



Supporting Information

for

1,5-Phosphonium betaines from *N*-triflylpropiolamides, triphenylphosphane, and active methylene compounds

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Experimental procedures, characterization data, NMR spectra (^1H , ^{13}C , ^{31}P , ^{19}F) and IR spectra for the synthesized compounds, and data for the X-ray crystal structure determinations

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1. General information

Melting points were determined in open capillaries with a Büchi B-540 instrument at a heating rate of 2 °C min⁻¹. IR spectra of solids were taken from solids as KBr pellets or from oils between NaCl plates and were recorded on a Bruker Vector 22 FT-IR instrument. NMR spectra: Bruker Avance 400 (¹H: 400.13 MHz; ¹³C: 100.62 MHz; ¹⁹F: 376.47 MHz; ³¹P: 161.98 MHz) and Bruker Avance 500 (¹H: 500.14 MHz; ¹³C: 125.77 MHz). ¹H and ¹³C spectra were referenced to the residual solvent signal [¹H: δ = 7.26 (CHCl₃) 1.94 (CD₃CN); ¹³C: δ = 77.16 (CDCl₃), 1.32/118.26 (CD₃CN) ppm. ¹⁹F NMR spectra were referenced to internal C₆F₆ (−162.90 ppm) and ³¹P spectra to external 85% aqueous H₃PO₄ (δ = 0.00 ppm). ¹³C NMR spectra were recorded in the broad-band ¹H-decoupled mode. Mass spectra: Finnigan-MAT SSQ-7000 (CI, 100 eV) and Solarix (HRMS, ESI). Elemental analyses: elemental Hanau vario MICRO cube analyser.

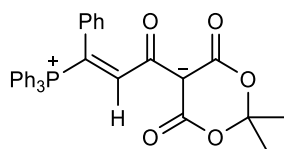
The synthesis of *N*-triflyl-propiolamides **1** has been published [1].

2. Syntheses and characterization of betaines **3**

2.1. General procedure

Triphenylphosphane (PPh₃, 1.00 equiv) and a methylene compound **2** (1.00 equiv) were dissolved in anhydrous dichloromethane (5 mL/mmol) at room temperature and an *N*-triflyl-propiolamide **1** (1.03 equiv) was added. The solution was stirred until the reaction was complete (0.5–6 hours, reaction control by TLC, ¹⁹F or ³¹P NMR spectroscopy), then slowly added to ice-cold diethyl ether (≈50 mL), whereupon betaines **3** precipitated. They were isolated by filtration and washed with a few milliliters of cold ether to furnish a product which in most cases was pure by elemental analysis. The ratio of *E* and *Z* diastereoisomers was determined by ³¹P NMR signal integration. A separation of the diastereoisomers, which can be distinguished by color and habit, was achieved in several cases by the solvent diffusion method from CH₂Cl₂/pentane at 8 °C. The *E* isomer, as the major component, crystallized first and was isolated by filtration. The *Z* isomer crystallized from the mother liquor, when the solvent diffusion procedure was continued.

2.1.1. (*E*)-2,2-Dimethyl-4,6-dioxo-5-(3-phenyl-3-(triphenylphosphonio)acryloyl)-1,3-dioxan-5-ide ((*E*)-**3a**)



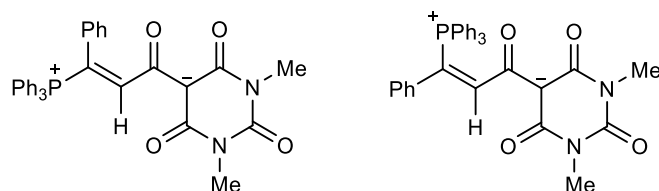
Prepared from triflamide **1** (361 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and Meldrum's acid (144 mg, 1.00 mmol); 2 hours. Yield: 518 mg (0.97 mmol, 97%). Pale yellow crystals of (*E*)-**3a**, m.p. 194 °C. – ¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 1.43 (s, 6H, CH₃), 6.98–7.00 (m, 2H, H_{Ph}), 7.08–7.12 (m, 2H, H_{Ph}), 7.18–7.21 (m, 1H, H_{Ph}), 7.56 (d, ³J_{H,P} = 23.52 Hz, 1H, C=CH), 7.59–7.66 (m, 12H, H_{Ph}), 7.76–7.80 (m, 3H, H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 26.41 (CH₃), 89.79 (d, ⁴J_{C,P} = 3.0 Hz, C), 100.65 (OCO), 114.55 (d, ¹J_{C,P} = 78.7 Hz, PC=CH), 118.35 (d, ¹J_{C,P} = 88.0 Hz, PC_{Ph}), 128.66

(d, $J_{C,P} = 1.8$ Hz, C_{sp^2}), 129.04 (d, $J_{C,P} = 1.9$ Hz), 130.15 (d, $^3J_{C,P} = 12.7$ Hz, C_{PPh}), 130.41 (d, $J_{C,P} = 12.8$ Hz), 130.83 (d, $J_{C,P} = 4.2$ Hz), 131.42 (d, $J_{C,P} = 10.1$ Hz), 134.84 (d, $J_{C,P} = 10.0$ Hz), 135.04 (d, $^2J_{C,P} = 10.0$ Hz, C_{PPh}), 135.13, 160.26 (d, $^2J_{C,P} = 7.7$ Hz, $C=\underline{CH}$), 183.64 (d, $^3J_{C,P} = 17.7$ Hz, $CH-\underline{C=O}$). – ^{31}P NMR ($CDCl_3$): δ [ppm] = 24.47.

IR (KBr): $\tilde{\nu}$ [cm^{-1}] = 1724 (s), 1659 (vs), 1560 (m), 1439 (m), 1384 (vs), 1312 (m), 1263 (m), 1197 (m), 1156 (m), 1107 (m), 1031 (m), 932 (m), 755 (m), 727 (m), 699 (m), 544 (m), 526 (m). – MS (CI, 100 eV): m/z (%) = 359 (28), 331 (24), 313 (15), 263 [$PPh_3 + H$]⁺ (39), 185 [$C_8H_8O_5 + H$]⁺ (100). – Anal. for $C_{33}H_{27}O_5P$ (534.55 g/mol): calcd. C 74.15, H 5.09 N 0.00; found C 74.26, H 5.18, N 0.02.

