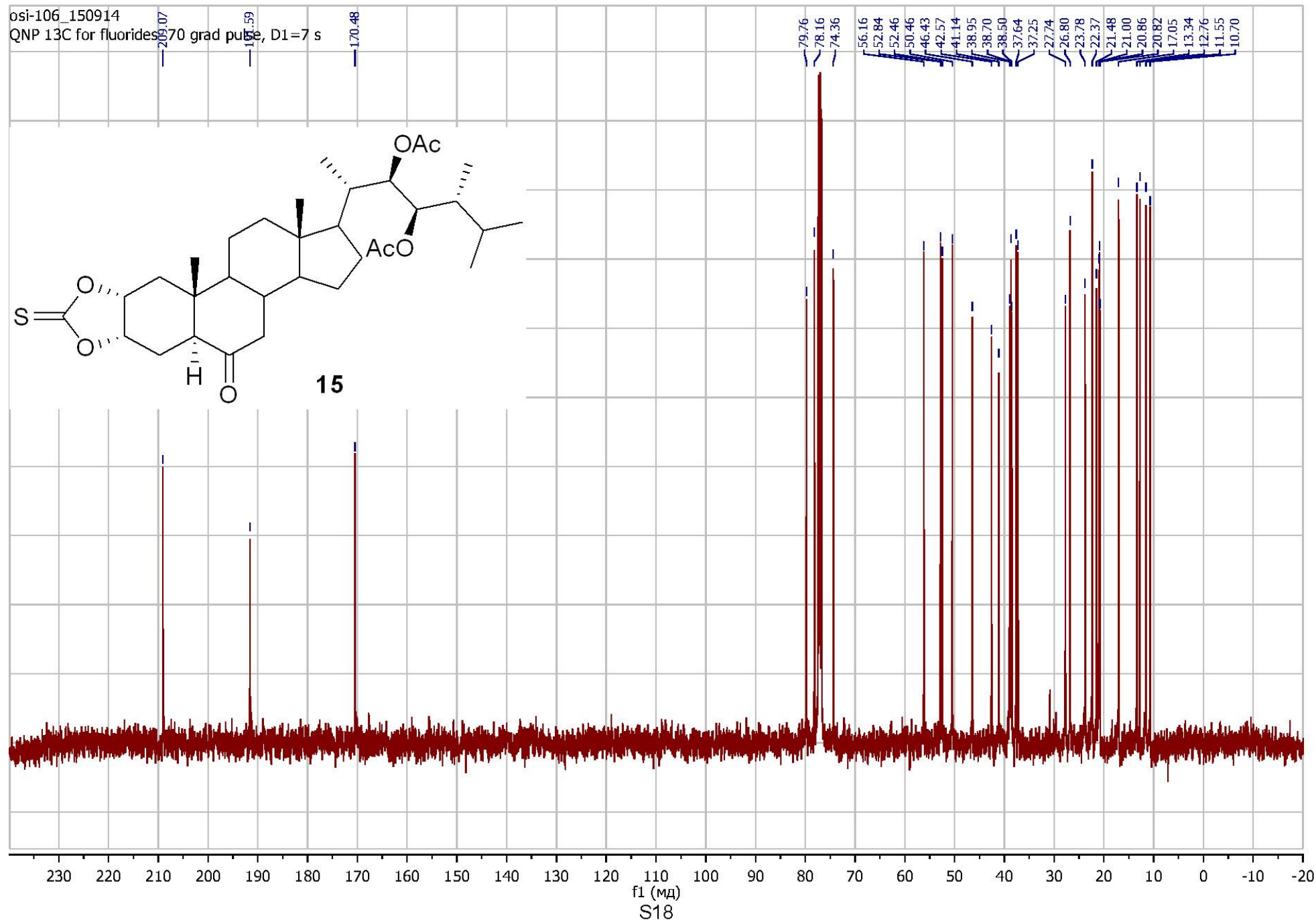
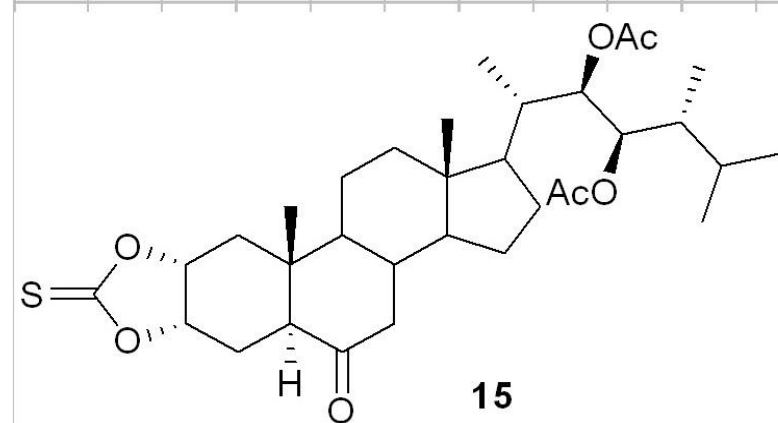


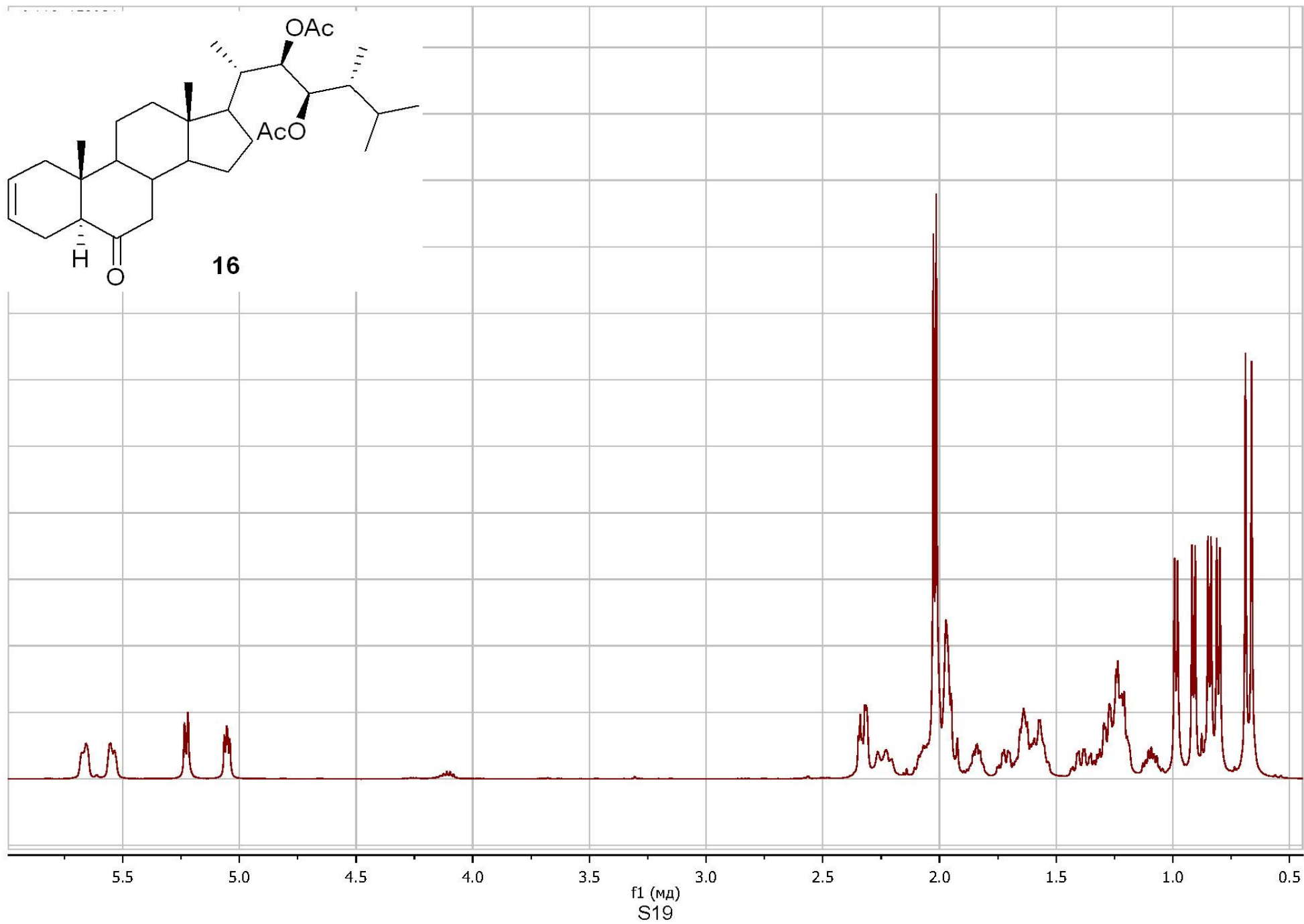




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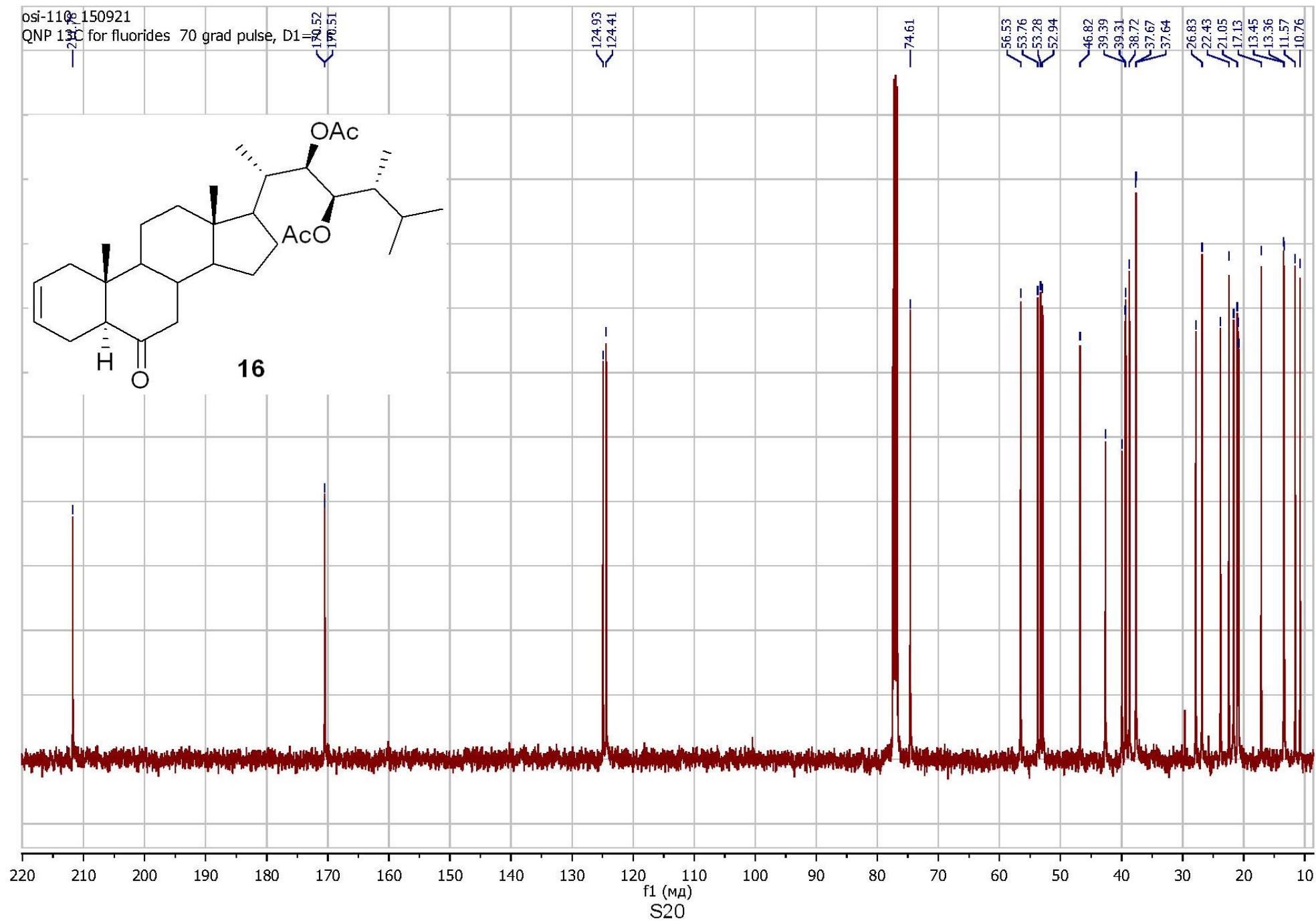
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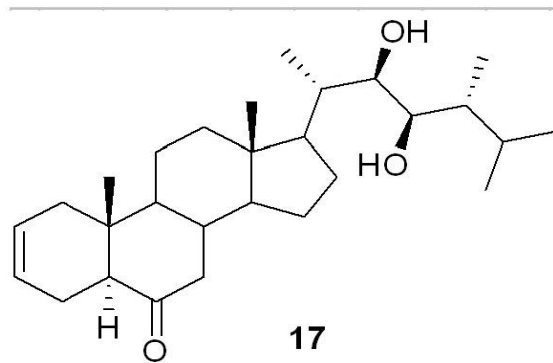




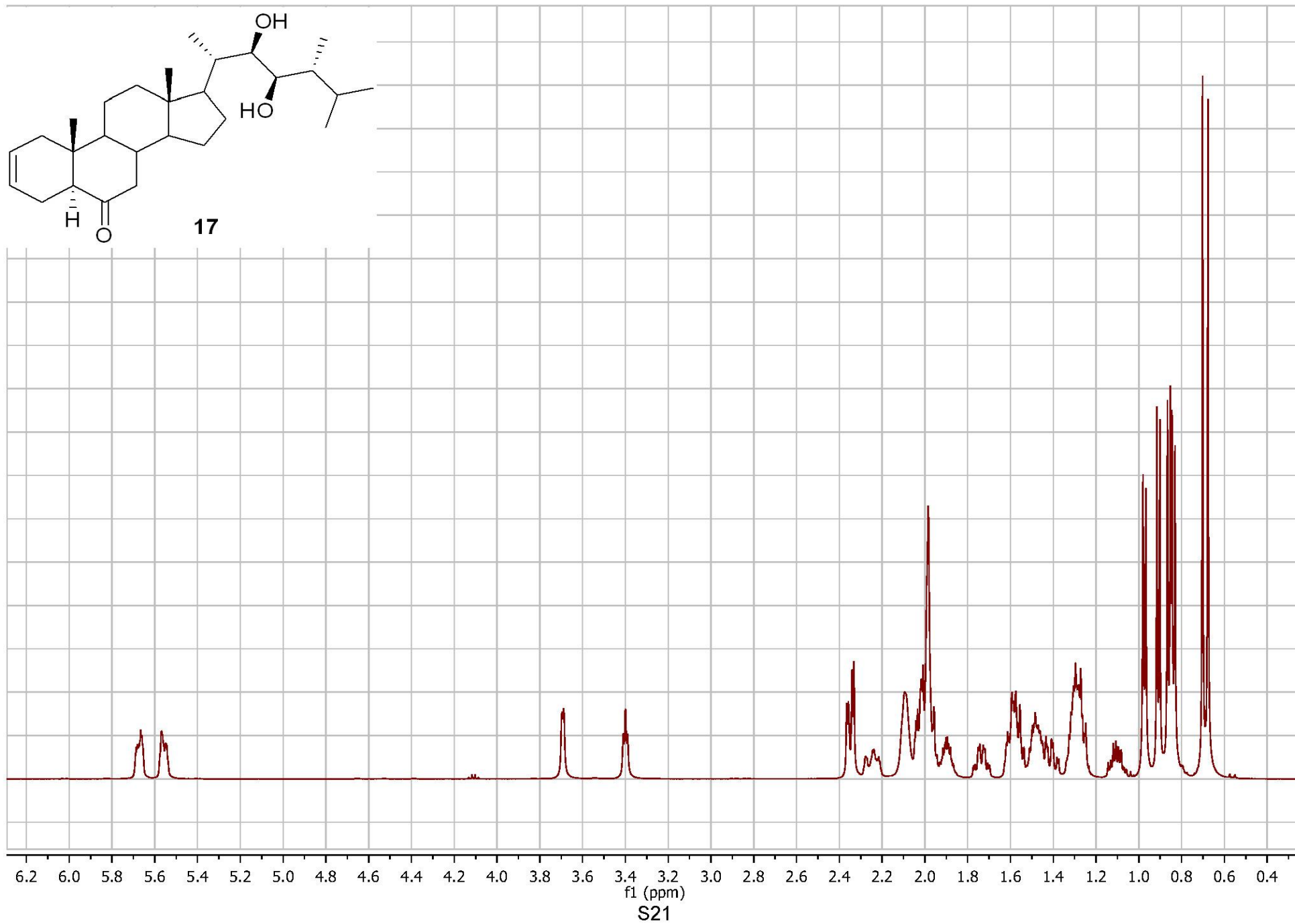
osi-110-150921

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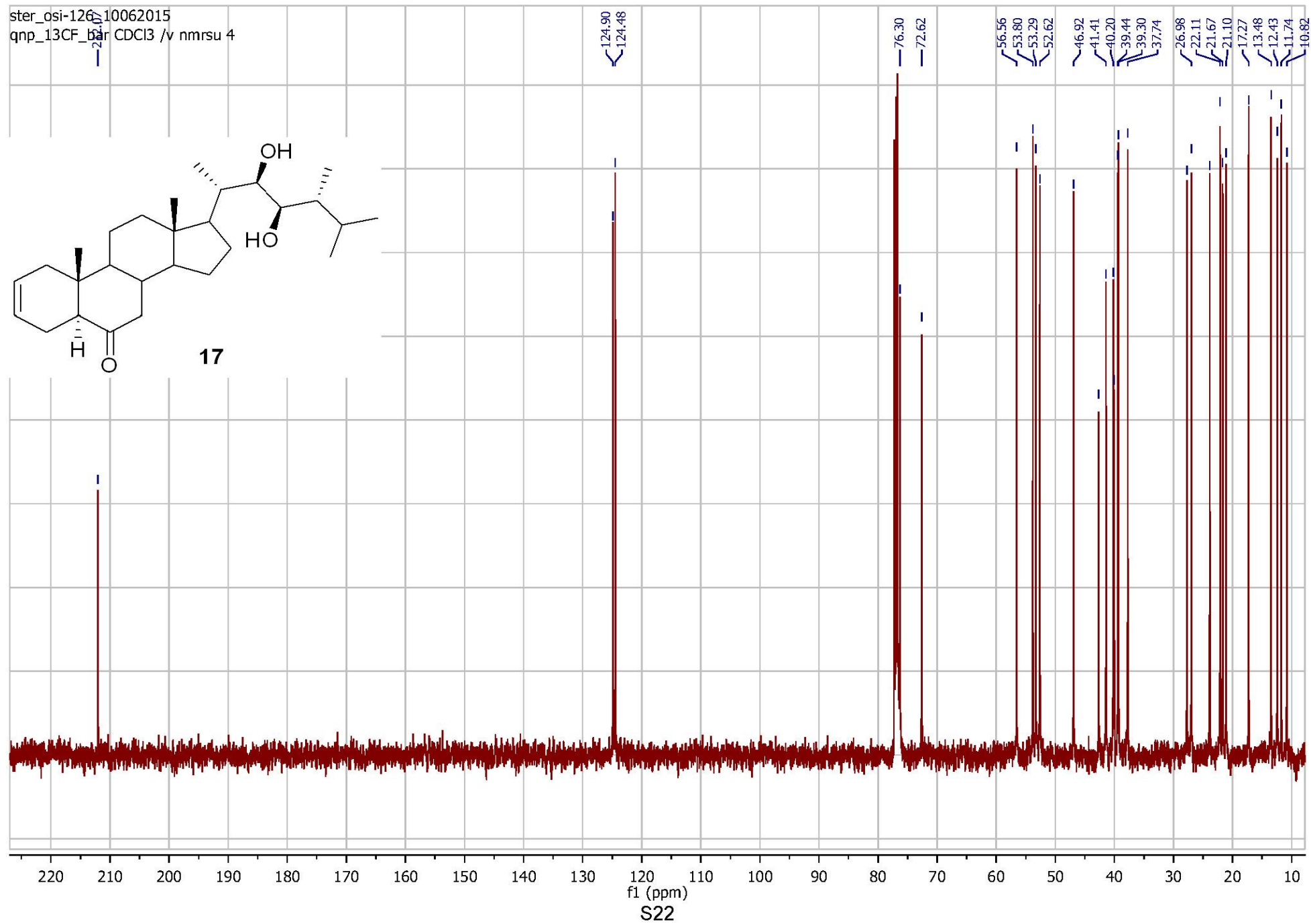
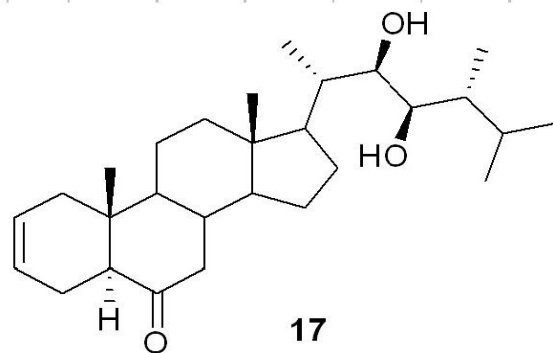


