



Supporting Information

for

A golden opportunity: benzofuranone modifications of aurones and their influence on optical properties, toxicity, and potential as dyes

Joza Schmitt and Scott T. Handy

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Experimental methods and spectra for all new compounds

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Instrumentation

The structures of the newly synthesized aurones were determined by NMR (JEOL ECA-500 and ECX-300) and GC–MS (Shimadzu QP2010S). UV-Vis analyses were performed on an HP 8452A Diode Array spectrophotometer. Spectrometry results from MTT assay were analyzed using Molecular Devices SpectraMax M5 Multi-Mode Microplate Reader.

General procedure for the condensation of non-hydroxy-substituted benzofuranones

In a round bottom flask, 1 mmol of the appropriate benzofuranone was mixed with 2 mmol of *p*-tolualdehyde in approximately 3.5 g of neutral alumina and 5 mL of dichloromethane. The reaction was stirred at room temperature for 24 hours. The alumina was filtered out and the mixture concentrated *in vacuo*. Two equivalents of isonicotinic acid hydrazide was added to the concentrated mixture in 5 mL/mmol of both dichloromethane and dimethylformamide and left to stir at room temperature for another 24 hours to remove any excess aldehyde. In some cases, further purification methods were used and/or the ratio of the hydrazide scavenger and the solvents were changed to increase the scavenger's effectiveness after air exposure. Exceptions are noted below. The aurone product was extracted using a mixture of ethyl acetate and 1M HCl except where noted. The organic layer was concentrated and subjected to NMR and GC/MS analysis for purity.

General synthetic procedure for hydroxyl-substituted benzofuranones

In a round bottom flask, 1 mmol of the appropriate benzofuranone was mixed with 2 mmol of *p*-tolualdehyde in 1.5 g/mmol of 1:1 (w:w) KOH/H₂O and 5 mL/mmol methanol. The mixture was left to stir for 6 hours at room temperature. The mixture was triturated with 1 M HCl until acidic and precipitates formed. The crystals were filtered out and washed with 100% hexane followed by 25% EtOAc in hexane. Finally, 50% EtOAc in hexane was used as a final wash. The product was analyzed via NMR and GC/MS.

2-(4-Methylbenzylidene)-1-(benzofuran-3-one) [1]: used 4 equivalents of the hydrazide scavenger and 5 mL/0.5 mmol of DCM and DMF to produce 0.15 g (62%) of the desired product as a pale yellow solid (mp = 97-99 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.29 (s, 3H), 6.76 (s, 1H), 7.08 (t, J=7.4 Hz, 1H), 7.13 (d, J=8.4 Hz, 2H), 7.19 (d, J=8.6 Hz, 1H), 7.51 (t, J=8.6 Hz, 1H), 7.68 (d, J=8.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.67, 112.93, 113.27, 121.74, 123.34, 124.48, 129.53, 129.70 (2C), 131.62 (2C), 136.68, 140.43, 146.47, 165.91, 184.51; GC RT: (24.81 minutes); MS (200 °C, 70 eV): 236 (M⁺, 95.1), 235 (96.3), 221 (100.0), 92 (46.7), 76 (58.6).

2-(4-Methylbenzylidene)-1-(5'-fluorobenzofuran-3-one) [2]: purified via column chromatography using silica gel and 10:90 EtOAc/hexane as eluent followed by extraction in 1M HCl and EtOAc to give 0.15 g (59%) of the product as a pale yellow solid (mp = 122-124 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.41 (s, 3H), 6.91 (s, 1H), 7.26 (d, J=8.2 Hz, 2H), 7.30 (dd, J=8.6, 3.4 Hz, 1H), 7.36 (td, J=8.6, 2.3 Hz, 1H), 7.45 (dd, J=6.3, 2.8 Hz, 1H), 7.80 (d, J=5.8 Hz, 2H); ¹³C NMR (125 MHz,

CDCl₃): δ = 21.69, 109.91 (d, J =95 Hz), 114.07 (d, J =33 Hz), 114.21, 122.39 (d, J =33 Hz), 123.93 (d, J =104 Hz), 129.47 (d, J =248 Hz), 129.77 (2C), 131.74 (2C), 140.85, 147.03, 157.78, 161.90, 183.86; GC RT: (24.00 minutes); MS (200 °C, 70 eV): 254 (M⁺, 47.9), 253 (49.0), 239 (100.0), 126 (53.9), 115 (41.7).

2-(4-Methylbenzylidene)-1-(6'-fluorobenzofuran-3-one) [3]: a new batch of the hydrazide scavenger was reintroduced after column chromatography giving 0.1 g (39%) of the product as a pale yellow solid (mp = 172-174 °C). ¹H NMR (500 MHz, CDCl₃): δ = 2.40 (s, 3H), 6.87 (s, 1H), 6.90 (td, J =14.9, 3.4 Hz, 1H), 7.0 (dd, J =14.3, 3.4 Hz, 1H), 7.25 (d, J =13.2 Hz, 2H), 7.77 (d, J =13.2 Hz, 2H), 7.80 (d, J =13.8 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ = 21.74, 100.77(d, J =104 Hz), 111.86 (d, J =95 Hz), 113.75, 118.47, 126.45 (d, J =48 Hz), 129.55 (d, J =286 Hz), 129.83 (2C), 131.70 (2C), 140.83, 146.89, 167.14 (d, J =33 Hz), 169.25, 182.97; GC RT: (23.88 minutes); MS (200 °C, 70 eV): 254 (M⁺, 56.3), 253 (65.4), 239 (100.0), 126 (61.2), 115 (50.6).

2-(4-Methylbenzylidene)-1-(4'-chlorobenzofuran-3-one) [4]: required 4 equivalents of the hydrazide scavenger and 5 mL/0.5 mmol of DCM and DMF to yield 0.2 g (74%) of the desired product as a pale yellow solid (mp = 172-173 °C). ¹H NMR (500 MHz, CDCl₃): δ = 2.33 (s, 3H), 6.77 (s, 1H), 7.04 (d, J =8.0 Hz, 1H), 7.12 (d, J =8.0 Hz, 1H), 7.15 (d, J =8.0 Hz, 2H), 7.43 (t, J =8.0 Hz, 1H), 7.67 (d, J =8.1 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ = 21.75, 111.39, 113.95, 119.03, 124.56, 129.24, 129.79 (2C), 131.69 (2C), 132.27, 136.76, 140.83,

146.11, 166.43, 181.77; GC RT: (26.50 minutes); MS (200 °C, 70 eV): 272 (M+2, 12.2), 270 (M⁺, 34.2), 269 (40.3), 257 (31.1), 255 (100.0), 134 (58.7), 115 (53.5).

2-(4-Methylbenzylidene)-1-(5'-chlorobenzofuran-3-one) [5]: another 2

equivalents of the hydrazide scavenger were reintroduced in 5 mL/mmol DCM and DMF to yield 0.2 g (72%) of the desired product as a pale yellow solid (mp = 143-146 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.39 (s, 3H), 6.86 (s, 1H), 7.23 (d, J=6.8 Hz, 2H), 7.55 (dd, J=8.6, 2.3 Hz, 1H), 7.71 (d, J=2.3 Hz, 1H), 7.75 (d, J=8.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.76, 114.25, 114.41, 122.91, 124.09, 129.06, 129.17, 129.79 (2C), 131.78 (2C), 136.40, 140.98, 146.57, 164.05, 183.21; GC RT: (25.93 minutes); MS (200 °C, 70 eV): 272 (M+2, 14.2), 270 (M⁺, 43.9), 269 (44.6), 257 (33.8), 255 (100.0), 134 (59.6), 115 (76.4).

2-(4-Methylbenzylidene)-1-(6'-chlorobenzofuran-3-one) [6]: another 2

equivalents of the hydrazide scavenger were reintroduced in 5 mL/mmol DCM and DMF to yield 0.19 g (70%) of the product as a pale yellow solid (mp = 165-166 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.37 (s, 3H), 6.83 (s, 1H), 7.13 (dd, J=8.1, 1.7 Hz, 1H), 7.21 (d, J=8.0 Hz, 2H), 7.29 (s, 1H), 7.66 (d, J=8.2 Hz, 1H), 7.72 (d, J=8.6 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.76, 113.65, 114.23, 120.48, 124.30, 125.38, 129.24, 129.84 (2C), 131.75 (2C), 140.94, 142.72, 146.59, 166.06, 183.21; GC RT: (25.81 minutes); MS (200 °C, 70 eV): 272 (M+2, 15.3), 270 (M⁺, 46.0), 269 (46.3), 257 (31.3), 255 (100.0), 134 (63.5), 115 (79.4).

2-(4-Methylbenzylidene)-1-(7'-chlorobenzofuran-3-one) [7]: another 2 equivalents of the hydrazide scavenger were reintroduced in 5 mL/mmol DCM and DMF to yield 0.14 g (50%) of the product as a pale yellow solid (mp = 172-174 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.37 (s, 3H), 6.90 (s, 1H), 7.11 (t, *J*=7.7 Hz, 1H), 7.23 (d, *J*=8.0 Hz, 2H), 7.59 (d, *J*=7.4 Hz, 1H), 7.65 (d, *J*=7.4 Hz, 1H), 7.80 (d, *J*=8.1 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.80, 115.01, 118.62, 122.86, 123.65, 124.14, 129.17, 129.93 (2C), 132.02 (2C), 136.34, 141.16, 146.26, 161.53, 183.79; GC RT: (25.54 minutes); MS (200 °C, 70 eV): 272 (M+2, 16.6), 270 (M⁺, 47.8), 269 (45.0), 257 (33.5), 255 (100.0), 134 (63.7), 115 (81.1).

2-(4-Methylbenzylidene)-1-(5'-bromobenzofuran-3-one) [8]: another 2 equivalents of the hydrazide scavenger were reintroduced in 5 mL/mmol DCM and DMF to yield 0.15 g (42%) of the desired product as a pale yellow solid (mp = 154-155 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.41 (s, 3H), 6.90 (s, 1H), 7.23 (d, *J*=8.6 Hz, 1H), 7.25 (d, *J*=8.2 Hz, 2H), 7.71 (dd, *J*=8.6, 1.7 Hz, 1H), 7.78 (d, *J*=8.6 Hz, 2H), 7.90 (d, *J*=2.3 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.63, 114.42, 114.57, 116.14, 123.36, 127.14, 129.04, 129.69 (2C), 131.66 (2C), 139.07, 140.91, 146.29, 164.41, 183.03; GC RT: (26.89 minutes); MS (200 °C, 70 eV): 316 ([M+2-1]⁺, 31.8), 315 (M⁺, 34.5), 314 ([M-1]⁺, 32.4), 301 (69.9), 299 (74.2), 115 (100.0), 91 (14.8).

2-(4-Methylbenzylidene)-1-(6'-bromobenzofuran-3-one) [9]: scavenger was reintroduced three times to produce 0.26 g (83%) of the desired product as a

pale yellow solid (mp = 157-158 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.37 (s, 3H), 6.84 (s, 1H), 7.20 (d, *J*=8.1 Hz, 2H), 7.29 (d, *J*=8.0 Hz, 1H), 7.47 (s, 1H), 7.59 (d, *J*=8.0 Hz, 1H), 7.72 (d, *J*=8.2 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.79, 114.30, 116.64, 120.86, 125.47, 127.11, 129.25, 129.85 (2C), 131.15, 131.77 (2C), 140.96, 146.45, 165.96, 183.36; GC RT: (26.79 minutes); MS (200 °C, 70 eV): 316 ([M+2-1]⁺, 29.3), 315 (M⁺, 35.2), 314 ([M-1]⁺, 30.2), 301 (53.6), 299 (58.9), 115 (100.0), 91 (16.8).

2-(4-Methylbenzylidene)-1-(7'-bromobenzofuran-3-one) [10]: scavenger was reintroduced two times to yield 0.18 g (53%) of the product as a pale yellow solid (mp = 187-192 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.40 (s, 3H), 6.93 (s, 1H), 7.08 (t, *J*=7.7 Hz, 1H), 7.26 (d, *J*=8.0 Hz, 2H), 7.72 (d, *J*=8.2 Hz, 1H), 7.77 (d, *J*=8.1 Hz, 1H), 7.84 (d, *J*=8.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ= 21.66, 106.08, 114.85, 123.39, 124.43, 129.06, 129.81 (2C), 131.91 (2C), 139.11, 141.02, 146.15, 162.75, 183.85; GC RT: (26.41 minutes); MS (200 °C, 70 eV): 316 ([M+2-1]⁺, 37.5), 315 (M⁺, 38.2), 314 ([M-1]⁺, 38.6), 301 (72.2), 299 (75.4), 115 (79.6), 91 (25.2).

2-(4-Methylbenzylidene)-1-(5'-methylbenzofuran-3-one) [11]: scavenger was reintroduced one more time to produce 0.14 g (56%) of the desired product as a pale yellow solid (mp = 117-119 °C). ¹H NMR (300 MHz, CDCl₃): δ= 2.37 (s, 3H), 2.38 (s, 3H), 6.83 (s, 1H), 7.17 (d, *J*=8.6 Hz, 1H), 7.22 (d, *J*=7.9 Hz, 2H), 7.41 (d, *J*=8.6 Hz, 1H), 7.55 (s, 1H), 7.77 (d, *J*=7.9 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃):

δ = 20.84, 21.71, 112.54, 113.04, 121.67, 124.20, 129.67, 129.73 (2C), 131.60 (2C), 133.09, 137.91, 140.39, 146.97, 164.51, 184.83; GC RT: (25.50 minutes); MS (200 °C, 70 eV): 250 (M^+ , 69.2), 249 (74.0), 235 (100), 134 (54.5), 124 (77.5), 89 (60.2).

2-(4-Methylbenzylidene)-1-(5',6'-dimethylbenzofuran-3-one) [12]: another 2 equivalents of the hydrazide scavenger were reintroduced in 5 mL/mmol DCM and DMF to yield 0.11 g (40%) of the product as a pale yellow solid (mp = 181-183 °C). ^1H NMR (500 MHz, CDCl_3): δ = 2.22 (s, 3H), 2.31 (s, 3H), 2.36 (s, 3H), 6.76 (s, 1H), 7.03 (s, 1H), 7.20 (d, J =8.0 Hz, 2H), 7.45 (s, 1H), 7.73 (d, J =8.1 Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3): δ = 19.28, 21.30, 21.48, 112.23, 113.18, 119.33, 124.16, 129.48 (2C), 129.59, 131.26 (2C), 132.01, 139.95, 146.93, 147.59, 164.98, 184.27; GC RT: (27.00 minutes); MS (200 °C, 70 eV): 264 (M^+ , 68.2), 263 (84.2), 249 (91.45), 148 (42.3), 131 (55.9), 117 (53.7), 91 (100.0).

2-(4-Methylbenzylidene)-1-(7'-methylbenzofuran-3-one) [13]: 0.13 g (52%) yield as a pale yellow solid (mp = 107-108 °C). ^1H NMR (500 MHz, CDCl_3): δ = 2.37 (s, 3H), 2.44 (s, 3H), 6.83 (s, 1H), 7.05 (t, J =7.4 Hz, 1H), 7.22 (d, J =8.0 Hz, 2H), 7.39 (d, J =6.9 Hz, 1H), 7.58 (d, J =8.0 Hz, 1H), 7.76 (d, J =8.0 Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3): δ = 14.38, 21.72, 113.11, 121.26, 121.90, 122.97, 123.29, 128.74 (3C- two are coincident), 131.62 (2C), 137.60, 140.42, 146.74, 164.79, 185.20; GC RT: (25.08 minutes); MS (200 °C, 70 eV): 250 (M^+ , 58.9), 249 (66.0), 235 (100), 124 (61.0), 106 (70.0), 89 (59.3).

2-(4-Methylbenzylidene)-1-(6'-hydroxybenzofuran-3-one) [14]: 0.05 g (20%)

yield as an off-white solid (mp > 210 °C). ¹H NMR (500 MHz, (CD₃)₂CO): δ= 2.36 (s, 3H), 6.68 (s, 1H), 6.76 (dd, J=8.6, 2.3 Hz, 1H), 6.82 (d, J=1.7 Hz, 1H), 7.28 (d, J=8.0 Hz, 2H), 7.60 (d, J=8.0 Hz, 1H), 7.83 (d, J=8.6 Hz, 2H); ¹³C NMR (125 MHz, (CD₃)₂CO): δ= 20.67, 98.79, 110.41, 112.82, 113.96, 125.79, 129.65 (2C), 129.88, 131.22 (2C), 139.93, 147.49, 166.08, 168.39, 181.58; GC RT: (28.08 minutes); MS (200 °C, 70 eV): 252 (M⁺, 58.5), 251 (54.8), 63 (76.1), 44 (100.0).

2-(4-Methylbenzylidene)-1-(4'-hydroxybenzofuran-3-one) [15]: 0.04 g (15%)

yield, as a pale yellow solid (mp = 156-158 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.41 (s, 3H), 6.61 (d, J=8.1 Hz, 1H), 6.79 (d, J=8.1 Hz, 1H), 6.83 (s, 1H), 7.25 (d, J=7.4 Hz, 2H), 7.49 (t, J=8.3 Hz, 1H), 7.80 (d, J=8.0 Hz, 2H), 7.83 (x, 1H, OH); ¹³C NMR (125 MHz, CDCl₃): δ= 21.77, 103.71, 109.77, 109.81, 113.59, 129.32, 129.85 (2C), 131.75 (2C), 139.19, 140.89, 146.28, 156.72, 164.90, 185.75; GC RT: (25.09 minutes); MS (200 °C, 70 eV): 252 (M⁺, 98.7), 251 (77.8), 237 (100.0).

2-(4-Methylbenzylidene)-1-(6'-hydroxy-4'-methylbenzofuran-3-one) [16]:

0.03 g (11%) yield as a pale yellow solid (mp >210 °C). ¹H NMR (500 MHz, CDCl₃): δ= 2.39 (s, 3H), 2.62 (s, 3H), 6.44 (s, 1H), 6.57 (s, 1H), 6.75 (s, 1H), 7.23 (d, J=8.2 Hz, 2H), 7.76 (d, J=8.6 Hz, 2H); ¹³C NMR (500 MHz, (CD₃)₂CO): δ= 16.98, 20.66, 96.34, 109.58, 112.35, 113.43, 129.61 (2C), 130.01, 131.08 (2C), 139.67, 141.38, 147.65, 165.36, 168.61, 182.43; GC RT: (28.46 minutes); MS

(200 °C, 70 eV): 266 (M^+ , 72.7), 265 (100.0), 251 (70.7), 132 (82.9), 77 (90.0), 51 (84.9).

2-(4-Methylbenzylidene)-1-(6'-hydroxy-7'-methylbenzofuran-3-one) [17]:

0.05 g (19%) yield as a pale yellow solid (mp >210 °C). ^1H NMR (500 MHz, $(\text{CD}_3)_2\text{CO}$): δ = 2.25 (s, 3H), 2.43 (s, 3H), 6.42 (d, J =2.3 Hz, 1H), 6.52 (s, 1H), 6.52 (d, J =1.7 Hz, 1H), 7.17 (d, J =7.4 Hz, 2H), 7.71 (d, J =8.1 Hz, 2H); ^{13}C NMR (125 MHz, $(\text{CD}_3)_2\text{CO}$): δ = 16.96, 20.65, 96.33, 109.55, 112.36, 113.43, 129.62 (2C), 130.02, 131.08 (2C), 139.67, 141.37, 148.57, 165.39, 168.49, 182.42; GC RT: (28.34 minutes); MS (200 °C, 70 eV): 266 (M^+ , 31.9), 265 (48.9), 251 (43.6), 77 (53.6), 44 (100.0).

General procedure for UV-vis spectroscopy

Stock solutions ranging from 87 μM to 134 μM of the aurones were made by dissolving a known mass of the aurones in acetonitrile. Each stock solution was used for subsequent dilutions to achieve a final concentration ranging from 1.5 μM to 3.3 μM . Using five different known concentration points for each aurone, the molar extinction coefficient was determined by measurement of the absorbance at the λ_{max} closest to the visible range. The absorbance measurement of the highest concentration point was used for the absorbance shift study. 2-(4-methylbenzylidene)-1-(6'-hydroxy-7'-methylbenzofuran-3-one (**17**) was the exception as the exact concentration of the solution could not be determined due to its poor solubility in acetonitrile.

Hep G2 cytotoxicity assay

Hep G2 human liver cancer cells were cultured in phenol red Dulbecco's Modified Eagle Medium (DMEM) with 10% fetal bovine serum (FBS) and 1% Pen-strep (PSG) under 37°C and 5% CO₂ atmospheric conditions. The cells were washed with phosphate buffered saline (PBS) and diluted with phenol red free DMEM to a concentration between 200,000 to 250,000 cells/mL. The cells were distributed in a 96-well plate with each well containing 100 µL of the cell solution and were allowed to settle for 2 hours in the incubator. The cells were then treated with 200 µM of the aurones in DMSO and the known dyes in 10:90 (v:v) DMSO/ultrapure water in triplicates and were left in the incubator for 3 days. The vehicles were used as the negative controls (DMSO for the aurones and 10:90 DMSO:ultrapure water for the known dyes). 20 µL of 5 mg/mL MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added to each well containing treated cells after the 3-day incubation period and was allowed to settle in the incubator for 3.5 more hours. Finally, the media was removed from the wells and replaced by 100 µL of DMSO to lyse the cells and dissolve the formazan crystals that were formed. The absorbance was then recorded on the SpectraMax M5 Multi-Mode Microplate Reader. The procedure was repeated two more times using different batches of cultured Hep G2 cells on different days.

Dyeing and mordanting processes [1]

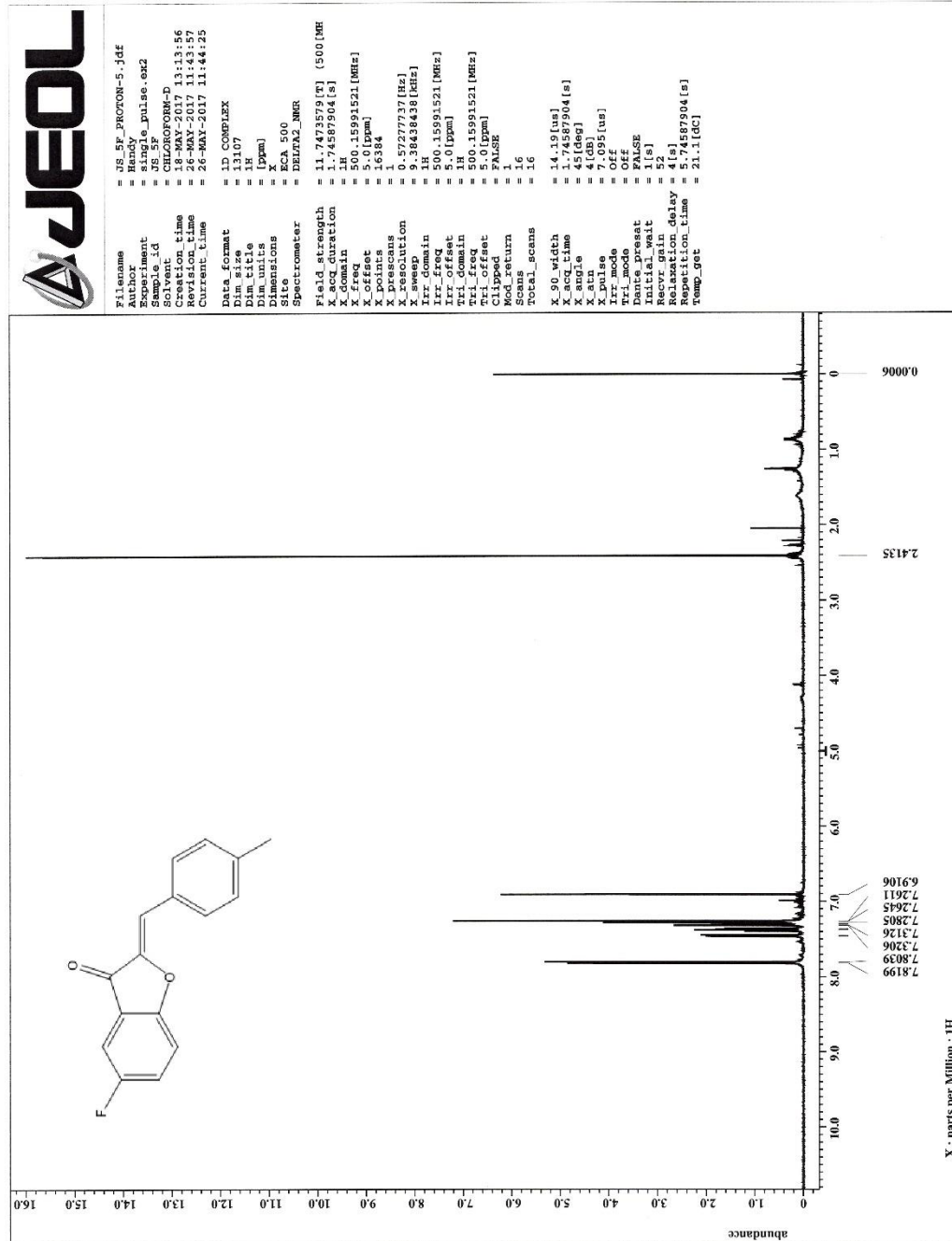
Multi-fiber fabric (Kimble 43, 13 fabric strip) was washed with warm water first then air dried. Mordanting solutions with FeSO₄ were made fresh at 1 g/25 mL of deionized H₂O for both the pre-mordanting and post-mordanting processes. Both (9) (2-

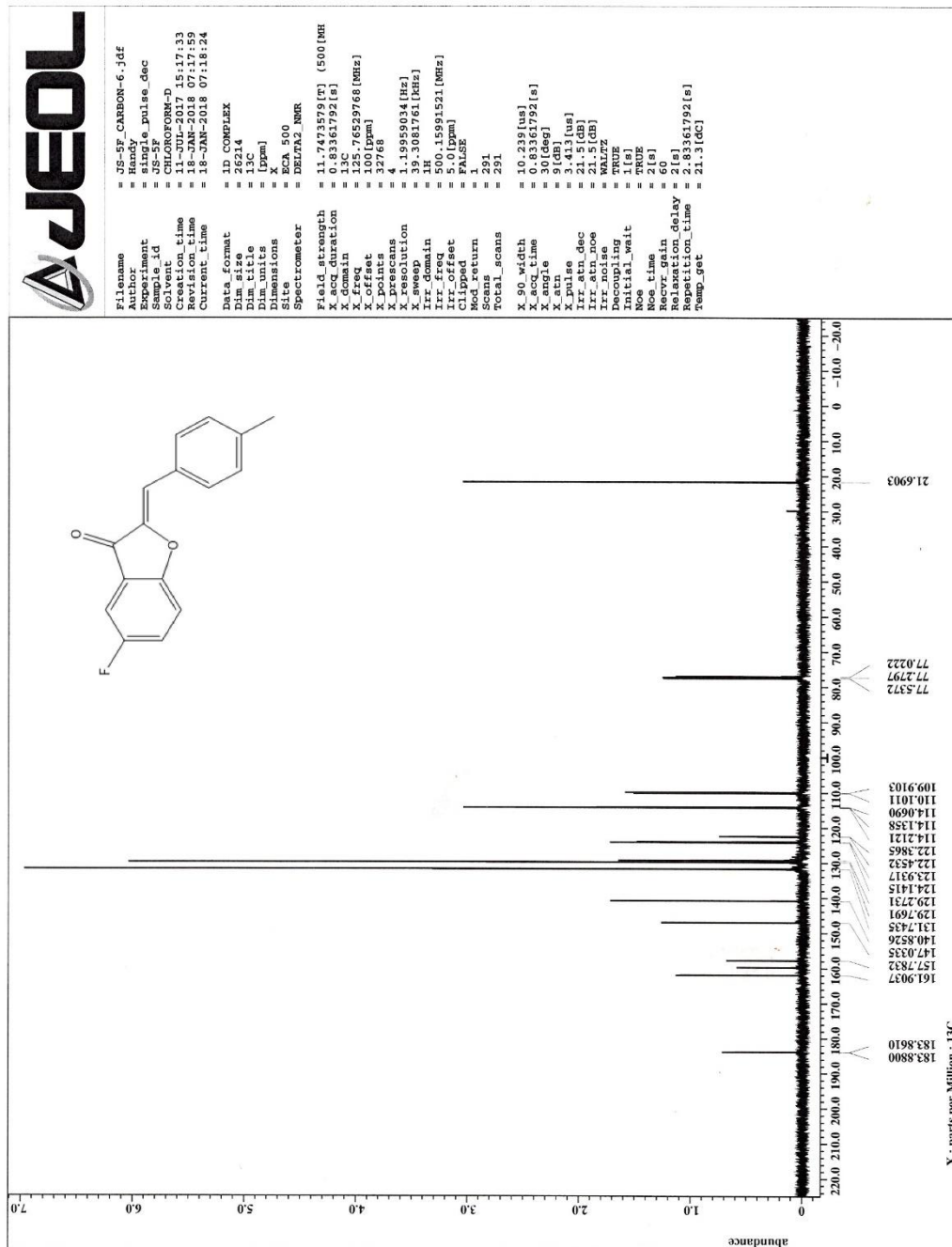
(4-methylbenzylidene)-1-(7'-bromobenzofuran-3-one)) and (**14**) (2-(4-methylbenzylidene)-1-(4'-hydroxybenzofuran-3-one)) were used as the dye samples with each undergoing pre, simultaneous, and post-mordanting processes.