In the 1H NMR spectra of the crude product, a trace of (*Z*)-**3a** was detected: $\delta = 8.32$ ppm ($^3J_{H,P} = 41.3$ Hz).

2.1.2. 1,3-Dimethyl-2,4,6-trioxo-5-(3-phenyl-3-(triphenylphosphonio)acryloyl)hexahydropyrimidin-5-ide ((*E*)- and (*Z*)-**3b**)



From triflamide **1** (R = Ph; 538 mg, 1.53 mmol), PPh_3 (393 mg, 1.50 mmol) and *N,N*-dimethylbarbituric acid (234 mg, 1.50 mmol); 2 hours. Yield: 795 mg (1.45 mmol, 97%) of a light yellow mixture of diastereoisomers, *E/Z* = 83:17, which could be separated by fractionating solvent diffusion crystallization.

(*E*)-3b: colorless, m.p. 242 °C. – 1H NMR ($CDCl_3$, 400.13 MHz): δ [ppm] = 3.24 (s, 6H, NCH_3), 6.93–6.96 (m, 2H, H_{Ph}), 7.06–7.10 (m, 2H, H_{Ph}), 7.16–7.20 (m, 1H, H_{Ph}), 7.58–7.68 (m, 13H, H_{Ph} , $C=CH$), 7.75–7.80 (m, 3H, H_{Ph}). – ^{13}C NMR ($CDCl_3$, 100.62 MHz): δ [ppm] = 27.24 (NCH_3), 96.98 (d, $^4J_{C,P} = 3.0$ Hz, C^-), 113.45 (d, $^1J_{C,P} = 79.6$ Hz, $PC=CH$), 118.80 (d, $^1J_{C,P} = 87.9$ Hz, PC_{Ph}), 128.60 (d, $J_{C,P} = 1.5$ Hz, C_{Ph}), 128.95 (d, $J_{C,P} = 2.3$ Hz, C_{Ph}), 130.09 (d, $^3J_{C,P} = 12.6$ Hz, C_{PPh}), 130.92 (d, $J_{C,P} = 4.5$ Hz, C_{Ph}), 131.71 (d, $J_{C,P} = 10.3$ Hz, C_{Ph}), 135.00 (d, $^4J_{CP} = 2.9$ Hz, C_{PPh}), 135.12 (d, $^2J_{C,P} = 10.0$ Hz, C_{PPh}), 153.28 ($N,N-C=O$), 161.68 (d, $^2J_{C,P} = 8.0$ Hz, $C=\underline{CH}$), 164.06 (br, $N-C=O$), 184.16 (d, $^3J_{C,P} = 18.0$ Hz, $CH-\underline{C=O}$). – ^{31}P NMR ($CDCl_3$): δ [ppm] = 24.43.

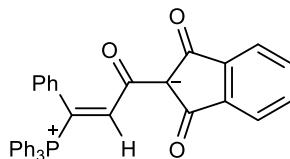
IR (KBr): $\tilde{\nu}$ [cm^{-1}] = 1701 (w), 1649 (s), 1615 (s), 1419 (s), 1386 (m). – HRMS (MALDI-TOF): m/z = 547.17828 [$M+H$]⁺; calcd. 547.17867 – Anal. for $C_{33}H_{27}N_2O_4P$ (546.56 g/mol): calcd. C 72.52, H 4.98, N 5.13; found C 72.50, H 4.97, N 5.01.

(*Z*)-3b: yellow, m.p. 244 °C. – 1H NMR ($CDCl_3$, 400.13 MHz): δ [ppm] = 3.24 (s, 6H, NCH_3), 6.99–7.01 (m, 2H, H_{Ph}), 7.07–7.10 (m, 2H, H_{Ph}), 7.13–7.17 (m, 1H, H_{Ph}), 7.41–7.45 (m, 6H, H_{Ph}), 7.55–7.63 (m, 9H, H_{Ph}), 8.36 (d, $^3J_{H,P} = 41.58$ Hz, 1H, $C=CH$). – ^{13}C NMR ($CDCl_3$, 125.77 MHz): δ [ppm] = 27.42 (NCH_3), 100.14 (C^-), 121.07 (d, $^1J_{C,P} = 80.4$ Hz, $P-\underline{C=CH}$), 121.67 (d, $^1J_{C,P} = 92.0$ Hz, PC_{Ph}), 128.41, 128.50, 129.20 (d, $^3J_{C,P} = 13.1$ Hz), 129.81 (d, $J_{CP} = 4.1$ Hz), 130.12 (d, $J_{C,P} = 12.6$ Hz), 133.47 (d, $^4J_{C,P}$

= 2.9 Hz), 134.42 (d, $^2J_{C,P}$ = 9.9 Hz), 135.14 (d, $J_{C,P}$ = 9.9 Hz), 153.06 (N,N-C=O), 160.13 (d, $^2J_{C,P}$ = 5.9 Hz, C=CH), 182.66 (d, $^3J_{C,P}$ = 5.8 Hz, CH-C=O). – ^{31}P NMR (CDCl_3): δ [ppm] = 23.17.

IR (KBr): $\tilde{\nu}$ [cm^{-1}] = 1698 (m), 1639 (s), 1618 (s), 1553 (m), 1477 (m), 1415 (s), 1387 (m), 1106 (m), 1009 (w). – HRMS (MALDI-TOF): m/z = 547.17828 $[\text{M}+\text{H}]^+$; calcd. 547.17867. – Anal. for $\text{C}_{33}\text{H}_{27}\text{N}_2\text{O}_4\text{P}$ (546.56 g/mol): calcd. C 72.52, H 4.98, N 5.13; found C 72.50, H 4.97, N 5.01.