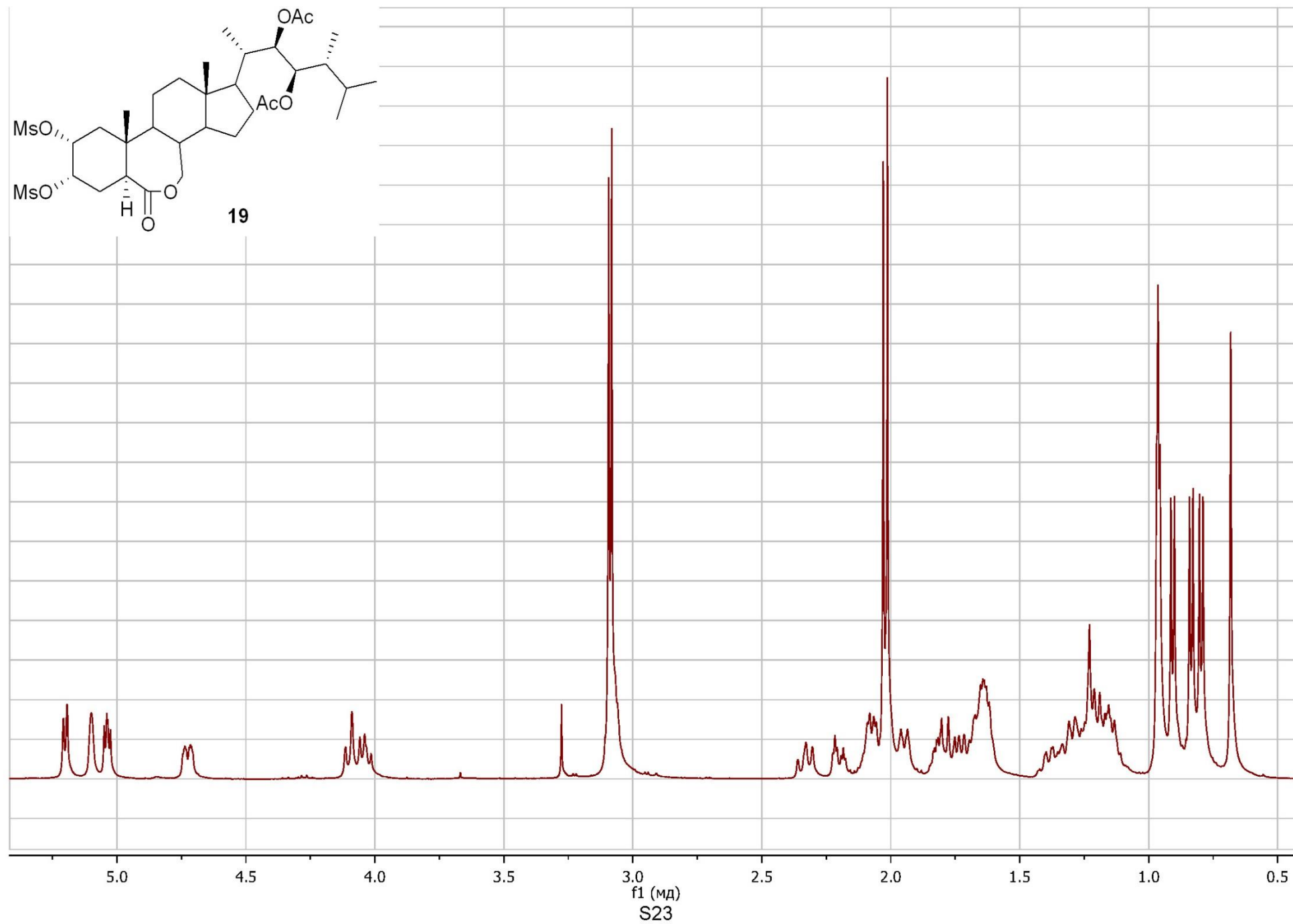


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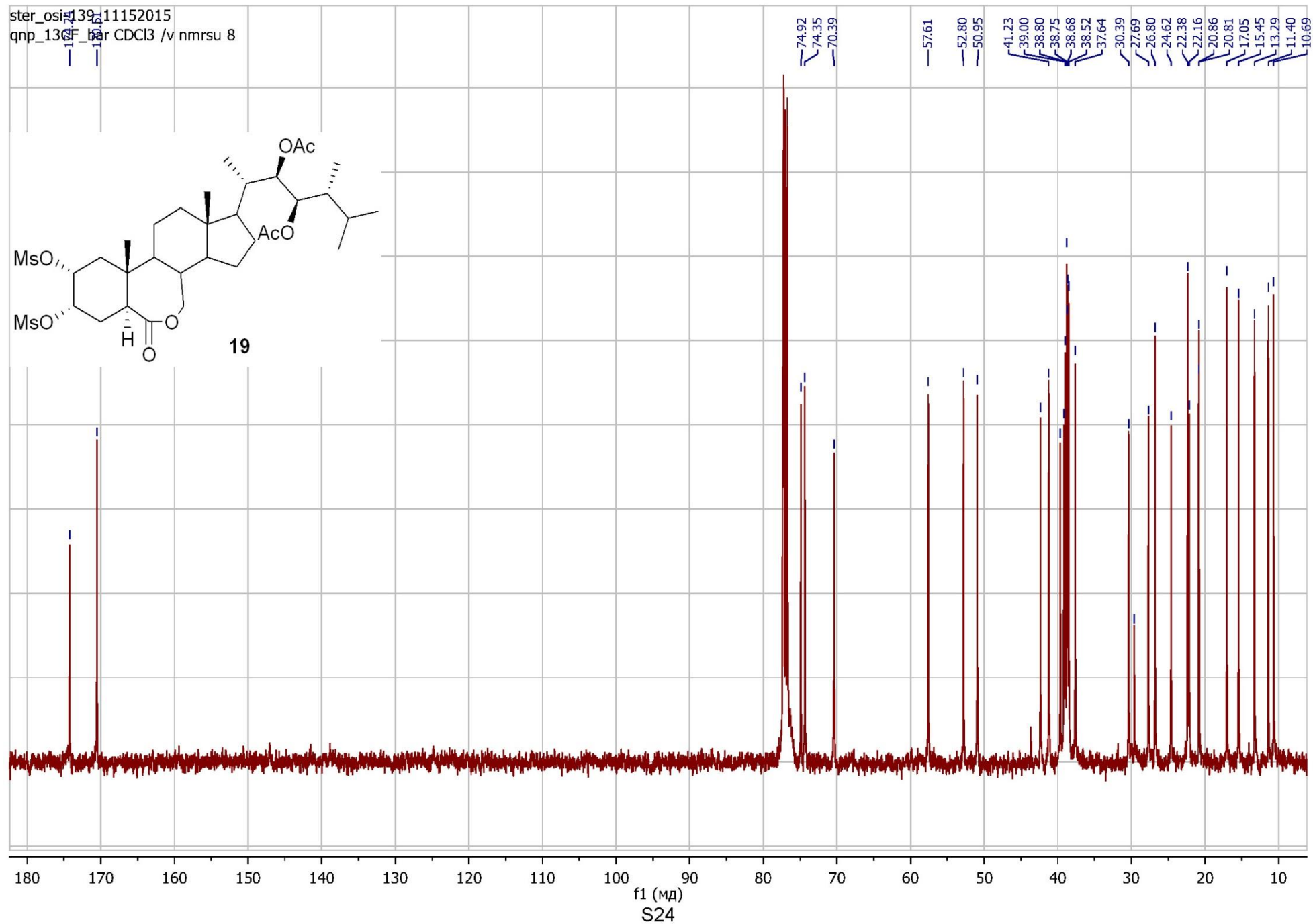
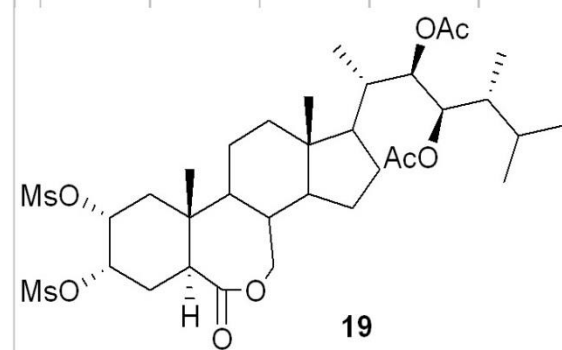


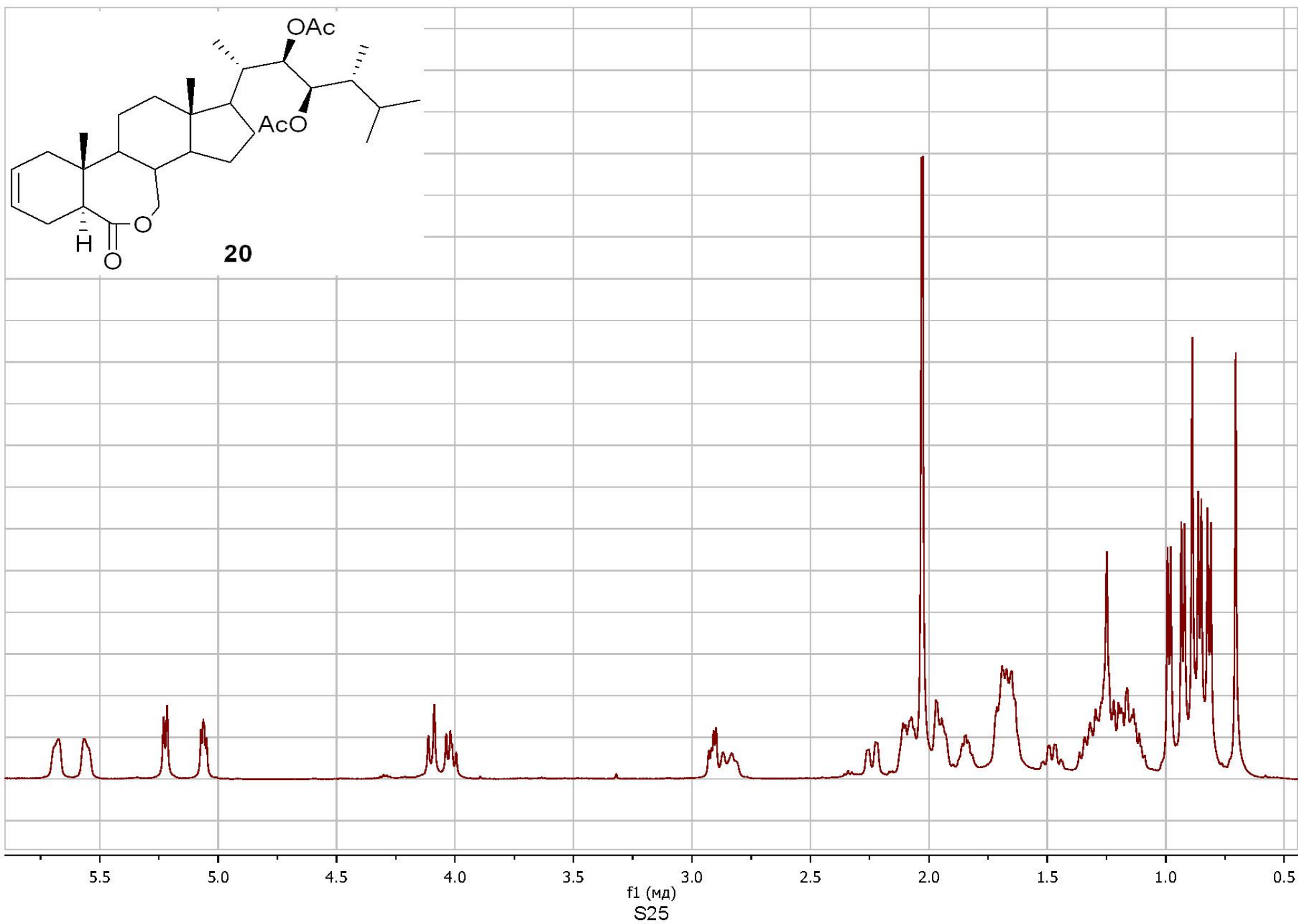
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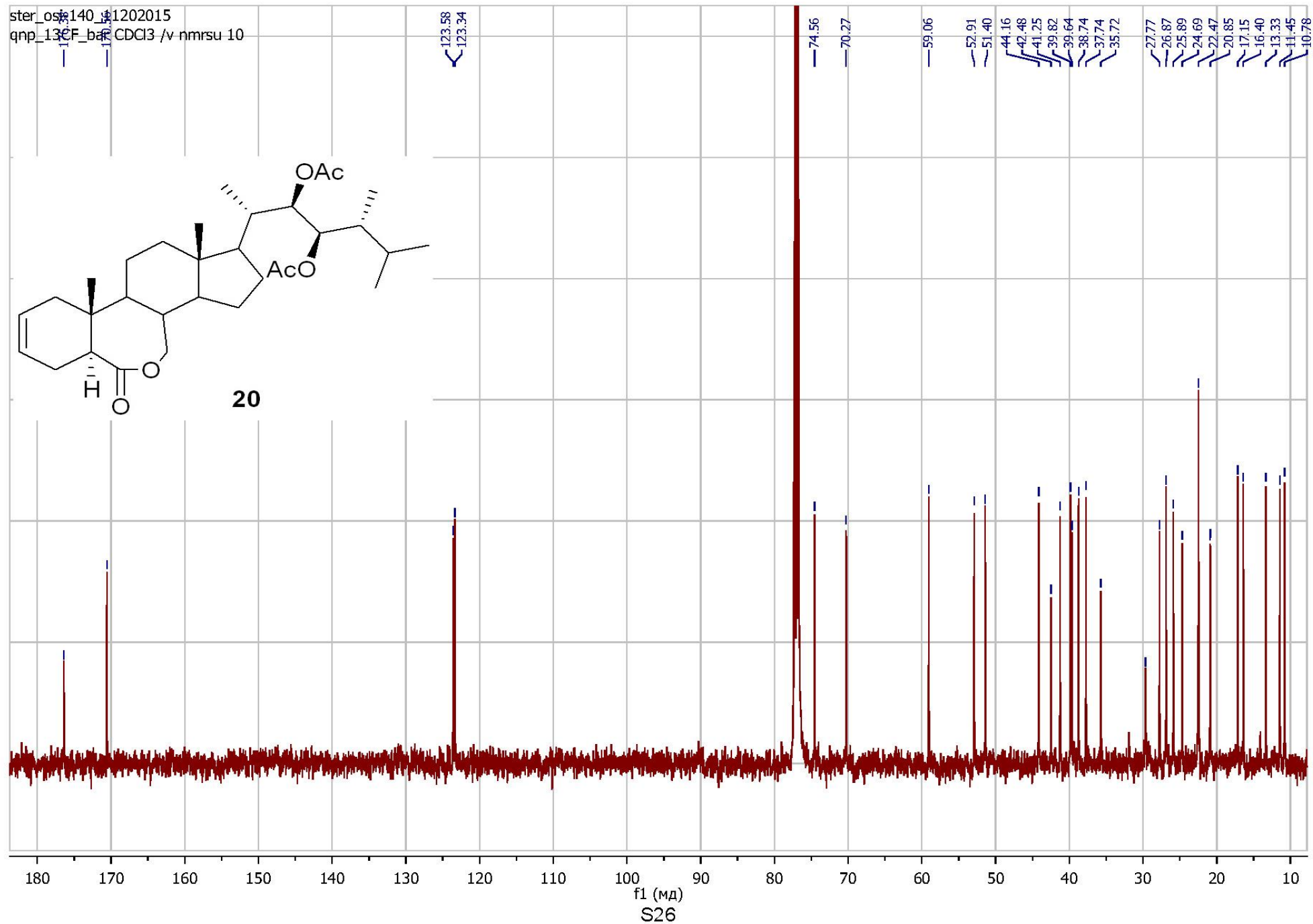
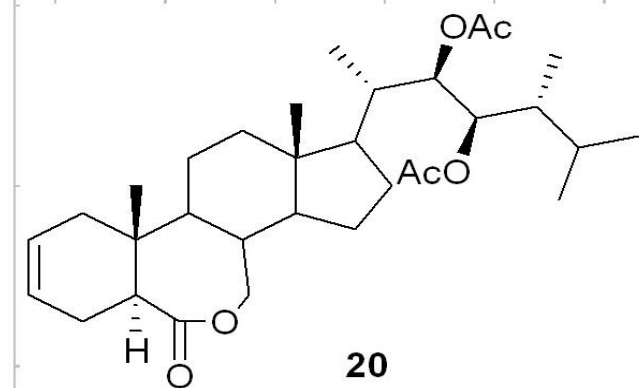


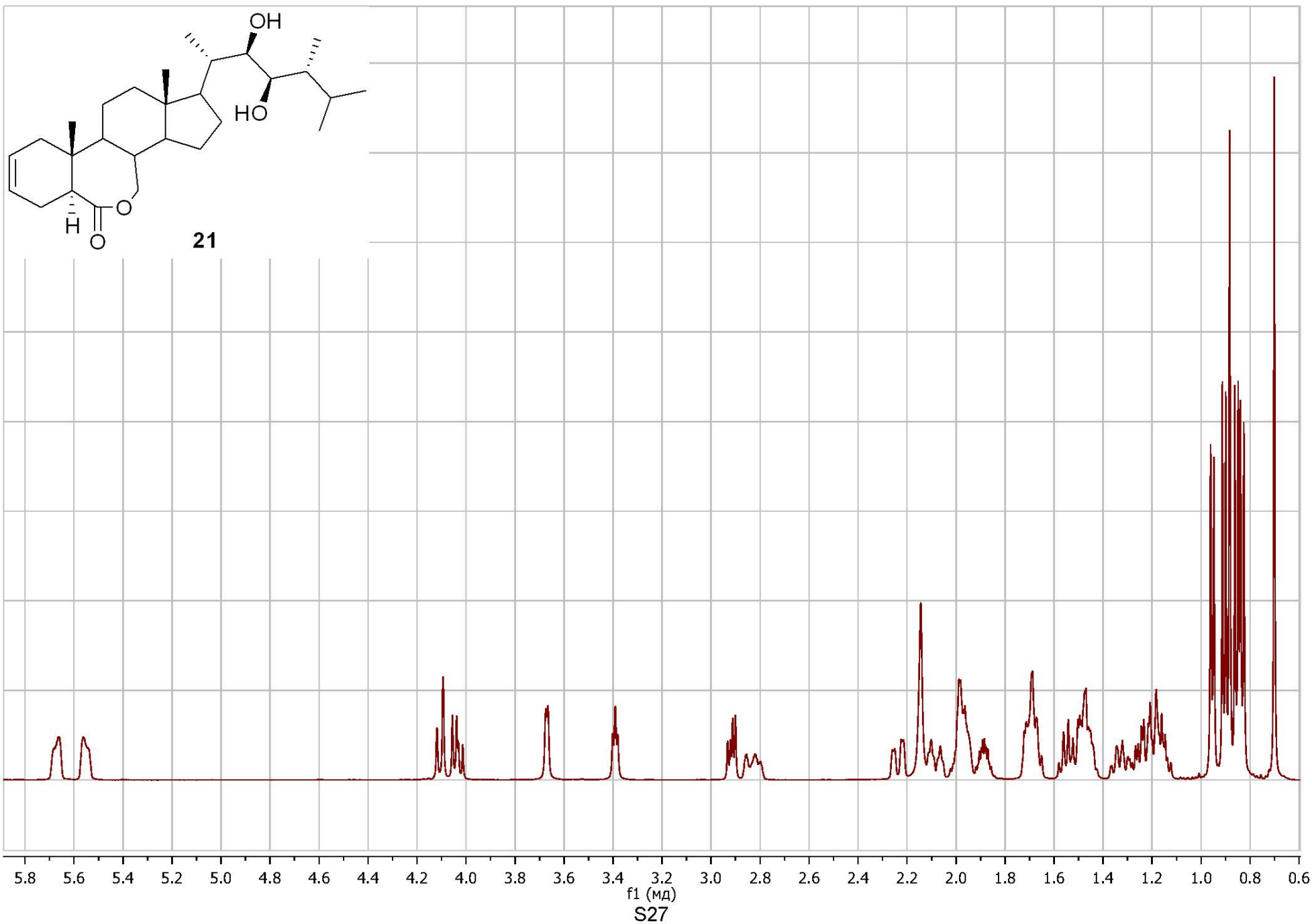
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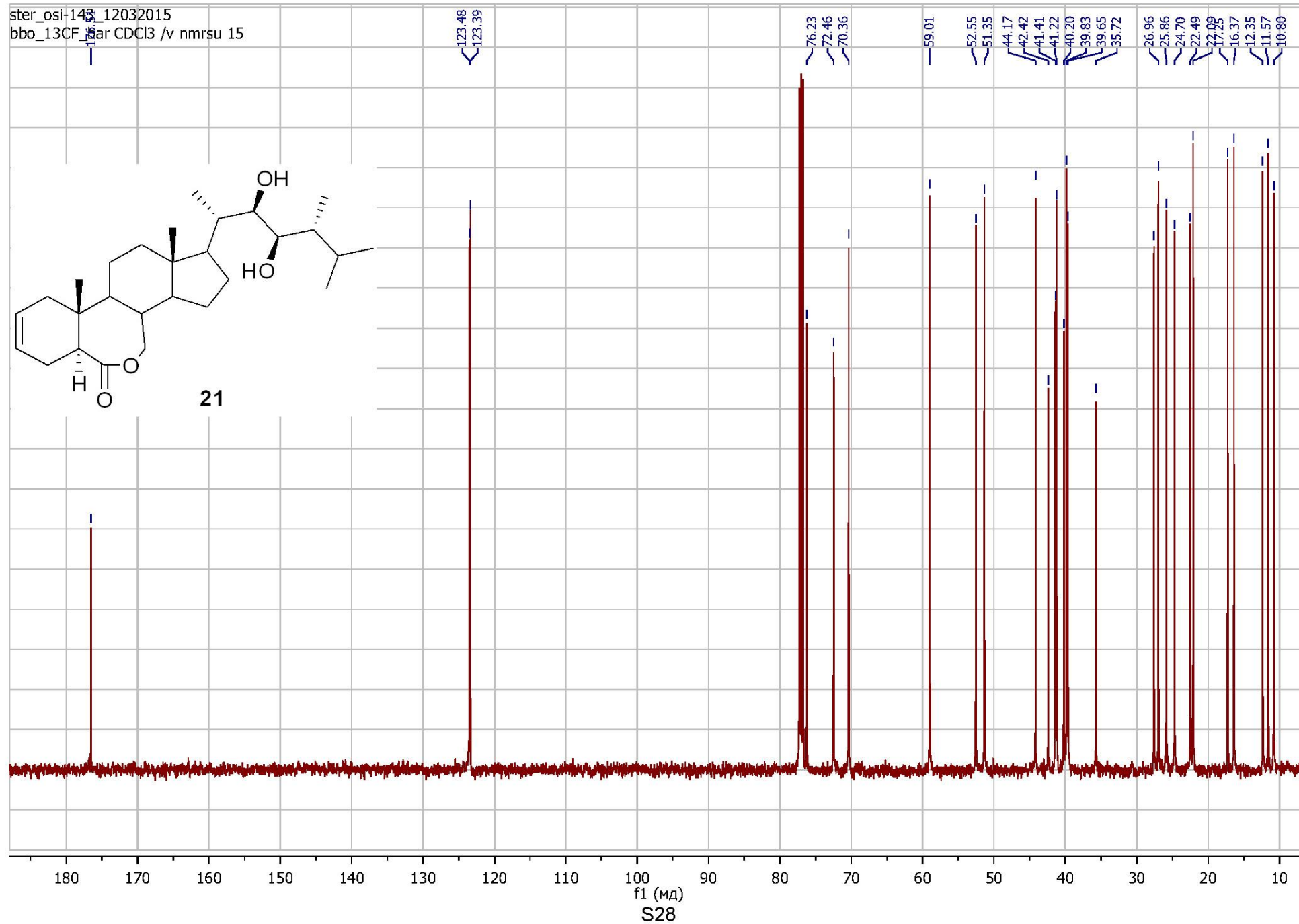