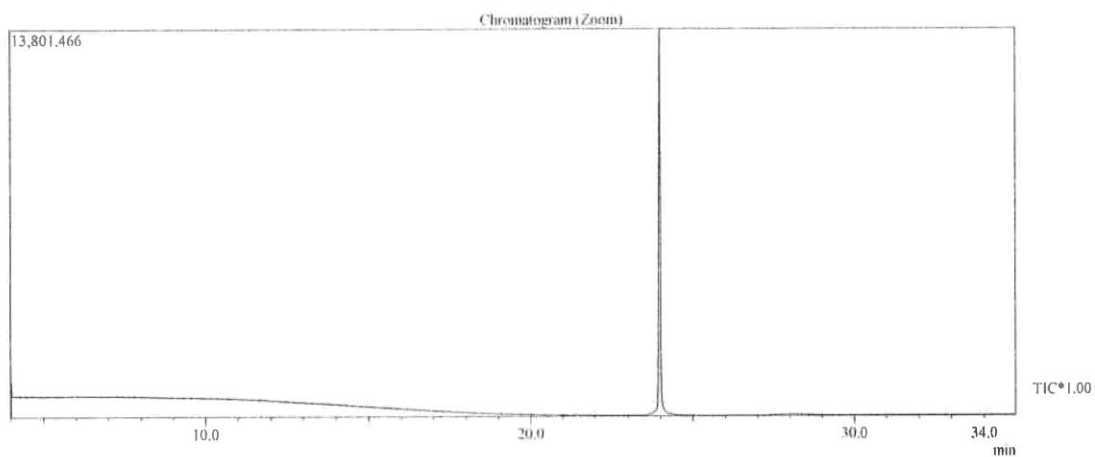
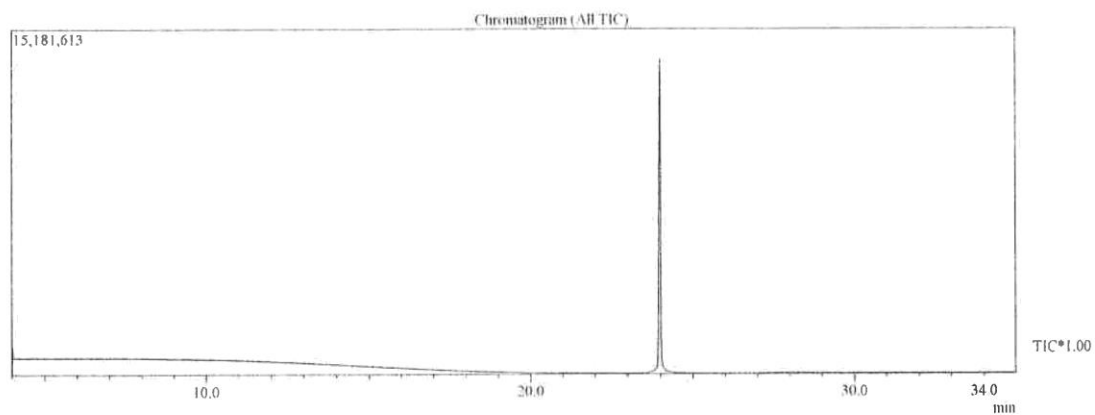
References

1. Praveena, R.; Deepa, V.; Sivakumar, R. Investigations on C-glycosyl flavonoids from *Rhynchosia capitata*, its antimicrobial and dyeing properties. *Fibers Polym.* **2014**, *15*, 525-533.

2-(4-methylbenzylidene)-1-(5'-fluorobenzofuran-3-one) [2]

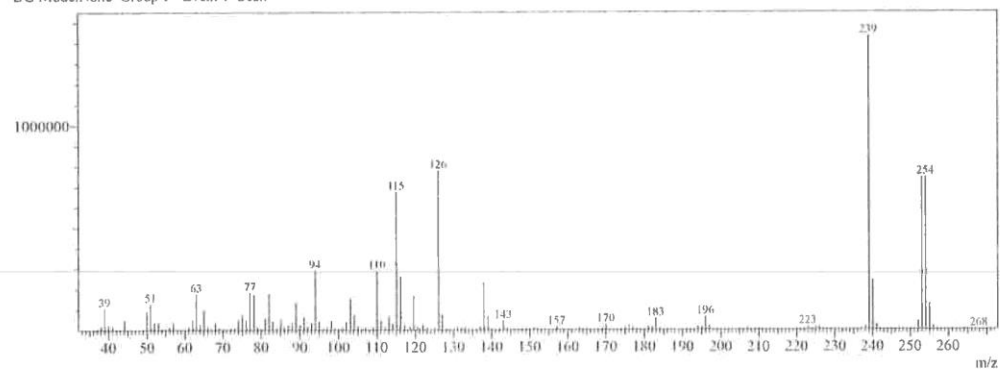




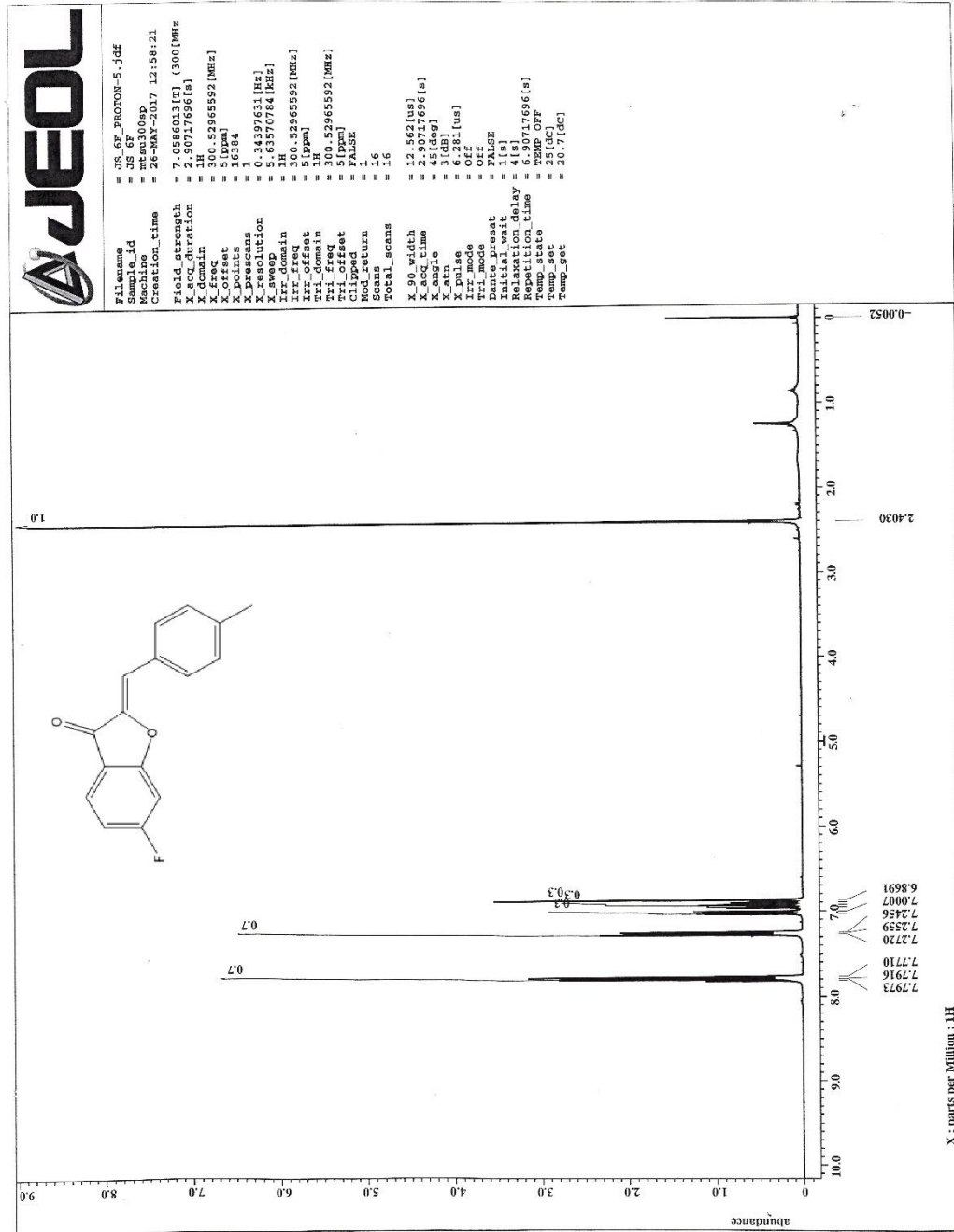


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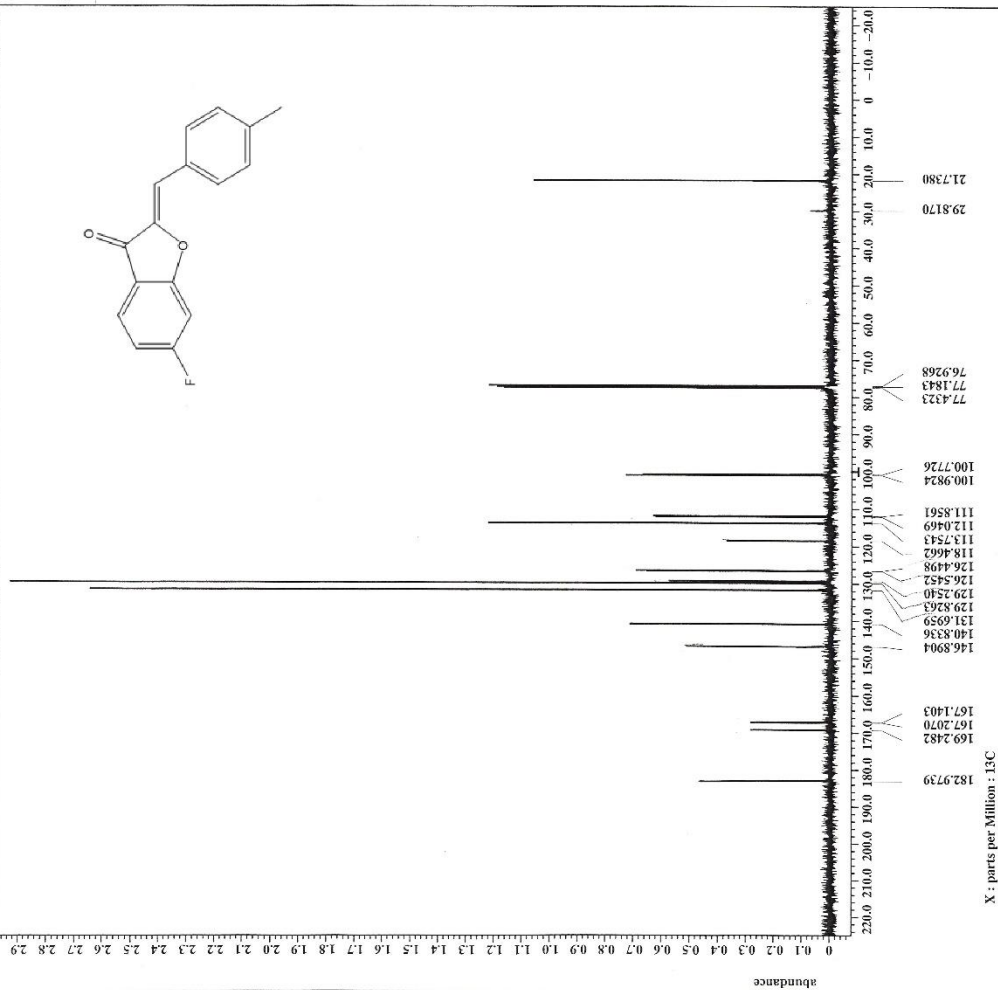
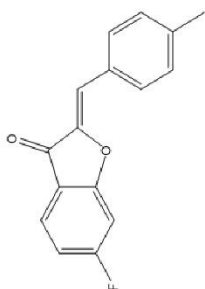


2-(4-methylbenzylidene)-1-(6'-fluorobenzofuran-3-one) [3]

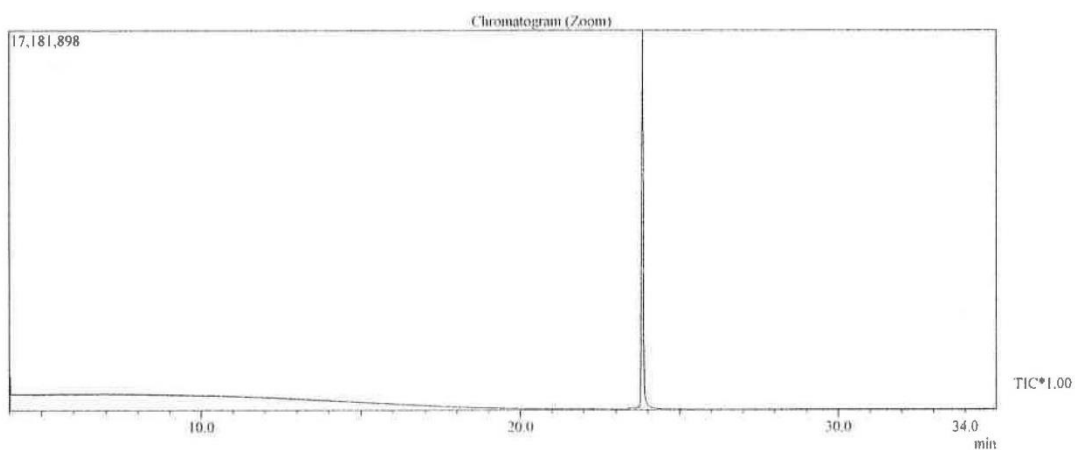
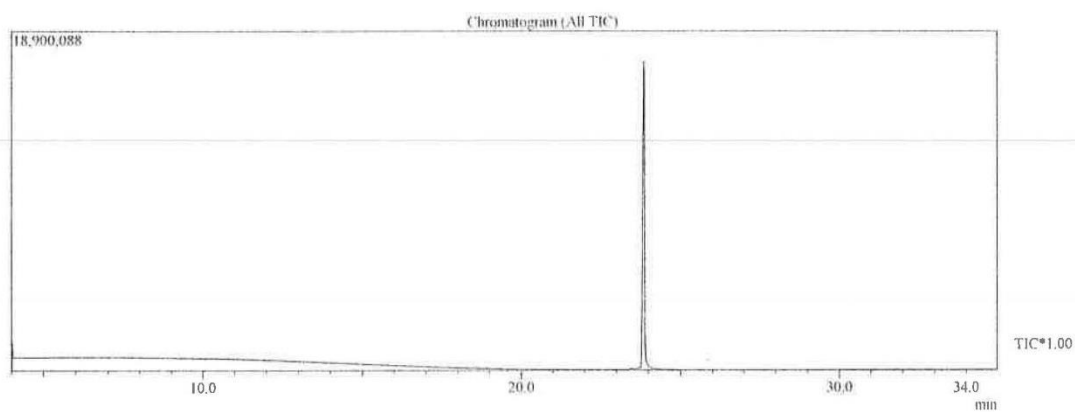




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Spectrometer = JEOL-500
Field_strength = 11.7473579 [T] (500 [MHZ])
X_acq_duration = 0.83361792 [s]
X_offset = 125.76529768 [MHz]
X_points = 32768
X_prescans = 1
X_sweep = 1.19959034 [Hz]
X_resolution = 39.3081761 [kHz]
Irr_domain = 1H
Irr_freq = 500.15991521 [MHz]
Irr_offset = 125.76529768 [MHz]
Mod_return = FALSE
Total_scans = 420
X_90_width = 10.239 [us]
X_acq_time = 0.83361792 [s]
X_angle = 30 [deg]
X_pulse = 3.413 [us]
Irr_atn_dec = 21.5 [dB]
Irr_atn_noe = 21.5 [dB]
X_resolution = 39.3081761 [kHz]
Decoupling = WALTZ
Initial_wait = 118
Noe_time = TRUE
Noe_delay = 2 [s]
Relaxation_delay = 2 [s]
Repetition_time = 2.83361792 [s]
Temp_get = 21.4 [degC]

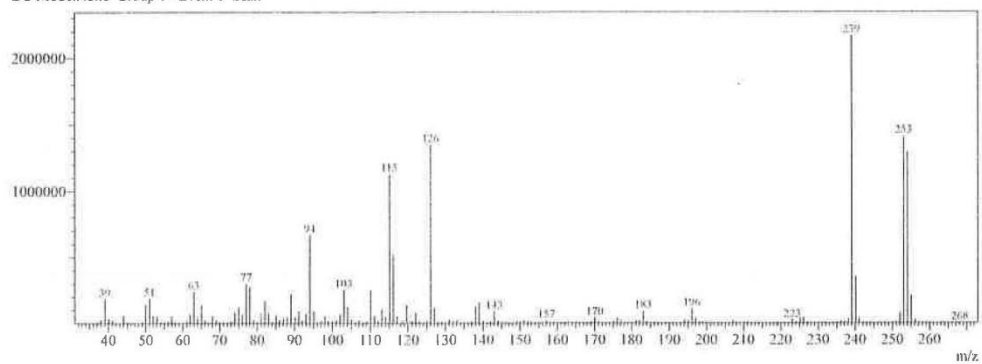


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Spectrum

Line#: 1 R.Time: 23.865 (Scan#: 3974)
MassPeaks: 197
RawMode: Single 23.865 (3974) BasePeak: 239.05 (2166300)
BG Mode: None Group 1 - Event 1 Scan

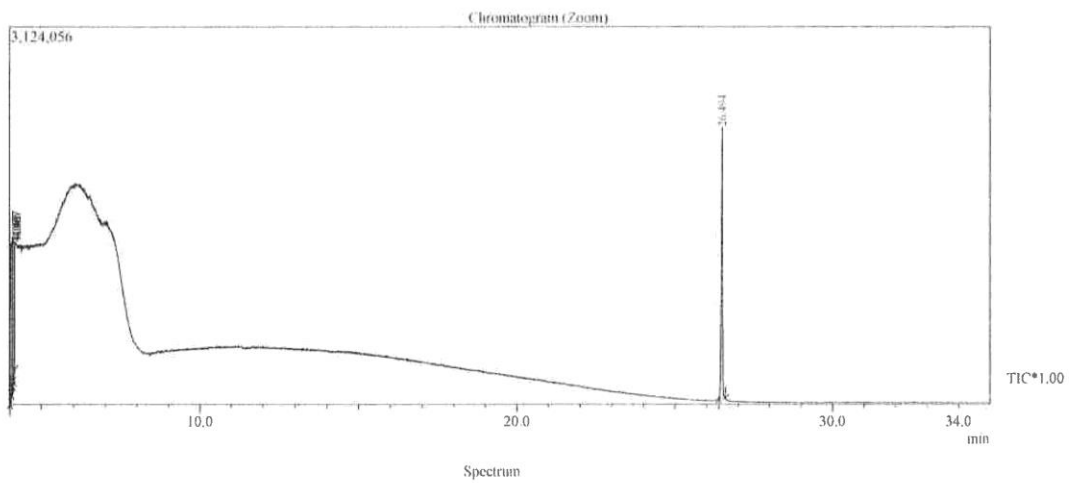
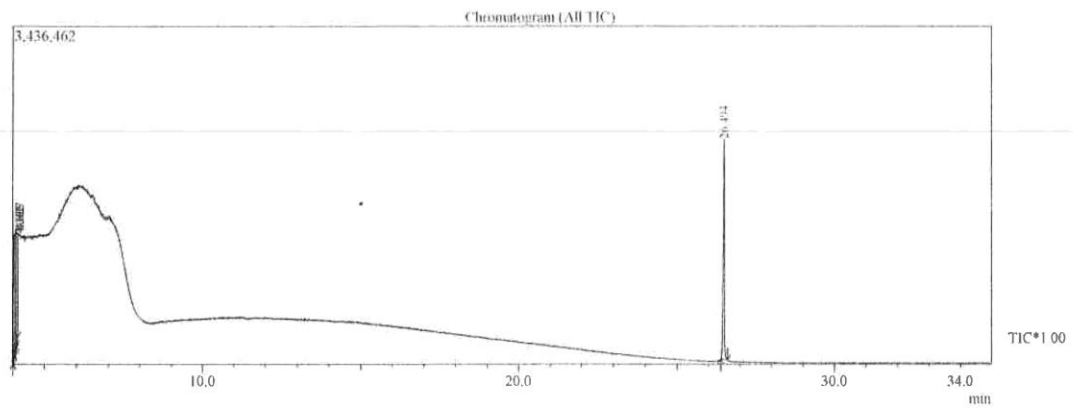