2.1.3. (*E*)-1,3-Dioxo-2-(3-phenyl-3-(triphenylphosphonio)acryloyl)-2,3-dihydro-1*H*-inden-2-ide ((*E*)-3c)

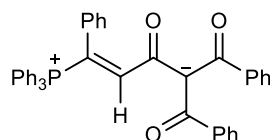


From triflamide **1** (R = Ph; 361 mg, 1.03 mmol), PPh_3 (262 mg, 1.00 mmol) and indane-1,3-dione (146 mg, 1.00 mmol); 2 hours. Only the *E* isomer was observed. Yield: 440 mg (0.81 mmol, 82%); orange solid, m.p. 234 °C. –

^1H NMR (CDCl_3 , 500.13 MHz): δ [ppm] = 6.90–6.92 (m, 2H, H_{Ar}), 7.01–7.4 (m, 2H, H_{Ar}), 7.12–7.15 (m, 1H, H_{Ar}), 7.30–7.62 (m, br, 16H, H_{Ar}), 7.69 (d, $^3J_{\text{H,P}}$ = 23.51 Hz, 1H, C=CH), 7.76–7.79 (m, 3H, H_{Ar}). – ^{13}C NMR (CDCl_3 , 125.77 MHz): δ [ppm] = 109.72 (d, $^4J_{C,P}$ = 2.8 Hz, C $^-$), 116.97 (d, $^1J_{C,P}$ = 88.2 Hz, PC_{Ph}), 119.68, 120.27 (d, $^1J_{C,P}$ = 75.9 Hz, $\text{PC}=\text{CH}$), 120.52, 128.39, 128.94, 130.00 (d, $^3J_{C,P}$ = 12.8 Hz), 130.47 (d, $J_{C,P}$ = 4.0 Hz), 130.66 (d, $J_{C,P}$ = 9.6 Hz), 131.66 (d, $J_{C,P}$ = 20.2 Hz), 134.47 (d, $J_{C,P}$ = 10.1 Hz), 134.84 (d, $^2J_{C,P}$ = 10.1 Hz), 135.16 (d, $^4J_{C,P}$ = 1.8 Hz), 138.95 (C_{indene}), 139.69 (C_{indene}), 158.34 (d, $^2J_{C,P}$ = 6.7 Hz, C=CH), 180.16 (d, $^3J_{C,P}$ = 16.8 Hz, CH-C=O), 191.14 ($\text{C}=\text{O}_{\text{indene}}$), 193.31 ($\text{C}=\text{O}_{\text{indene}}$). – ^{31}P NMR (CDCl_3): δ [ppm] = 24.79.

IR (KBr): $\tilde{\nu}$ [cm^{-1}] = 1683 (w), 1631 (s), 1587 (s), 1550 (m), 1437 (s), 1417 (s), 1201 (m), 1150 (m), 1106 (m), 695 (m), 600 (m), 529 (m). – HRMS (MALDI-TOF): m/z = 537.16133 $[\text{M} + \text{H}]^+$; calcd. 537.16196. – Anal. for $\text{C}_{36}\text{H}_{25}\text{O}_3\text{P}$ (536.57 g/mol): calcd. C 80.59, H 4.70, N 0.00; found C 80.51, H 4.62, N 0.00.

2.1.4. (*E*)-2-Benzoyl-1,3-dioxo-1,5-diphenyl-5-(triphenylphosphonio)pent-4-en-2-ide ((*E*)-3d)



From triflamide **1** (R = Ph; 361 mg, 1.03 mmol), PPh_3 (262 mg, 1.00 mmol) and 1,3-diphenylpropane-1,3-dione (242 mg, 1.00 mmol); 2 hours. In contrast to all other preparations, the solution turned black after addition of the triflamide and by-products were formed, which could not be removed completely even after repeated recrystallization. The purification efforts ensued a considerably reduced yield of the desired product: 132 mg (0.21 mmol, 21%) of still impure product; beige solid, m.p. 116 °C. Only the *E* isomer was observed. – ^{13}C NMR (CDCl_3 , 100.62 MHz): δ [ppm] = 93.24 (C $^-$), 115.11 (d, $^1J_{C,P}$ = 71.4

Hz), 118.15 (d, $^1J_{C,P}$ = 87.5 Hz, PC_{Ph}), 122.90, 123.14, 127.24, 128.53, 128.73 (d, $J_{C,P}$ = 1.4 Hz), 128.79, 129.03 (d, $J_{C,P}$ = 4.5 Hz), 130.24 (d, $^3J_{C,P}$ = 12.5 Hz), 130.27, 130.34, 130.39, 130.92 (d, $J_{C,P}$ = 12.8 Hz), 132.16 (d, $J_{C,P}$ = 10.1 Hz), 132.58, 134.69 (d, $^2J_{C,P}$ = 9.8 Hz), 134.92, 135.05 (d, $J_{C,P}$ = 5.6 Hz), 135.17, 135.22 (d, $J_{C,P}$ = 3.1 Hz), 135.57, 135.91 (d, $J_{C,P}$ = 3.0 Hz), 136.39 (d, $J_{C,P}$ = 3.3 Hz), 156.06 (C=C_H), 166.97 (d, $^2J_{C,P}$ = 20.1 Hz, CH-C=O), 185.85 (Ph-C=O); some of the reported data may result from impurities, see spectrum. – ^{31}P NMR (CDCl₃): δ [ppm] = 24.90.

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 1760 (m), 1655 (s), 1593 (s), 1488 (s), 1440 (s), 1303 (vs), 1198 (vs), 1153 (vs), 1107 (vs), 1001 (m), 754 (s), 726 (s), 699 (vs), 599 (m), 525 (s). – HRMS (MALDI-TOF): m/z = 615.20843 [M+H]⁺; calcd. for C₄₂H₃₂O₃P⁺ 615.20836.

2.1.5. 1,1-Dicyano-2-oxo-4-phenyl-4-(triphenylphosphonio)but-3-en-1-ide ((*E*)- and (*Z*)-3e)



From triflamide **1** (R = Ph; 538 mg, 1.53 mmol), PPh₃ (393 mg, 1.50 mmol) and malononitrile (99 mg, 1.50 mmol); 30 minutes. Yield: 644 mg (1.41 mmol, 94%) of a mixture of diastereoisomers, *E/Z* = 67:33, which could be separated by solvent diffusion crystallization.