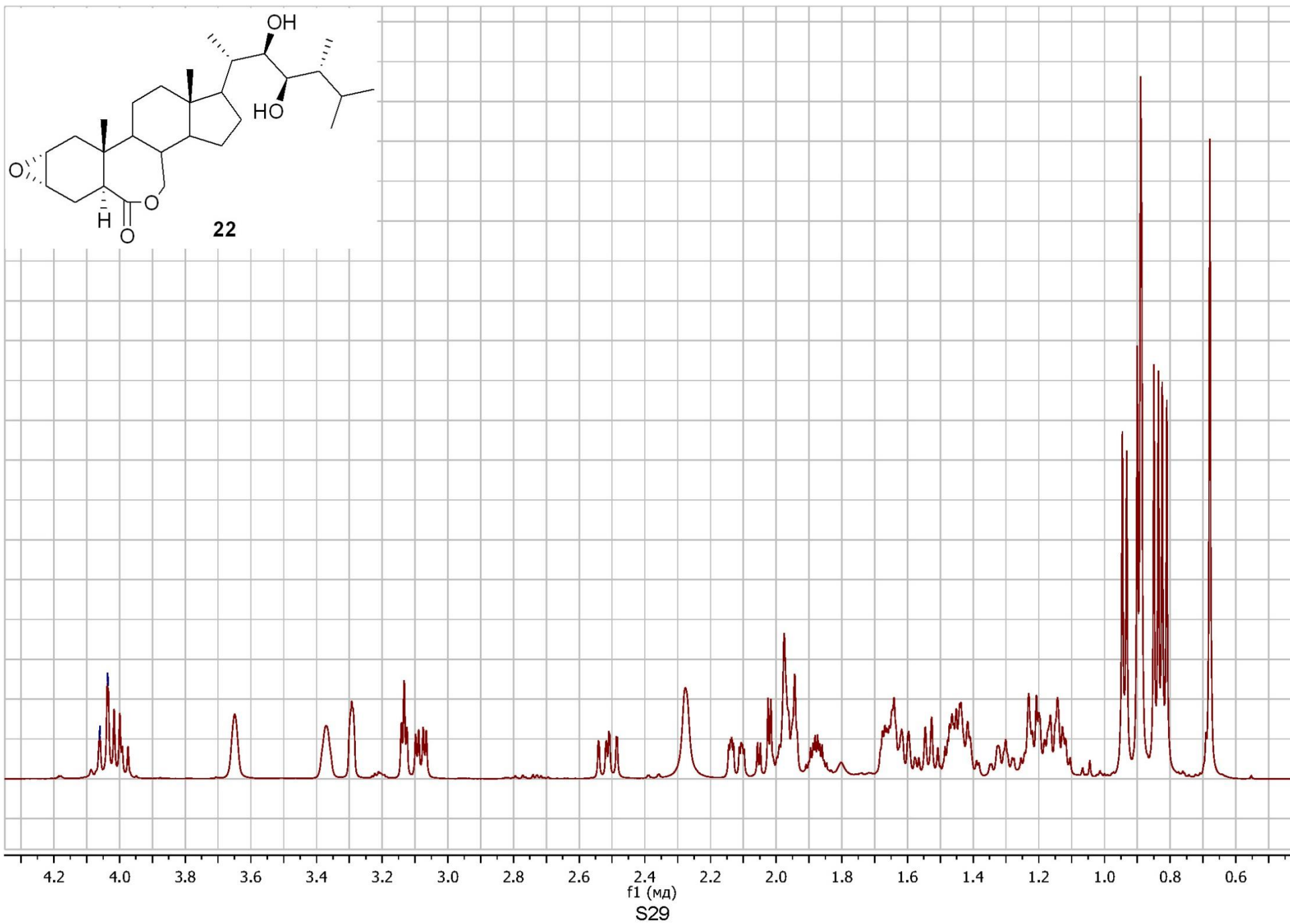
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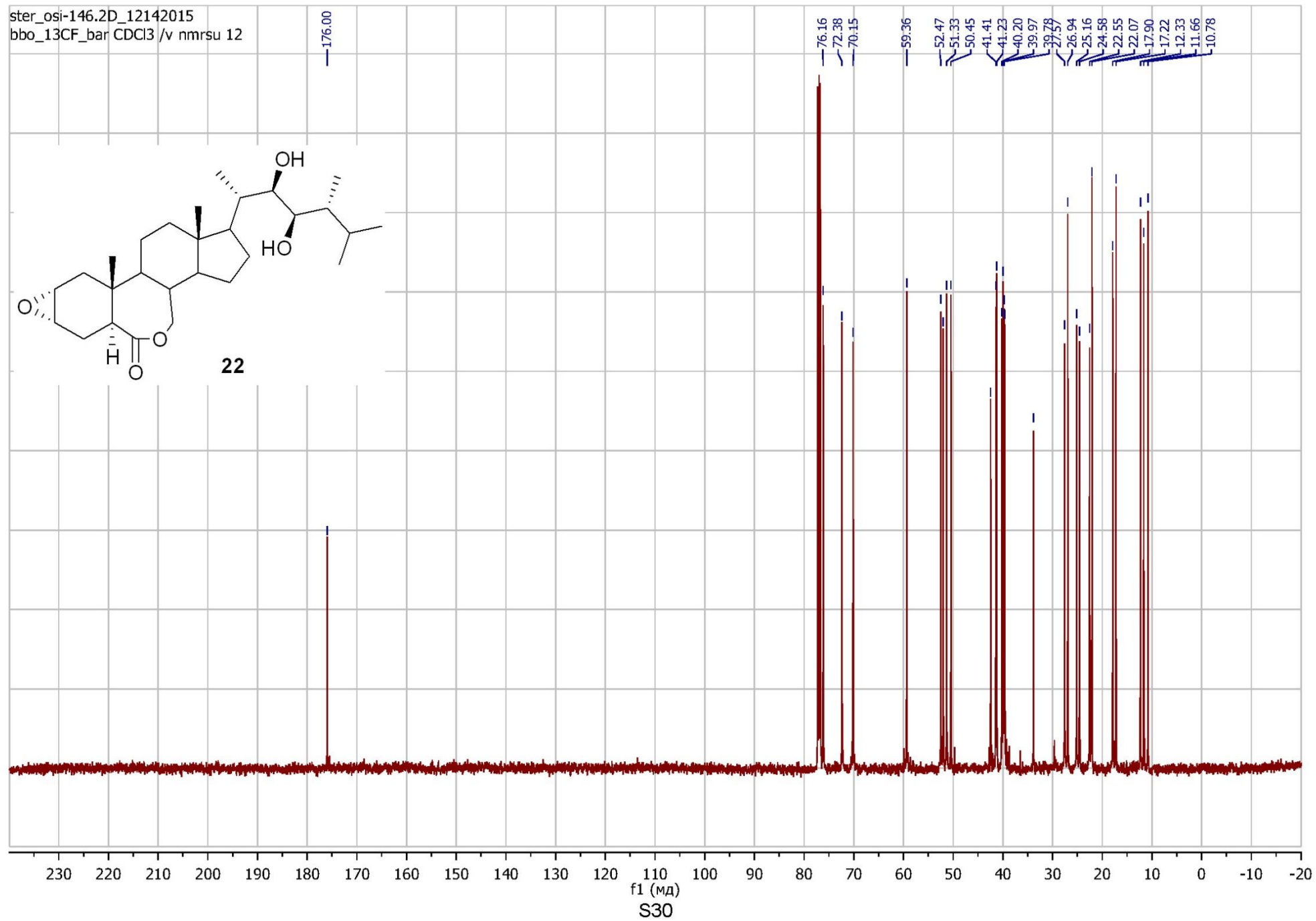


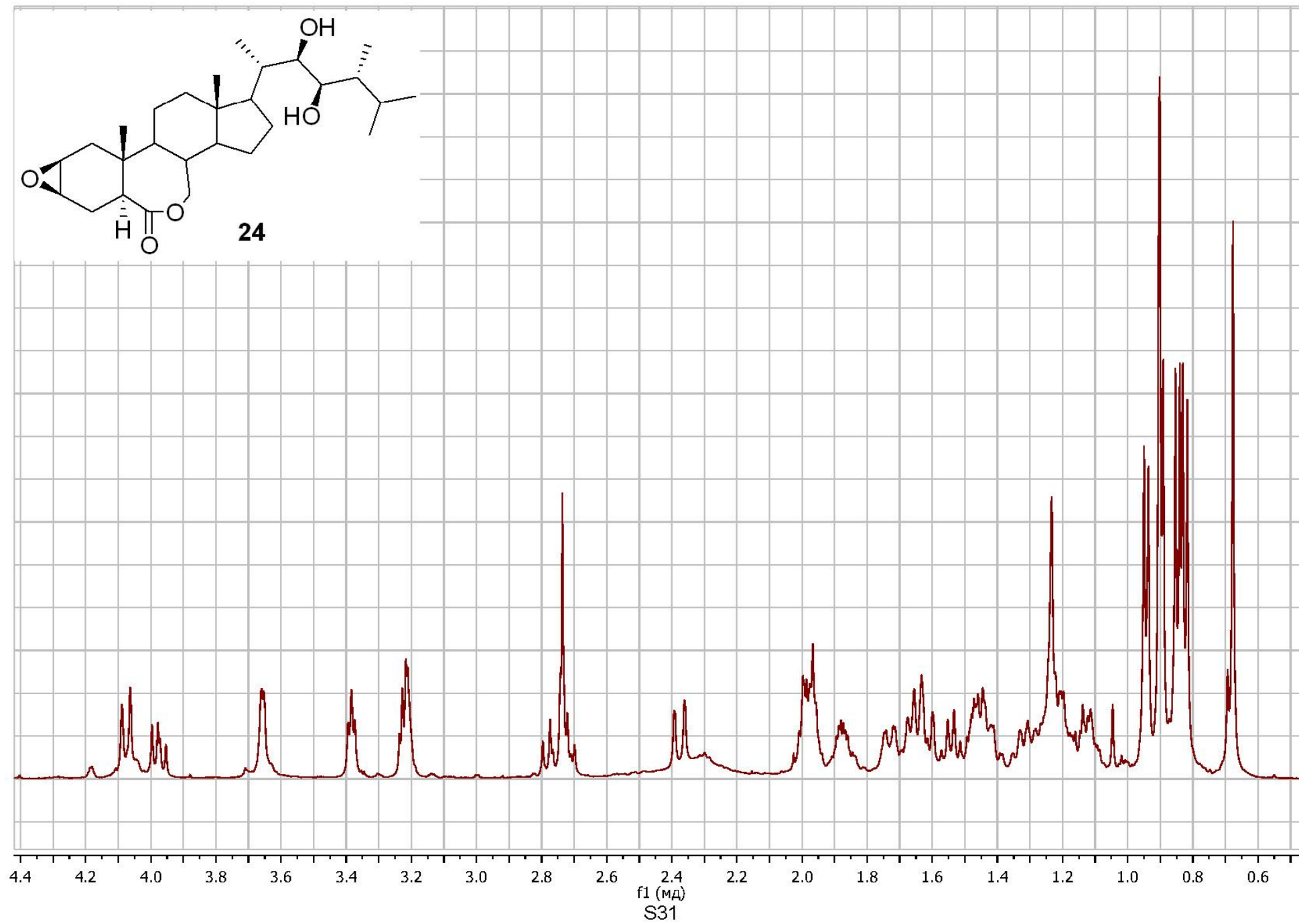
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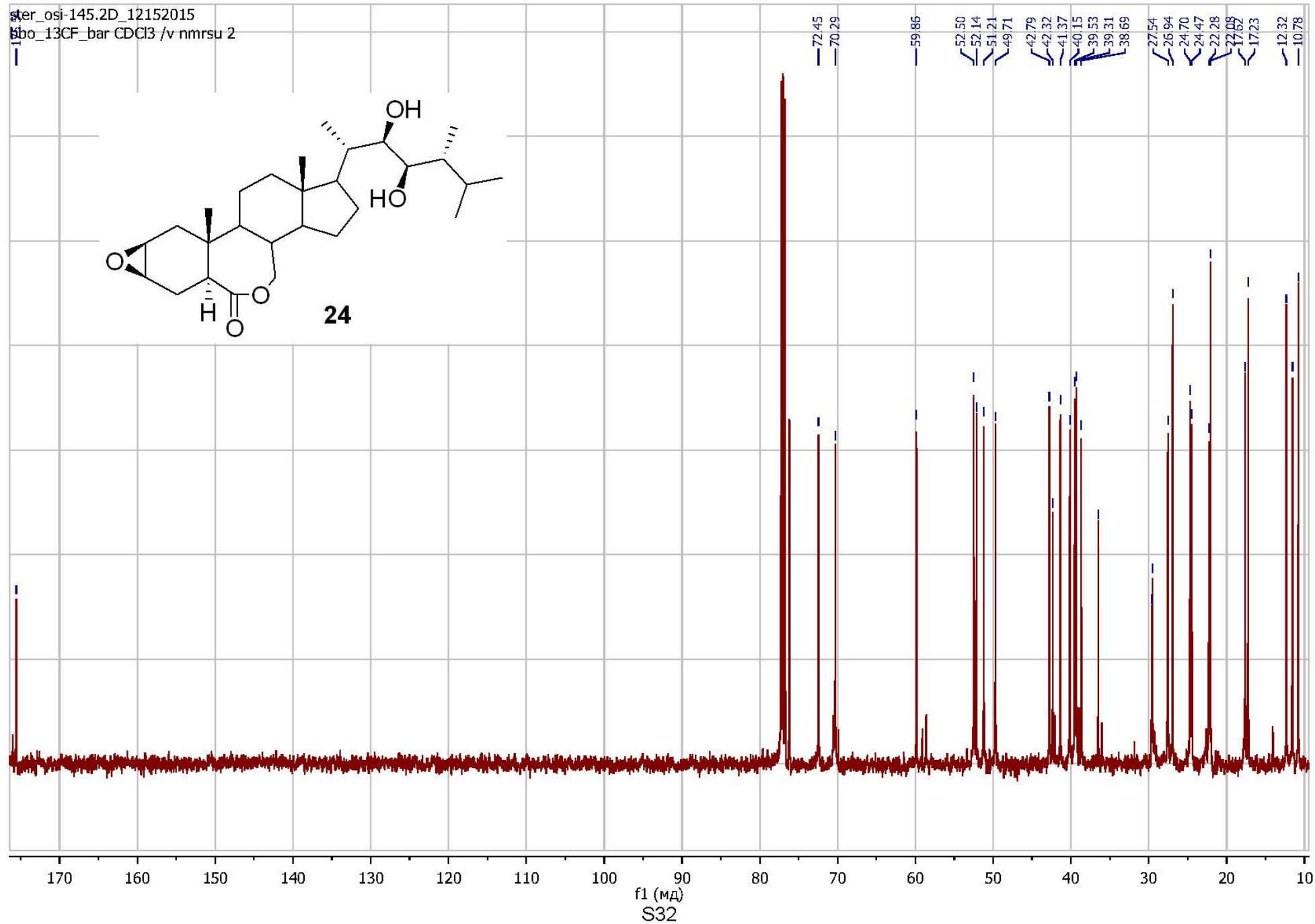
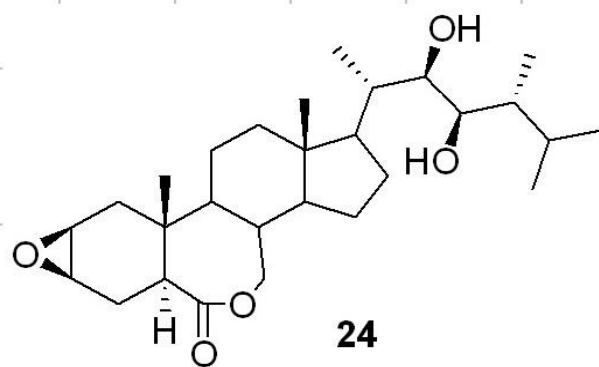