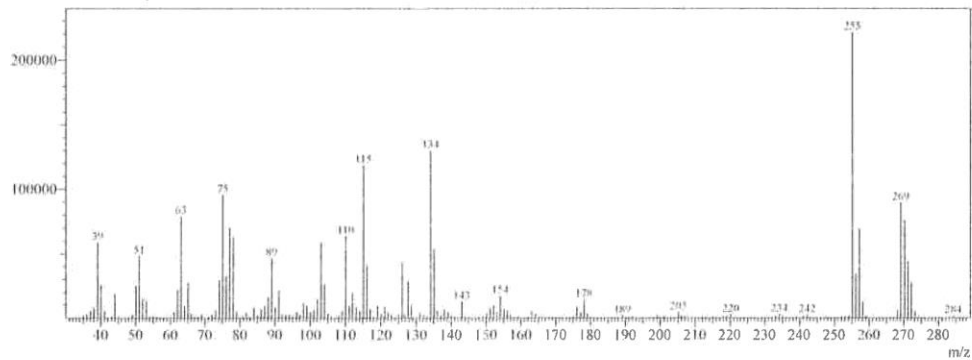
JEOL



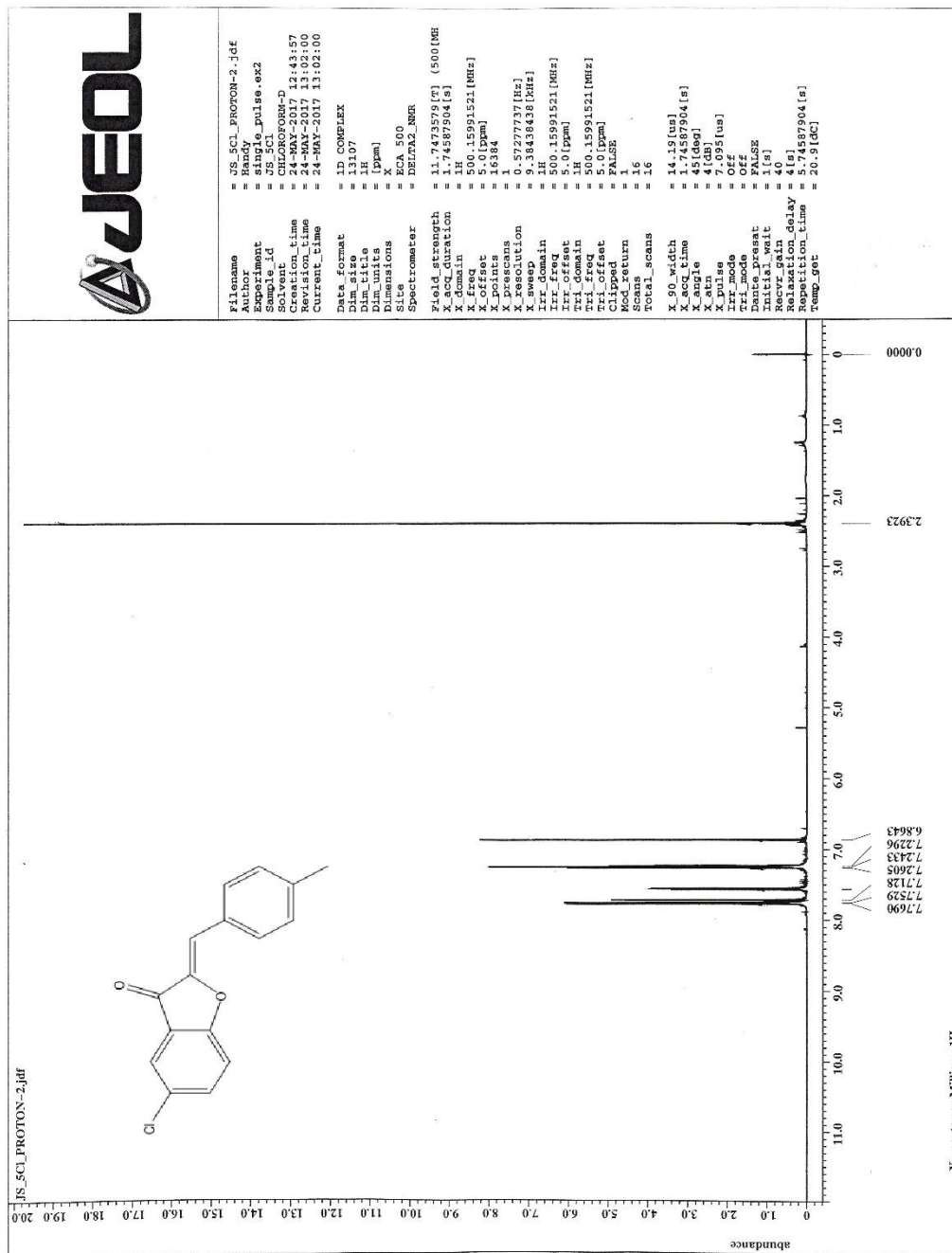
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Line#1 R.Time:26.495(Scan#:4500)
MassPeaks:156
RawMode:Single 26.495(4500) BasePeak:255.15(221250)
BG Mode:None Group 1 - Event 1 Scan

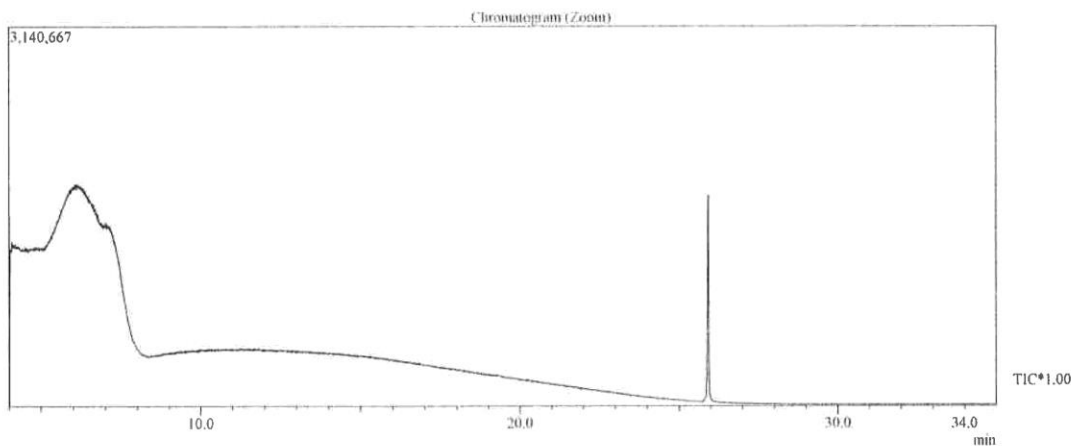
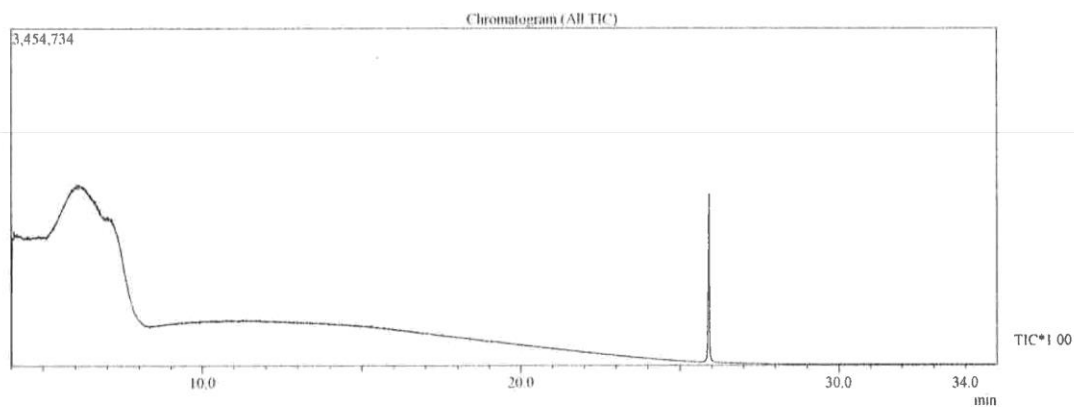


2-(4-methylbenzylidene)-1-(5'-chlorobenzofuran-3-one) [5]





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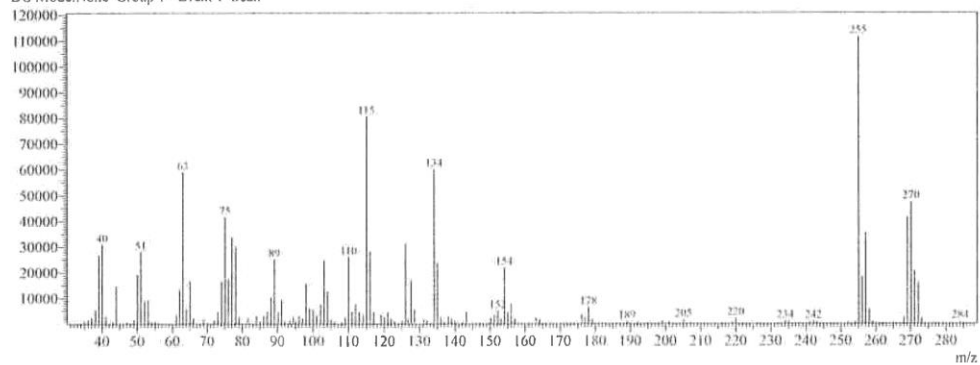
Spectrum

Line#1 R.Time:25.910(Scan#:4383)

MassPeaks:128

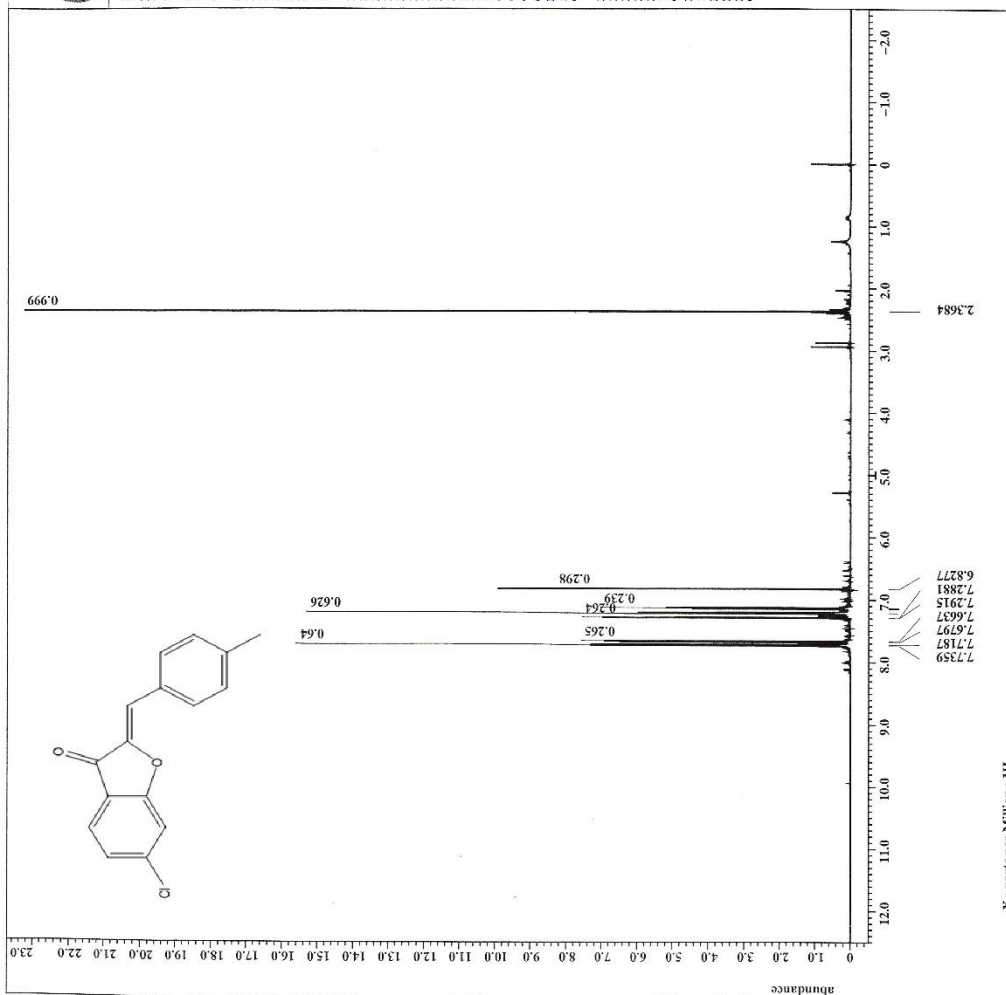
RawMode:Single 25.910(4383) BasePeak:255.10(111482)

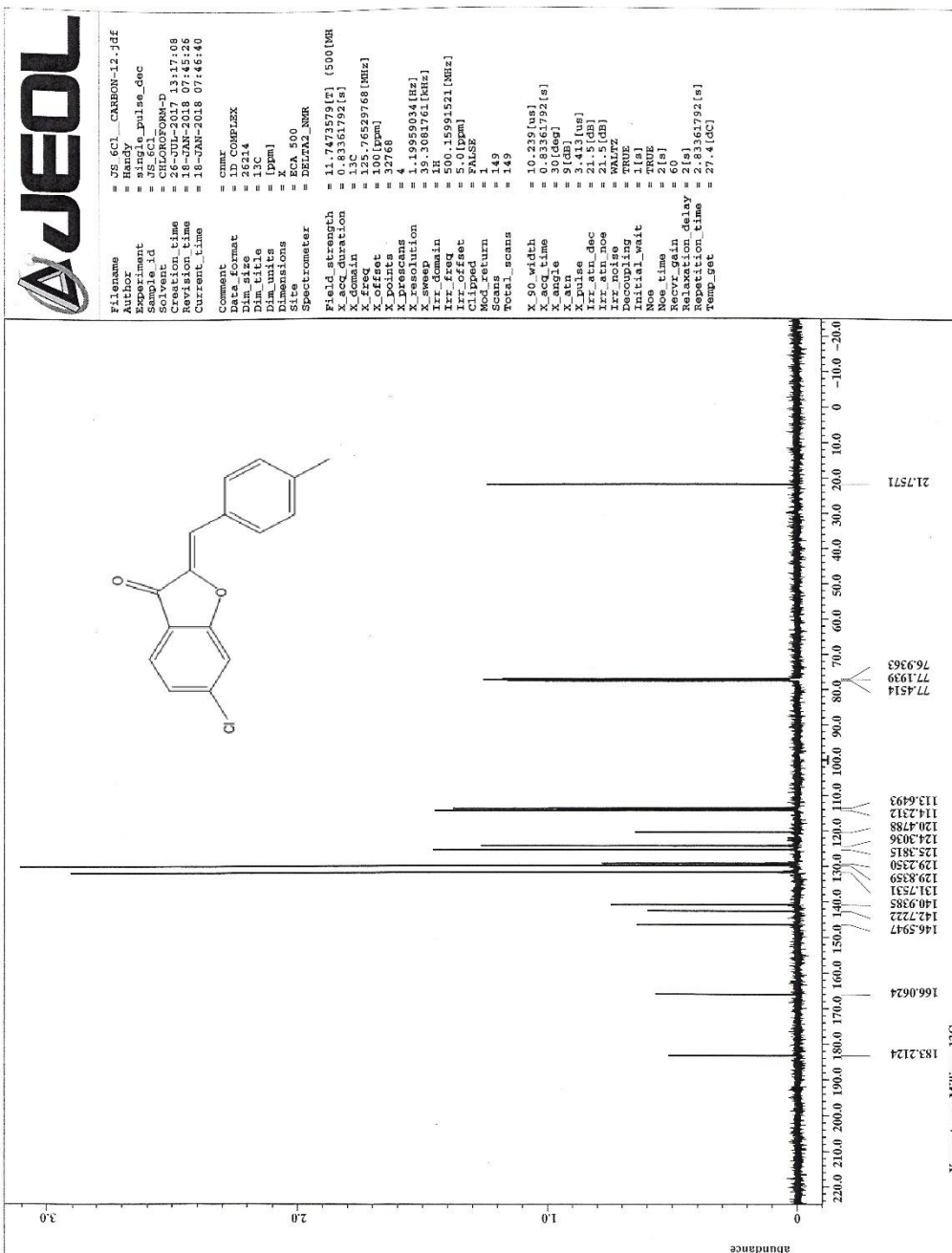
BG Mode:None Group 1 - Event 1 Scan



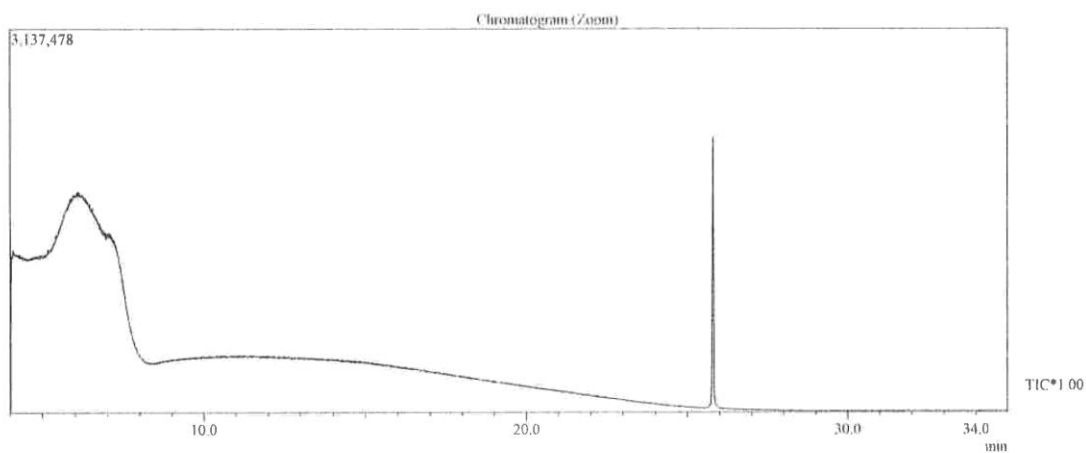
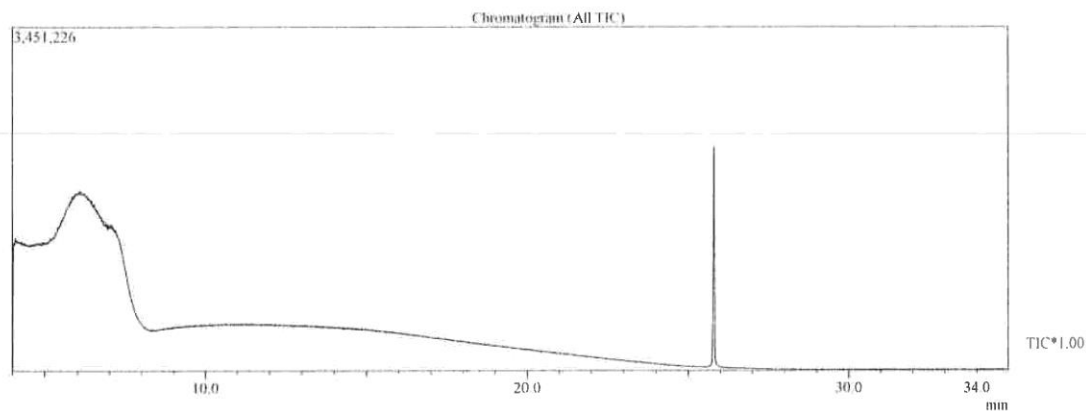


| | |
|------------------|-----------------------------|
| Filename | = JS_SCL_PROTON-15_jd1 |
| Author | = Handy |
| Experiment | = single_pulse_eh2 |
| Sample_id | = SCL |
| Sample_name | = SCL-PROTON-D |
| Creation time | = 26-JUN-2017 13:07:10 |
| Revision time | = 19-JAN-2018 07:44:42 |
| Current time | = 19-JAN-2018 07:44:49 |
| Comment | = cmnr |
| Data format | = 1D COMPLEX |
| Dim size | = 13107 |
| Dim title | = 1H |
| Dimensions | = X [ppm] |
| Site | = EKA 500 |
| Spectrometer | = DELTA500 |
| Acq duration | = 11.7473579 [s] (500 [MHz] |
| X domain | = 1.74507904 [s] |
| X freq | = 500.15991521 [MHz] |
| X fref | = 500.15991521 [MHz] |
| X points | = 16384 |
| X prescans | = 1 |
| X resolution | = 0.57227773 [Hz] |
| X rdomain | = 1.74507904 [MHz] |
| Irq freq | = 5.0 [Dpm] |
| Irq offset | = 500.15991521 [MHz] |
| Trf domain | = 5.0 [Dpm] |
| Trf offset | = 500.15991521 [MHz] |
| Clipped | = 5.0 [Dpm] |
| Mod return | = FALSE |
| Total scans | = 16 |
| X 90 width | = 14.19 [us] |
| X 90 time | = 1.74507904 [s] |
| X angle | = 45 [deg] |
| X attn | = 4 [db] |
| X pulse | = 7.095 [us] |
| Irq mode | = OFF |
| Distance preset | = FALSE |
| Initial wait | = 1 [s] |
| Relax gain | = 38 |
| Relaxation delay | = 5.74507904 [s] |
| Relaxation time | = 5.74507904 [s] |
| Temp get | = 27 [C] |



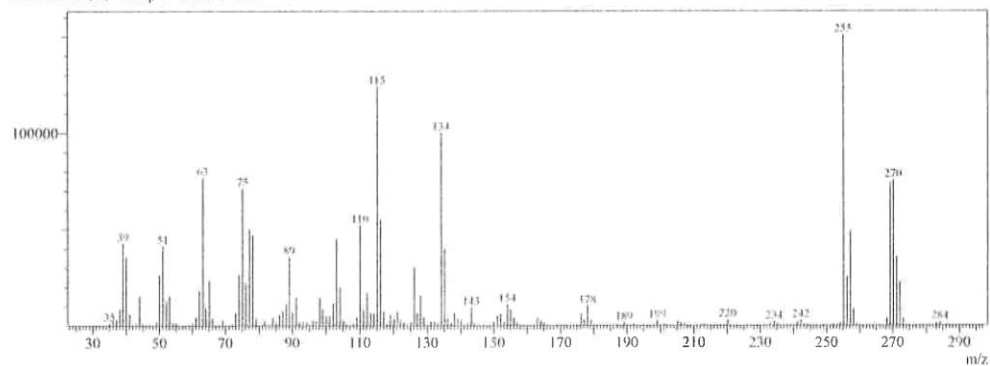


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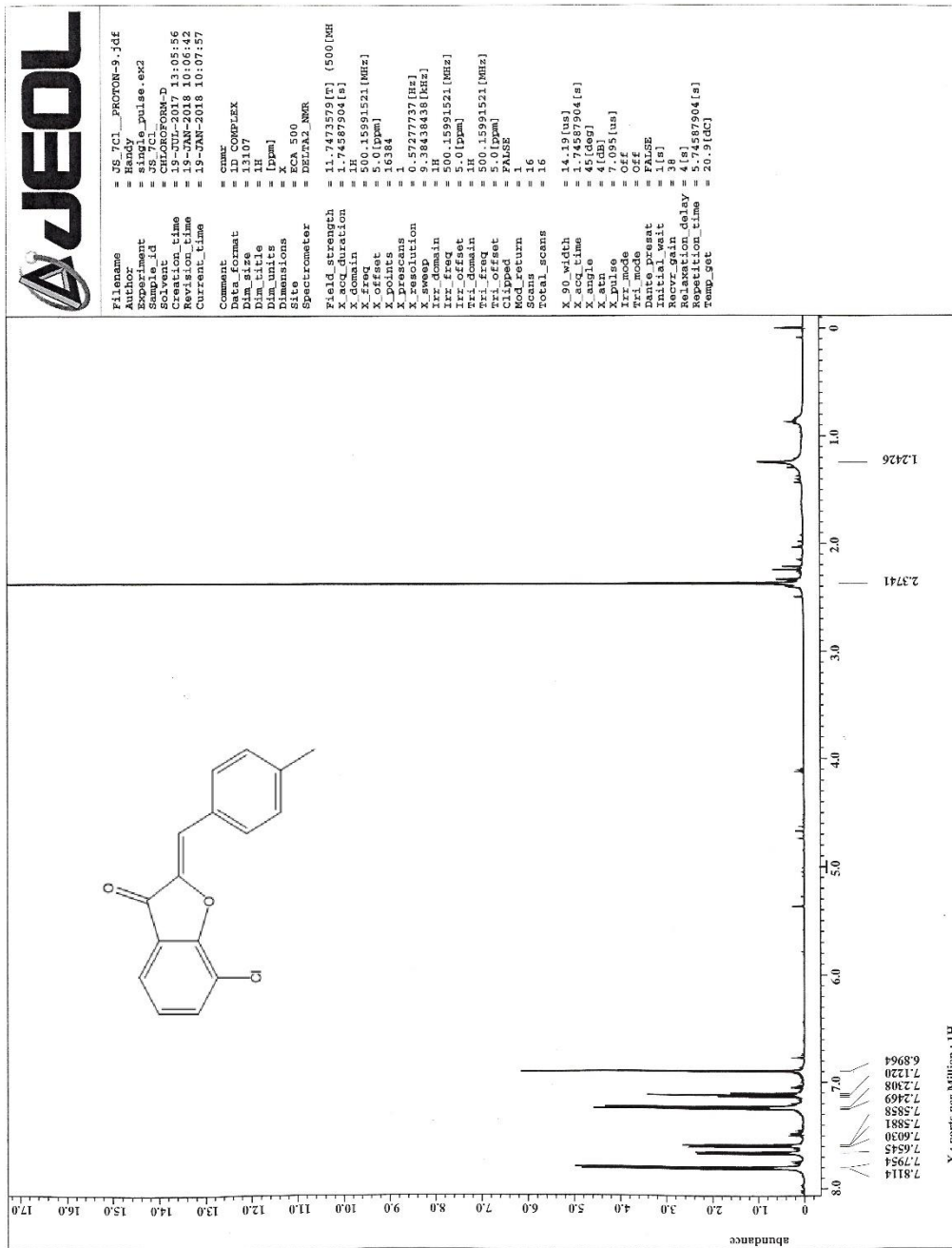


Spectrum

Line#:1 R Time:25.800(Scan#:4361)
MassPeaks:147
RawMode:Single 25.800(4361) BasePeak:255.10(150625)
BG Mode:None Group 1 - Event 1 Scan