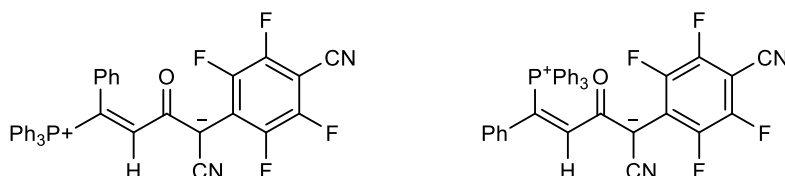
(*E*)-3e: colorless, m.p. 292 °C. – ^1H NMR (CDCl₃, 400.13 MHz): δ [ppm] = 7.03–7.06 (m, 2H, H_{Ph}), 7.17–7.23 (m, 2H, H_{Ph}), 7.20 (d, $^3J_{HP}$ = 22.7 Hz, 1H, C=CH), 7.28–7.33 (m, 1H, H_{Ph}), 7.47–7.53 (m, 6H, H_{Ph}), 7.61–7.66 (m, 6H, H_{Ph}), 7.79–7.84 (m, 3H, H_{Ph}). – ^{13}C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 52.20 (d, $^4J_{C,P}$ = 2.4 Hz, C⁻), 117.01 (d, $^1J_{C,P}$ = 87.9 Hz, PC_{Ph}), 118.02 (C≡N), 122.31 (C≡N), 125.41 (d, $^1J_{C,P}$ = 72.6 Hz, PC=CH), 128.89 (d, $J_{C,P}$ = 1.7 Hz), 129.81 (d, $J_{C,P}$ = 2.5 Hz), 129.97 (d, $J_{C,P}$ = 4.4 Hz), 130.47 (d, $^3J_{C,P}$ = 12.8 Hz), 130.78 (d, $J_{C,P}$ = 9.1 Hz), 134.83 (d, $^2J_{C,P}$ = 10.1 Hz), 135.65 (d, $^4J_{C,P}$ = 3.0 Hz), 153.71 (d, $^2J_{C,P}$ = 7.9 Hz, C=C_H), 182.60 (d, $^3J_{C,P}$ = 18.2 Hz, CH-C=O). – ^{31}P NMR (CDCl₃): δ [ppm] = 24.78.

UV (acetonitrile, 4·10⁻⁴ mol L⁻¹): λ_{max} [nm] (lg ϵ) = 207 (3.64), 368 (2.42). – IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2201 (m), 2176 (s), 1634 (m), 1549 (s), 1486 (w), 1437 (m), 1367 (s), 1106 (m), 754 (m), 728 (m), 696 (m), 518 (m), 501 (m). – HRMS (MALDI-TOF): m/z = 457.14658 [M + H]⁺; calcd. 457.14698. – Anal. for C₃₀H₂₁N₂OP (456.48 g/mol): calcd. C 78.94, H 4.64, N 6.14; found C 79.09, H 4.73, N 6.01.

(*Z*)-3e: yellow, m.p. 272 °C. – ^1H NMR (CDCl₃, 400.13 MHz): δ [ppm] = 6.82–6.84 (m, 2H, H_{Ph}), 7.04–7.08 (m, 2H, H_{Ph}), 7.13–7.19 (m, 1H, H_{Ph}), 7.37–7.45 (m, 12H, H_{Ph}), 7.52–7.59 (m, 3H, H_{Ph}), 7.91 (d, $^3J_{HP}$ = 38.76 Hz, 1H, C=CH). – ^{13}C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 56.26 (d, $^4J_{C,P}$ = 2.2 Hz, C⁻), 117.40 (C≡N), 119.93 (C≡N), 123.70 (d, $^1J_{C,P}$ = 95.1 Hz, PC_{Ph}), 128.45 (d, $J_{C,P}$ = 1.6 Hz), 128.75 (d, $J_{C,P}$ = 2.3 Hz), 129.22 (d, $J_{C,P}$ = 13.1 Hz), 129.40 (d, $J_{C,P}$ = 4.5 Hz), 129.95 (probably one branch of the doublet for PC=CH, the other one being covered by the signal at 129.22), 133.08 (d, $^2J_{C,P}$ = 9.7 Hz), 133.08 (d, $^4J_{C,P}$ = 3.3 Hz), 134.87 (d, $J_{C,P}$ = 10.1 Hz), 135.88 (d, $J_{C,P}$ = 10.6 Hz), 148.25 (d, $^2J_{C,P}$ = 5.0 Hz, C=C_H), 176.93 (d, $^3J_{C,P}$ = 4.1 Hz, CH-C=O). – ^{31}P NMR (CDCl₃): δ [ppm] = 22.80.

UV (acetonitrile, $4 \cdot 10^{-4}$ mol L⁻¹): λ_{max} [nm] (lg ϵ) = 215 (3.56), 390 (2.94). – IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2198 (s), 2175 (m), 1602 (m), 1543 (s), 1438 (m), 1376 (m), 1102 (m), 750 (m), 705 (m), 690 (m), 583 (m). – HRMS (MALDI-TOF): m/z = 457.14658 [M + H]⁺; calcd. 457.14698. – Anal. for C₃₀H₂₁N₂OP (456.48 g/mol): calcd. C 78.94, H 4.64, N 6.14; found C 79.09, H 4.73, N 6.01.

2.1.5. Cyano-1-(4-cyano-2,3,5,6-tetrafluorophenyl)-2-oxo-4-phenyl-4-(triphenylphosphonio)but-3-en-1-ide ((E)- and (Z)-3f)



From triflamide **1** (R = Ph; 361 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and 4-cyanomethyl-2,3,5,6-tetrafluorobenzonitrile (214 mg, 1.00 mmol); 1 hour. Yield: 562 mg (0.93 mmol, 93%) of a deep orange mixture of diastereoisomers, E/Z = 87:13, which could be separated by solvent diffusion crystallization. **(E)-3f**: 480 mg (0.80 mmol, 80%); dark red needle-shaped crystals, m.p. 190 °C.