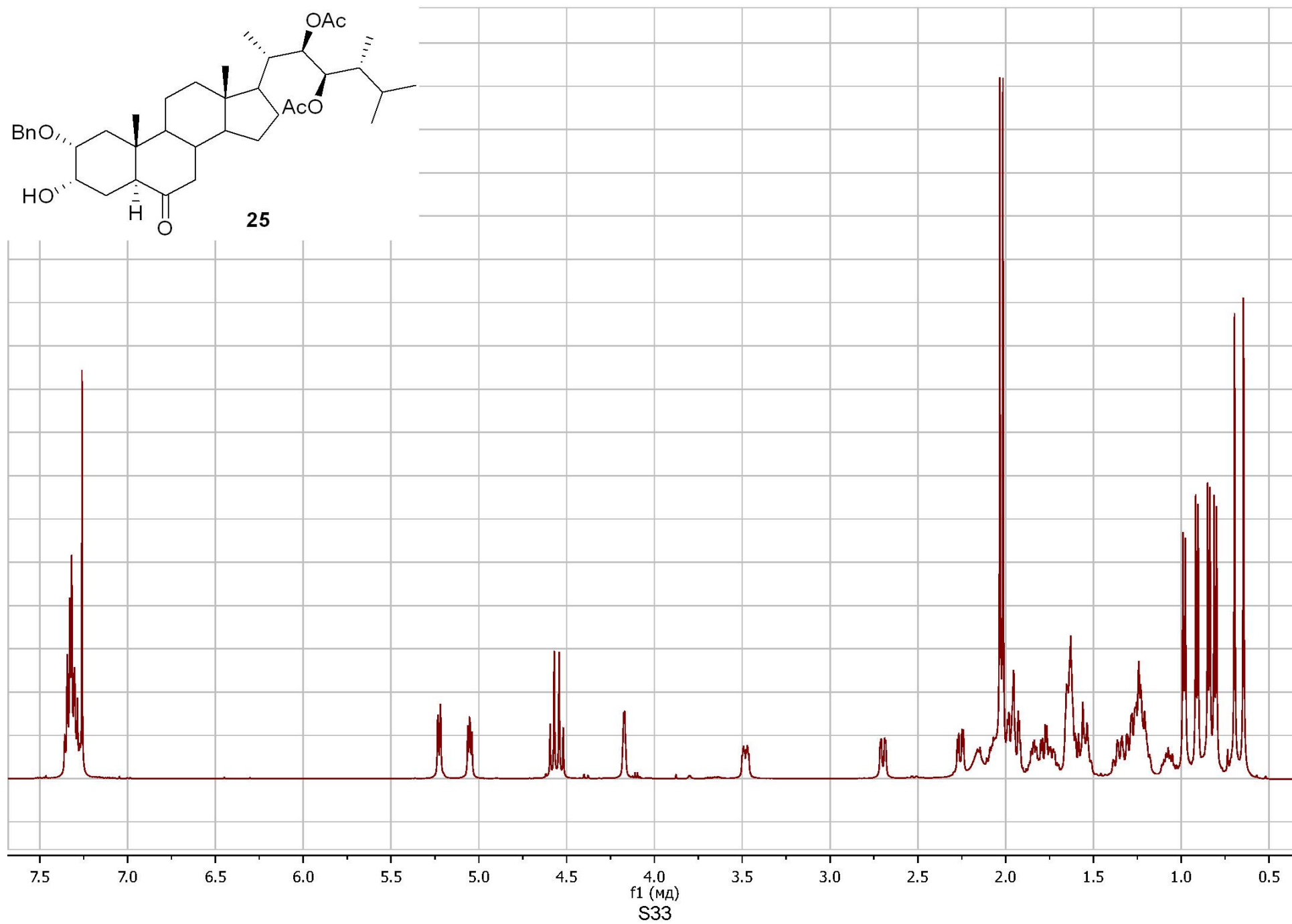
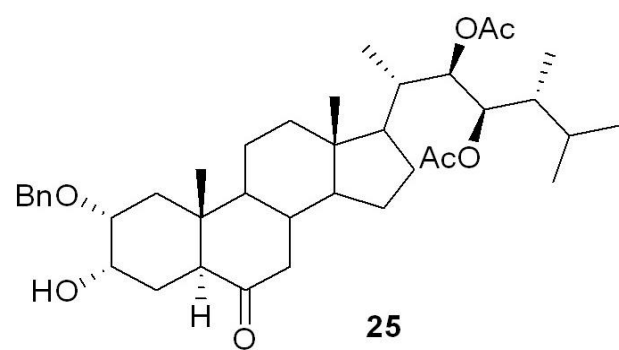
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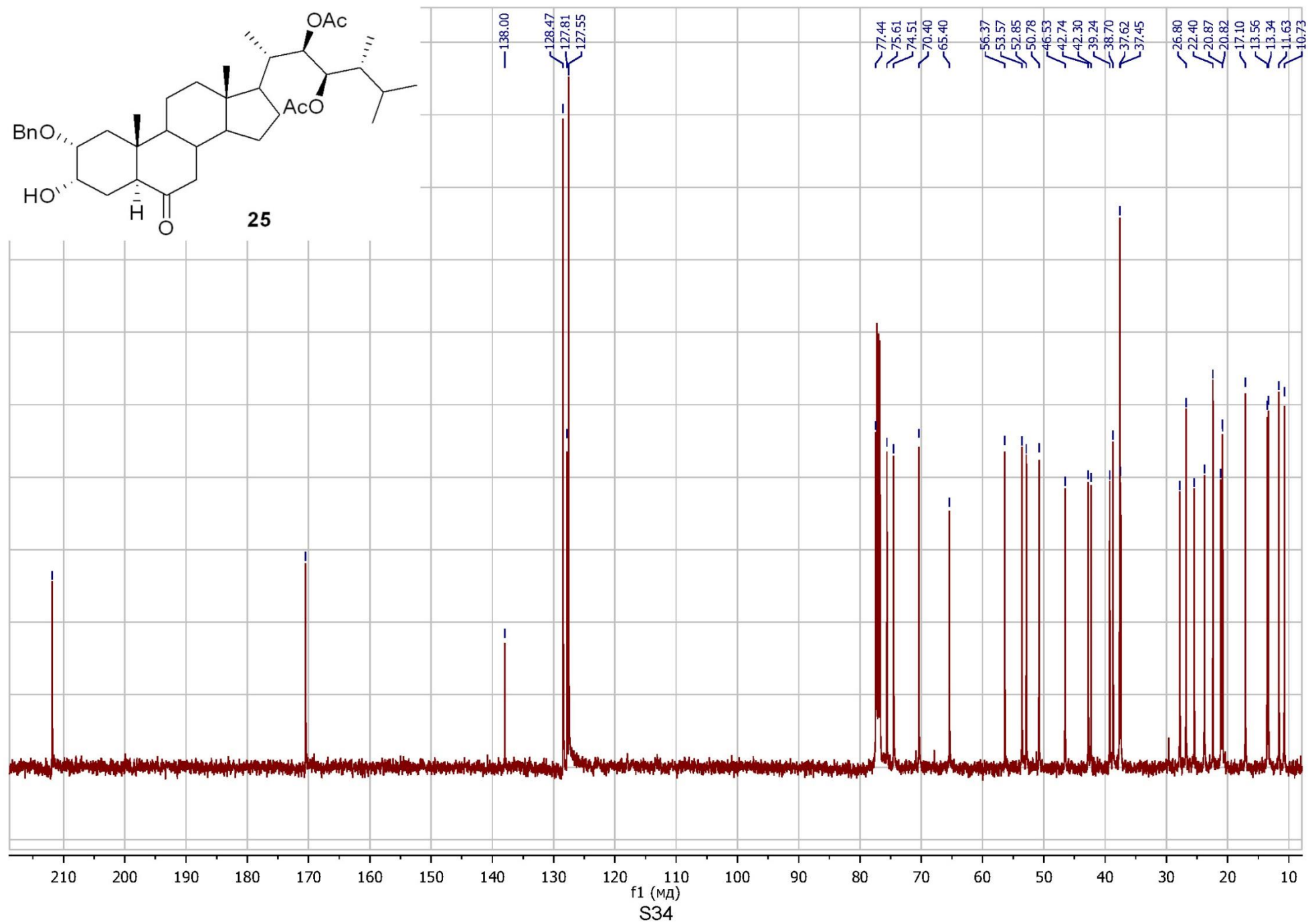
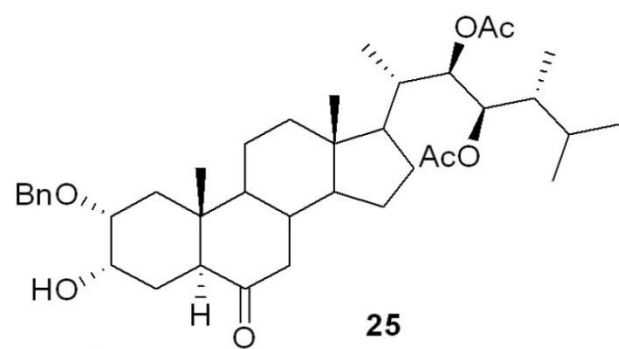


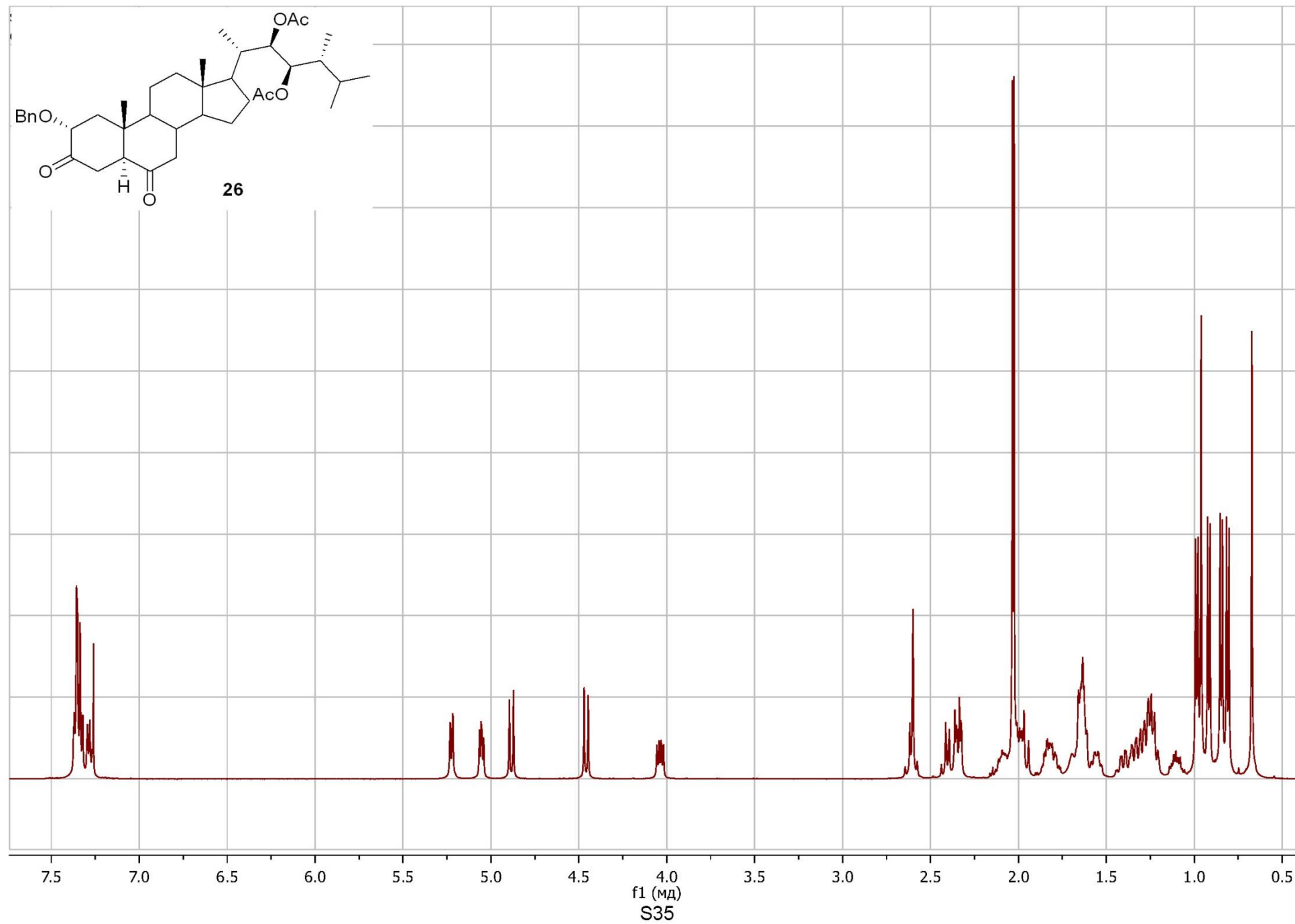


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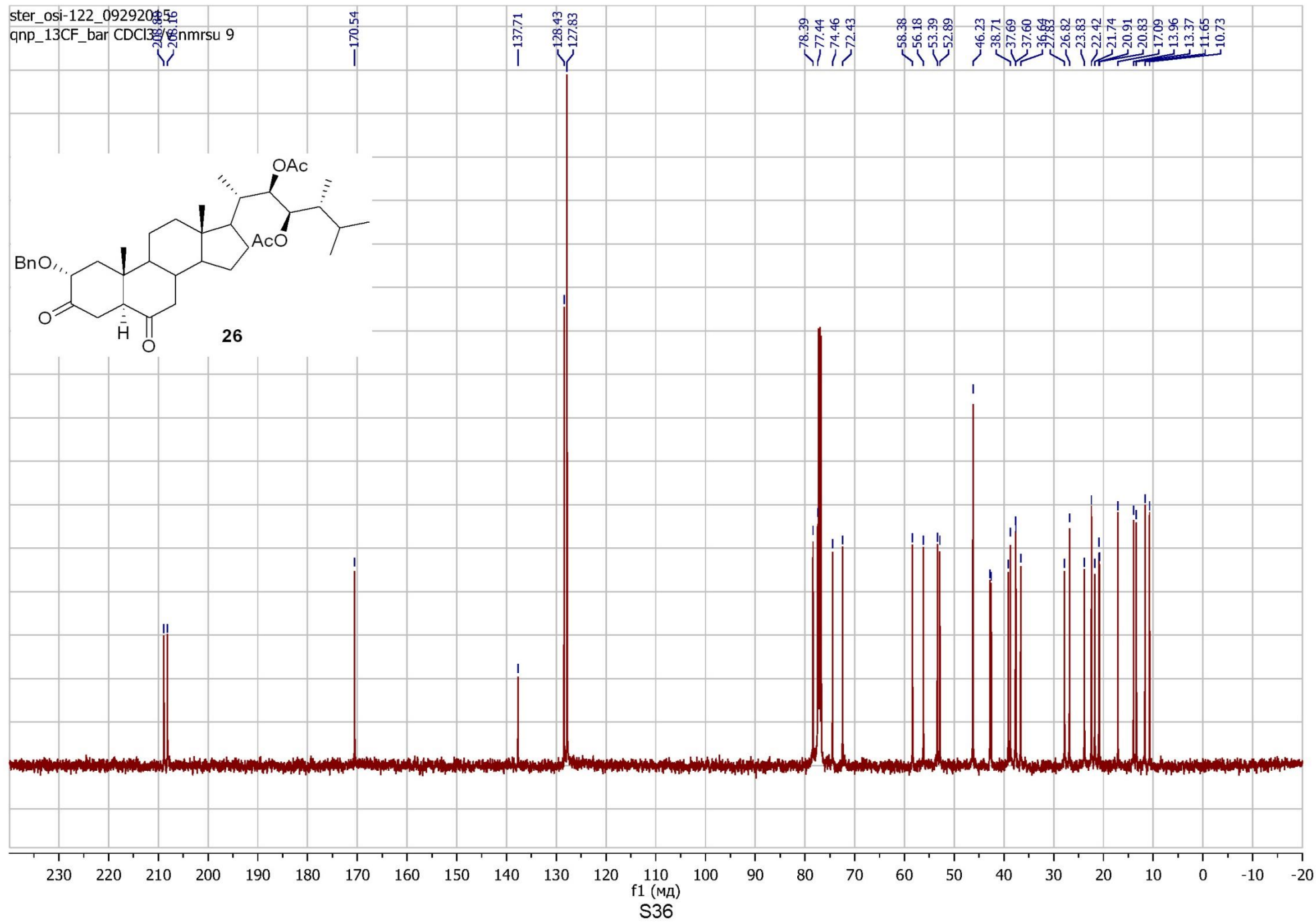
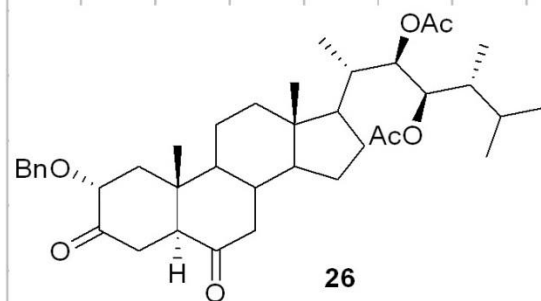


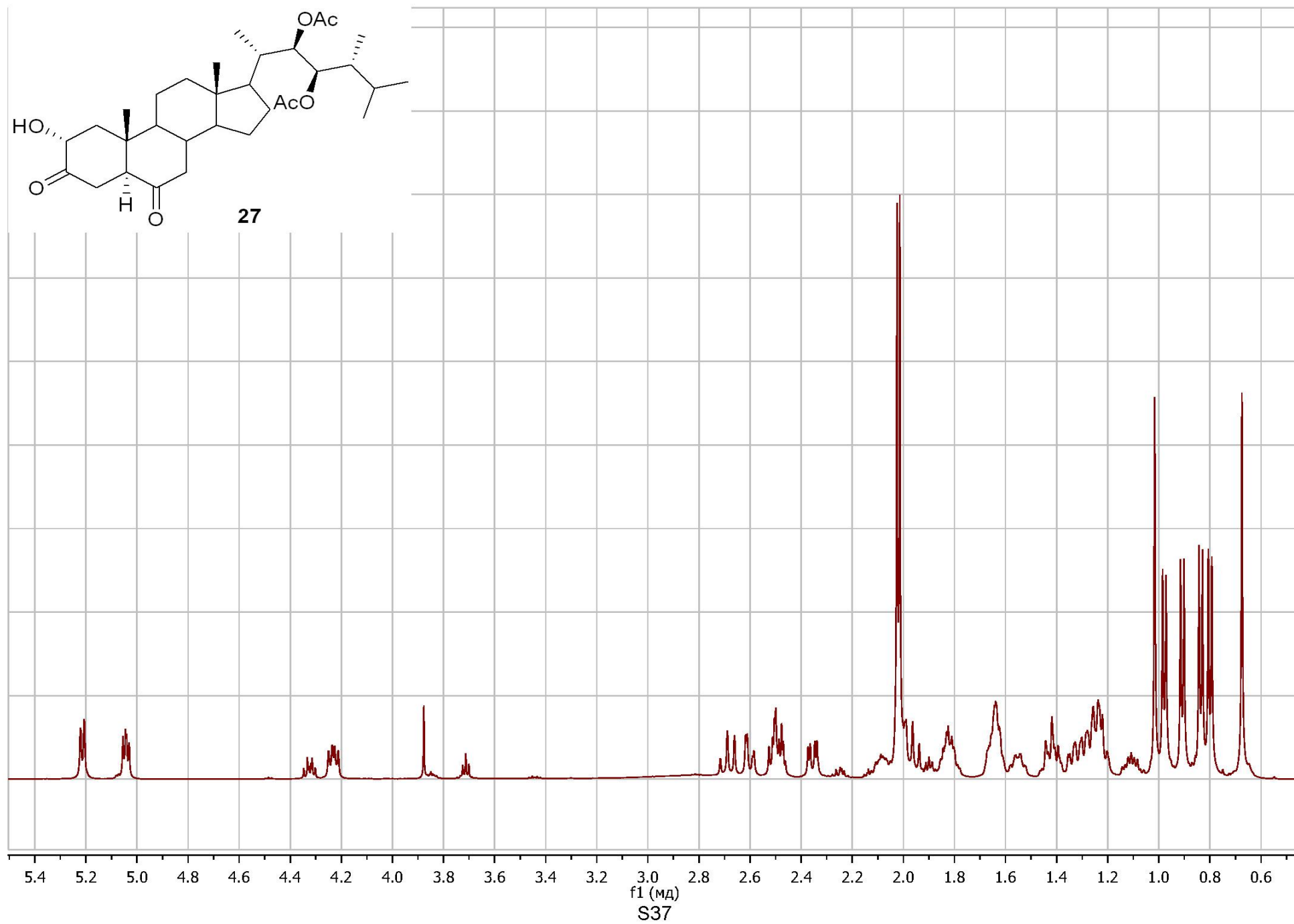
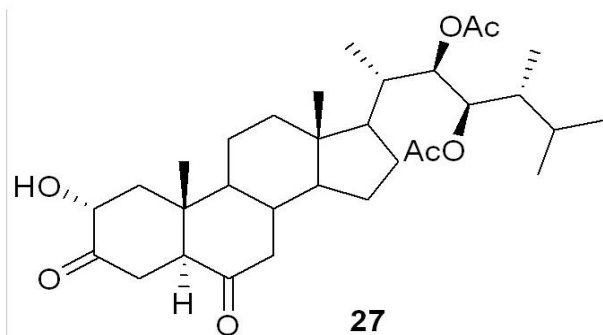