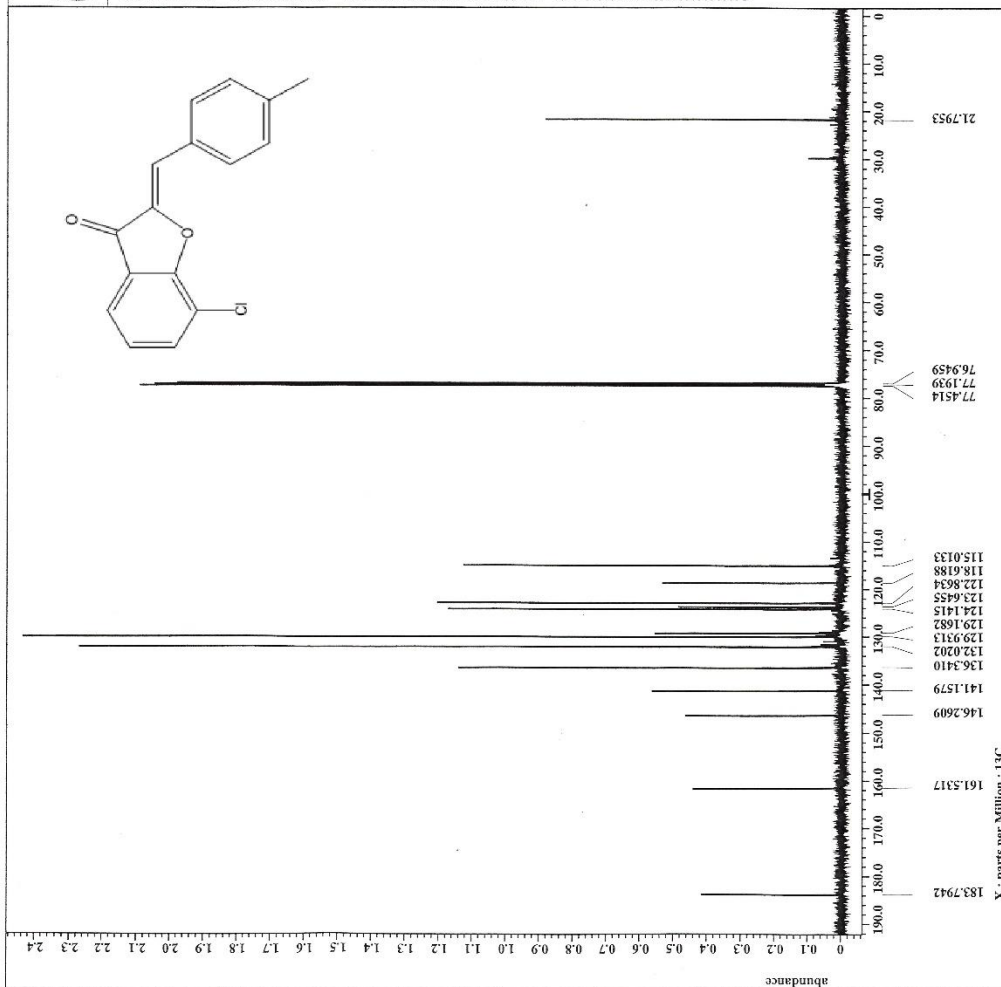
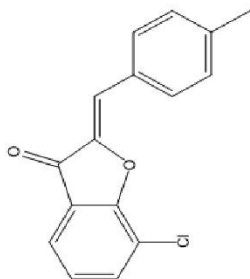


2-(4-methylbenzylidene)-1-(7'-chlorobenzofuran-3-one) [7]

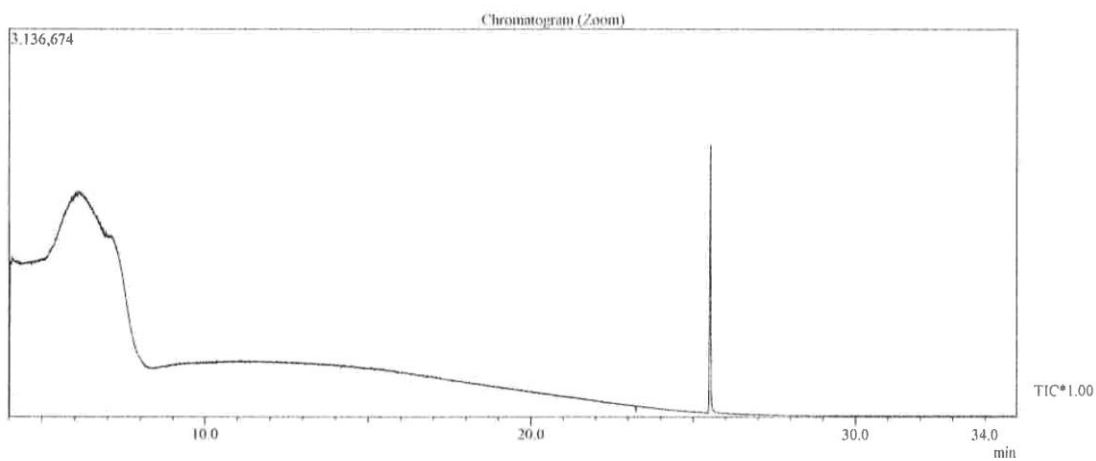
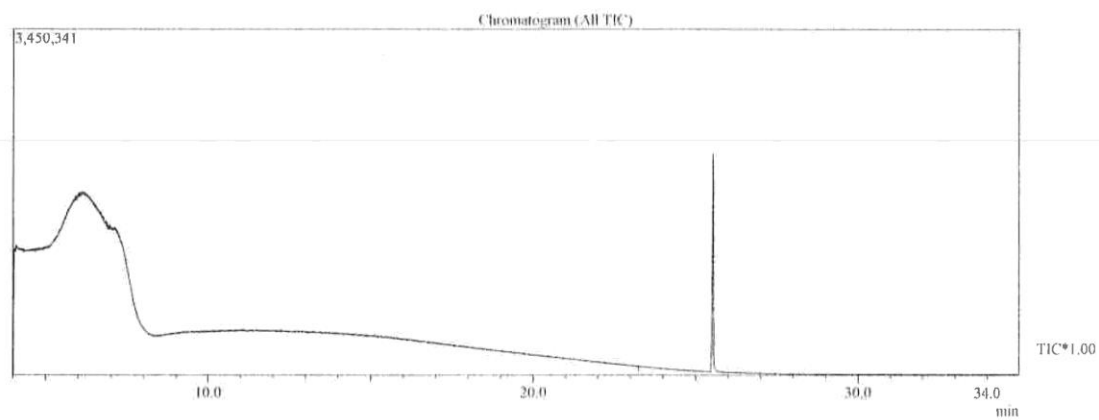




Filename = US_7C1_CARBON-6.jdf
Date_ = 18-JAN-2018 09:36:22
Experiment = single_pulse_dec
Sample_id = US_7C1
Solvent = CHLOROFORM-D
Acq_time = 18-JAN-2018 09:35:45
Revision_time = 18-JAN-2018 09:35:45
Current_time = 18-JAN-2018 09:36:22
Comment =
Data_format = 2D COMPLEX
Data_size = 26214
Dim_title = 13C
Dim_units = [ppm]
Dimensions = 1
Size = ECA 500
Spectrometer = DELTA2 NMR
Yield_strength = 11.747359 [T] (500 [MH]
X_acq_time = 0.3361792 [s]
X_domain = 13C
X_freq = 125.76529768 [MHz]
X_offset = 100 [ppm]
X_resolution = 4
X_sweep = 1.19959034 [Hz]
X_domain = 39.3081761 [MHz]
X_offset = 500.15891521 [MHz]
X_resolution = 5.0 [ppm]
Mod_return = FALSE
Total_scans = 1
X_90_width = 10.239 [us]
X_acq_time = 0.3361792 [s]
X_delay = 90 [us]
X_atn = 9 [dB]
X_pulse = 3.413 [us]
X_atn_dec = 21.5 [dB]
X_atn_inc = 21.5 [dB]
X_atn_iso = 21.5 [dB]
Decoupling = TRUE
Initial_wait = 1 [s]
Recvr_gain = 20 [dB]
Relaxation_delay = 2 [s]
Repetition_time = 2.3361792 [s]
Temp_get = 21.3 [C]

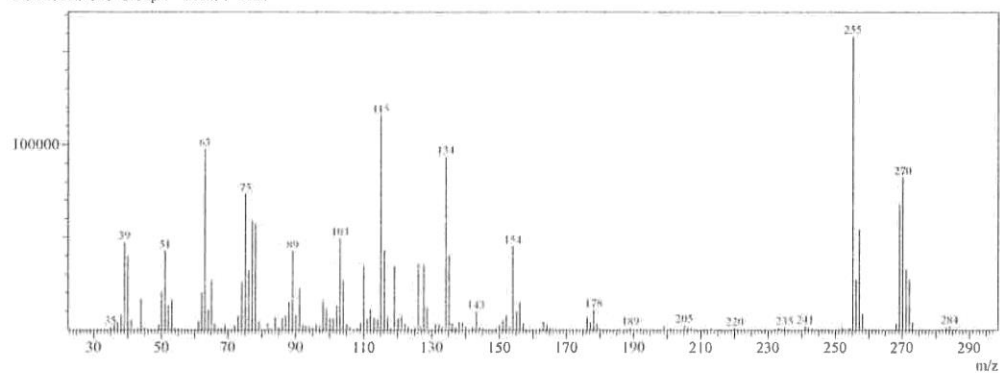


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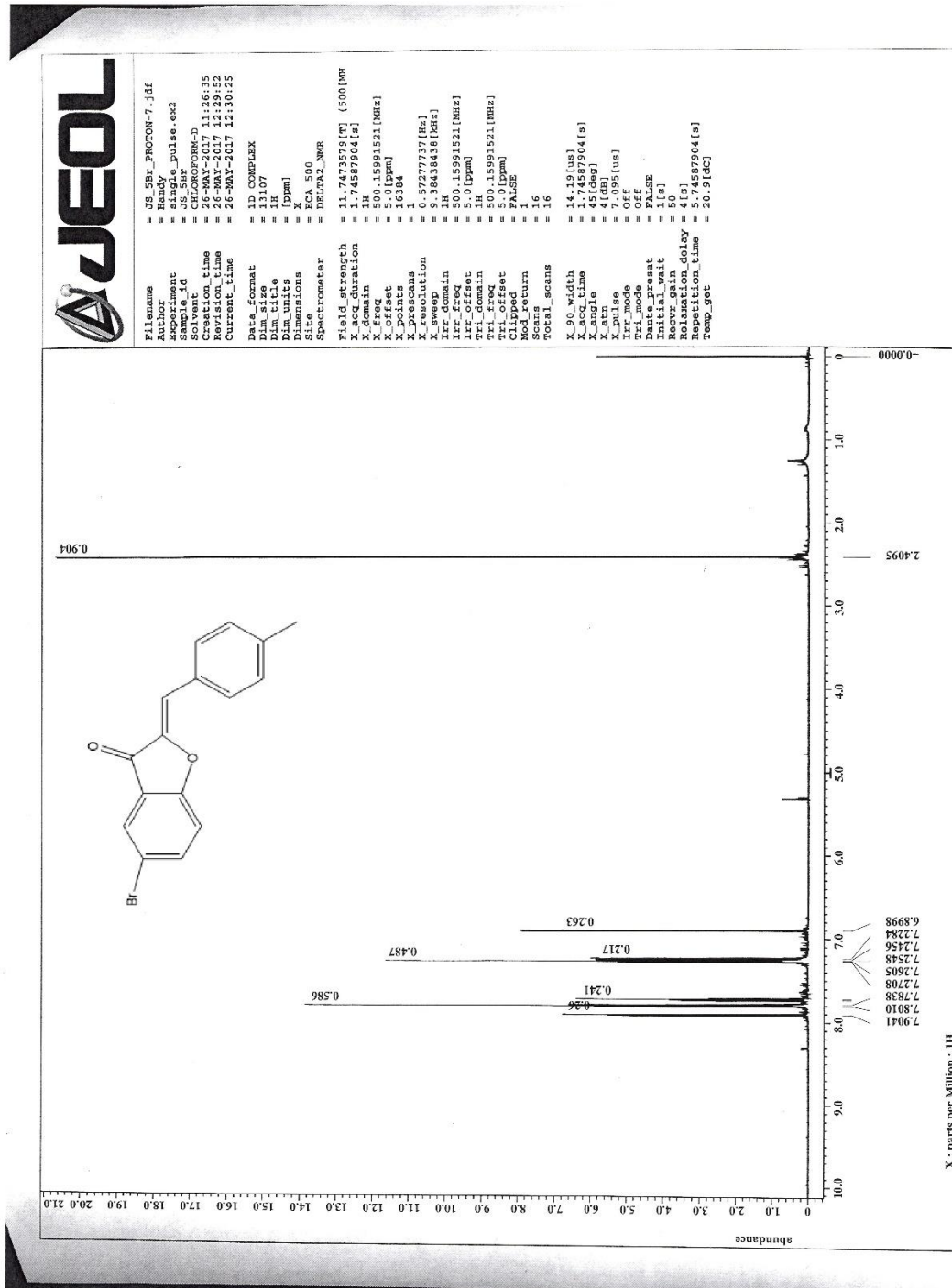


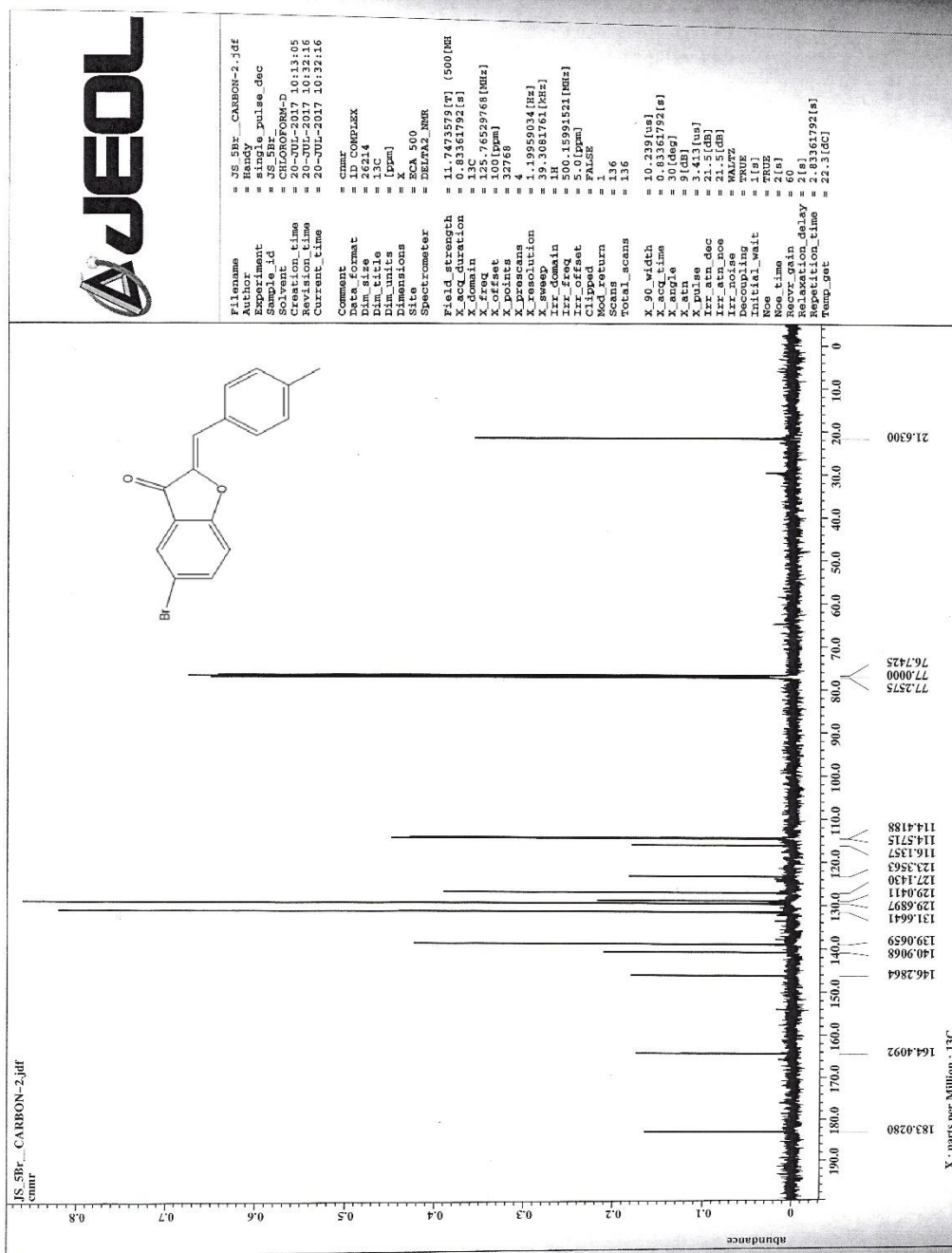
Spectrum

Line#:1 R.Time:25.530(Scan#:4307)
MassPeaks:145
RawMode:Single 25.530(4307) BasePeak:255.15(158383)
BG Mode:None Group 1 - Event 1 Scan

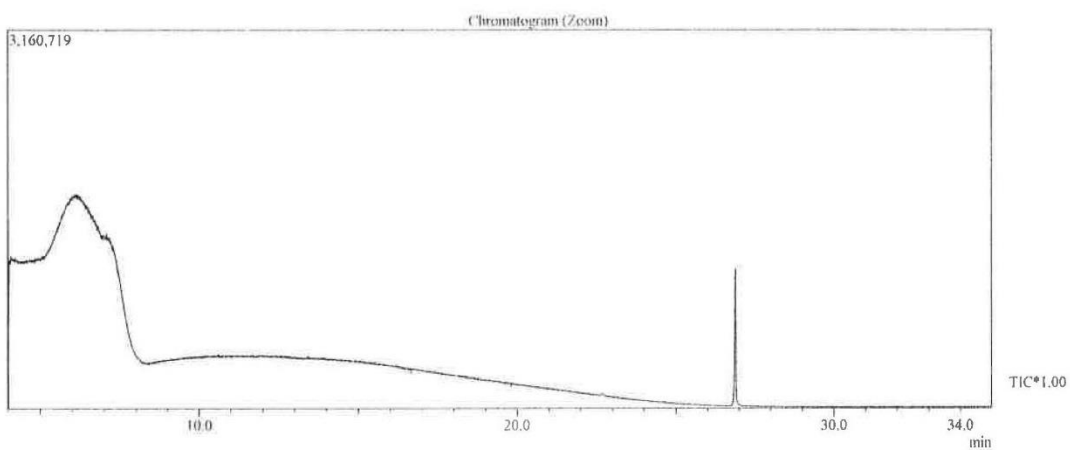
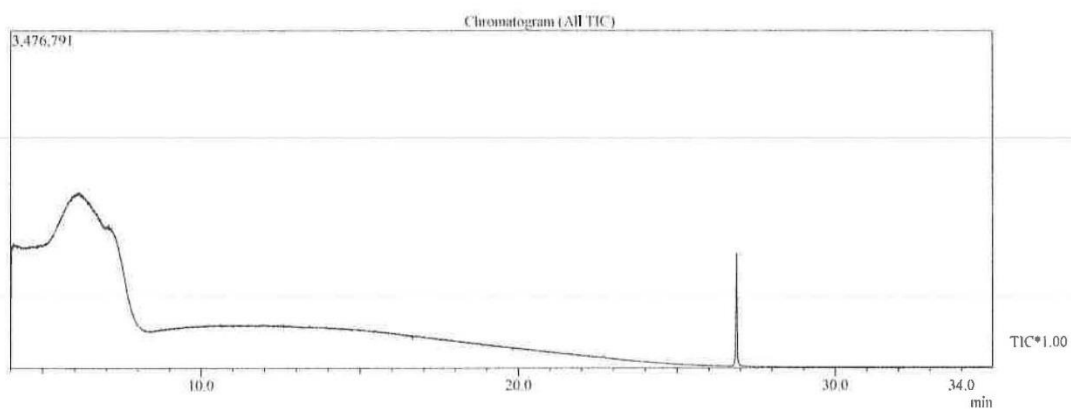


2-(4-methylbenzylidene)-1-(5'-bromobenzofuran-3-one) [8]



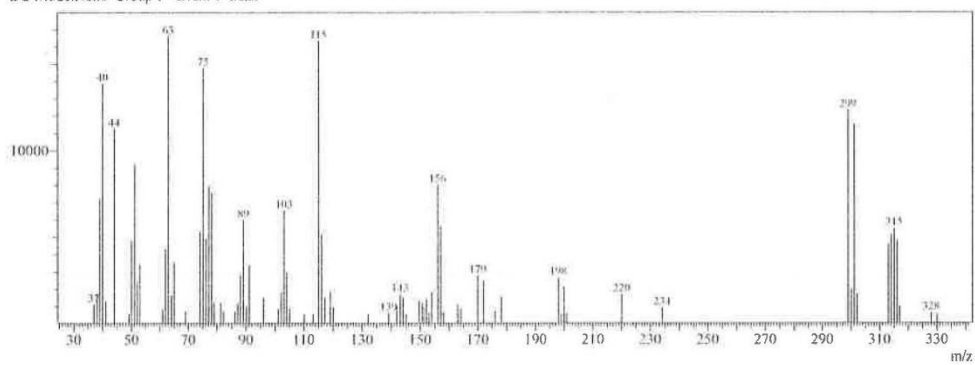


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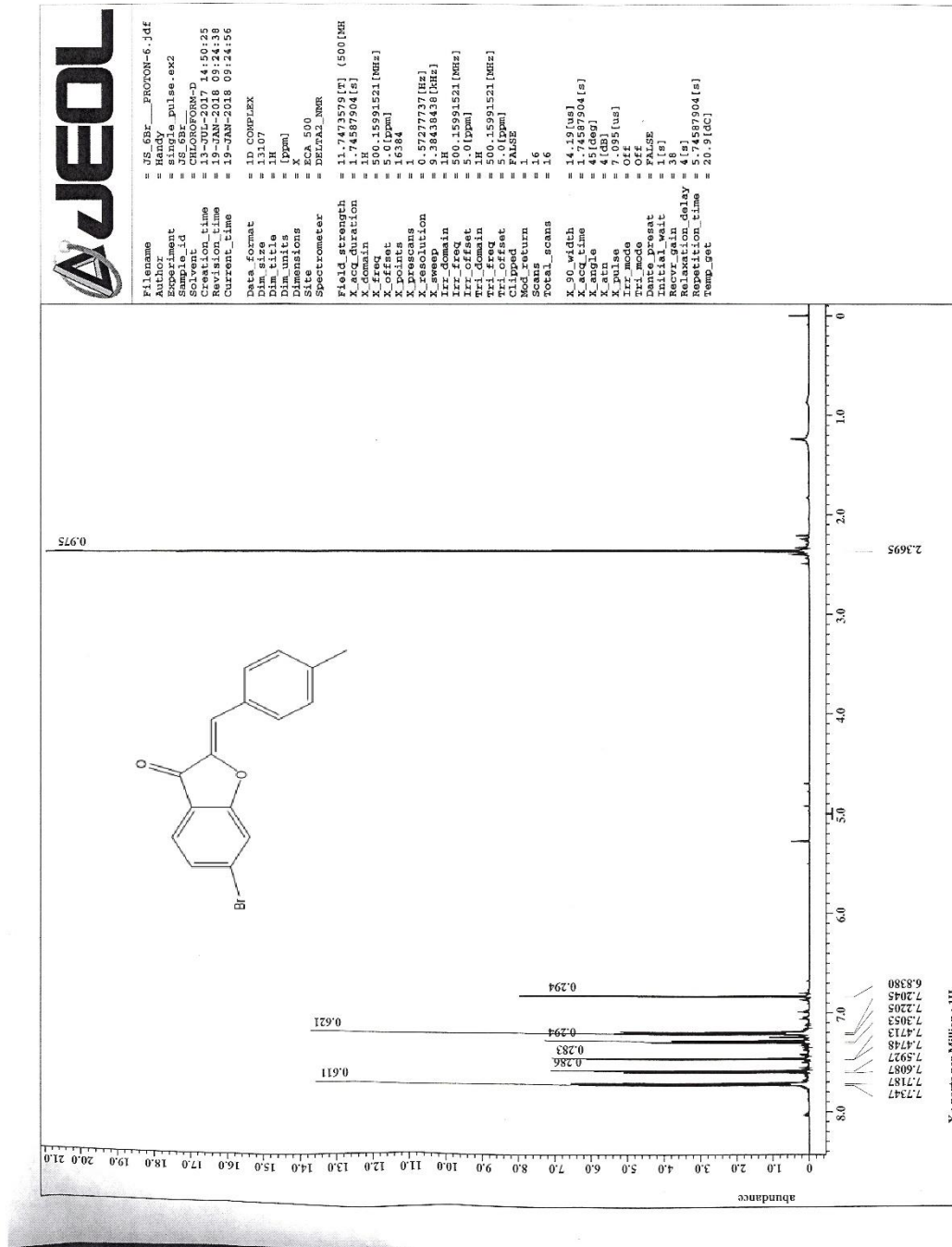


Spectrum

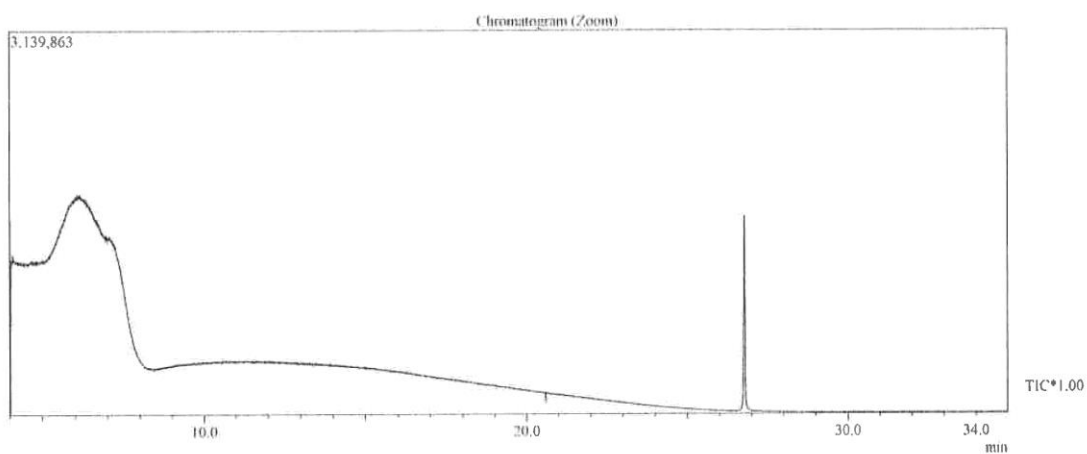
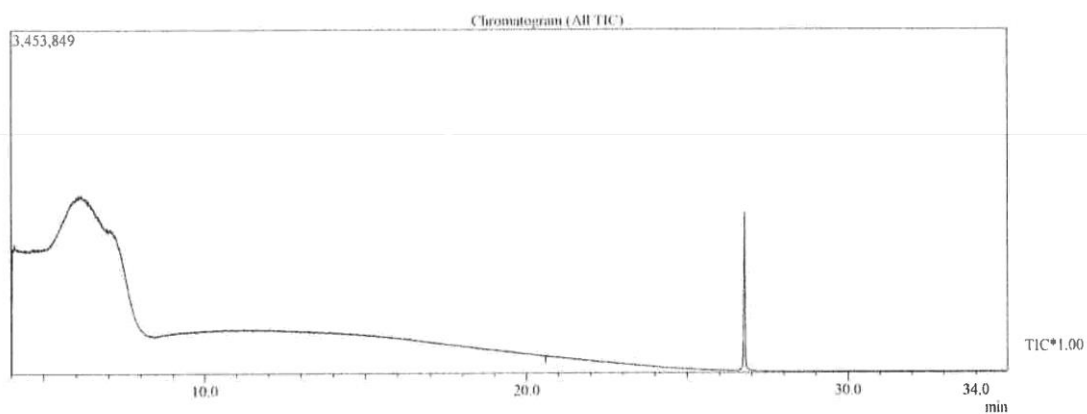
Line#:1 R.Time:26.900(Scan#:4581)
MassPeaks:82
RawMode:Single 26.900(4581) BasePeak:63.00(16604)
BG Mode:None Group 1 - Event 1 Scan