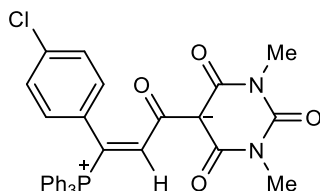
¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 7.11–7.14 (m, 2H, H_{Ph}), 7.17–7.21 (m, 2H, H_{Ph}), 7.27–7.31 (m, 1H, H_{Ph}), 7.48 (d, ³J_{H,P} = 22.79 Hz, 1H, C=CH), 7.53–7.68 (m, 12H, H_{Ph}), 7.80–7.85 (m, 3H, H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 65.55 (d, ⁴J_{C,P} = 2.8 Hz, C⁻), 108.96 (C≡N), 117.42 (d, ¹J_{C,P} = 87.9 Hz, PC_{Ph}), 122.69 (C≡N), 124.04 (d, ¹J_{C,P} = 73.1 Hz, PC=CH), 124.62, 124.82 (d, J = 1.4 Hz, C_{Ar}), 128.58, 129.46 (d, J = 2.3 Hz, C_{Ar}), 130.03 (d, J = 4.5 Hz, C_{Ar}), 130.34 (d, ³J_{C,P} = 12.7 Hz, C_{PPh}), 131.17 (d, J = 9.5 Hz, C_{Ar}), 134.80 (d, ²J_{C,P} = 10.0 Hz, C_{PPh}), 135.43 (d, ⁴J_{C,P} = 3.0 Hz, C_{PPh}); 142.5, 144.5, 145.7, 147.8 (4 weak m, CF and CCF), 155.65 (d, ²J_{C,P} = 7.3 Hz, C=CH), 176.19 (d, ³J_{C,P} = 17.7 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 24.55. – ¹⁹F NMR (CDCl₃): δ [ppm] = -131.64 (m), -138.65 (m). – IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2234 (w-m), 2171 (m), 1645 (m), 1561 (m), 1480 (s), 1438 (m), 1420 (m), 1276 (m), 1105 (m), 694 (m). – HRMS (MALDI-TOF): m/z = 605.14012 [M + H]⁺; calcd. 605.14059. – Anal. for C₃₆H₂₁F₄N₂OP (604.54 g/mol): calcd. C 71.52, H 3.5, N 4.63; found C 71.81, H 3.91, N 4.41.

(Z)-3f: 70 mg (12%); light yellow blocs, m.p. 214 °C.

¹H NMR (CDCl₃, 500.13 MHz, T = 294 K): δ [ppm] = 6.84–6.86 (m, 2H, H_{Ph}), 7.03–7.07 (m, 2H, H_{Ph}), 7.11–7.17 (m, 1 H), 7.26–7.57 (two broad unstructured m, 15H, H_{Ph}), 8.22 (d, ³J_{H,P} = 40.03 Hz, 1H, C=CH). – ¹³C NMR (CDCl₃, 100.62 MHz, T = 294 K): δ [ppm] = 108.64 (C≡N), 122.46 (C≡N), 128.24 (d, ¹J_{C,P} = 85.1 Hz, PC_{Ph}), 128.29 (d, J = 1.2 Hz, C_{Ar}), 128.35 (d, J = 2.2 Hz, C_{Ar}), 128.94 (d, ³J_{C,P} = 13.2 Hz, C_{PPh}), 129.60 (d, J = 4.6 Hz, C_{Ar}), 130.84 (d, ¹J_{C,P} = 56.7 Hz, PC=CH), 132.41 (d, ⁴J_{C,P} = 2.9 Hz, C_{PPh}), 132.80 (d, ²J_{C,P} = 9.0 Hz, C_{PPh}), 136.47 (d, $J_{C,P}$ = 11.6 Hz), 149.50 (d, ²J_{C,P} = 5.5 Hz, C=CH), 170.09 (d, ³J_{C,P} = 3.7 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 19.26. – ¹⁹F NMR (CDCl₃): δ [ppm] = -134.45 (m), -136.25 (m).

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2176 (m), 1644 (m), 1531 (s), 1482 (s), 1438 (m), 694 (m), 534 (m), 504 (m). – HRMS (MALDI): m/z = 604.13036 [M]⁺; calcd. 604.13276. – Anal. for C₃₆H₂₁F₄N₂OP (604.54 g/mol): calcd. C 71.52, H 3.50, N 4.63; found C 71.52, H 3.51, N 4.72.

2.1.6. (*E*)-5-(3-(4-Chlorophenyl)-3-(triphenylphosphonio)acryloyl)-1,3-dimethyl-2,4,6-trioxohexahydropyrimidin-5-ide ((*E*)-3g)

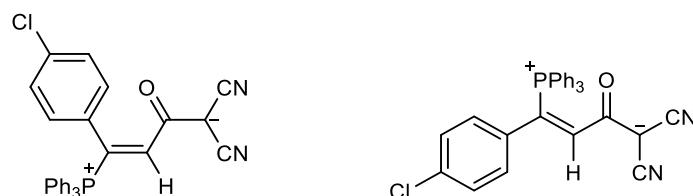


From triflamide **1** (R = 4-chlorophenyl; 399 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and *N,N*-dimethylbarbituric acid (156 mg, 1.00 mmol); 1 hour. Yield: 482 mg (0.83 mmol, 83%) of a pale yellow solid, m.p. 212 °C. Only the *E* isomer was observed.

¹H NMR (CDCl₃, 400.13 MHz): δ [ppm] = 3.25 (s, 6H, NCH₃), 6.88 (dd, ³*J*_{H,H} = 8.0 Hz, 2 H_{Ar}), 7.06 (broadened d, ³*J*_{H,H} = 8.0 Hz, 2 H_{Ar}), 7.60–7.70 (m, 12 H, H_{Ph}), 7.69 (d, ³*J*_{H,P} = 23.11 Hz, 1H, C=CH), 7.77–7.82 (m, 3 H). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 27.08 (NCH₃), 96.80 (d, ⁴*J*_{C,P} = 2.6 Hz, C⁻), 112.50 (d, ¹*J*_{C,P} = 81.0 Hz, PC=CH), 118.10 (d, ¹*J*_{C,P} = 88.2 Hz, PC_{Ph}), 128.68 (d, *J*_{C,P} = 1.8 Hz), 130.08 (d, ³*J*_{C,P} = 12.7 Hz), 130.25, 132.09 (d, *J*_{C,P} = 4.3 Hz), 134.87 (d, ²*J*_{C,P} = 10.0 Hz), 134.98 (d, *J*_{C,P} = 2.9 Hz), 135.14 (d, ⁴*J*_{C,P} = 3.0 Hz), 153.03 (N,N-C=O), 162.19 (d, ²*J*_{C,P} = 7.7 Hz, C=C_H), 163.83 (N-C=O), 183.77 (d, ³*J*_{C,P} = 17.6 Hz, CH-C=O). ³¹P NMR (CDCl₃): δ [ppm] = 24.38.