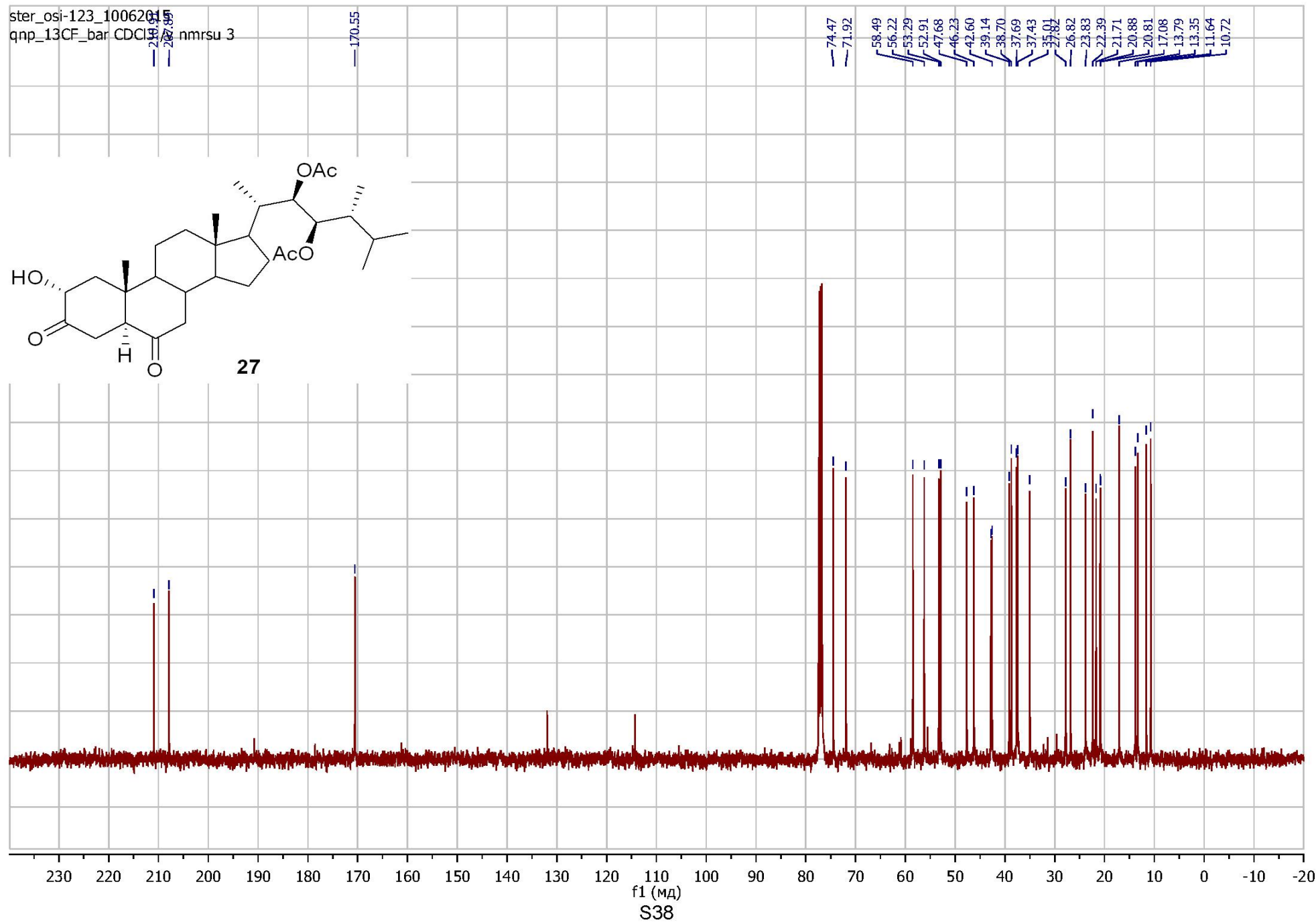
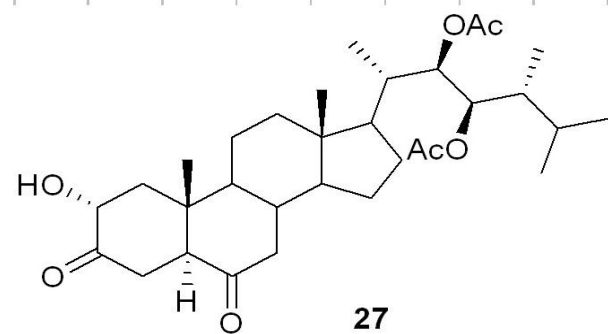


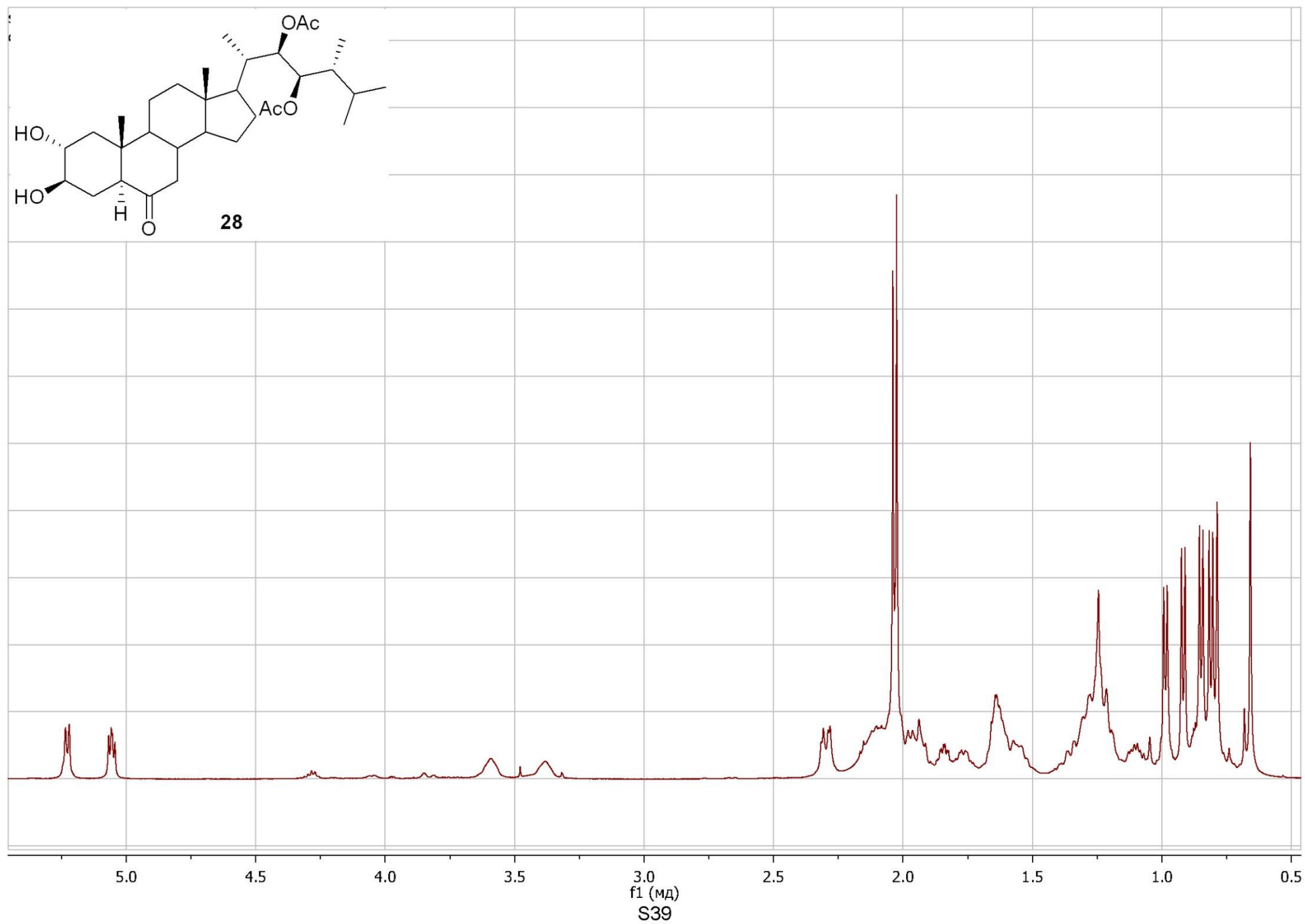
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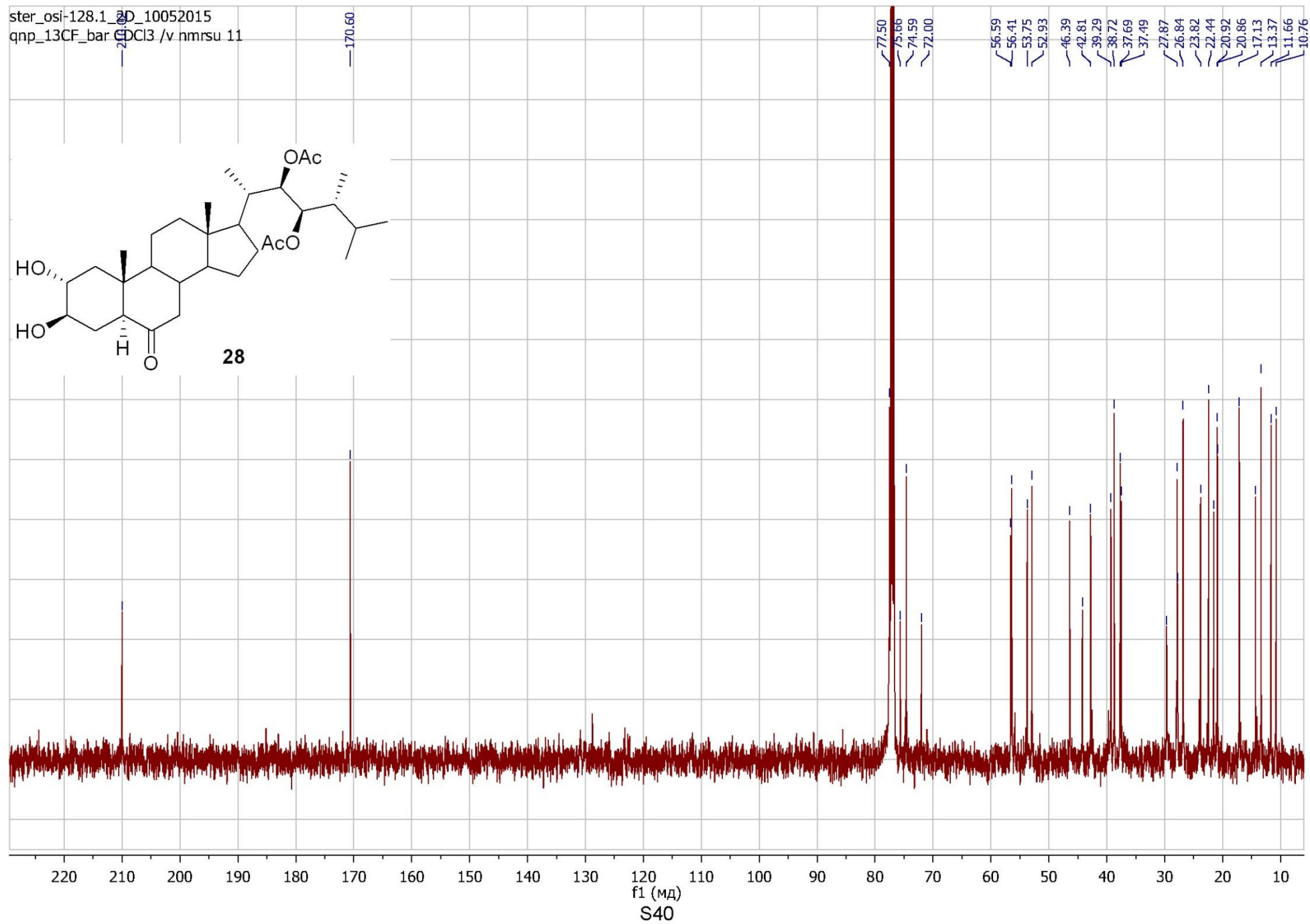
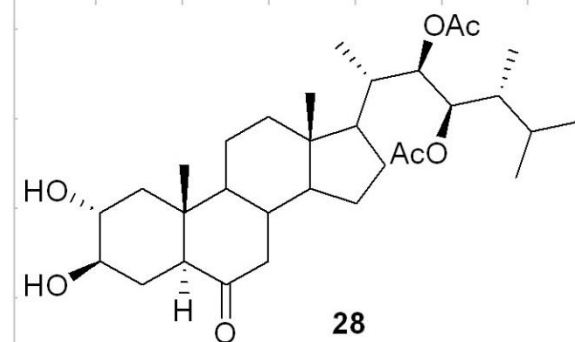


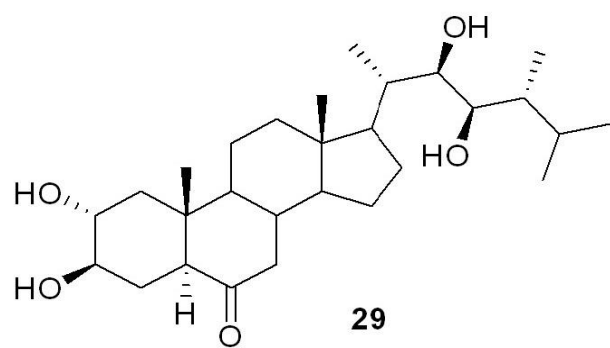
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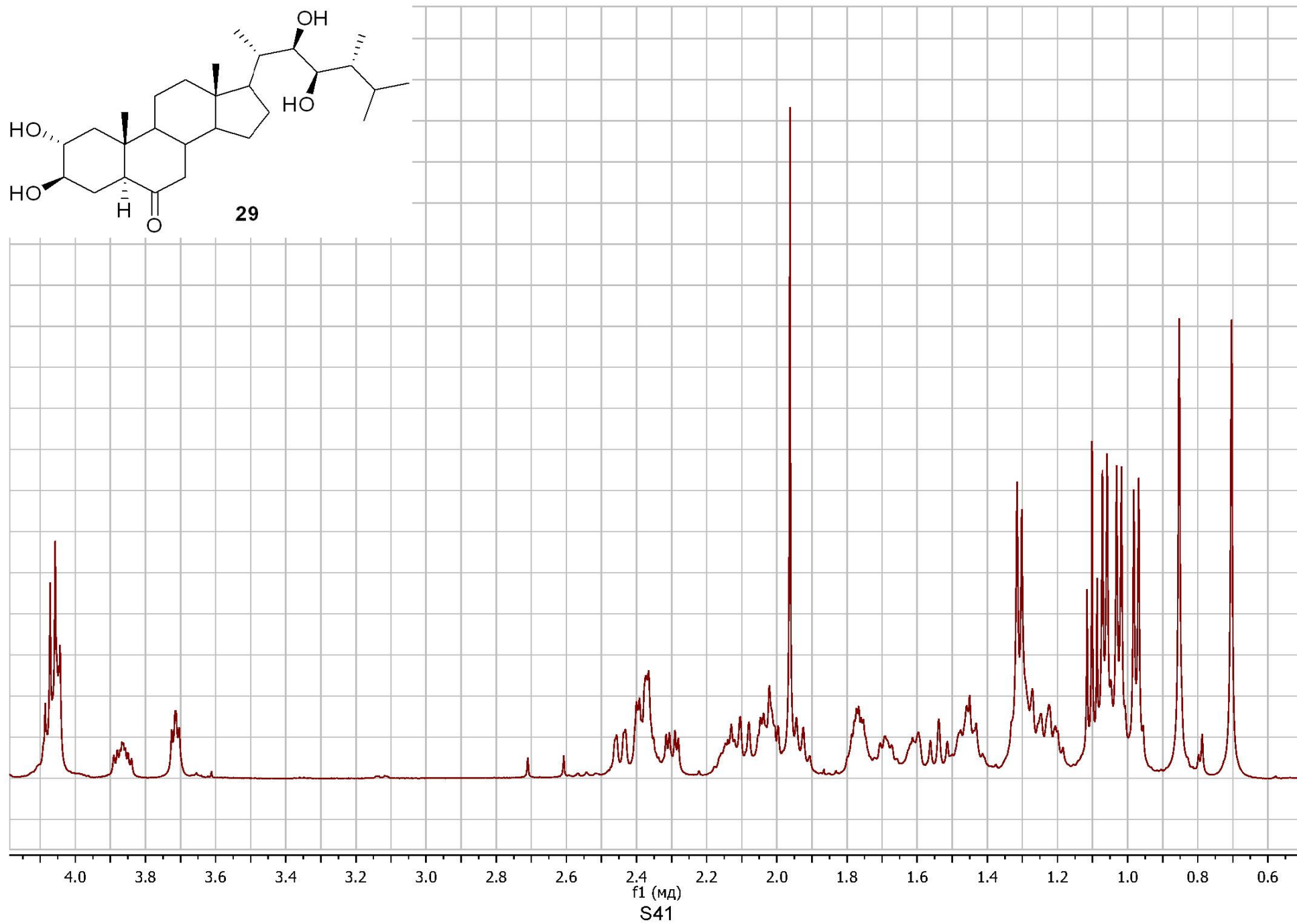


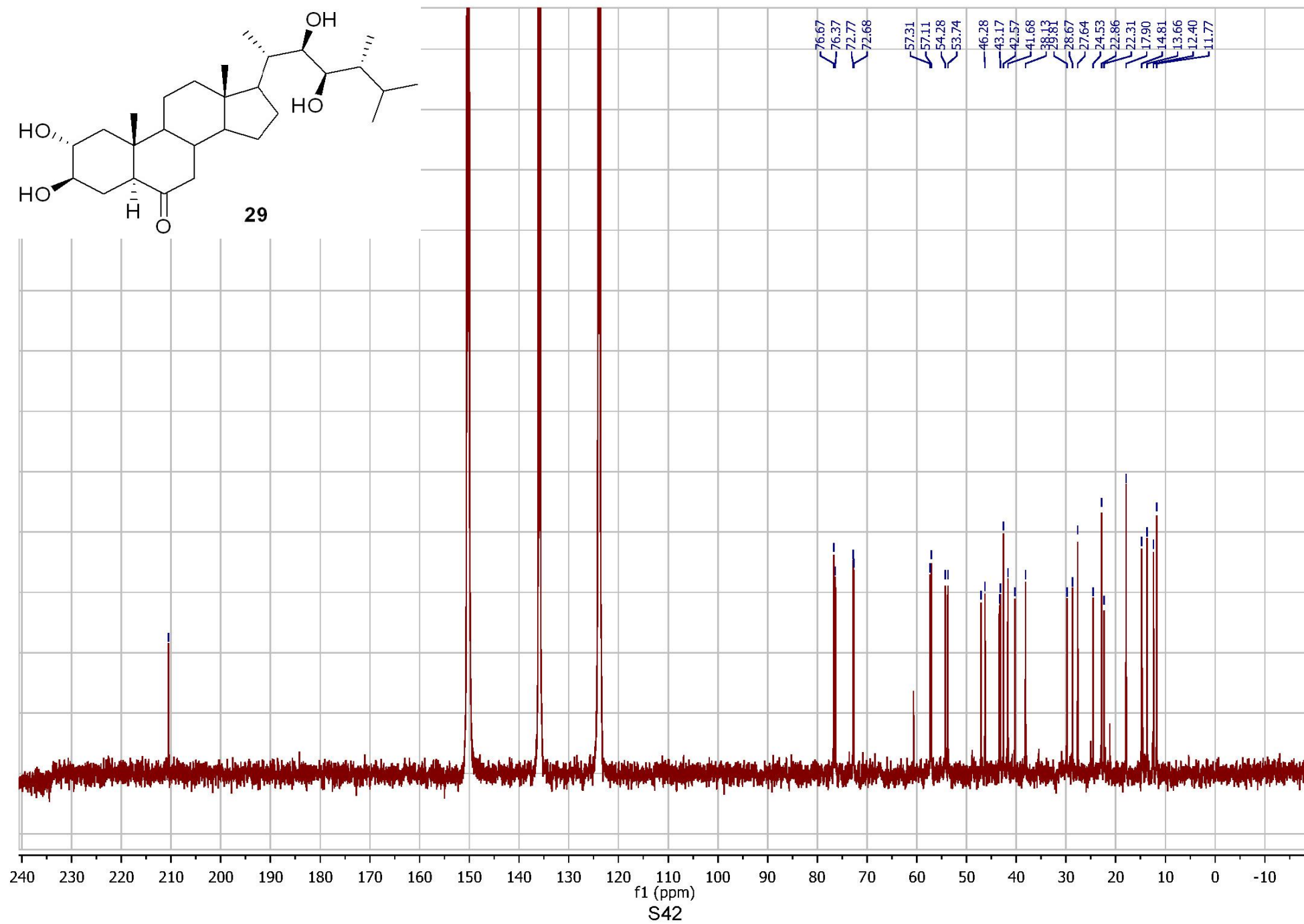
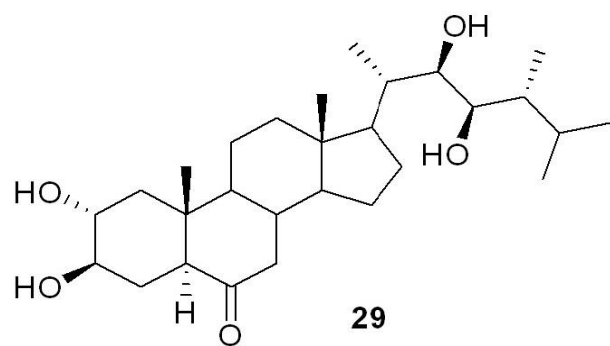
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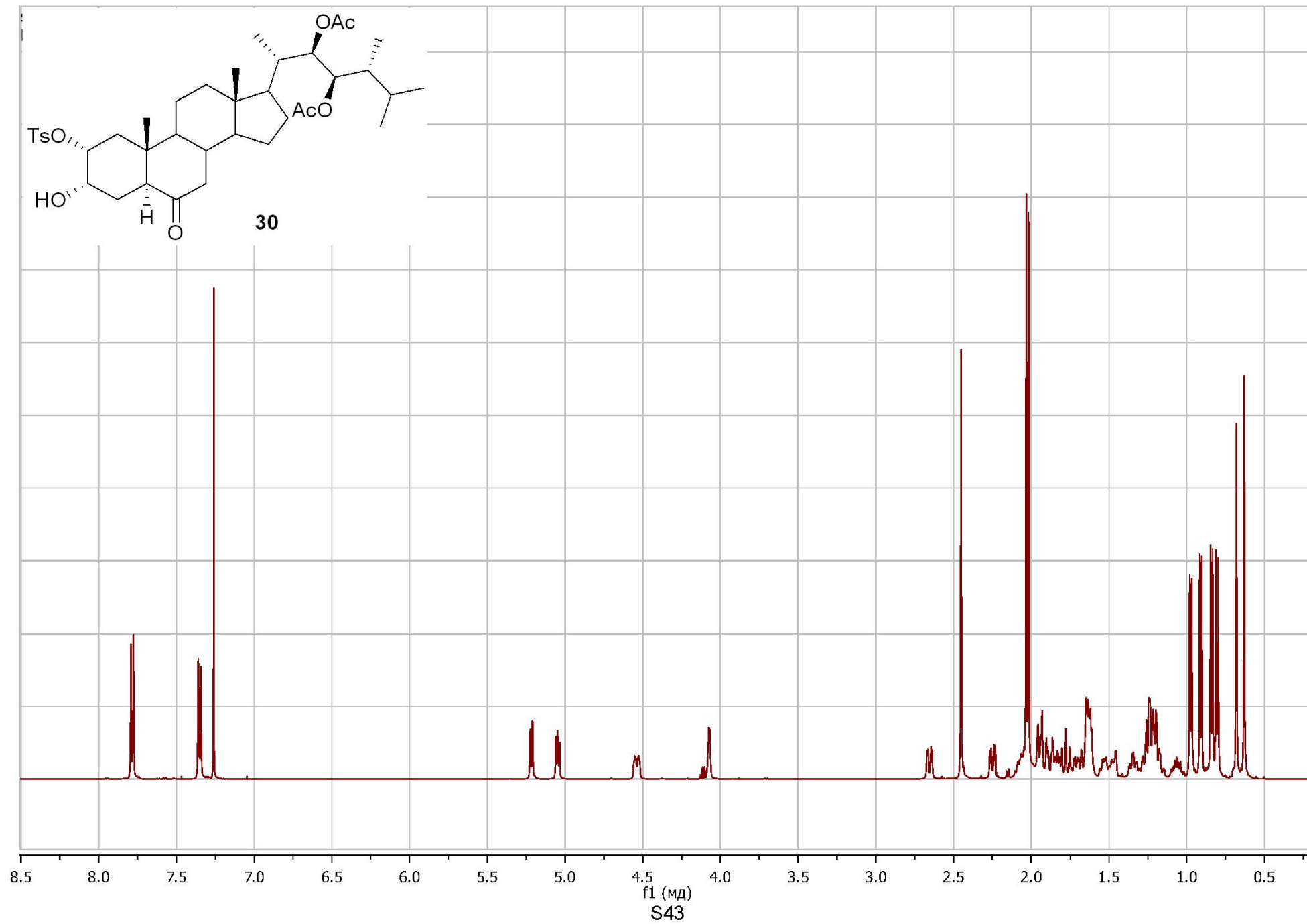
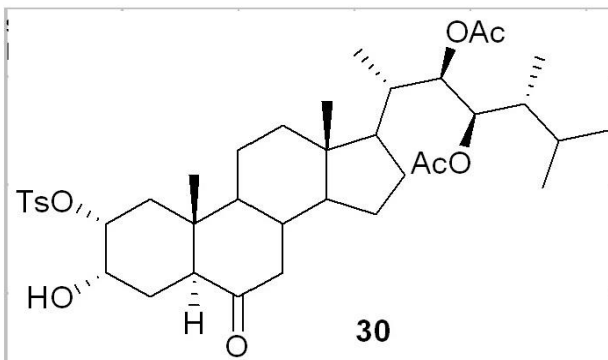