2-(4-methylbenzylidene)-1-(6'-bromobenzofuran-3-one) [9]

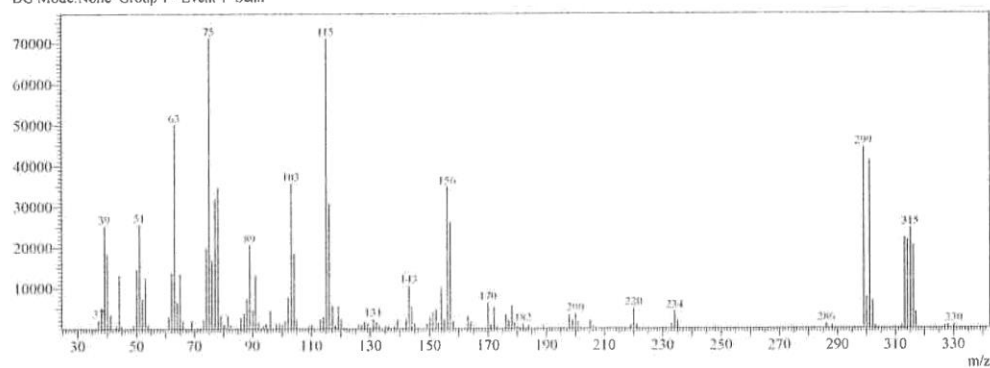


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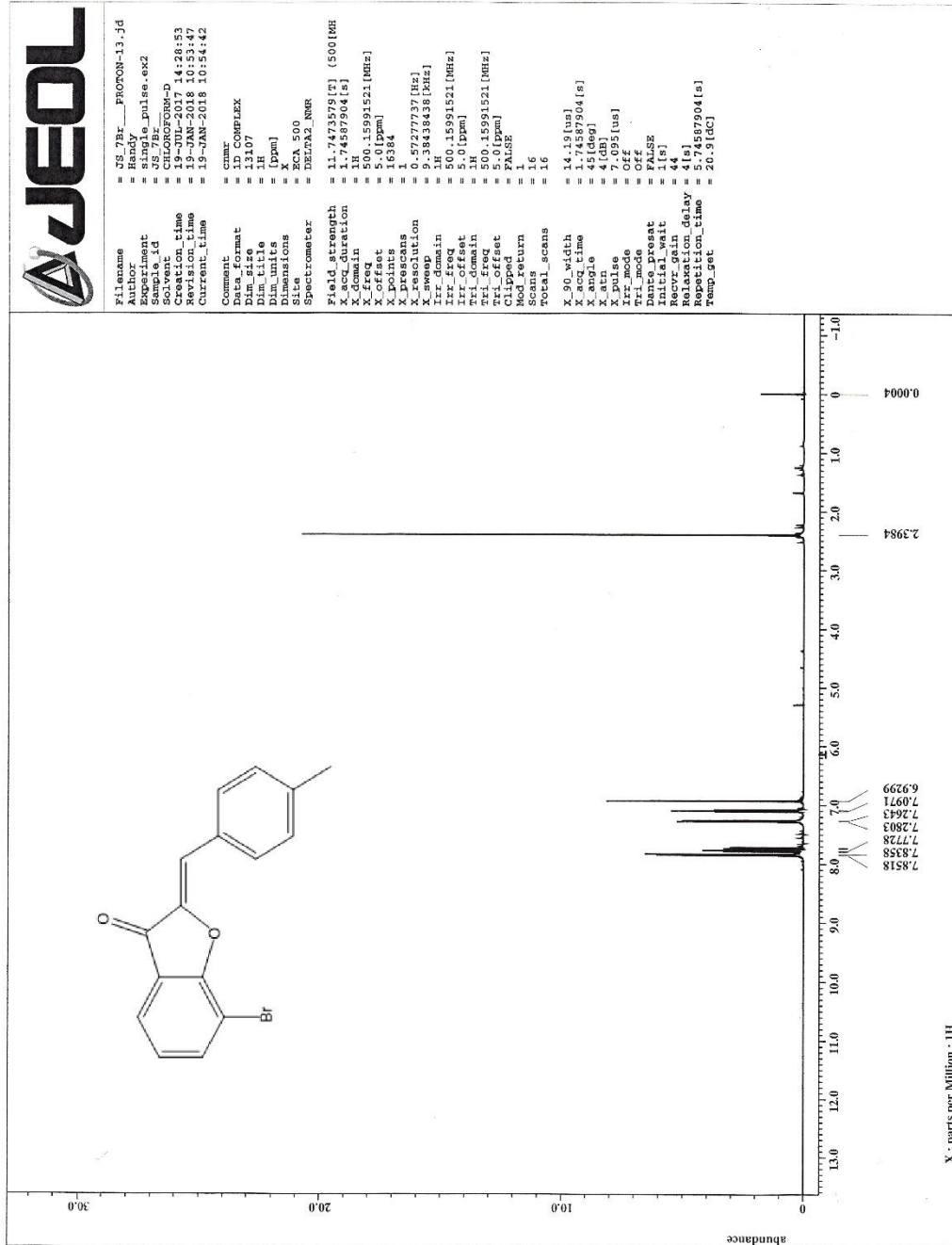


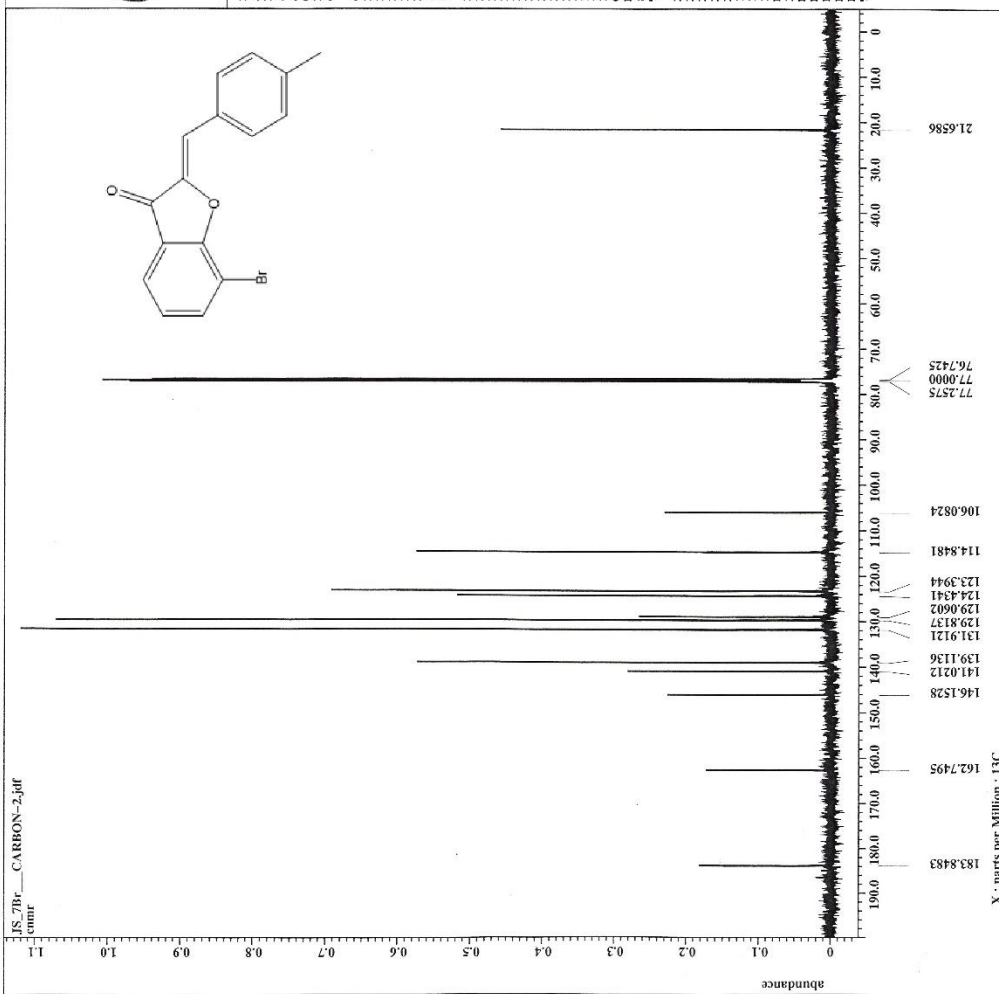
Spectrum

Line#:1 R.Time:26.765(Scan#:4554)
MassPeaks:127
RawMode:Single 26.765(4554) BasePeak:75.05(71248)
BG Mode:None Group 1 - Event 1 Scan



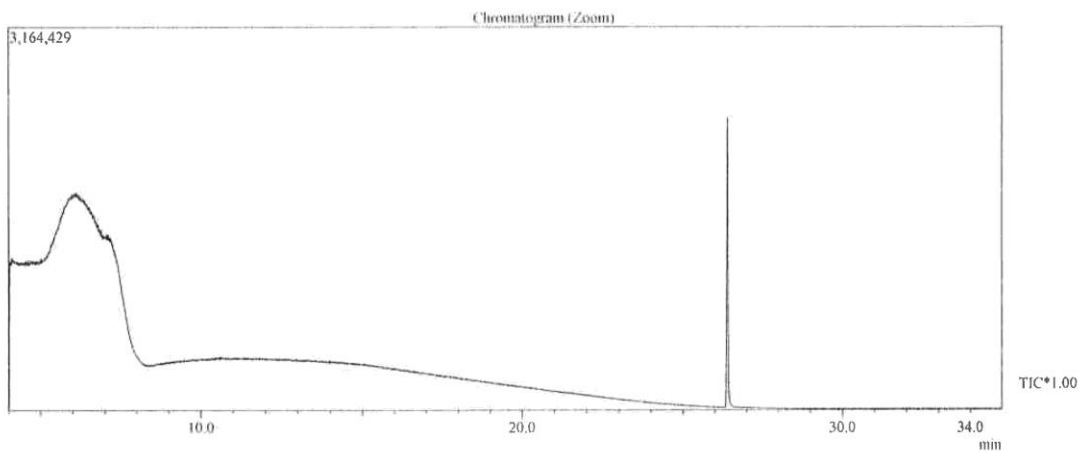
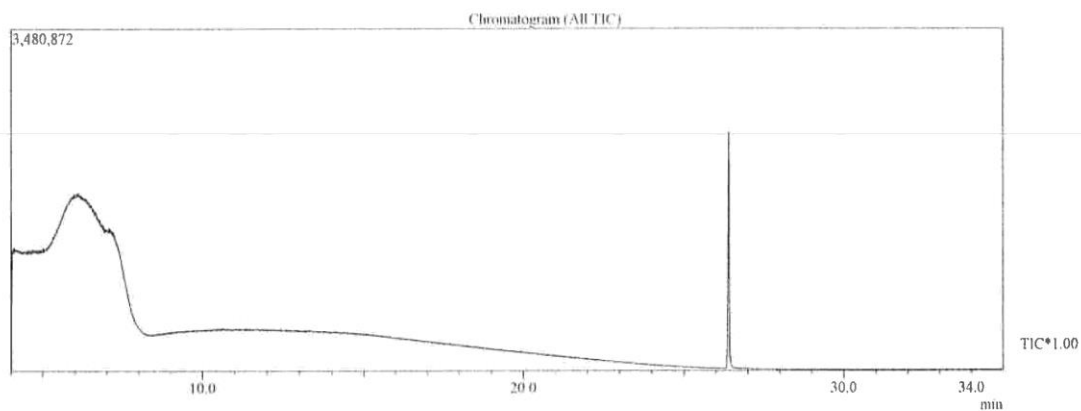
2-(4-methylbenzylidene)-1-(7'-bromobenzofuran-3-one) [10]





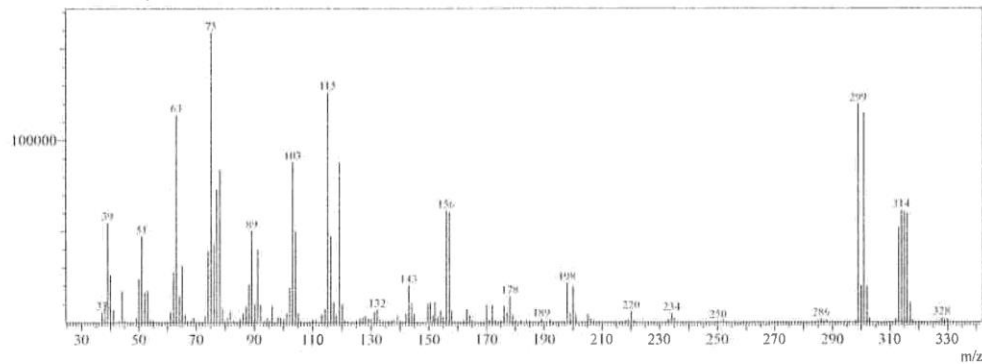
Filename = JS-7Br_CARBON-2.jdf
Author = Randy
Experiment = pulse_dec
Sample_id = JS-7Br
Solvent = CHLOROFORM-D
Creation_time = 19-JUL-2017 14:48:15
Acquisition_time = 19-JUL-2017 15:17:18
Current_time = 19-JUL-2017 15:07:26
Comment = C13C
Data_format = 1D COMPLEX
Dim_1 = 64
Dim_2 = 13C
Dim_units = [ppm]
Dimensions = X
Site = XA 500
Spectrometer = ECP32_NMR
Field_strength = 11.7473579[G] (500[MH
X_acq_duration = 0.83361792[s]
X_acq_time = 0.83361792[s]
X_freq = 125.76529768[MHz]
X_offset = 100[ppm]
X_points = 32768
X_resolution = 1.19959034[Hz]
X_sweep = 39.3081761[MHz]
X_domain = 1H
X_freq = 500.15991521[MHz]
X_offset = 0[ppm]
Clipped = TRUE
Mod_return = 1
Scans = 337
Total_scans = 337
X_90_width = 10.239[us]
X_acq_time = 0.83361792[s]
X_angle = 30[deg]
X_pulse = 3.413[us]
Irr_atn_dec = 21.5[db]
Irr_atn_noe = 21.5[db]
Decoupling = TRUE
Initial_wait = 1[s]
Nuc = 13C
Nuc_freq = 125.76529768[MHz]
Repetitions = 20
Relaxation_delay = 2[s]
Repetition_time = 2.83361792[s]
Temp_get = 21.3[deg]

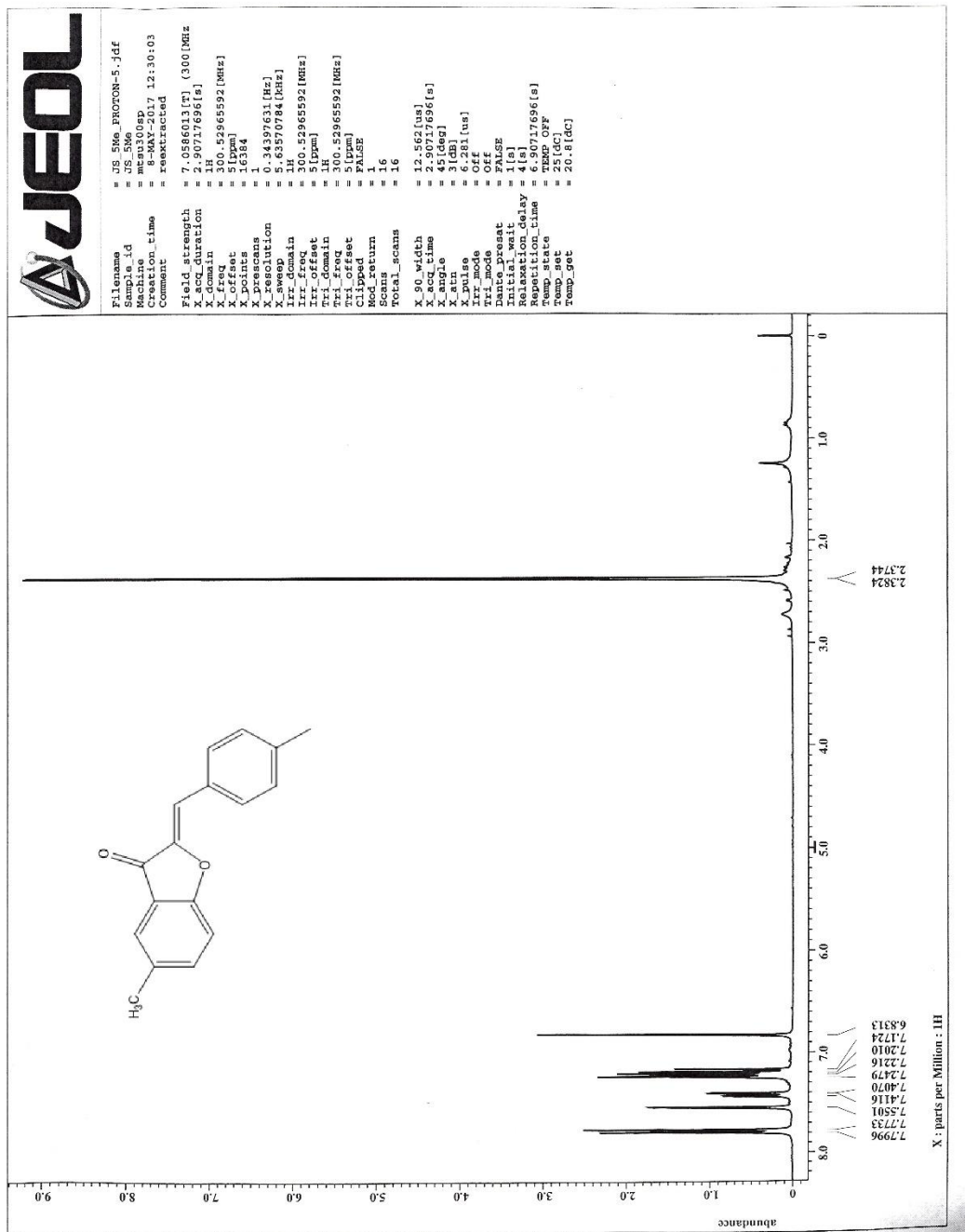
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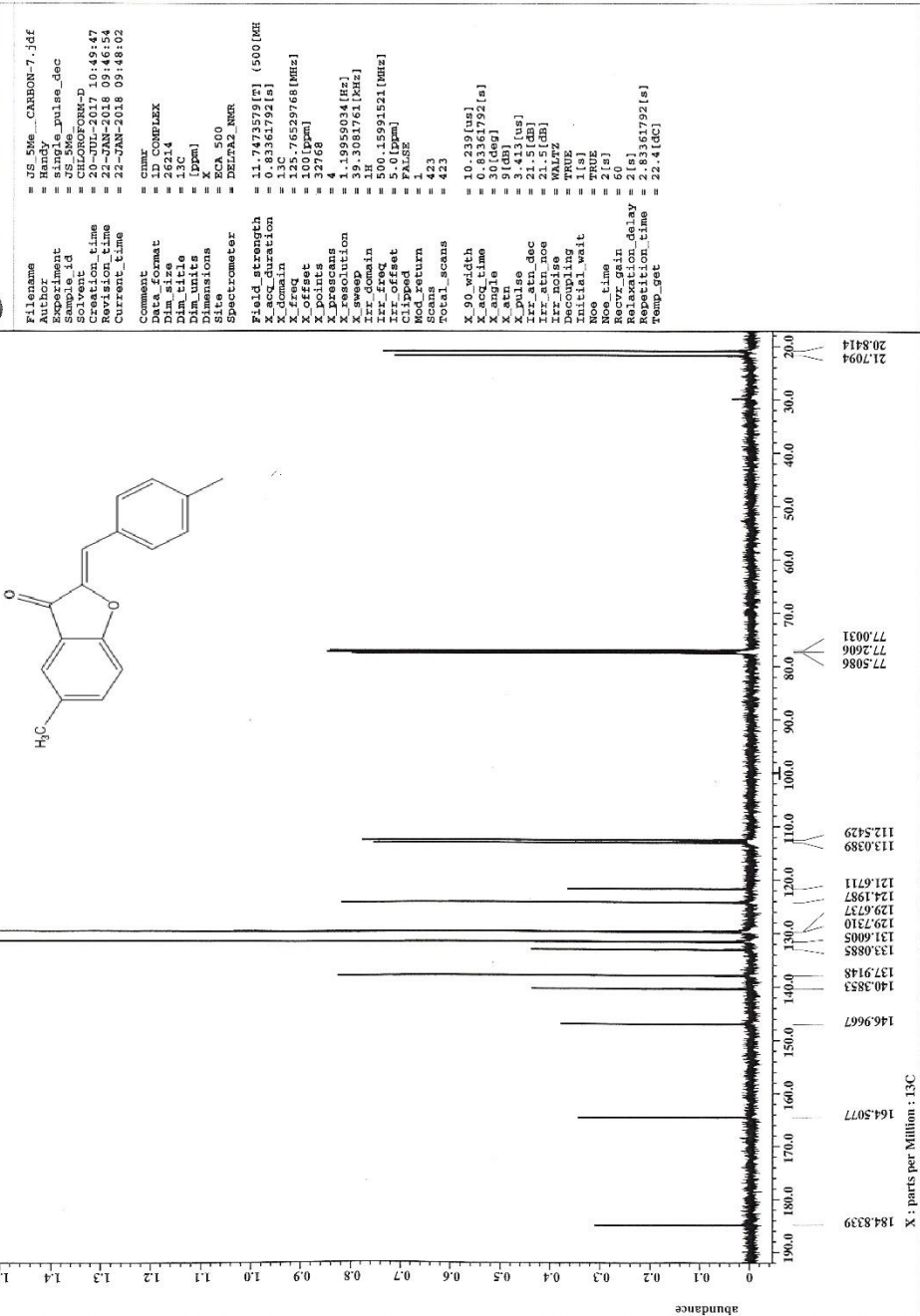


Spectrum

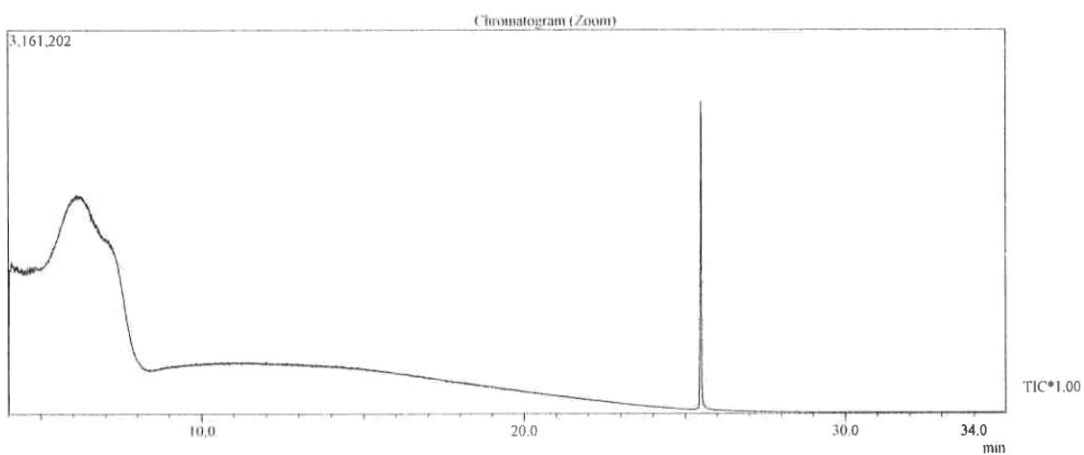
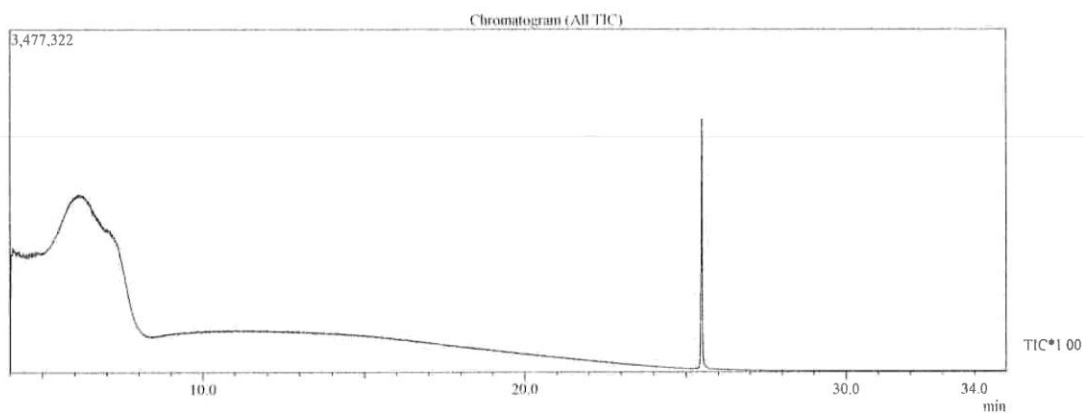
Line1:1 R.Time:26.405(Scan#:4482)
MassPeaks:157
RawMode:Single 26.405(4482) BasePeak:75.05(158668)
BG Mode:None Group 1 - Event 1 Scan