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 1701 (m), 1647 (vs), 1612 (s), 1482 (m), 1415 (s), 1387 (m), 1107 (m), 724 (m). – HRMS (MALDI): m/z = 581.13791 [M + H]⁺; calcd. 581.13699. – Anal. for C₃₃H₂₆ClN₂O₄P (581.00 g/mol): calcd. C 68.22, H 4.51, N 4.82; found C 68.43, H 4.38, N 4.81.

2.1.7. 4-(4-Chlorophenyl)-1,1-dicyano-2-oxo-4-(triphenylphosphonio)but-3-en-1-ide ((*E*)- and (*Z*)-3h)

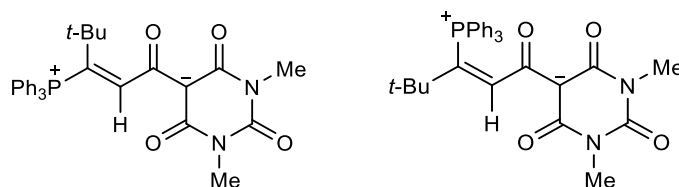


From triflamide **1** (R = 4-chlorophenyl; 399 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and malononitrile (66 mg, 1.00 mmol); 2 hours. Yield: dark yellow mixture of diastereoisomers (388 mg, 0.79 mmol, 79%, which were not separated. M.p. 283 °C (decomp.). The *E/Z* ratio was determined from the ¹H and ³¹P NMR spectra and exhibited a small solvent-dependent variation; values between 54:46 and 60:40 were found.

¹H NMR (CDCl₃, 500.13 MHz): *E/Z* = 54:46, δ [ppm] = 6.77 and 7.05 (AA'BB'X spin system, X = P, ³*J*_{H,H} = 8.43 Hz, 2H_{Ar}, *E*), 6.99 and 7.19 (AA'X,X', ³*J*_{H,H} = 8.43 Hz, 2H_{Ar}, *Z*), 7.24 (d, ³*J*_{H,P} = 22.21 Hz,

1H, C=CH, *E*), 7.36–7.47 (m, 13H, H_{Ph}), 7.50–7.61 (m, 9H, H_{Ph}), 7.64–7.69 (m, 5H, H_{Ph}), 7.82–7.87 (m, 3H, H_{Ph}), 7.89 (d, ³J_{H,P} = 38.26 Hz, 1H, C=CH, *Z*). – ¹H NMR (DMSO-*d*₆, see spectrum) *E*:*Z* = 60:40; olefinic proton signal of *Z* isomer covered by aromatic proton signals. – ¹³C NMR (CDCl₃, 100.62 MHz) of *E*/*Z* mixture; δ [ppm] = 52.55 (d, ⁴J_{C,P} = 2.2 Hz, C[–]), 56.55 (d, ⁴J_{C,P} = 2.4 Hz, C[–]), 116.81 (d, ¹J_{C,P} = 88.0 Hz, PC_{Ph}); 117.20–124.62 (4 C≡N, 4 C_{sp2}), 128.23–136.28 (14 C), 148.68 (d, ²J_{C,P} = 5.0 Hz, C=CH), 154.61 (d, ²J_{C,P} = 7.6 Hz, C=CH), 176.57 (d, ³J_{C,P} = 4.0 Hz, CH-C=O), 182.23 (d, ³J_{C,P} = 17.6 Hz, CH-C=O). – ³¹P NMR (DMSO-*d*₆): *E*/*Z* = 55:45, δ [ppm] = 22.50 (⁺PPh₃, *Z*), 24.87 (⁺PPh₃, *E*). IR (KBr): $\tilde{\nu}$ [cm^{–1}] = 2198 (s), 2175 (s), 1570 (s), 1547 (s), 1485 (m), 1438 (m), 1358 (m), 1105 (s), 723 (m), 692 (m), 541 (m), 514 (m). – HRMS (MALDI): *m/z* = 491.10746 [M + H]⁺; calcd. 491.10800. – Anal. for C₃₀H₂₀ClN₂OP (490.93 g/mol): calcd. C 73.40, H 4.11, N 5.71; found C 73.41, H 4.10, N 5.77.

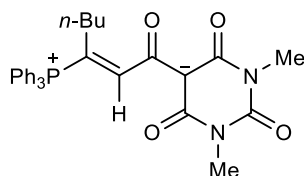
2.1.8. 5-(4,4-Dimethyl-3-(triphenylphosphonio)pent-2-enoyl)-1,3-dimethyl-2,4,6-trioxohexahydropyrimidin-5-ide ((*E*)- and (*Z*)-3i)



From triflamide **1** (R = *tert*-butyl; 343 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and *N,N*-dimethylbarbituric acid (156 mg, 1.00 mmol), 6 hours. A mixture of diastereoisomers was obtained (490 mg, 0.93 mmol, 93%, *E*/*Z* = 2:1), which were not separated. M.p. 204 °C.

¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] for *E* isomer: 1.17 (s, 9H, CMe₃), 3.27 (s, 6H, NMe), 7.18 (d, ³J_{H,P} = 30.8 Hz, 1H, C=CH); for *Z* isomer: 1.13 (s, 9H, CMe₃), 3.19 (s, 6H, NMe), 8.18 (d, ³J_{H,P} = 46.06 Hz, 1H, C=CH); for both isomers: 7.45–7.99 (several m, all H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] for *E* isomer: 27.19 (NMe), 31.33 (d, ³J_{C,P} = 5.1 Hz, CMe₃), 38.51 (d, ²J_{C,P} = 11.0 Hz, CMe₃), 96.51 (d, ⁴J_{C,P} = 2.2 Hz, C[–]), 119.07 (d, ¹J_{C,P} = 65.2 Hz, PC=CH), 121.38 (d, ¹J_{C,P} = 85.9 Hz, PC_{Ph}), 130.02 (d, ³J_{C,P} = 12.5 Hz, C_{PPh}), 134.51 (d, ⁴J_{C,P} = 3.0 Hz, C_{PPh}), 134.93 (d, ²J_{C,P} = 9.8 Hz, C_{PPh}), 153.16 (N,N-C=O), 163.46 (d, ¹J_{C,P} = 9.0 Hz, C=CH), 186.80 (d, ³J_{C,P} = 21.4 Hz, CH-C=O); for *Z* isomer: 27.23 (NMe), 31.33 (d, ³J_{C,P} = 3.6 Hz, CMe₃), 39.19 (d, ²J_{C,P} = 10.3 Hz, CMe₃), 97.63 (C[–]), 122.54 (d, ¹J_{C,P} = 88.0 Hz, PC_{Ph}), 125.69 (d, ¹J_{C,P} = 66.3 Hz, PC=CH), 128.98 (d, ³J_{C,P} = 12.7 Hz, C_{PPh}), 133.57 (d, ⁴J_{C,P} = 3.1 Hz, C_{PPh}), 135.25 (d, ²J_{C,P} = 10.0 Hz, C_{PPh}), 152.91 (N,N-C=O), 158.34 (d, ¹J_{C,P} = 6.9 Hz, C=CH), 185.04 (d, ³J_{C,P} = 7.4 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 28.96 (⁺PPh₃, *E*); 23.98 (⁺PPh₃, *Z*). – IR (KBr): $\tilde{\nu}$ [cm^{–1}] = 1699 (m), 1643 (vs), 1552 (m), 1479 (m), 1414 (vs), 1385 (vs), 1103 (m), 759 (m), 730 (m), 693 (m), 517 (m). – HRMS (MALDI): *m/z* = 527.20926 [M + H]⁺; calcd. 527.20997. – Anal. for C₃₁H₃₁N₂O₄P (526.57 g/mol): calcd. C 70.71, H 5.93 N 5.32; found C 70.72, H 5.92, N 5.22.

2.1.9. (*E*)-1,3-Dimethyl-2,4,6-trioxo-5-(3-(triphenylphosphonio)hept-2-enoyl)hexahydropyrimidin-5-ide ((*E*)-3j)

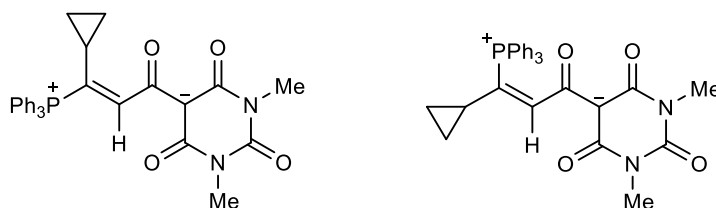


From triflamide **1** (R = *n*-Bu; 343 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and *N,N*-dimethylbarbituric acid (156 mg, 1.00 mmol); 2.5 hours. *E*-**3j** was obtained as a yellow solid (500 mg, 0.95 mmol, 95%), m.p. 220 °C.

¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 0.54 (t, ³J_{H,H} = 7.25 Hz, 3H, CH₂CH₃), 1.00 (h, ³J_{H,H} = 7.12 Hz, 2H, CH₂), 1.09–1.16 (m, 2H, CH₂), 2.40–2.49 (m, 2H, CH₂), 3.26 (s, 6H, NCH₃), 7.42 (d, ³J_{H,P} = 25.24 Hz, 1H, C=CH), 7.63–7.68 (m, 6H, H_{Ph}), 7.77–7.84 (m, 9H, H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 13.49 (CH₃), 22.48 (CH₂), 27.23 (NCH₃), 28.94 (d, ²J_{C,P} = 11.0 Hz, CH₂), 31.98 (d, ³J_{C,P} = 1.3 Hz, CH₂), 96.91 (d, ⁴J_{C,P} = 3.4 Hz, C[−]), 114.11 (d, ¹J_{C,P} = 74.4 Hz, PC=CH), 118.78 (d, ¹J_{C,P} = 87.1 Hz, PC_{Ph}), 130.16 (d, ³J_{C,P} = 12.5 Hz, C_{PPh}), 134.79 (d, ²J_{C,P} = 10.0 Hz, C_{PPh}), 134.99 (d, ⁴J_{C,P} = 3.0 Hz, C_{PPh}), 153.21 (N,N-C=O), 161.04 (d, ²J_{C,P} = 7.4 Hz, C=C_H), 164.10 (N-C=O), 184.98 (d, ³J_{C,P} = 19.8 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 26.24.

IR (KBr): $\tilde{\nu}$ [cm^{−1}] = 1697 (w), 1642 (s), 1481 (m), 1437 (m), 1414 (s), 1387 (s), 1107 (m), 999 (m), 726 (s), 694 (m). – Anal. for C₃₁H₂₁N₂O₄P (526.57 g/mol): calcd. C 70.71, H 5.93, N 5.32; found C 70.41, H 5.94, N 5.27.

2.1.10. 5-(3-Cyclopropyl-3-(triphenylphosphonio)acryloyl)-1,3-dimethyl-2,4,6-trioxohexahydropyrimidin-5-ide ((*E*)- and (*Z*)-3k)



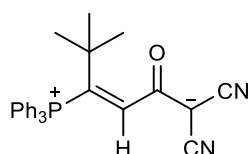
From triflamide **1** (R = cyclopropyl; 327 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and *N,N*-dimethylbarbituric acid (156 mg, 1.00 mmol); 2 hours. A mixture of diastereoisomers was obtained (454 mg, 0.89 mmol, 89%; *E/Z* = 83:17), which were not separated. M.p. 199 °C. –