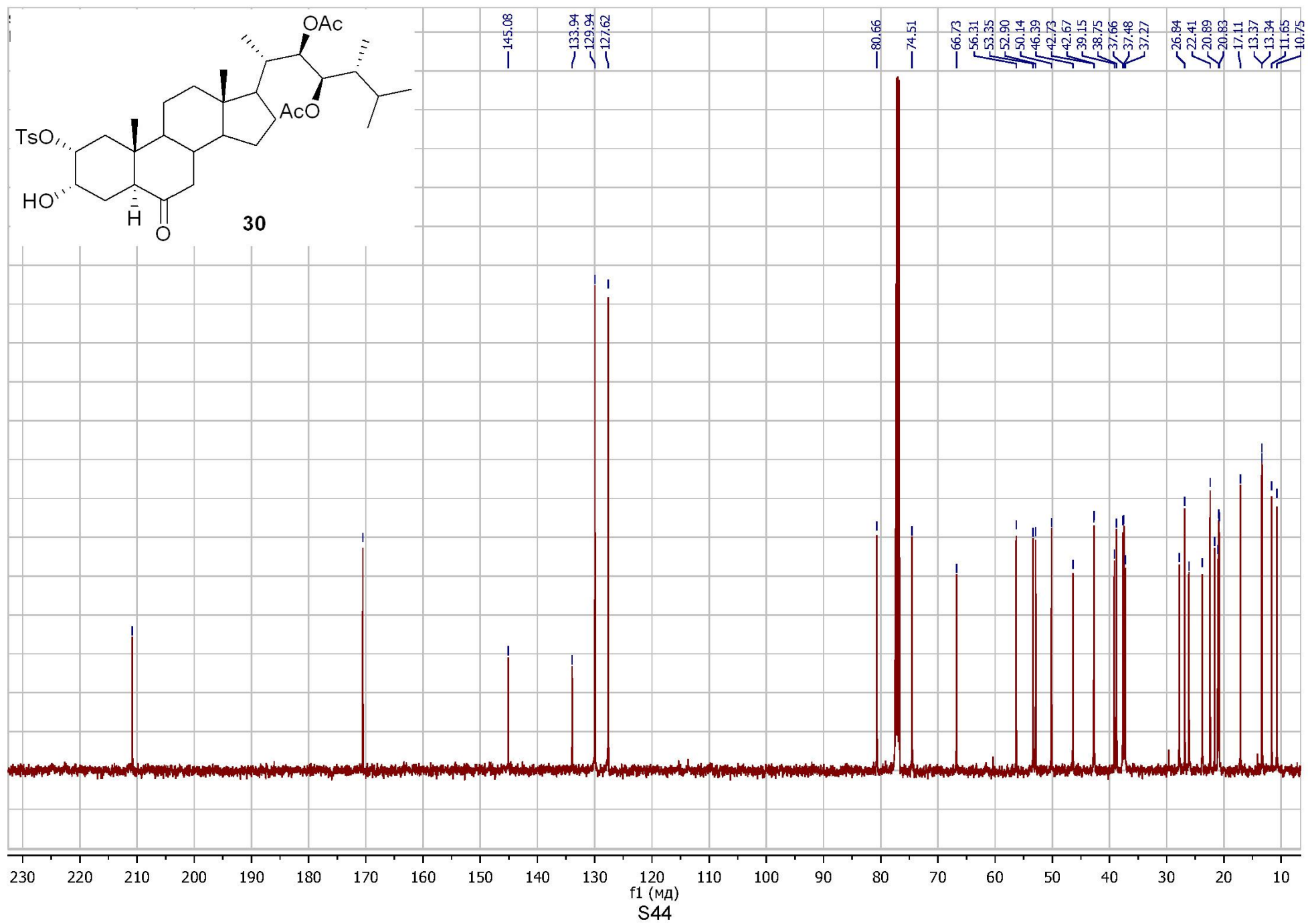


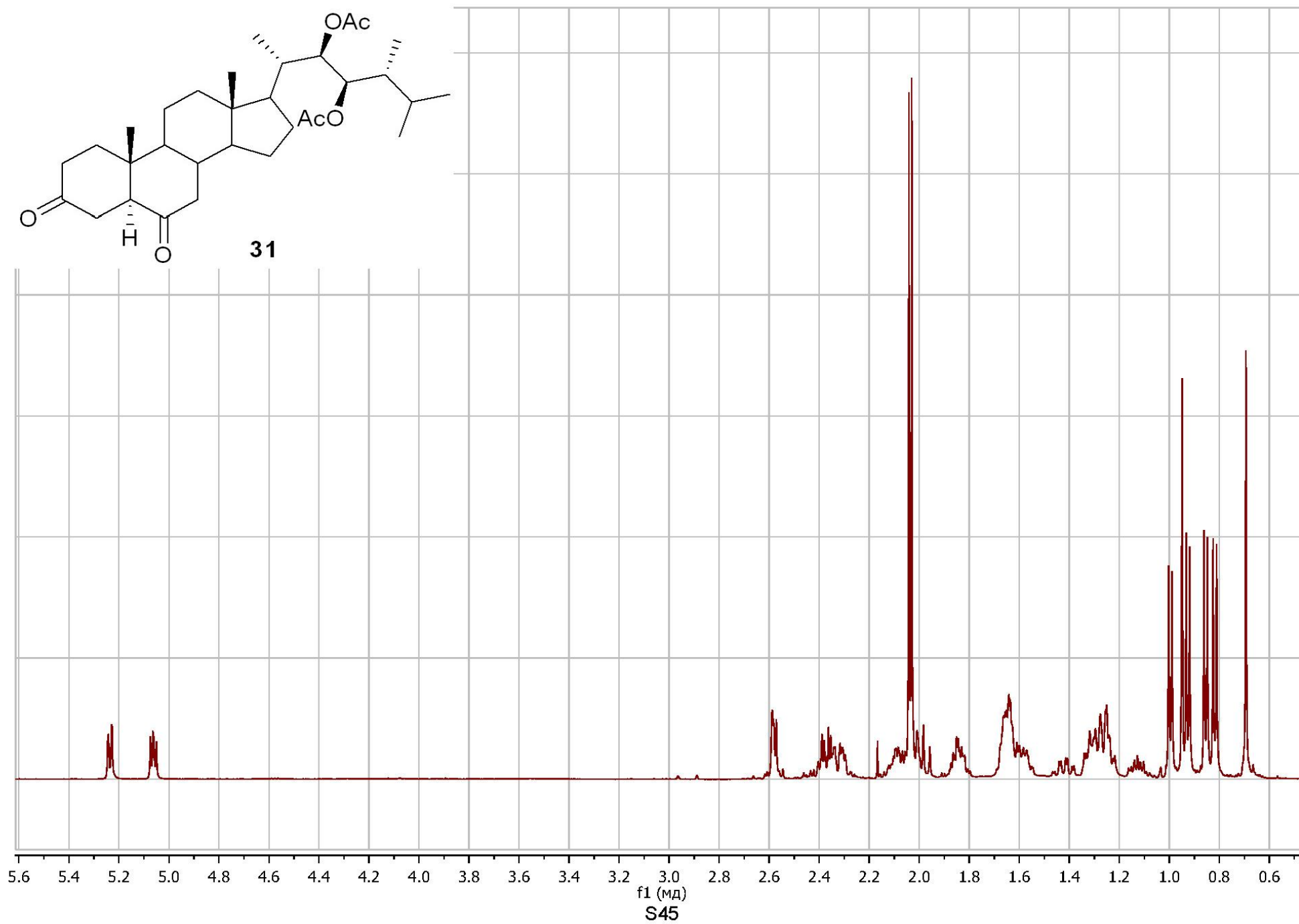
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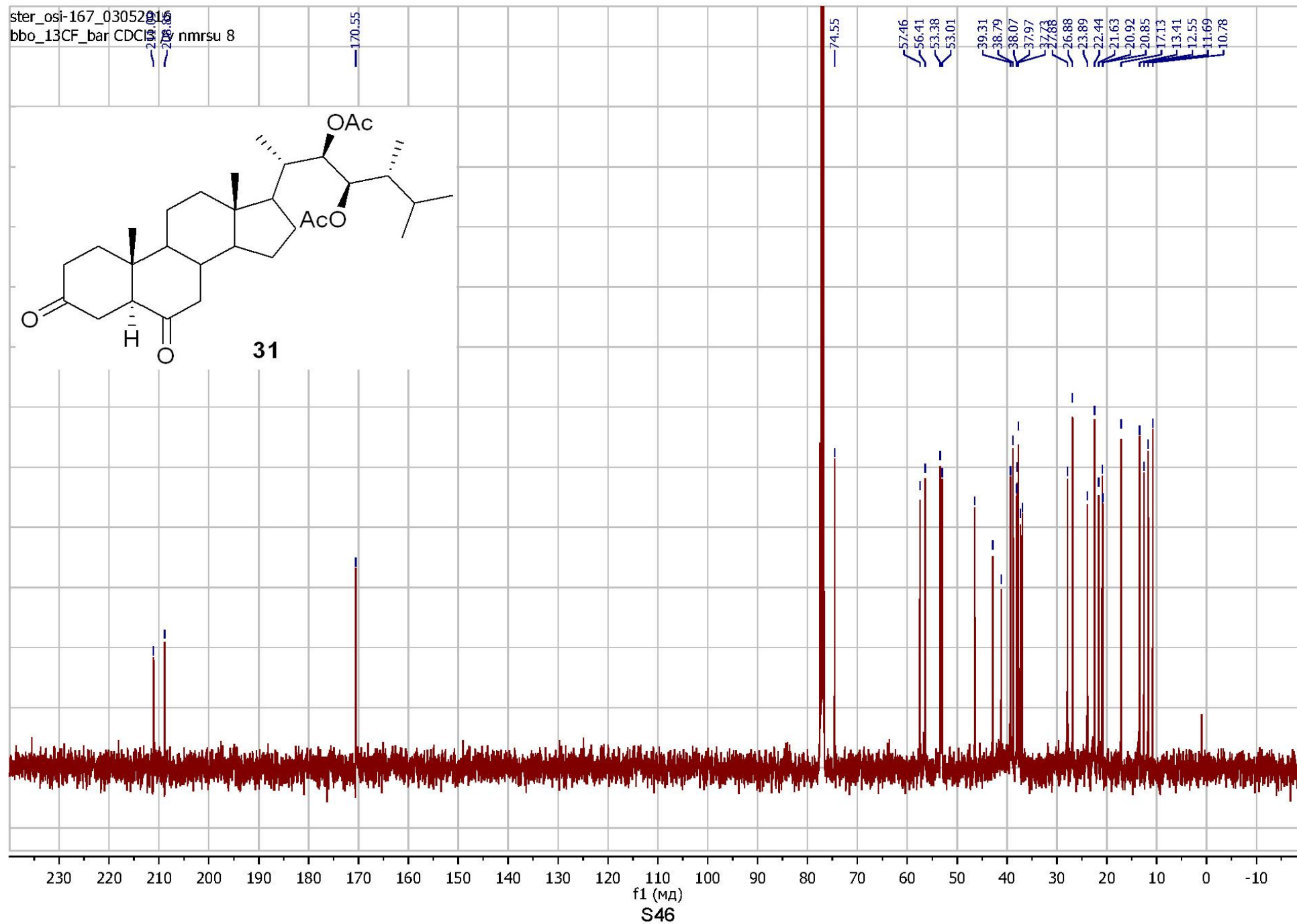
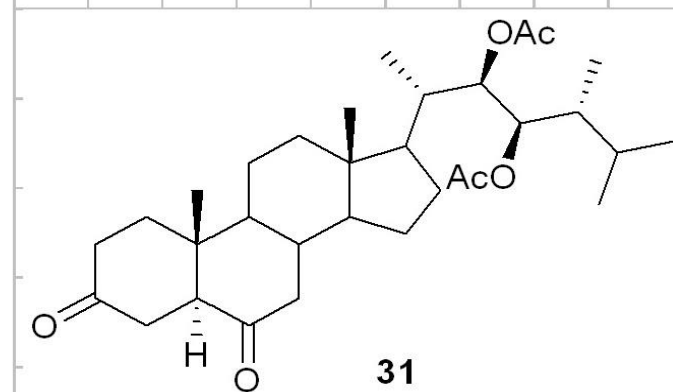


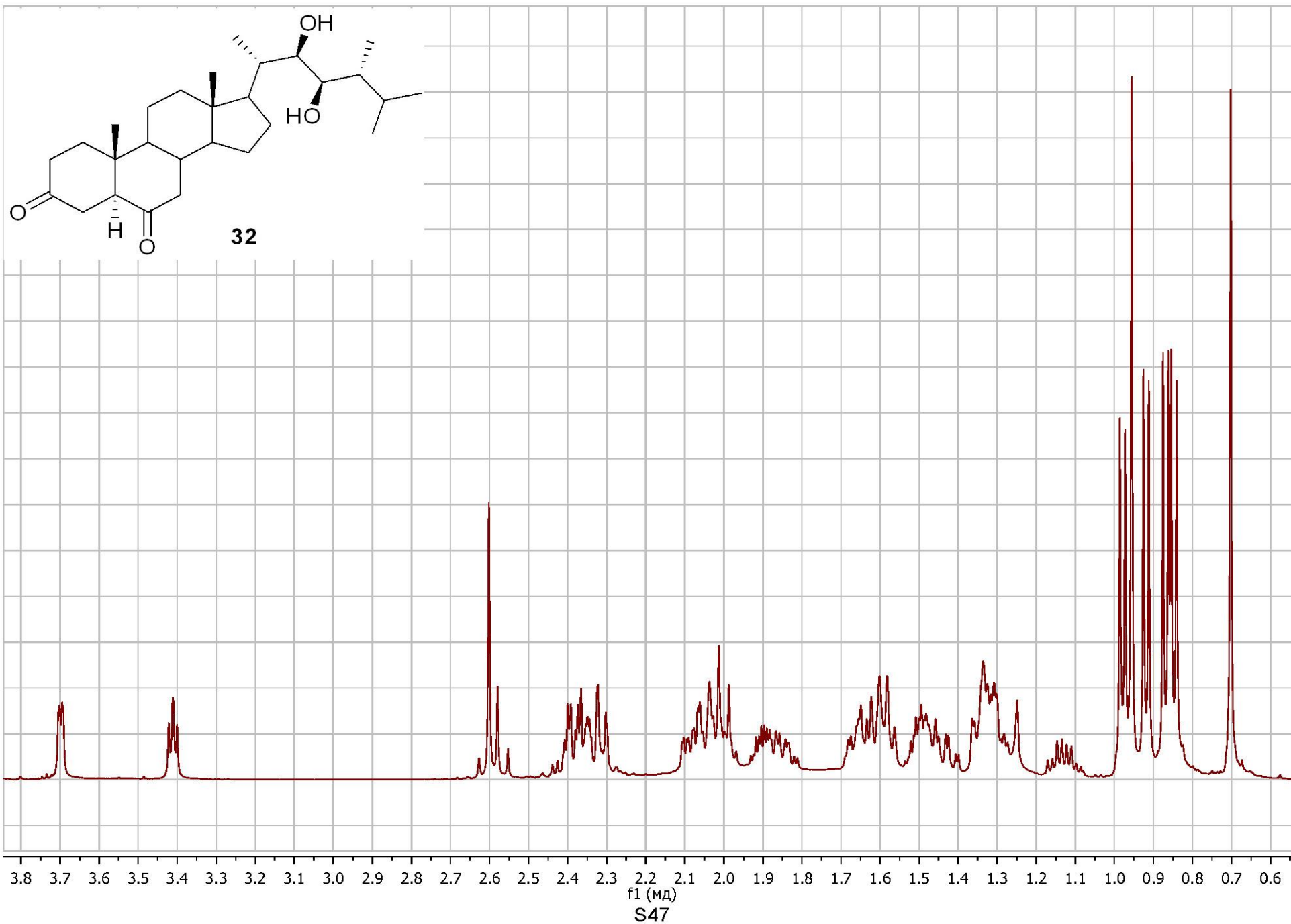




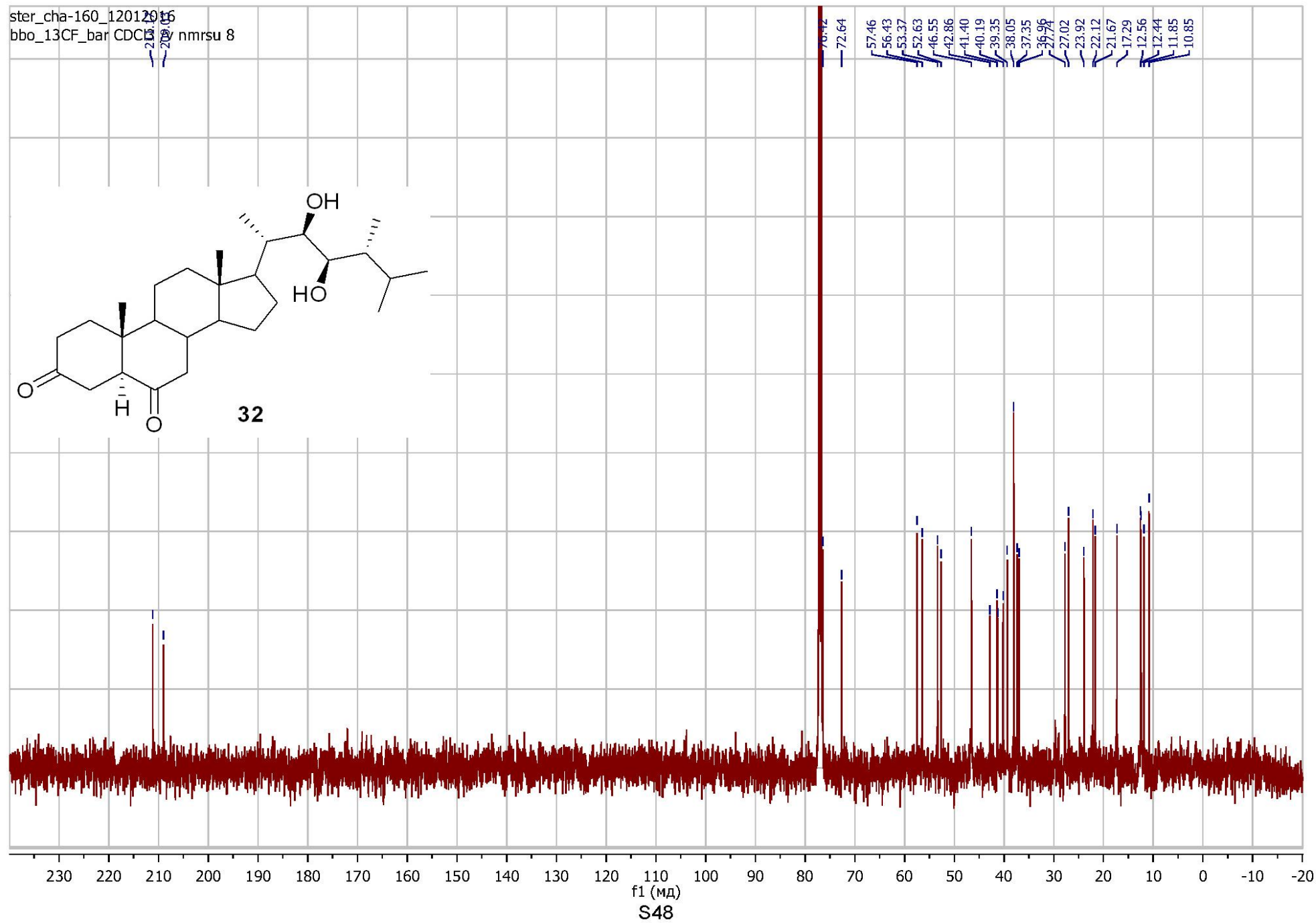
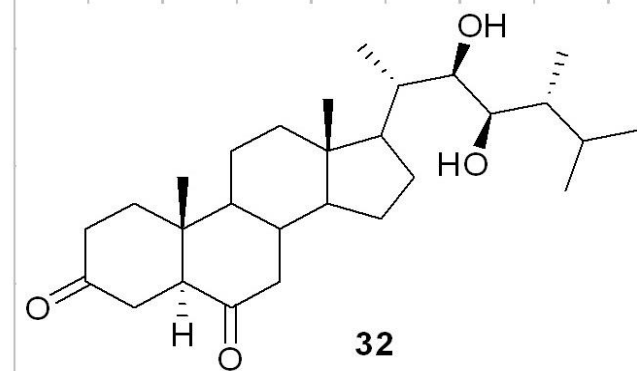