2-(4-methylbenzylidene)-1-(5'-methylbenzofuran-3-one) [**11**]

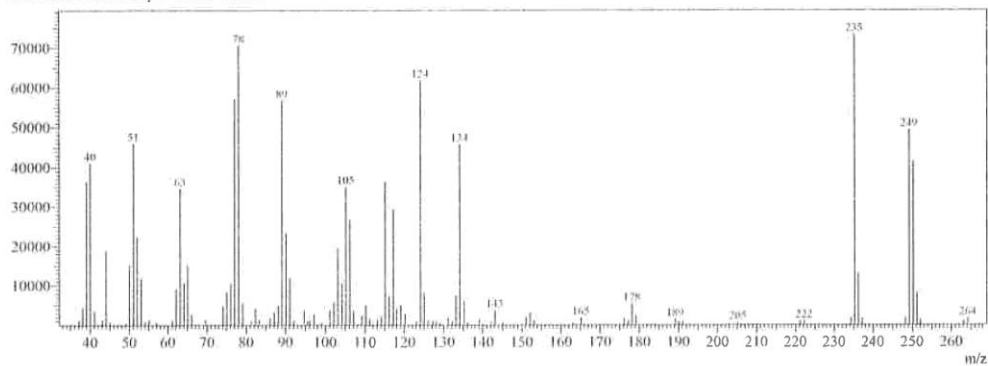


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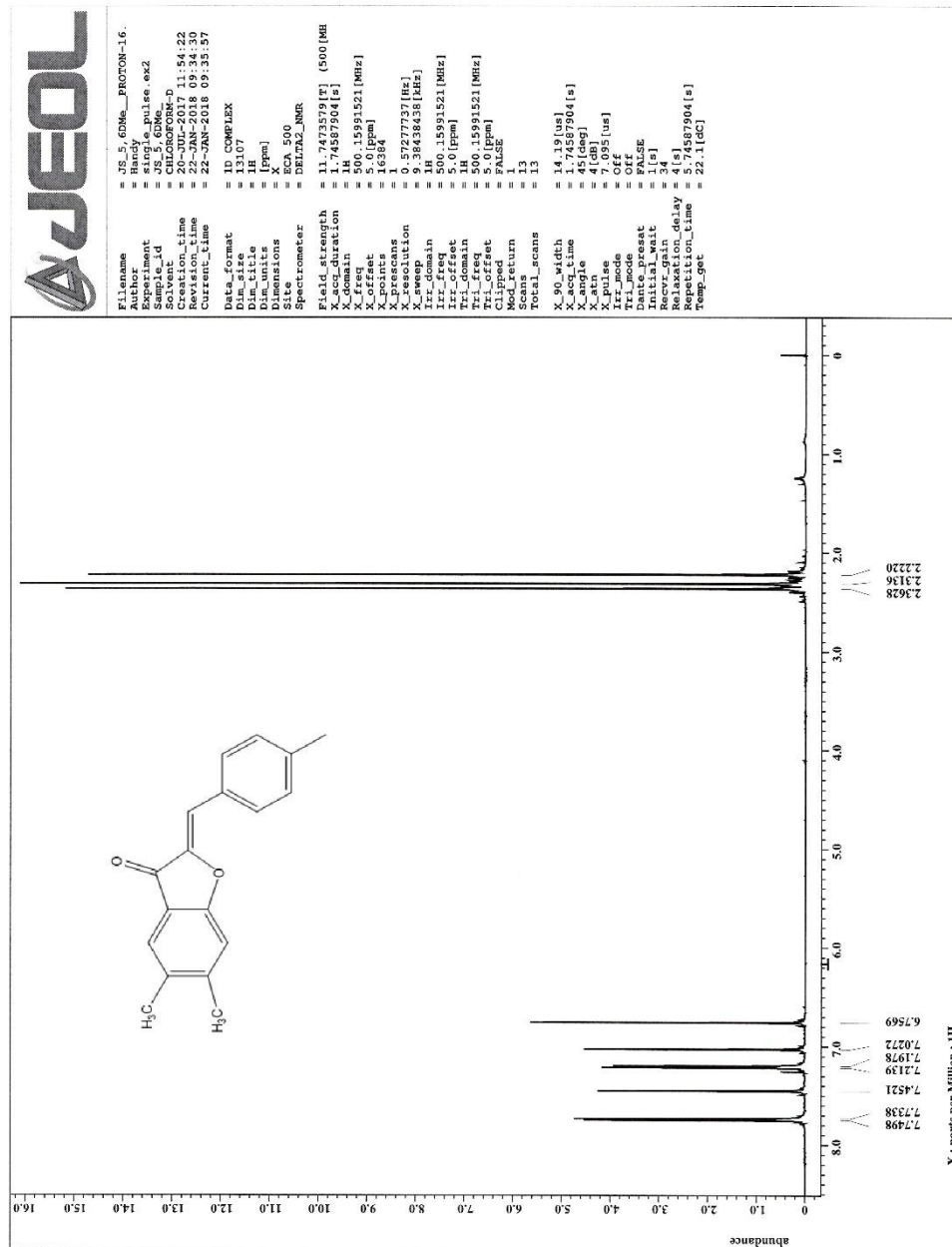


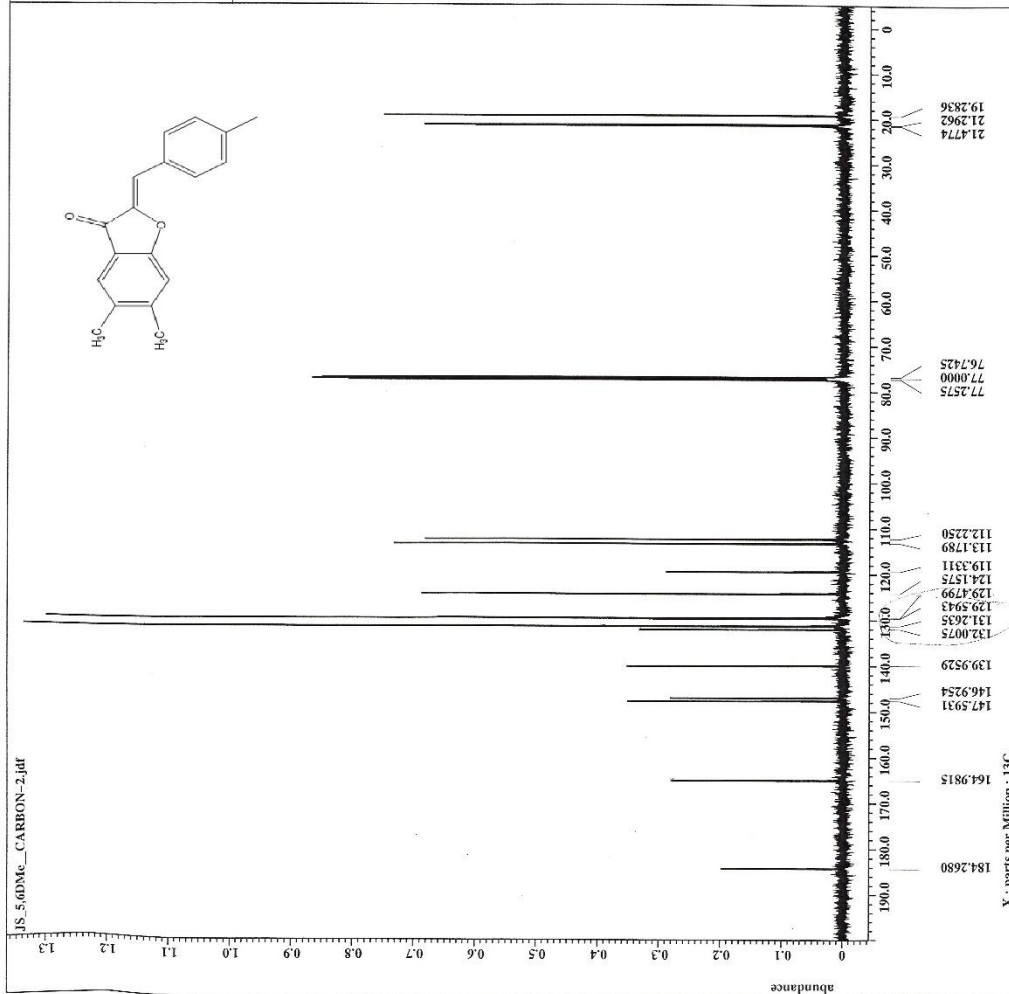
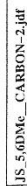
Spectrum

Line#:1 R.Time:25.505(Scan#:4302)
MassPeaks:105
RawMode:Single 25.505(4302) BasePeak:235.20(73345)
BG Mode:None Group 1 - Event 1 Scan



2-(4-methylbenzylidene)-1-(5',6'-dimethylbenzofuran-3-one) [12]



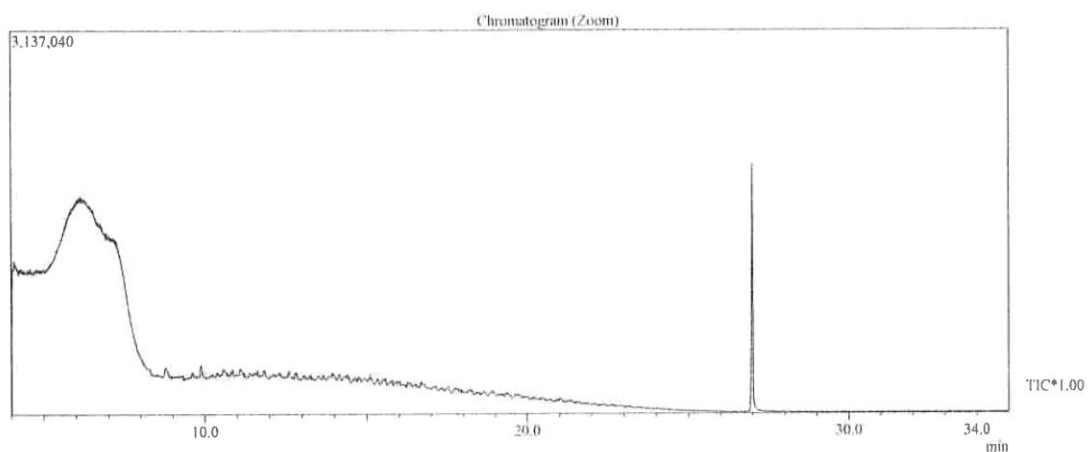
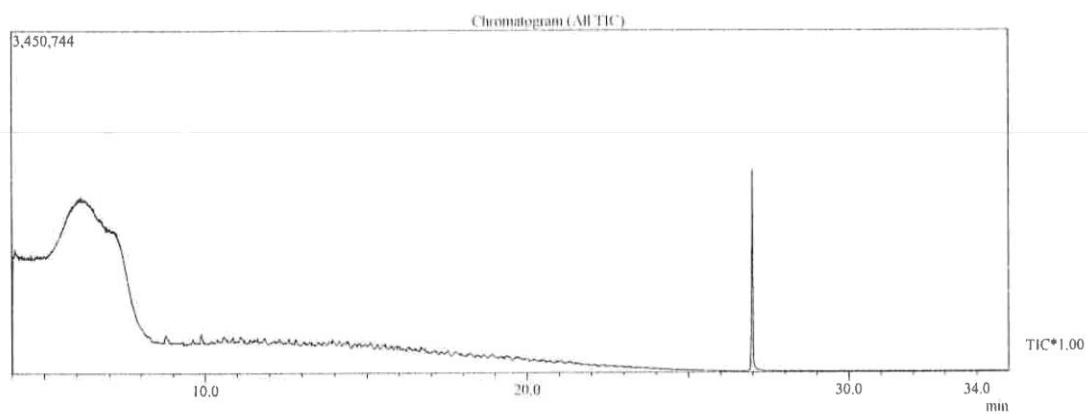


```

J5 = J5_6MHz_CARBON-2.3
Filesource
= Handy
Experiment
= single_pulse_dec
Sample_id
= J5_5_6MHz
Solvent
= CHLOROFORM-d
Time
= 12.04:07
Revision_time
= 20-YUG-2017 12:23:17
Current_time
= 20-YUG-2017 12:23:17
Data_format
= 1D_COMPLEX
Dim_size
= 262144
Fid
= 133C
Dimensions
= [ppm]
Dim0_units
= X
Dim1_units
= X
Spectrometer
= DELTA2X_NMR
Freq
= 18.7493579[7] (500[MH
Acq_duration
= 0.83361792[s]
Sweep_start_freq
= 125.76529768[MHz]
Sweep_end_freq
= 125.76529768[MHz]
X_freq
= 100[ppm]
Y_points
= 3768
Z_points
= 1
Prescans
= 1
Prg_name
= 13959034[Huz]
X_sweep
= 39.3081761[kHz]
Irz_domain
= 1H
Irr_freq
= 500.15991521[MHz]
Clamp_level
= 0.00[ppm]
Clipped_wet
= FALSE
Mod_return
= 1
Scans
= 131
Total_scans
= 131
X30_width
= 10.239[us]
X_acq_time
= 0.83361792[s]
X_angle
= 30[deg]
X_atn
= 9[db]
Xir_atn_dec
= 21.5[db]
Xir_atn_poc
= 21.5[db]
Xir_noise
= WAUTZ
Decoupling
= 1UPUR
Nucleus1_name
= TRUE
Nuc1_gamma
= 2[S]
Recvr_gain
= 60
Temp
= 22.4[degC]
Repetition_time
= 22.4[degC]
Reset_time
= 22.4[degC]

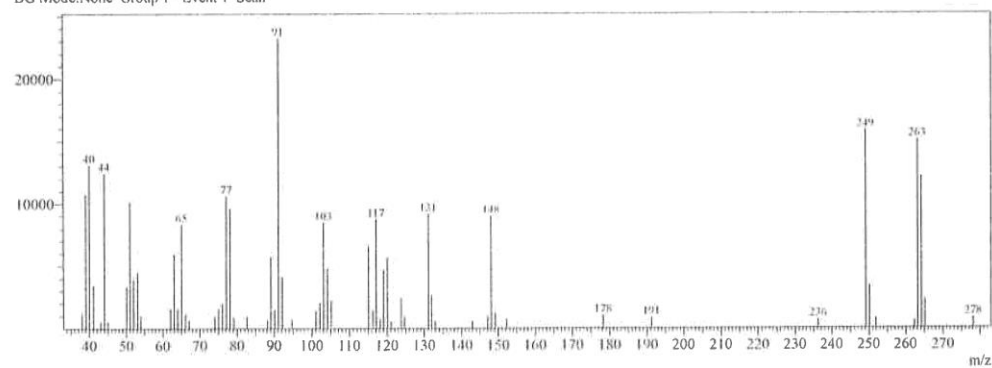
```

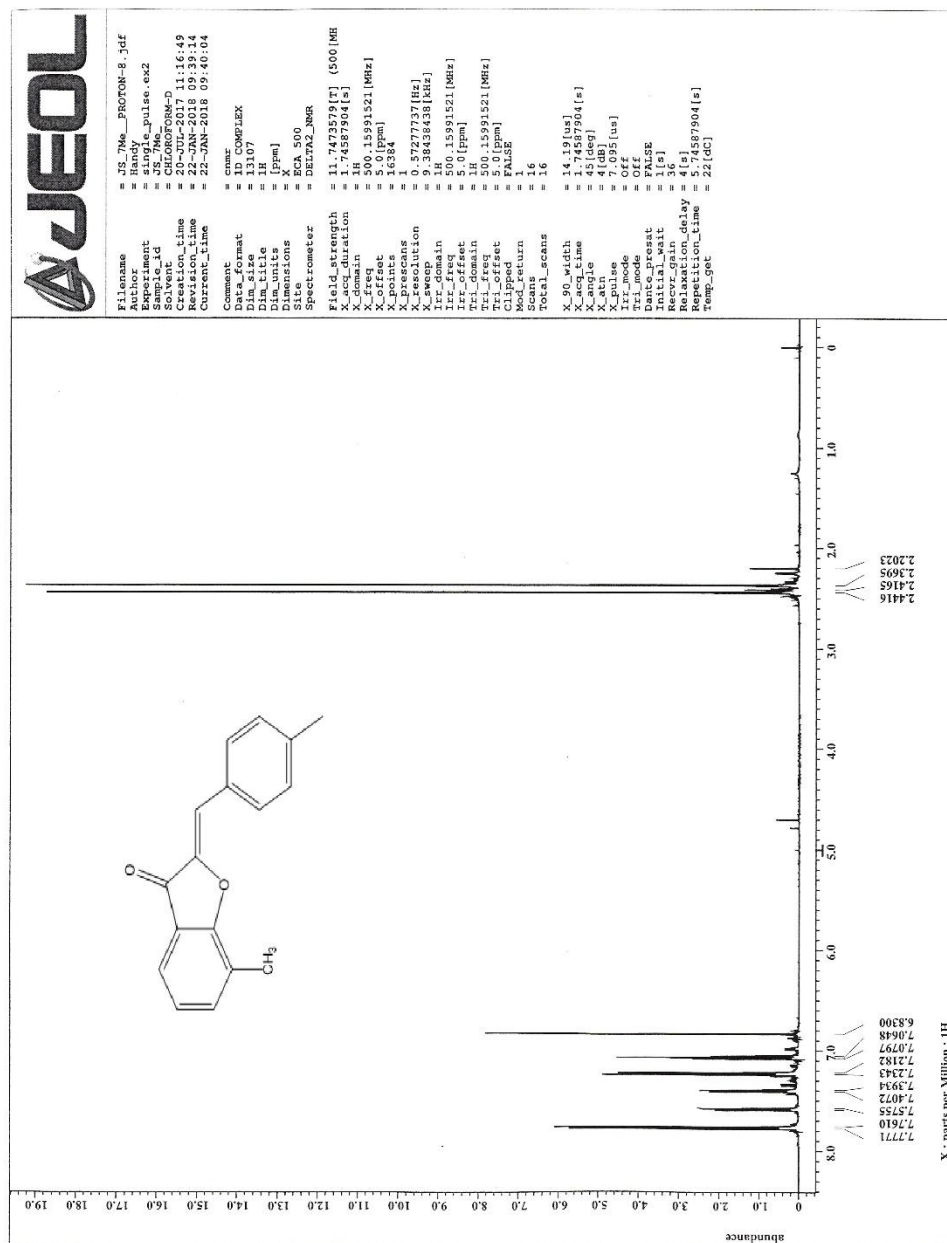
C:\Sync\ShimadzuGCMS\Documents\Handy\Schmitt\5,6 dMe.qgd

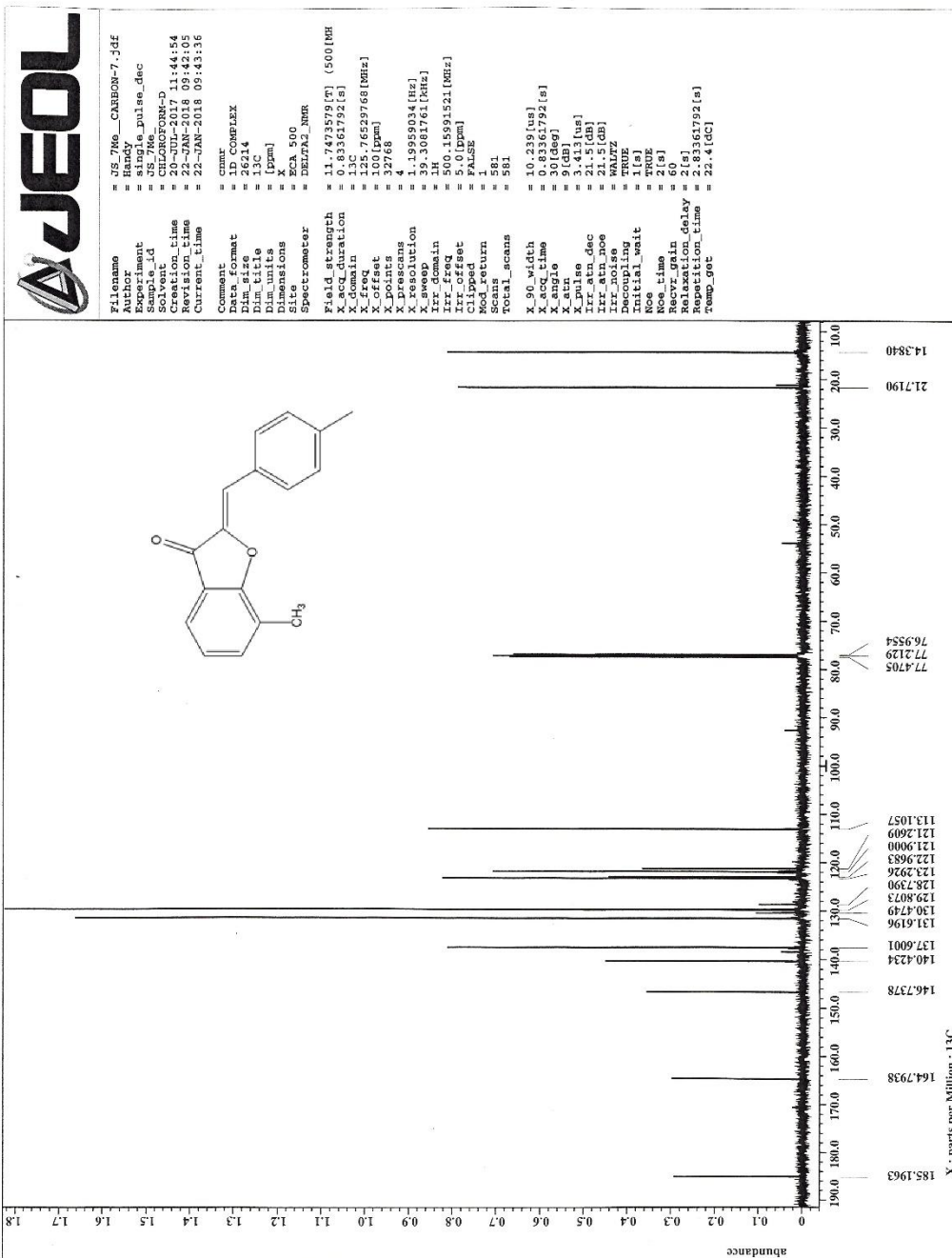


Spectrum

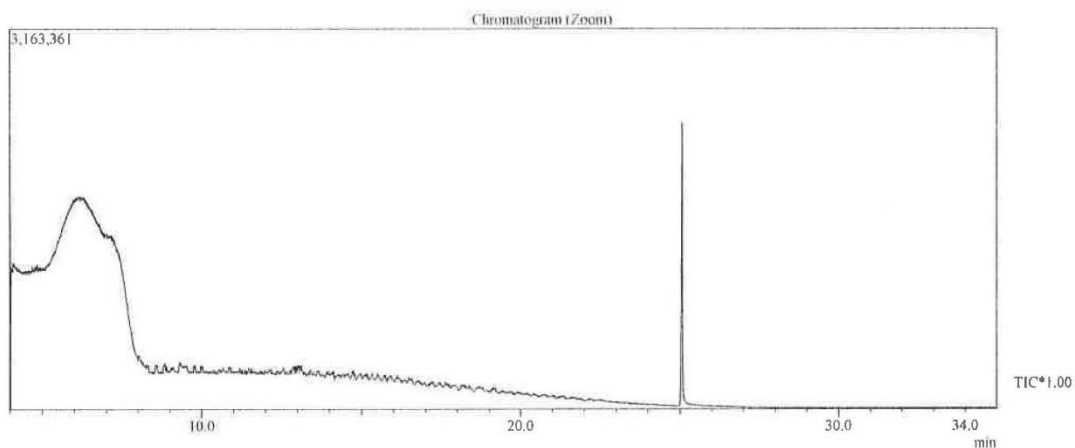
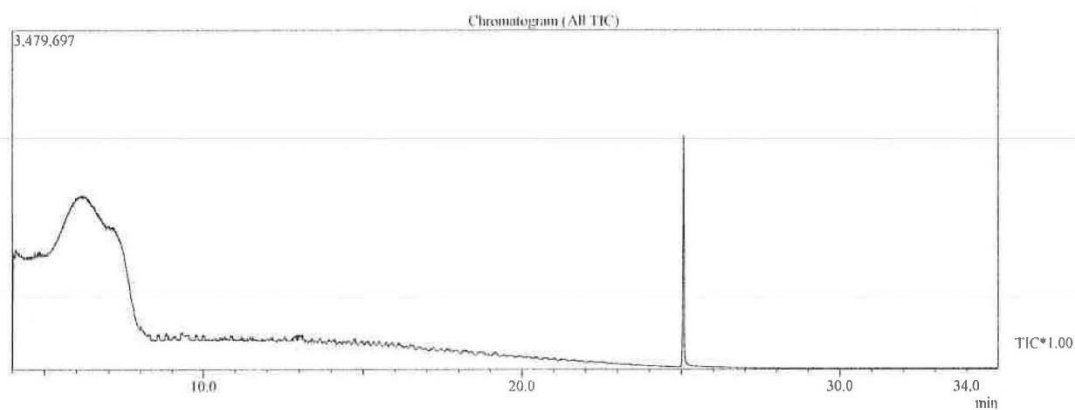
Line#:1 R Time:27.015(Scan#:4604)
MassPeaks:64
RawMode:Single 27.015(4604) BasePeak:91.10(23252)
BG Mode:None Group 1 - Event 1 Scan



2-(4-methylbenzylidene)-1-(7'-methylbenzofuran-3-one) [**13**]

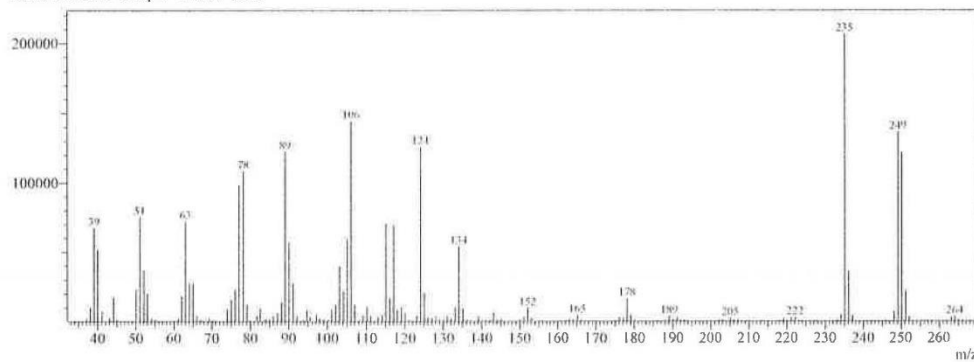


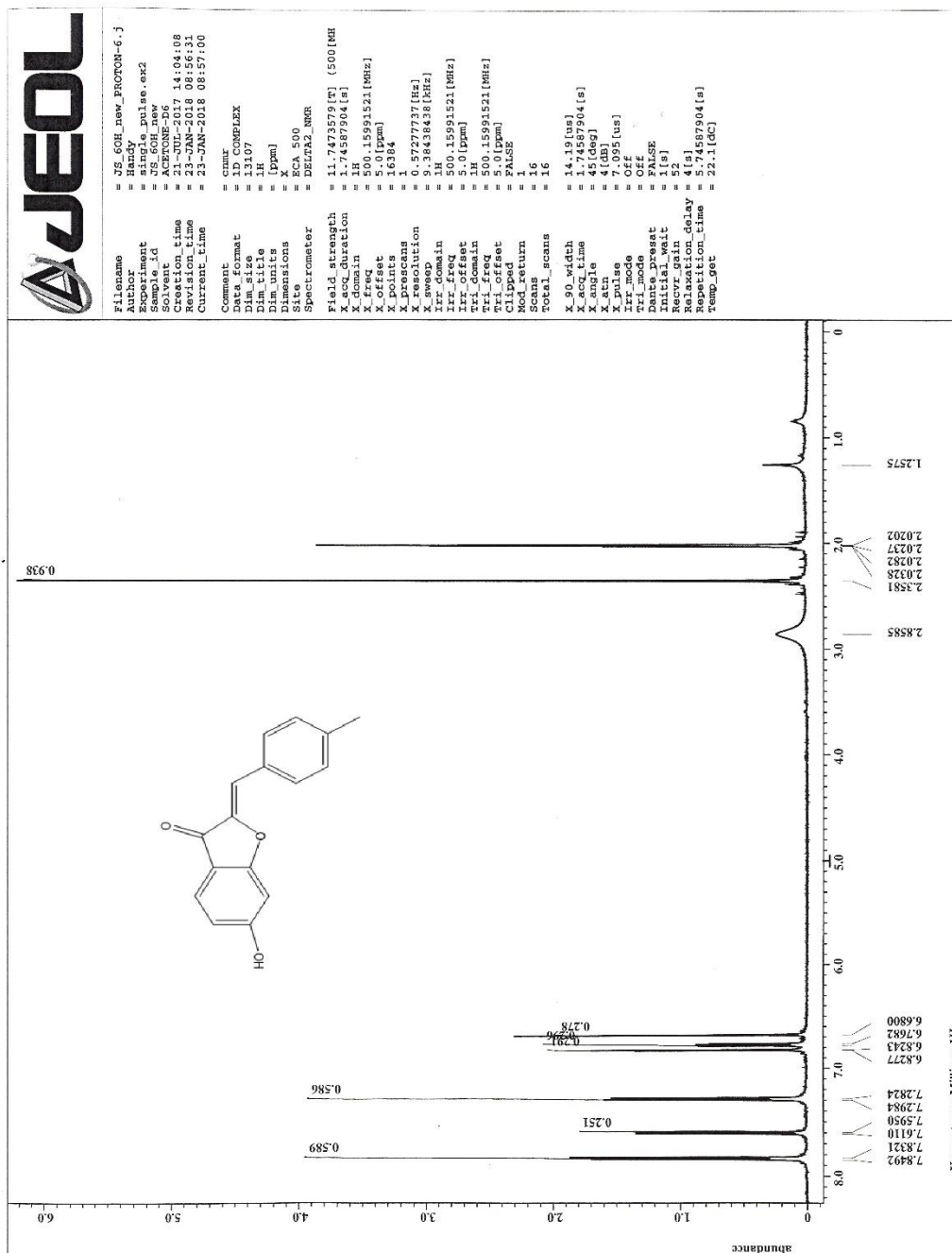
C:\Sync\ShimadzuGCMS\Documents\Handy\Schmitt\7Me.gpd

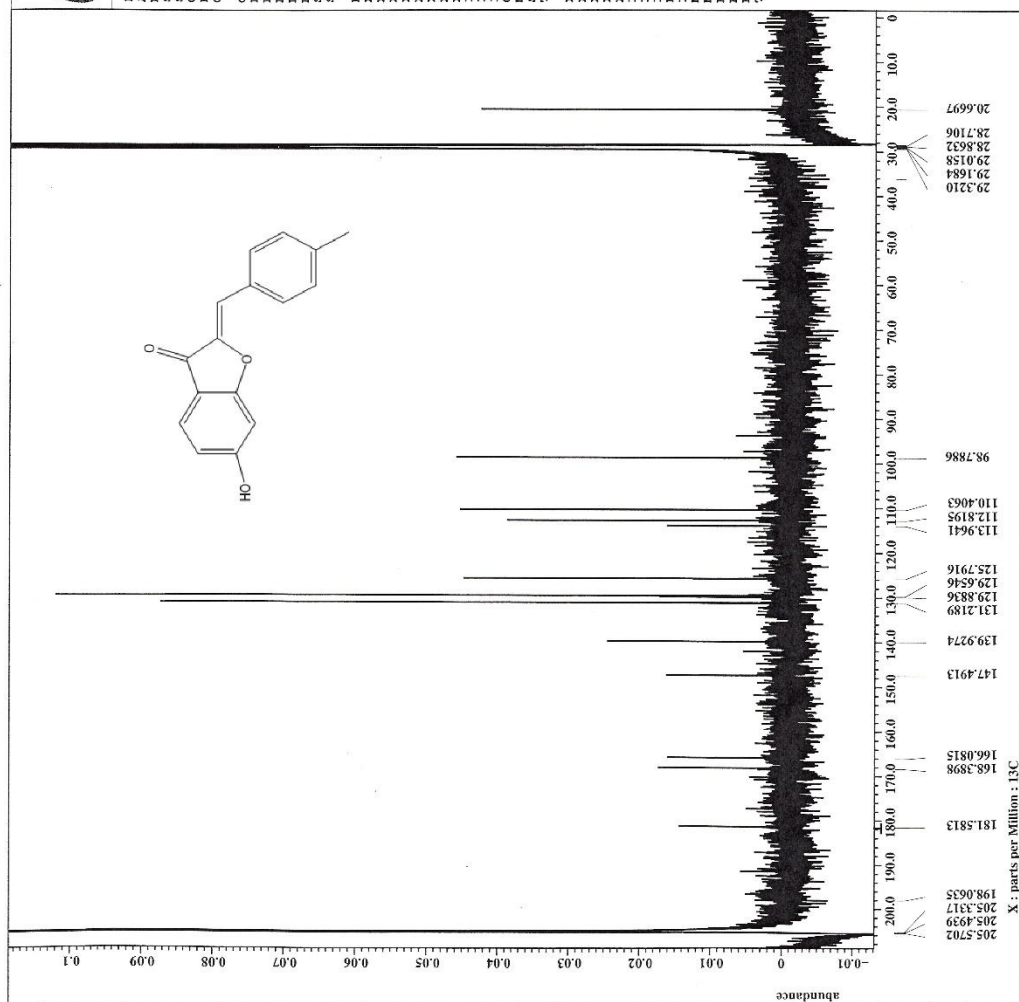


Spectrum

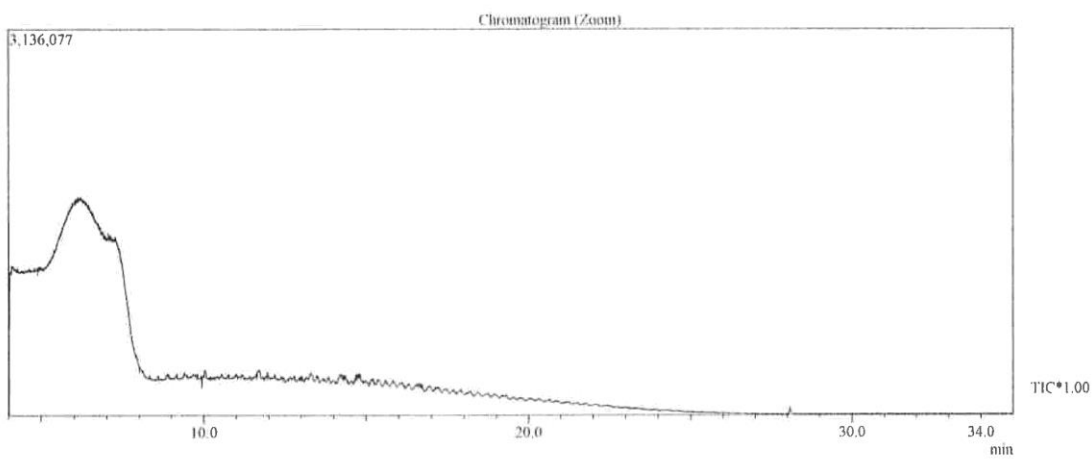
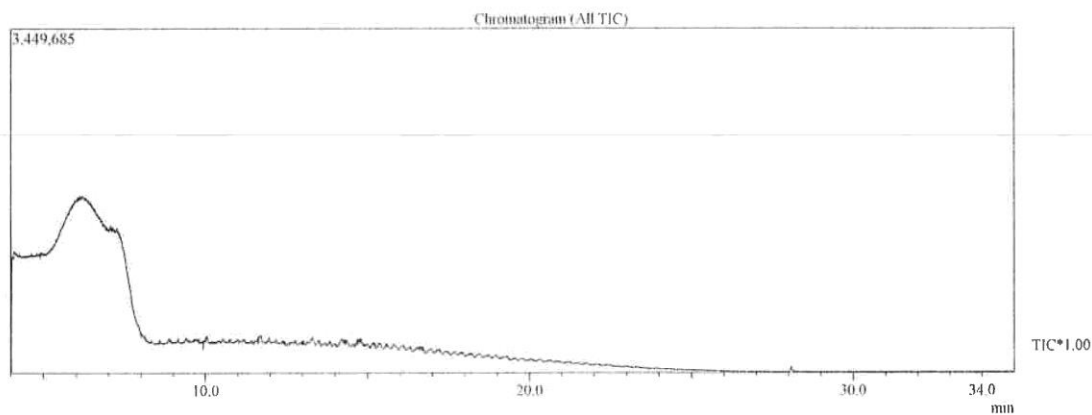
Line#:1 R.Time:25.080(Scan#:4217)
MassPeaks:135
RawMode:Single 25.080(4217) BasePeak:235.10(206370)
BG Mode:None Group 1 - Event 1 Scan



2-(4-methylbenzylidene)-1-(6'-hydroxybenzofuran-3-one) [**14**]

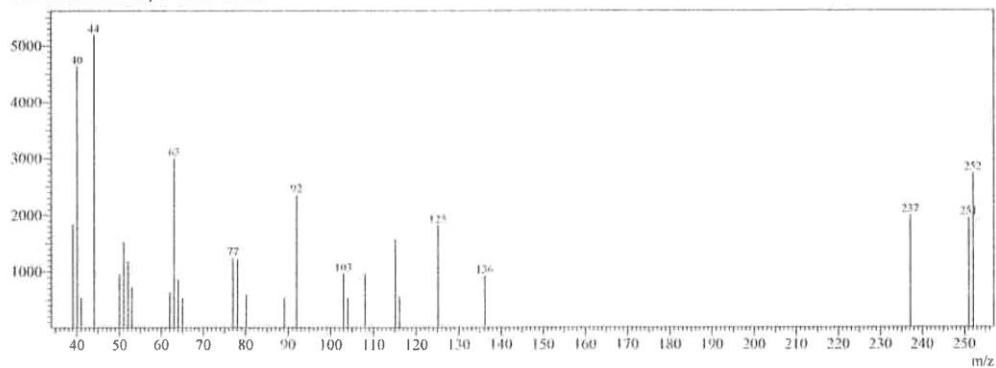


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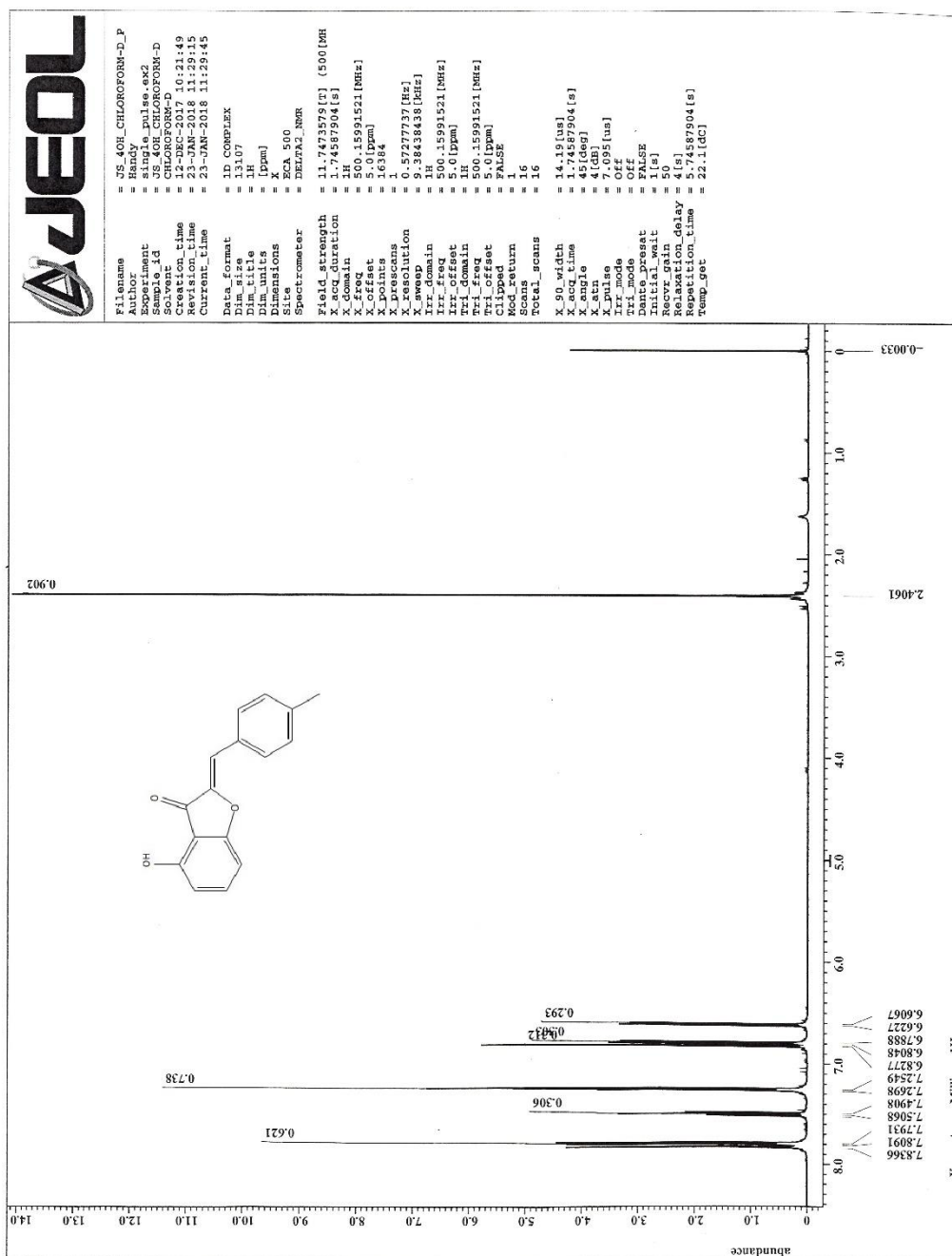


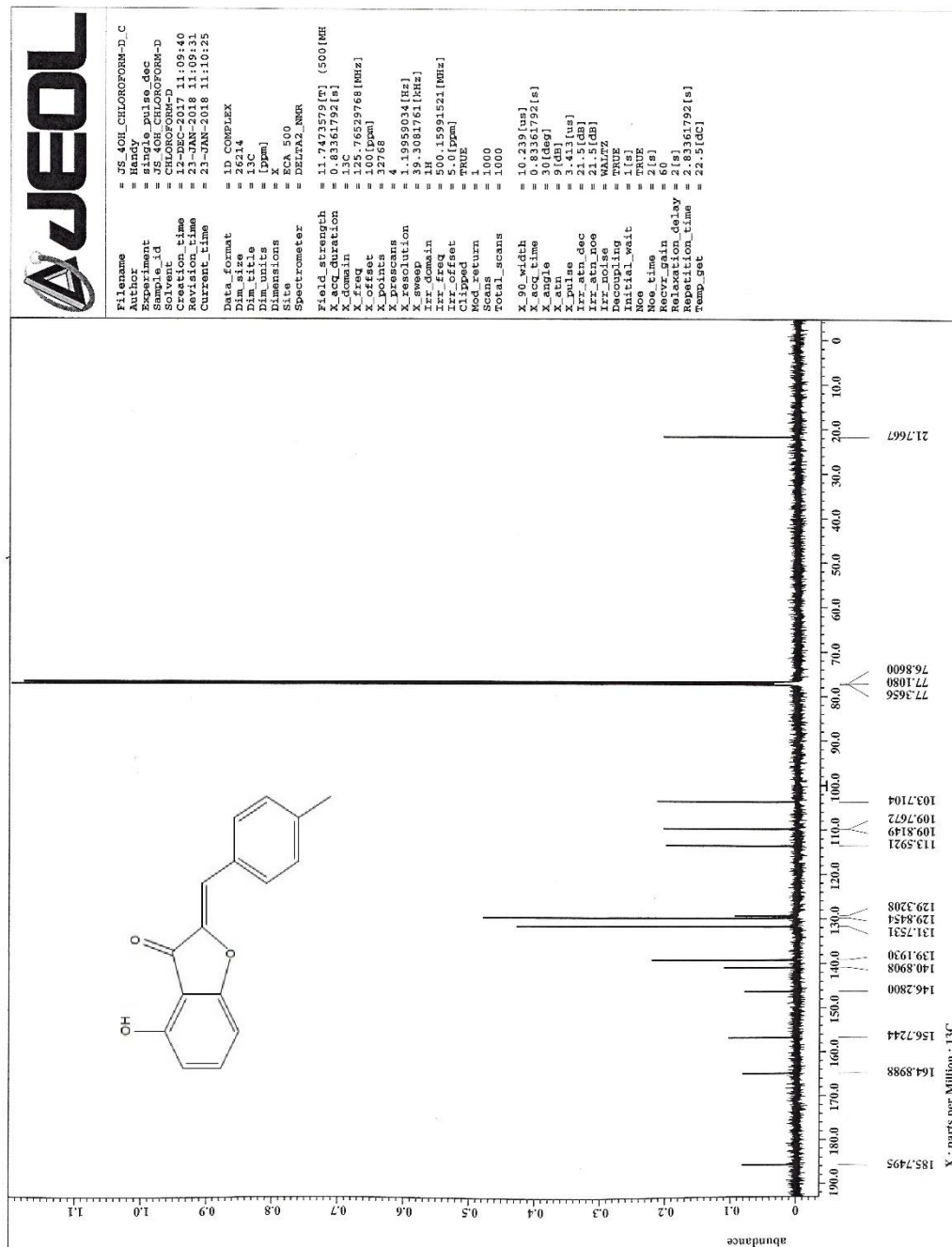
Spectrum

Line#:1 R.Time:28.090(Scan#:4819)
MassPeaks:27
RawMode:Single 28.090(4819) BasePeak 44.05(5186)
BG Mode:None Group 1 - Event 1 Scan

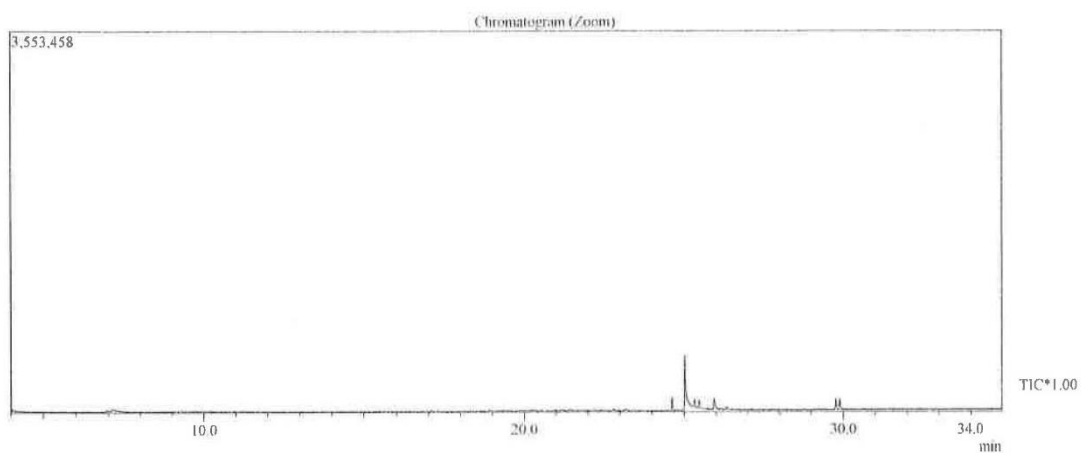
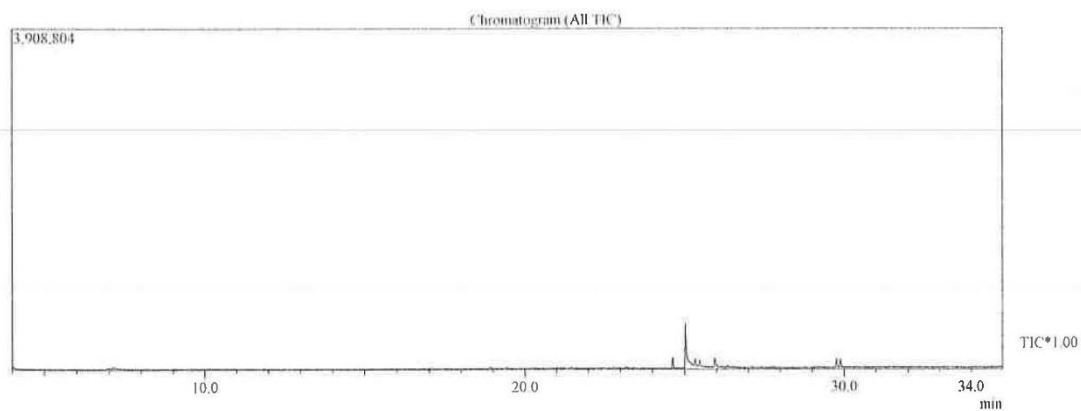


2-(4-methylbenzylidene)-1-(4'-hydroxybenzofuran-3-one) [15]