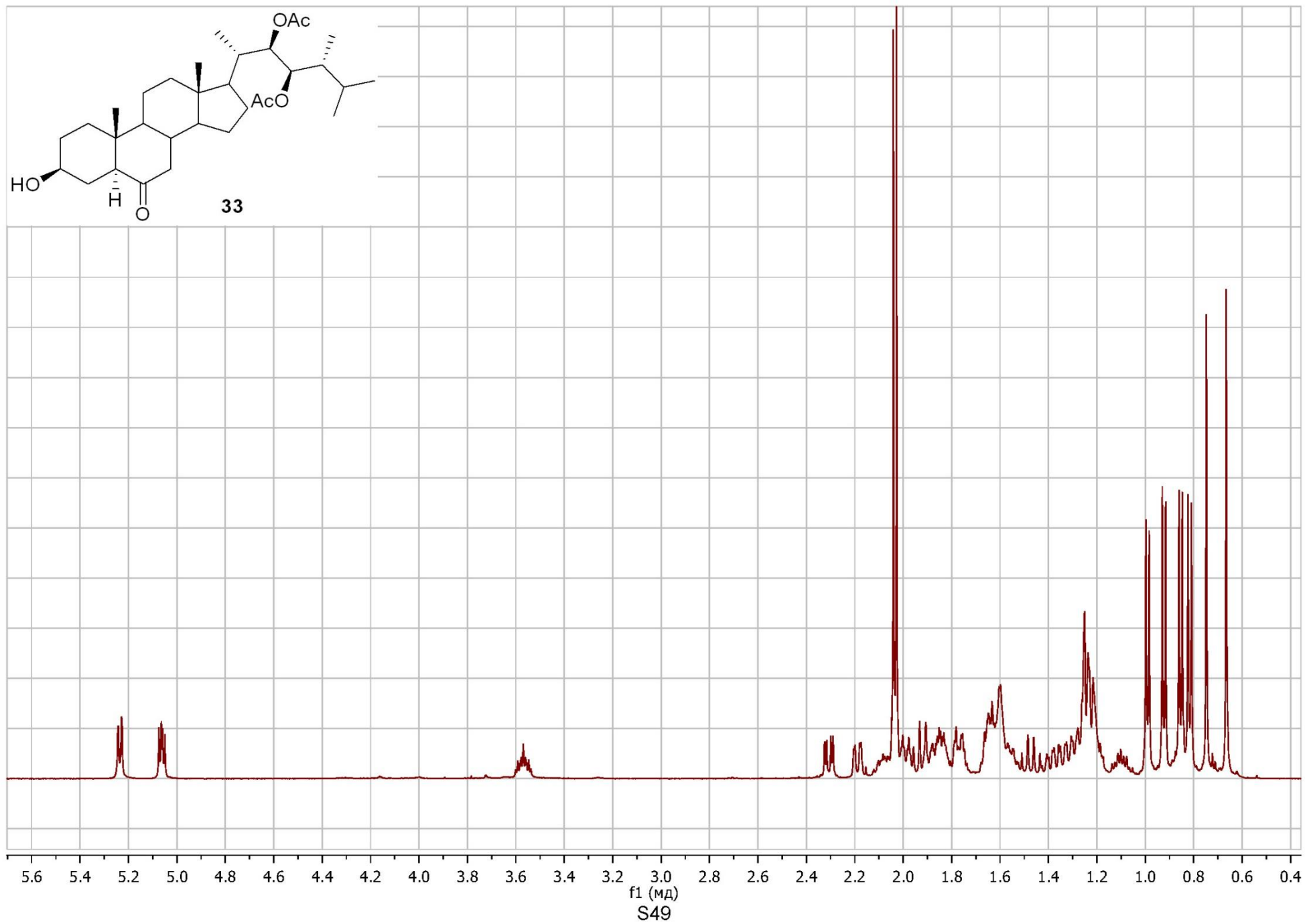
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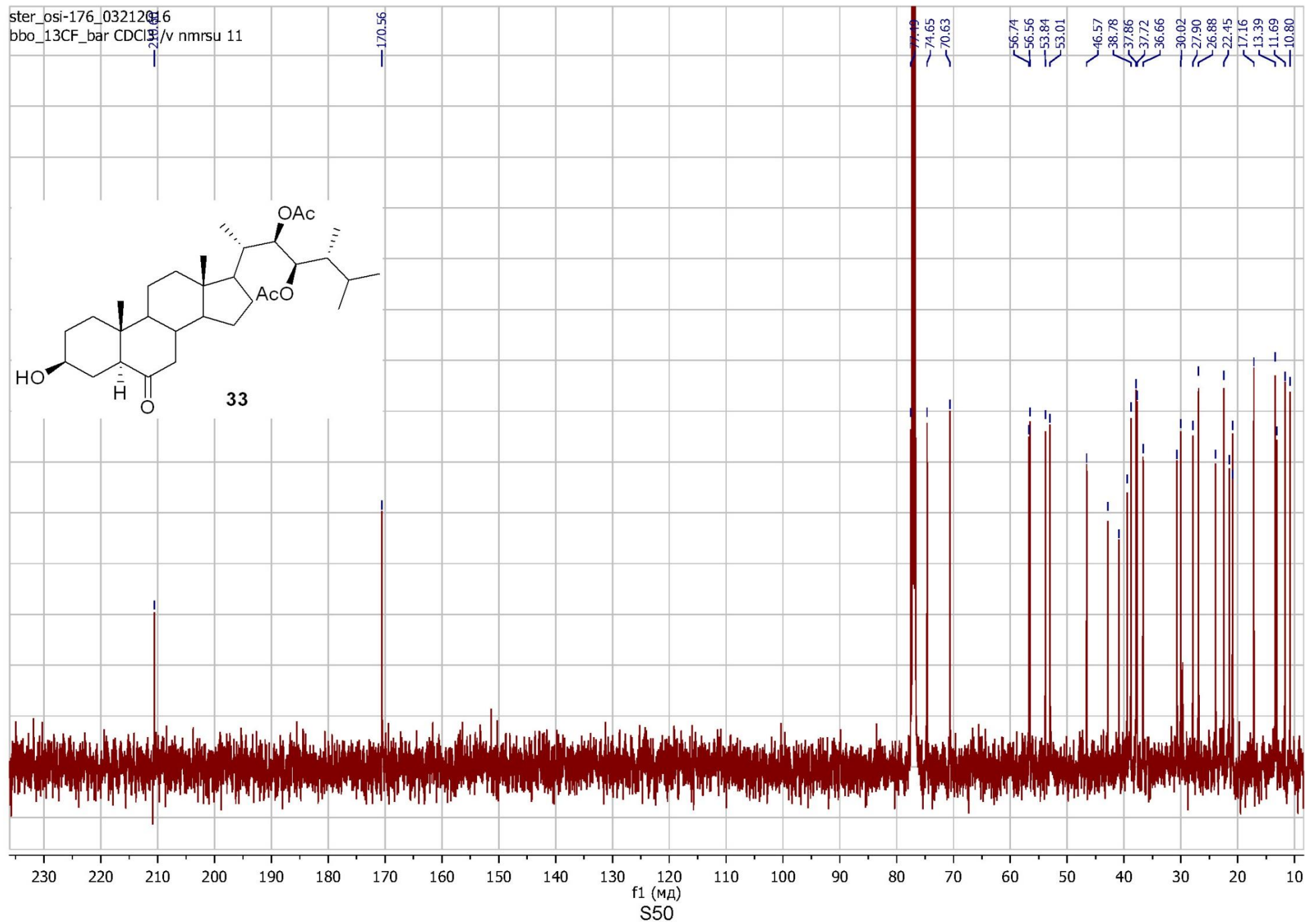
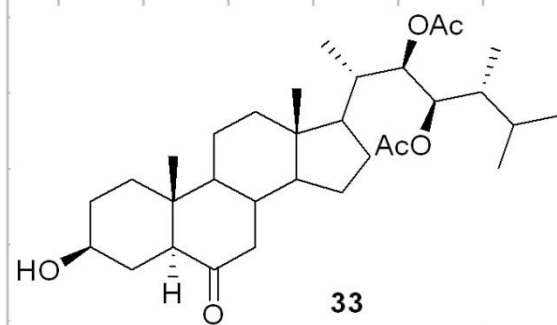


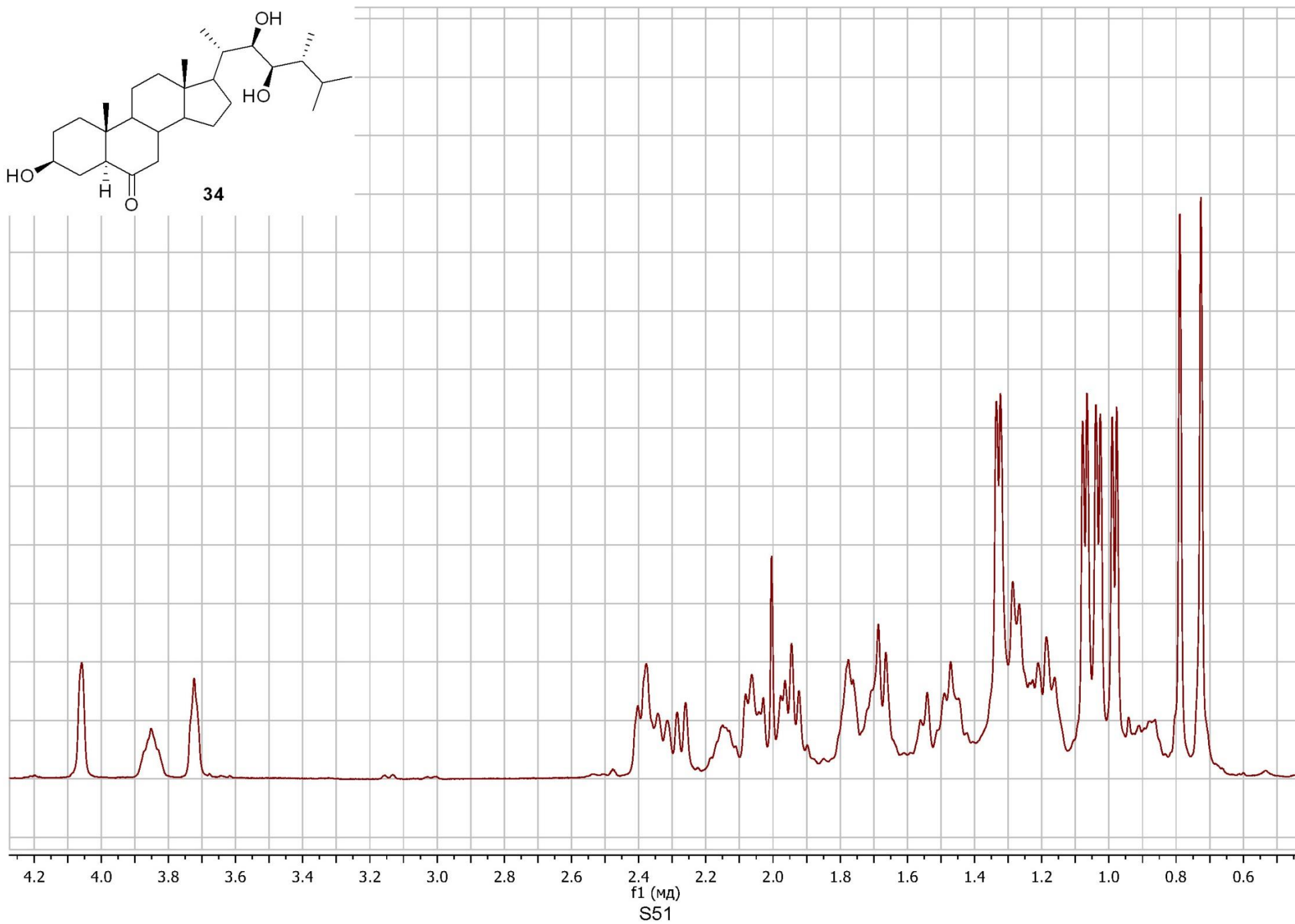
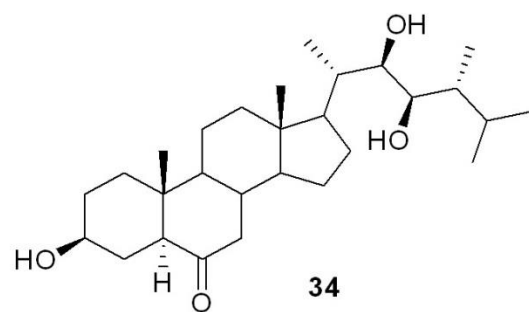
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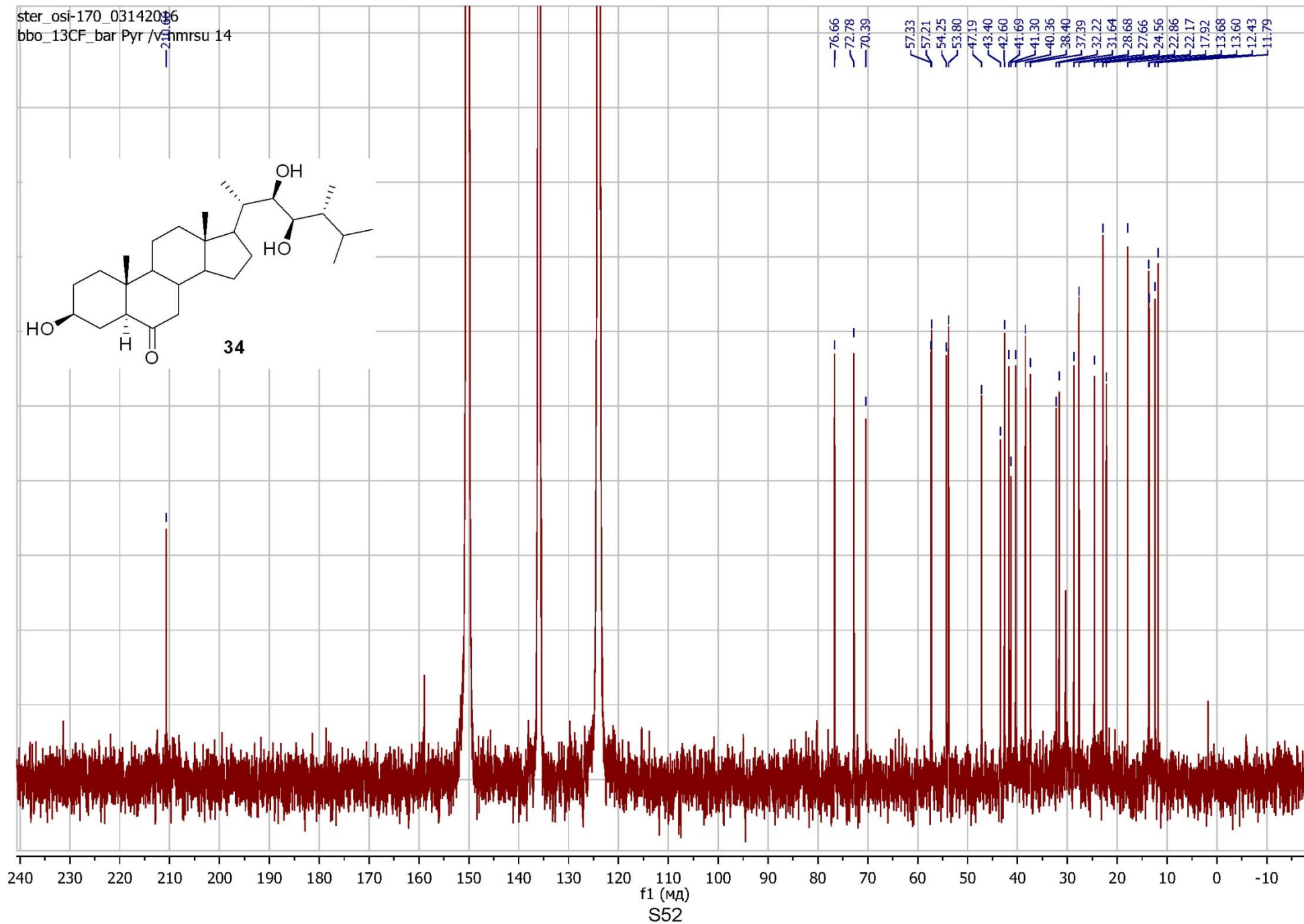
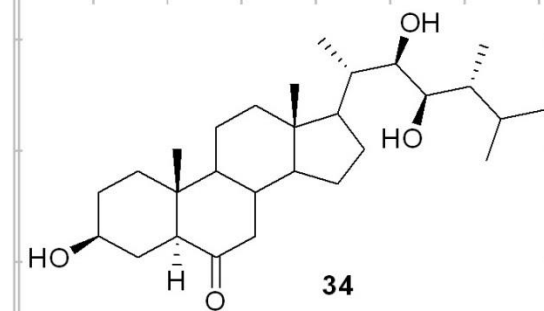