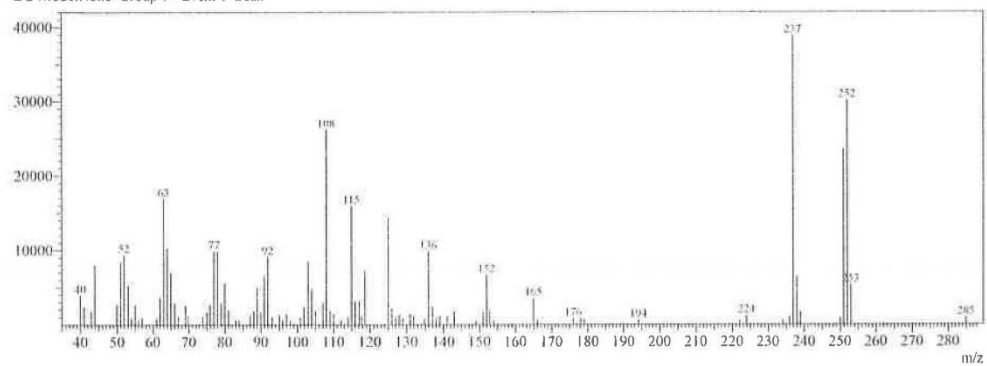


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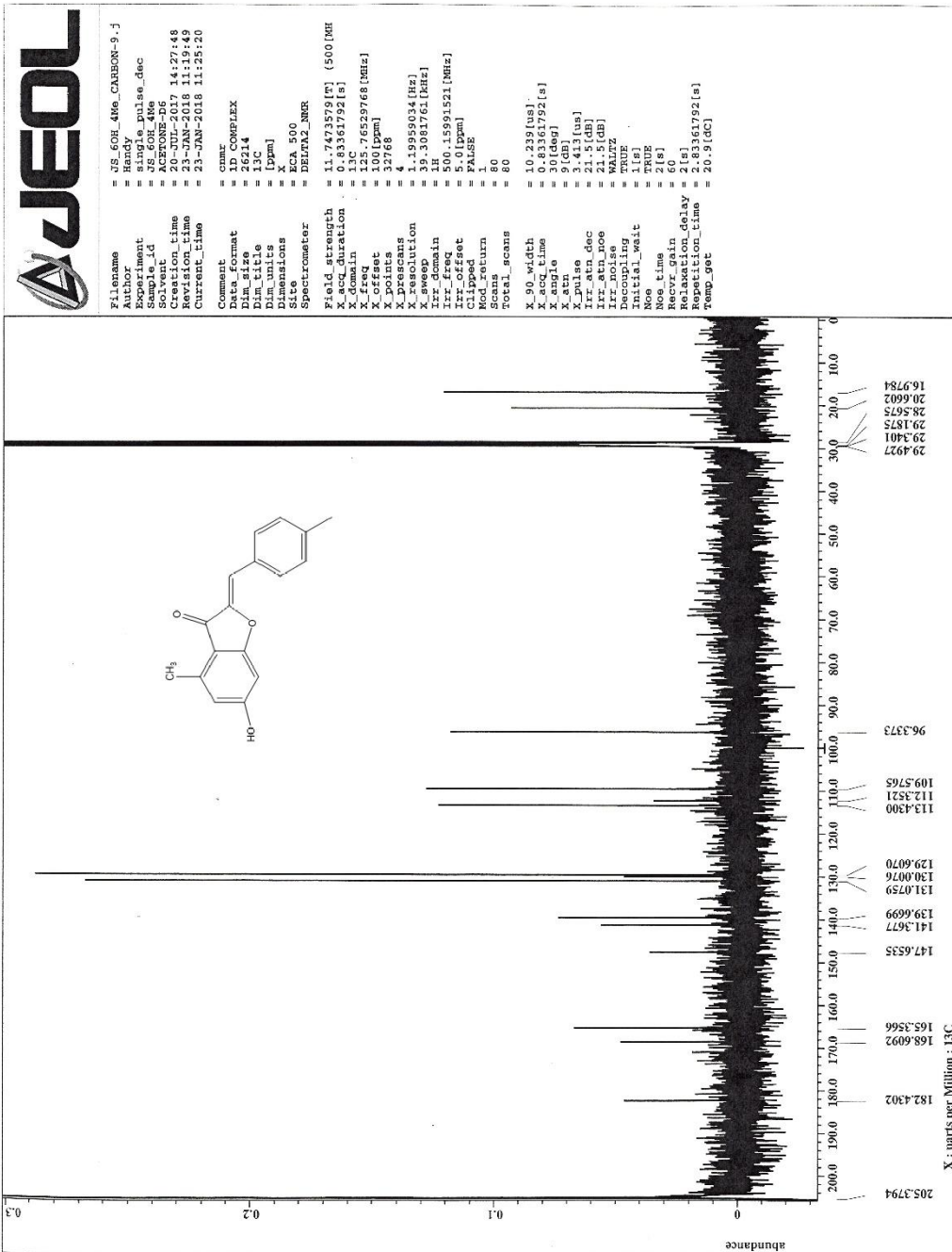
Spectrum

Line#:1 R.Time:25.035(Scan#:4208)
MassPeaks:95
RawMode:Single 25.035(4208) BasePeak:236.90(38782)
BG Mode:None Group 1 - Event 1 Scan

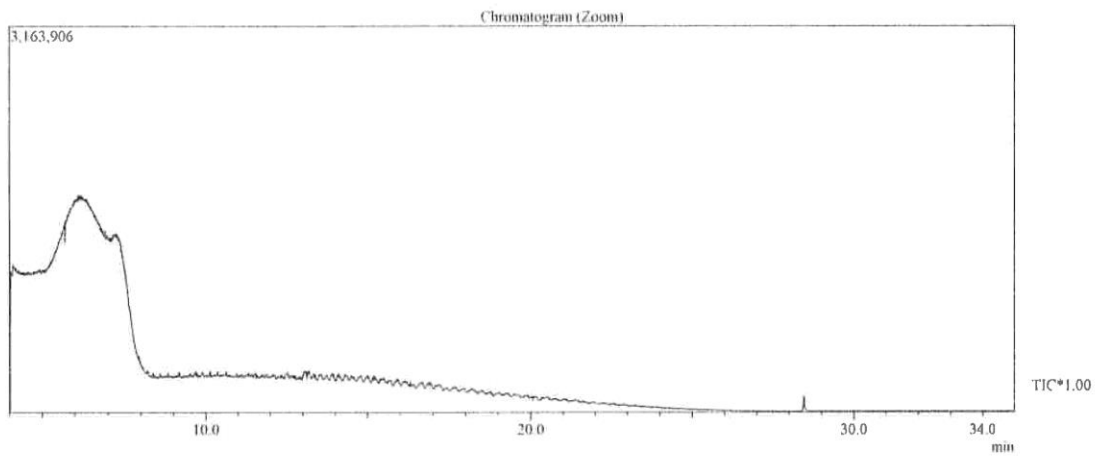
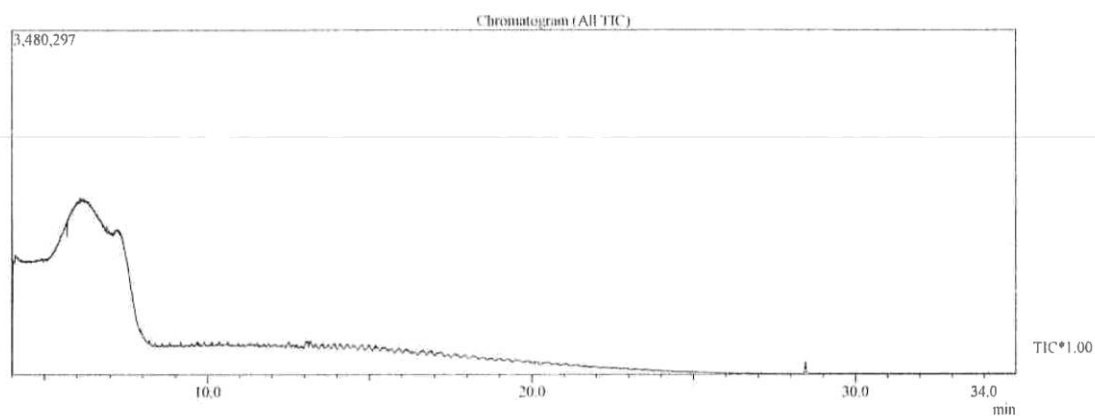


JOE



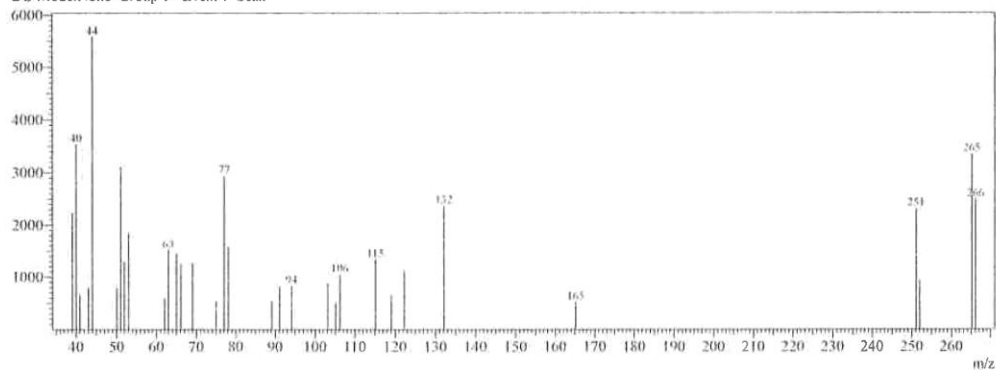


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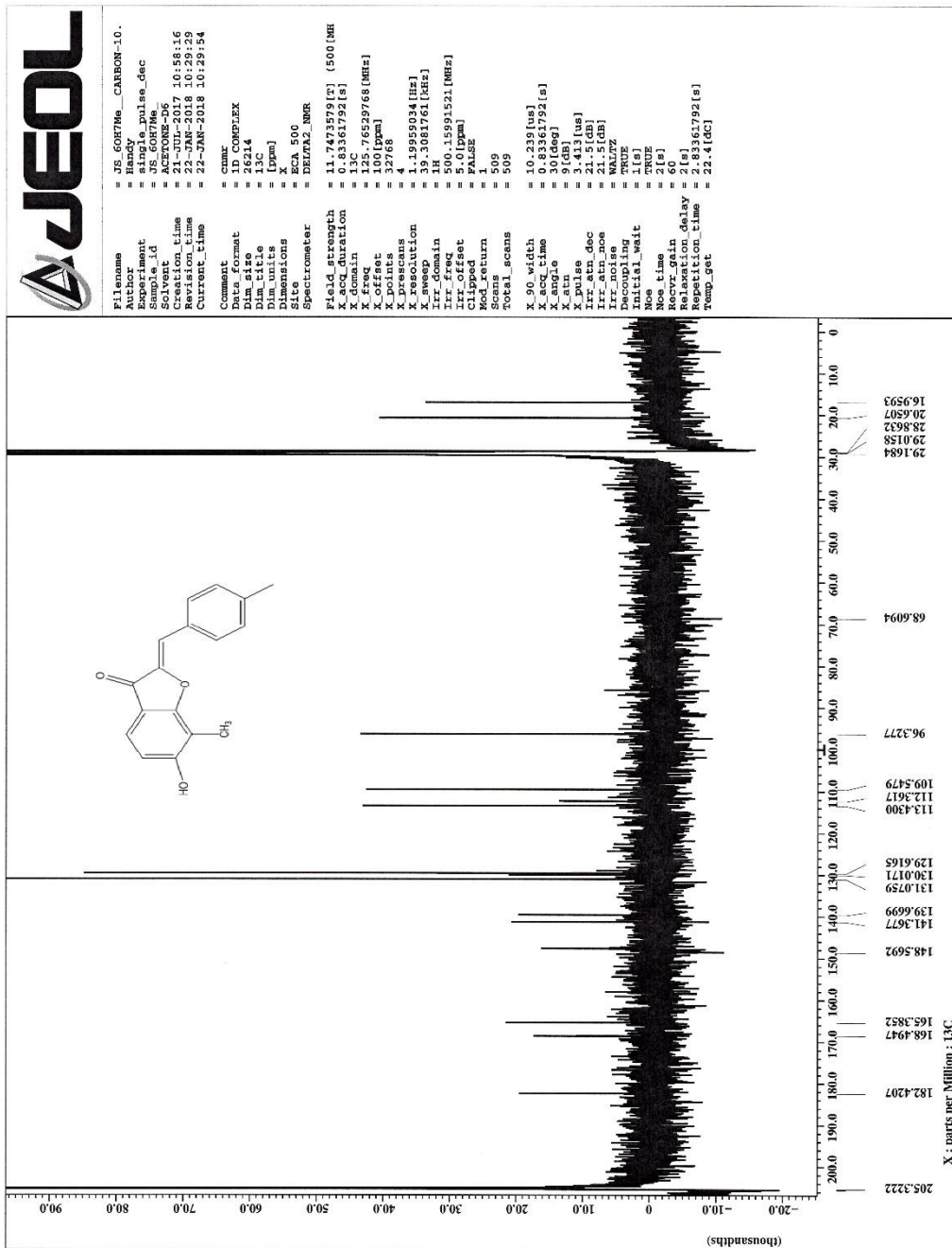


Spectrum

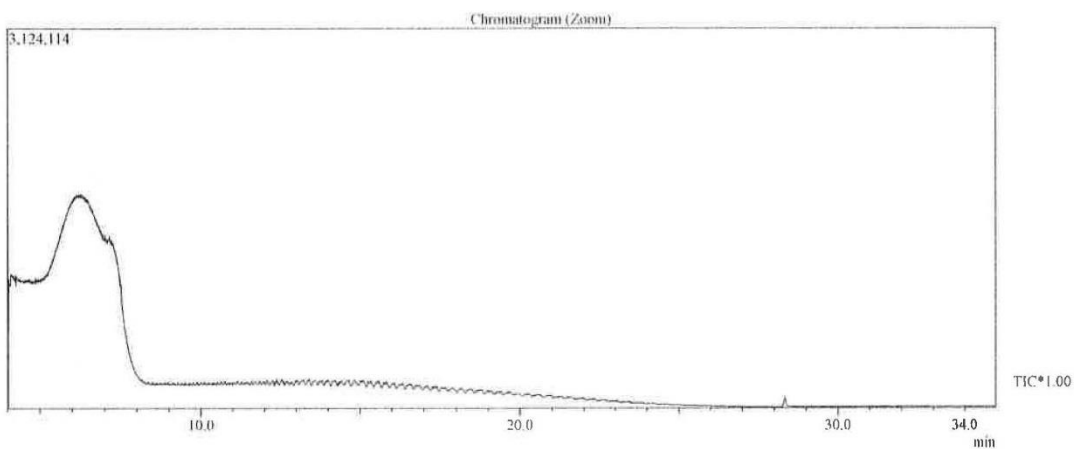
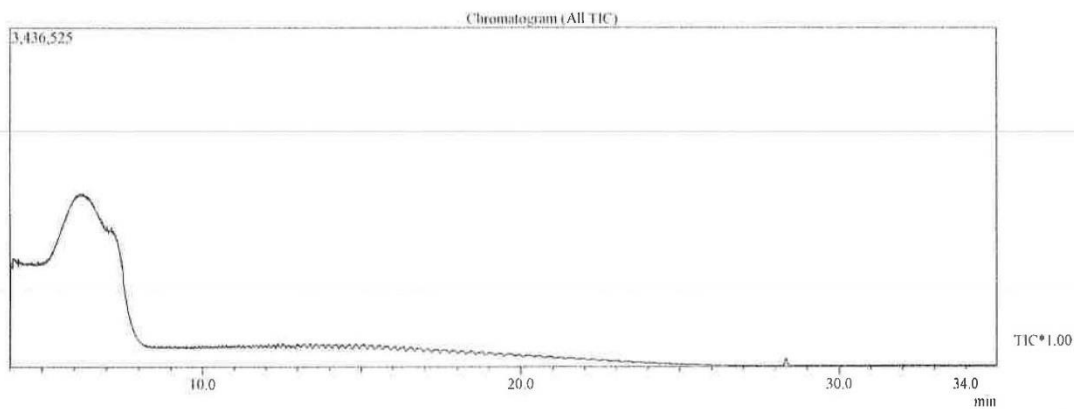
Line#:1 R.Time:28.475(Scan#:4896)
MassPeaks:32
RawMode:Single 28.475(4896) BasePeak:44.05(5575)
BG Mode:None Group 1 - Event 1 Scan



[illegible]

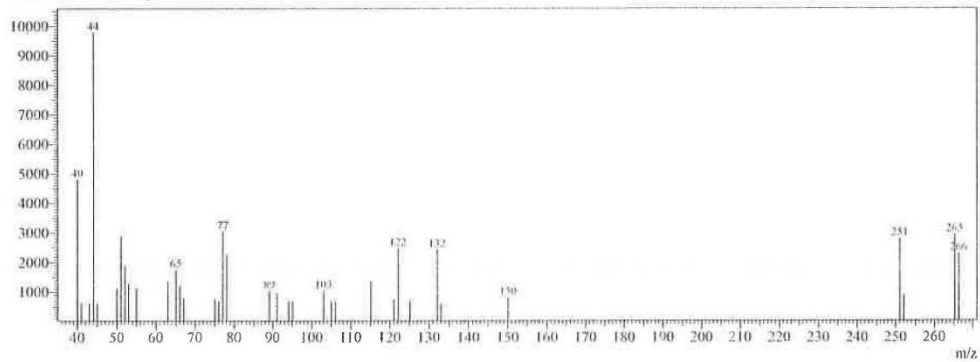


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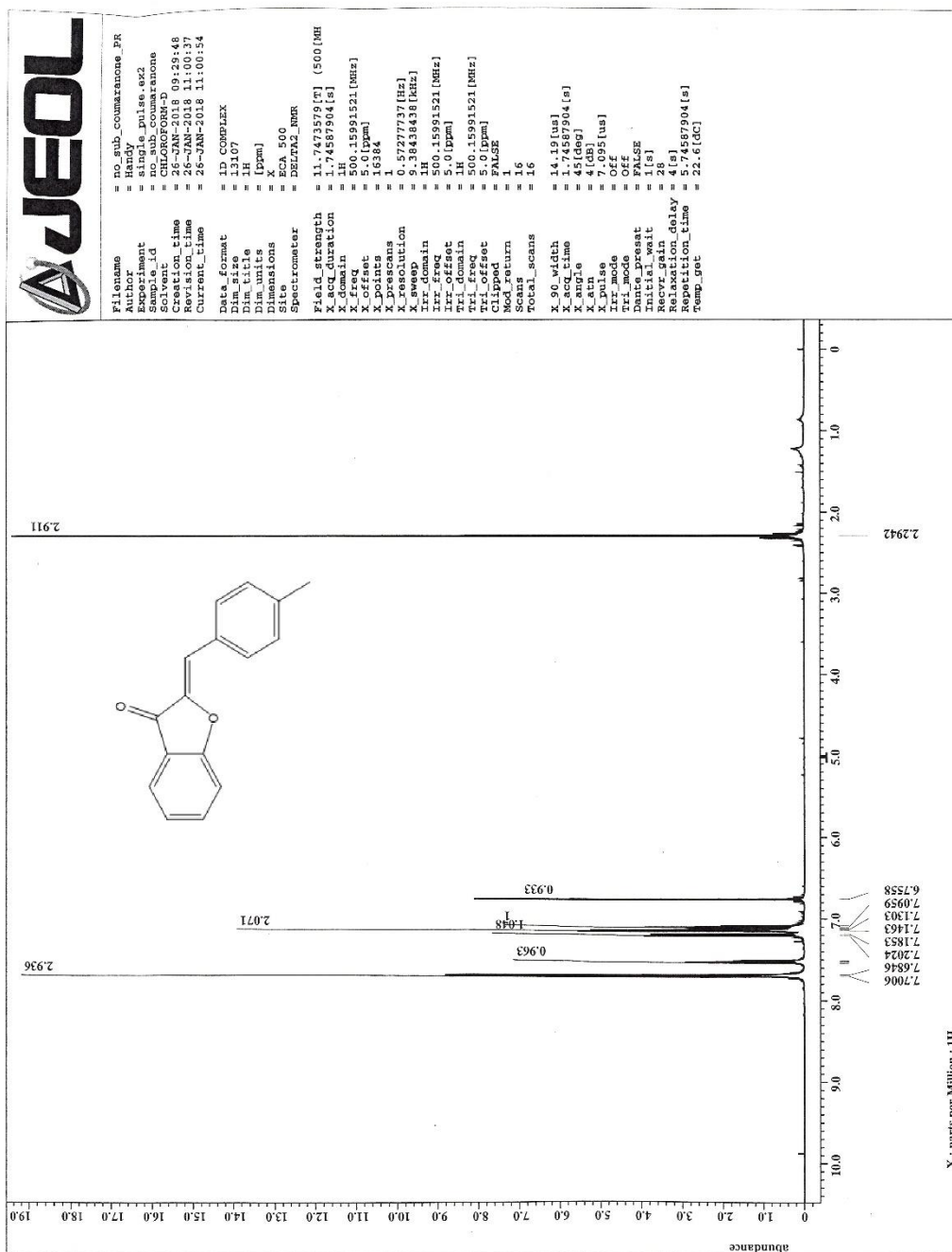


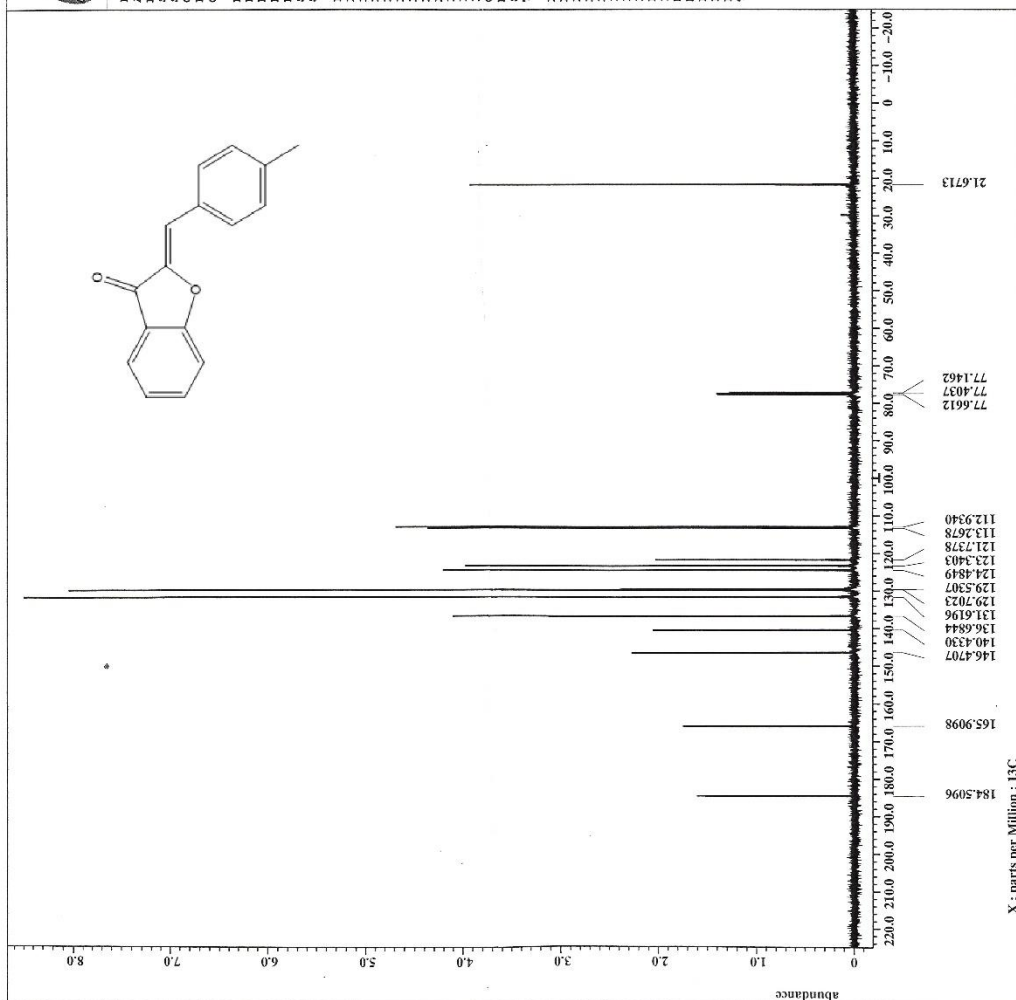
Spectrum

Line#:1 R.Time:28.360(Scan#:4873)
MassPeaks:36
RawMode:Single 28.360(4873) BasePeak:44.05(9778)
BG Mode:None Group I - Event I Scan

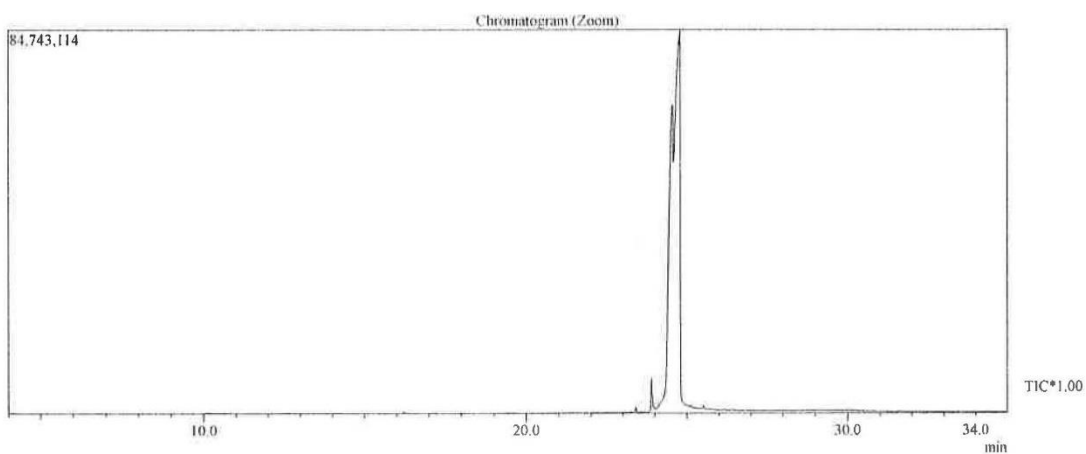
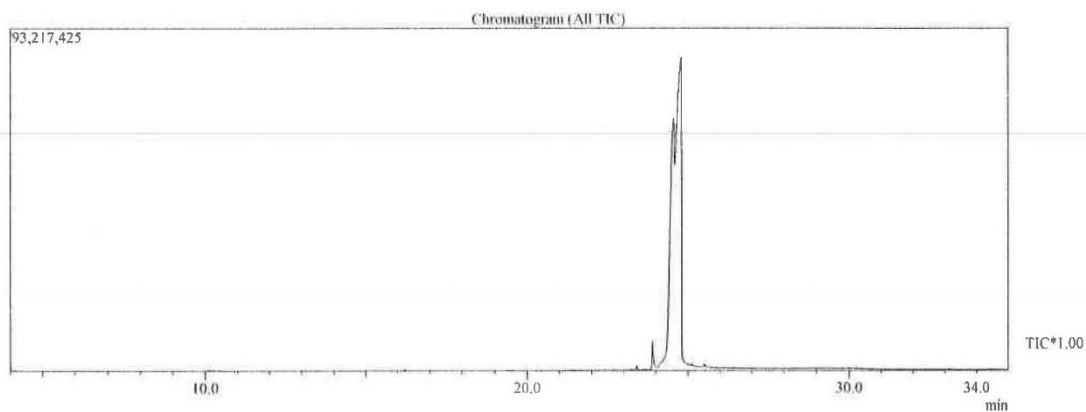


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Spectrum

Line#:1 R.Time:24.745(Scan#:4150)
MassPeaks:185
RawMode:Single 24.745(4150) BasePeak:220.95(8428992)
BG Mode:None Group 1 - Event 1 Scan