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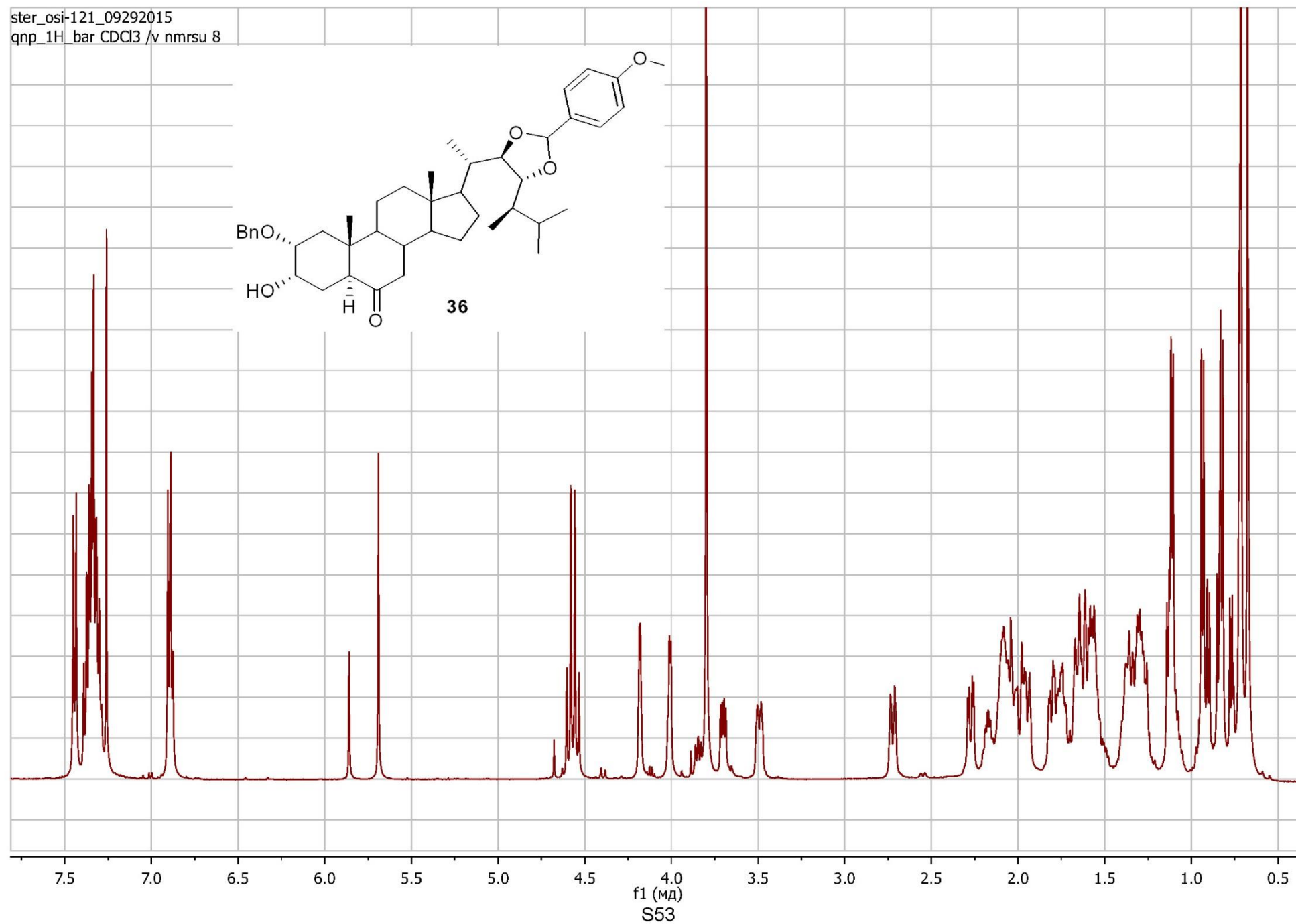
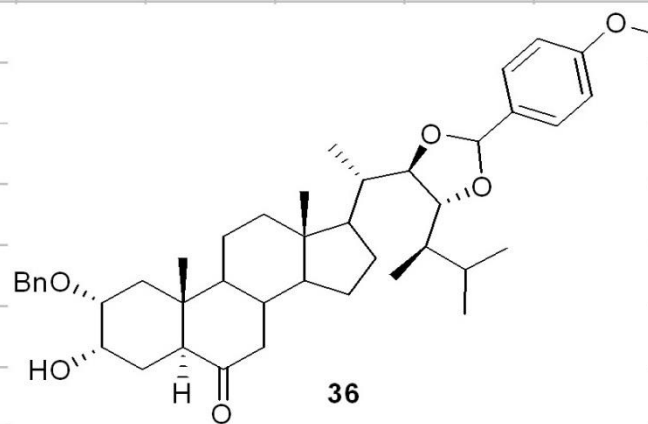




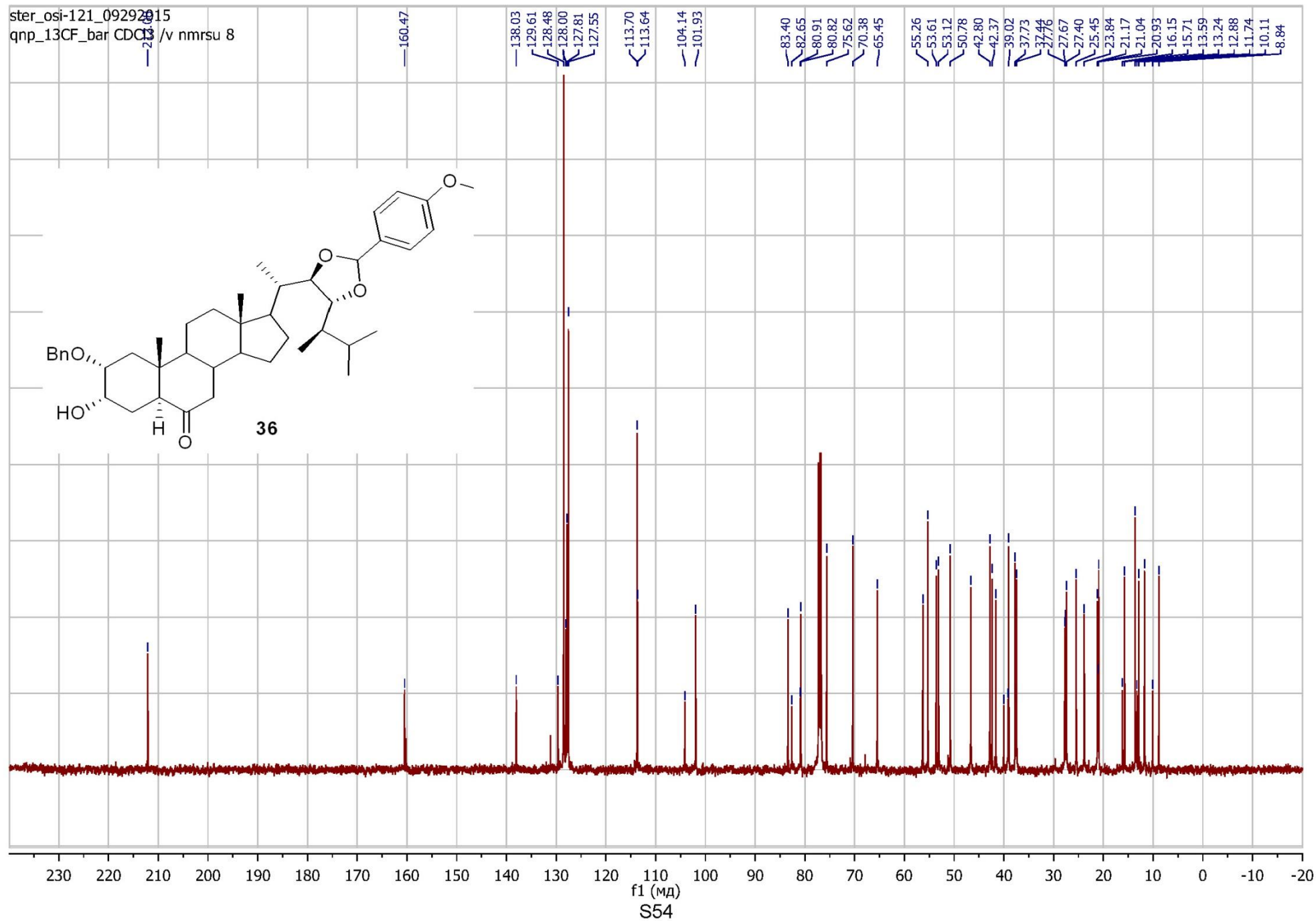
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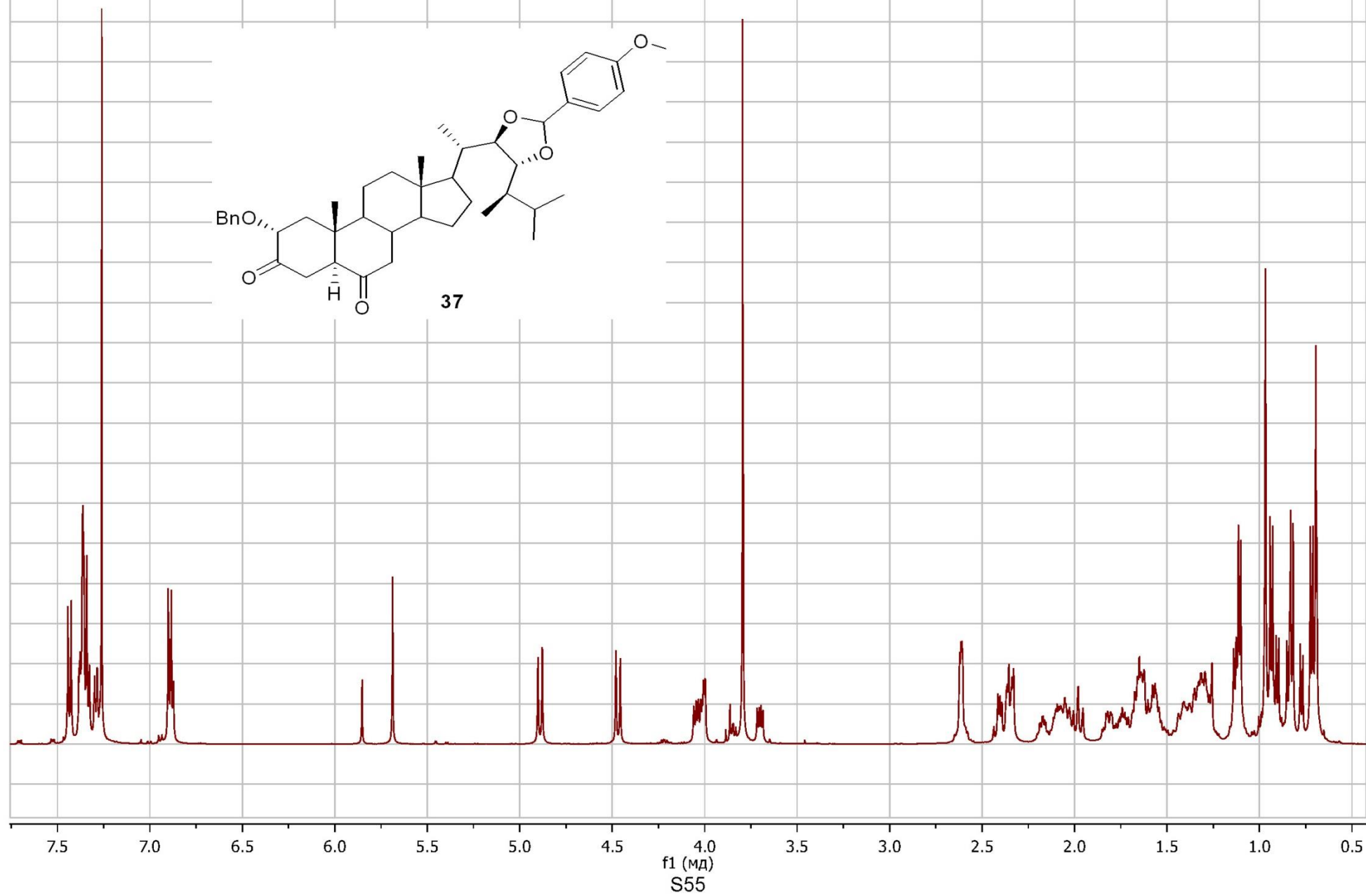
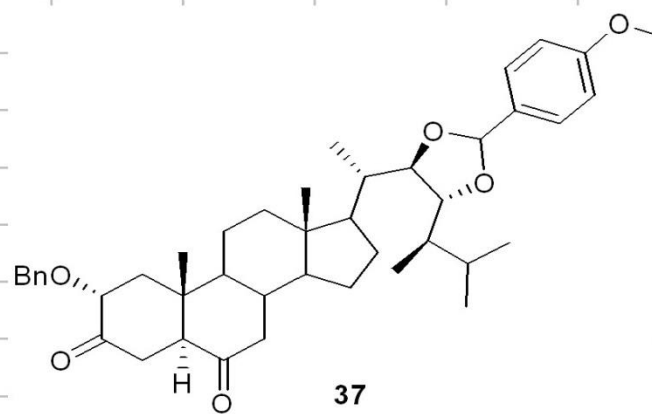
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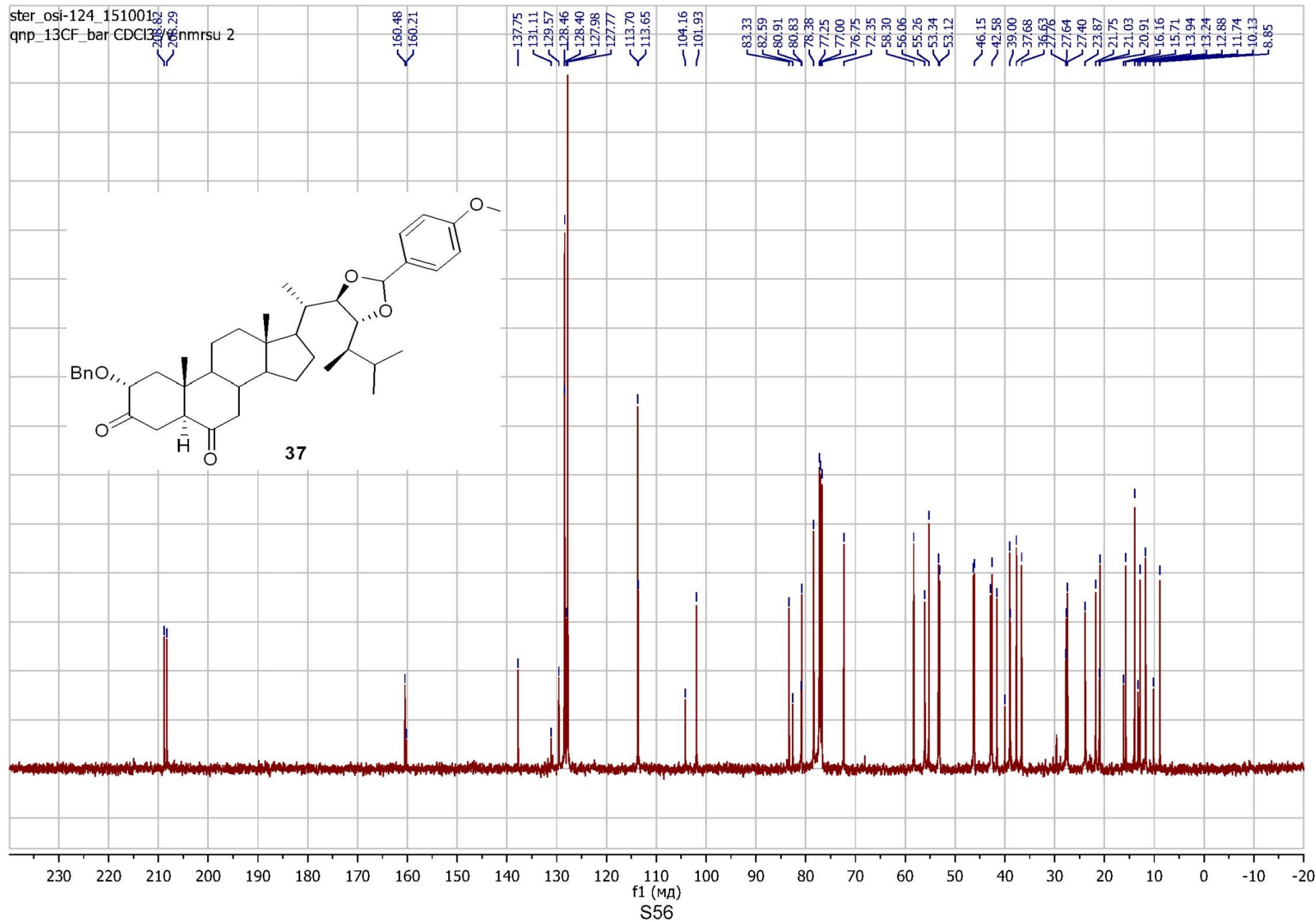
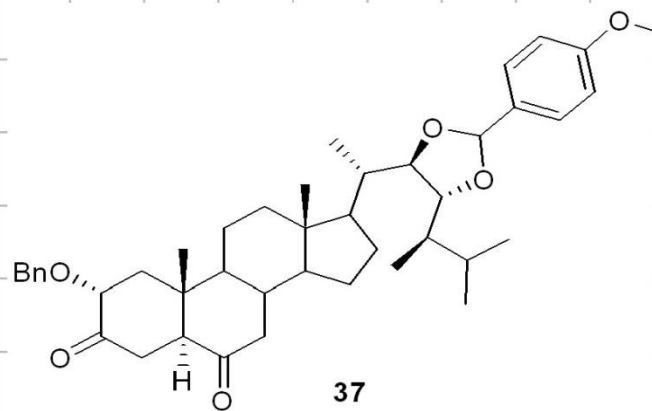
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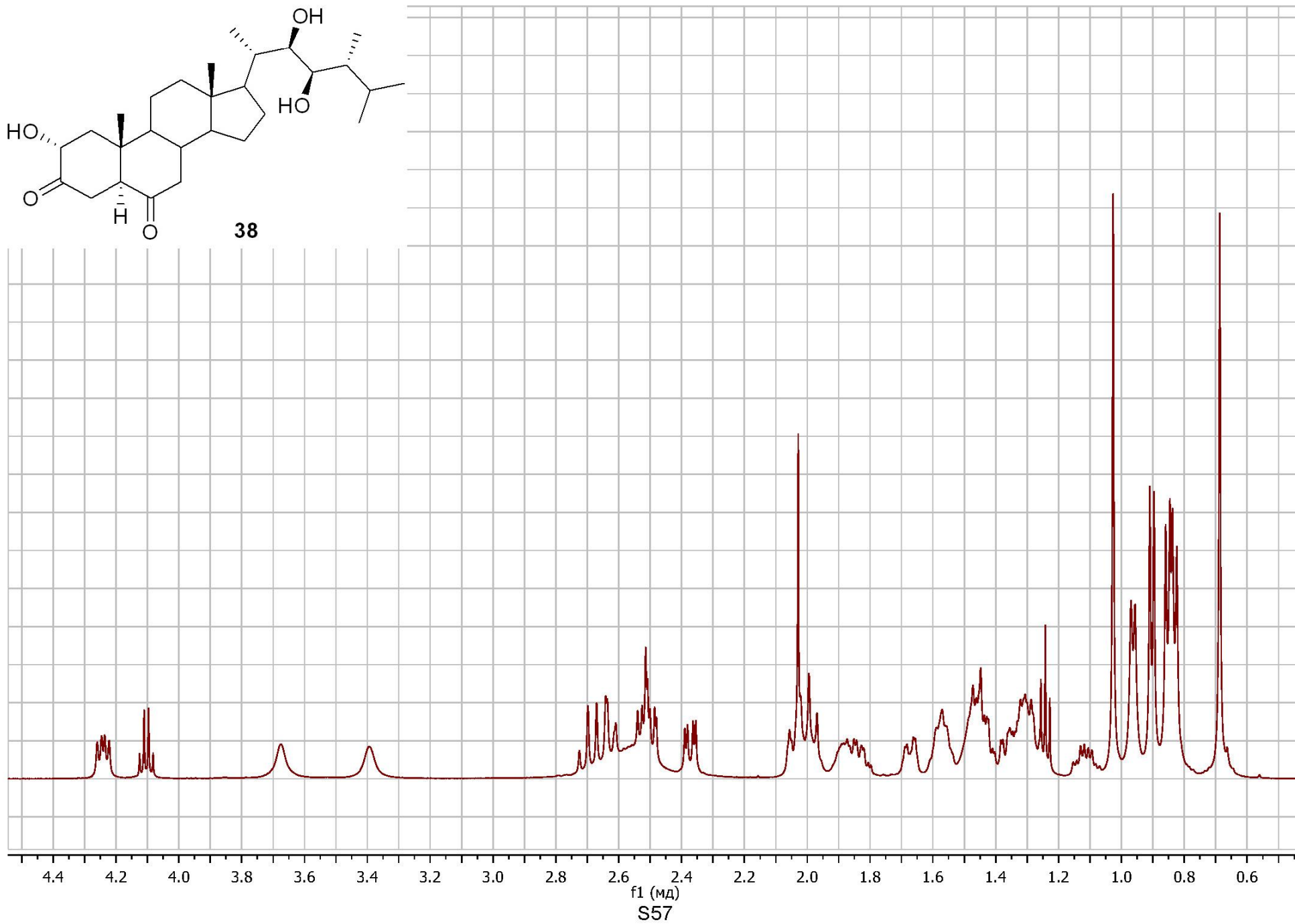
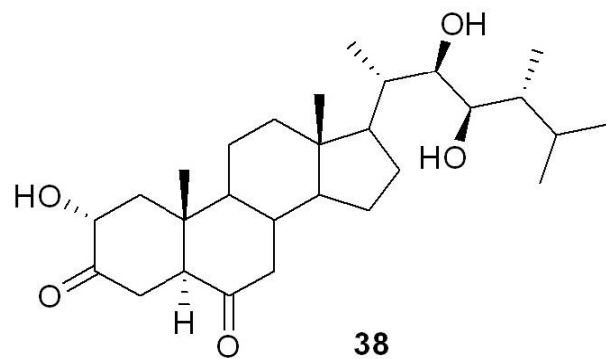


ster_osi-124_151001
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ster_osi-124_1510012
qnp_13CF_bar CDCl₃ nmsu 2





osi-134_151010

QNP 13C for fluoride 0 grad pulse, D1=7 s