¹H NMR (CDCl₃, 500.13 MHz): δ [ppm], *E* isomer: 0.62–0.83 (m, 4H, CH₂), 1.43–1.51 (m, 1H, CH), 3.30 (s, 6H, NCH₃), 7.44 (dd, *J*_{H,P} = 23.87 Hz, 1.83 Hz, 1H, C=CH); *Z* isomer: 0.62–0.83 (m, 4H, CH₂), 1.00–1.07 (m, 1H, CH), 3.24 (s, 6H, NCH₃), 8.30 (dd, *J*_{H,P} = 42.16 Hz, 1.62 Hz, 1H, C=CH); both isomers: 7.49–7.89 (several m, all H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm], *E* isomer: 8.66 (d, ³J_{C,P} = 5.8 Hz, CH₂), 12.58 (d, ²J_{C,P} = 14.9 Hz, CH), 27.27 (NCH₃), 96.66 (d, ⁴J_{C,P} = 3.4 Hz, C[−]), 114.26 (d, ¹J_{C,P} = 79.0 Hz, PC=CH), 119.36 (d, ¹J_{C,P} = 88.0 Hz, PC_{Ph}), 130.11 (d, ³J_{C,P} = 12.5 Hz, C_{PPh}), 134.87 (d, ²J_{C,P} = 10.4 Hz, C_{PPh}), 134.96 (C_{PPh}), 153.15 (N,N-C=O, both isomers?), 154.43 (NC=O), 163.33 (d, ²J_{C,P} = 9.9 Hz, C=C_H), 185.22 (d, ³J_{C,P} = 18.8 Hz, CH-C=O); *Z* isomer: 9.32 (d, ³J_{C,P} = 4.8 Hz, CH₂),

17.65 (d, $^2J_{C,P}$ = 17.8 Hz, CH), 27.38 (NCH₃), 97.16 (C⁻), 122.64 (d, $^1J_{C,P}$ = 92.1 Hz, C_{PPh}), 123.77 (d, $^1J_{C,P}$ = 81.0 Hz, PC=CH), 129.17 (d, $^3J_{C,P}$ = 12.9 Hz, PC_{Ph}), 133.33 (d, $^4J_{C,P}$ = 3.1 Hz, C_{PPh}), 134.05 (d, $^2J_{C,P}$ = 9.9 Hz, C_{PPh}), 153.15 (N,N-C=O, coinciding with *E* isomer?), 154.50 (N-C=O?), 164.11 (C=C_H), 182.75 (d, $^3J_{C,P}$ = 15.3 Hz, CH-C=O). – ^{31}P NMR (CDCl₃): δ [ppm] = 25.39 (⁺PPh₃, *E*); 24.21 (⁺PPh₃, *Z*).

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 1698 (s), 1644 (s), 1615 (vs), 1480 (m), 1418 (s), 1387 (vs), 1107 (s), 998 (m), 757 (s), 724 (s), 696 (s). – HRMS (MALDI): m/z = 511.17820 [M + H]⁺; calcd. 511.17867. – Anal. for C₃₀H₂₇N₂O₄P (510.53 g/mol): calcd. C 70.58, H 5.33, N 5.49; found C 70.45, H 5.22, N 5.39.

2.1.11. (*E*)-1,1-Dicyano-5,5-dimethyl-2-oxo-4-(triphenylphosphonio)hex-3-en-1-ide ((*E*)-3l)



From triflamide **1** (R = *tert*-butyl; 343 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and malononitrile (66 mg, 1.00 mmol); 3 hours. **E-3l** was obtained as a light yellow solid (419 mg, 0.96 mmol, 96%), which decomposed at 272 °C.

^1H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 1.28 (s, 9H, CH₃), 6.83 (d, $^3J_{H,P}$ = 30.77 Hz, 1H, C=CH), 7.69–7.74 (m, 6H, H_{Ph}), 7.80–7.85 (m, 9H, H_{Ph}). – ^{13}C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 31.81 (d, $^3J_{C,P}$ = 4.9 Hz, CCH₃), 39.55 (d, $^2J_{C,P}$ = 10.2 Hz, CCH₃), 51.40 (d, $^4J_{C,P}$ = 2.6 Hz, C⁻), 117.93 (C≡N), 120.09 (d, $^1J_{C,P}$ = 85.8 Hz, C_{PPh}), 122.36 (C≡N), 130.56 (d, $^3J_{CP}$ = 12.6 Hz, C_{PPh}), 131.87 (d, $^1J_{C,P}$ = 58.6 Hz, PC=CH), 134.66 (d, $^2J_{C,P}$ = 9.8 Hz, PC_{Ph}), 135.20 (d, $^4J_{C,P}$ = 3.0 Hz, C_{PPh}), 158.12 (d, $^5J_{C,P}$ = 8.5 Hz, C=C_H), 185.45 (d, $^3J_{C,P}$ = 21.4 Hz, CH-C=O). – ^{31}P NMR (CDCl₃): δ [ppm] = 30.23.

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2194 (s), 2166 (s), 1588 (s), 1565 (s), 1439 (m), 1346 (s), 1321 (m), 1100.62 (m), 721 (m), 694 (m), 522 (s). – Anal. for C₂₈H₂₅N₂OP (436.49 g/mol): calcd. C 77.05, H 5.77 N 6.42; found C 77.30, H 6.00, N 6.22.

2.1.12. 1,1-Dicyano-2-oxo-4-(triphenylphosphonio)oct-3-en-1-ide ((*E*)- and (*Z*)-3m)



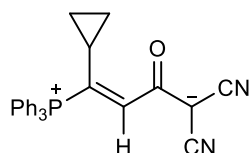
From triflamide **1** (R = *n*-butyl; 327 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and malononitrile (66 mg, 1.00 mmol); 2 hours. A mixture of diastereoisomers was obtained (388 mg, 0.93 mmol, 93%; *E/Z* = 83:17), which could be partly separated by solvent diffusion crystallization. The *E* isomer was obtained in pure form (310 mg, 0.71 mmol, 71%) as a light yellow solid, m.p. 185 °C (decomp.). The *Z* isomer was obtained only in admixture with (**E**)-**3m**.

(E)-3m: ^1H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 0.61 (t, $^3J_{H,H}$ = 7.22 Hz, 3H, CH₃), 1.04–1.11 (m, 2H, CH₂), 1.13–1.22 (m, 2H, CH₂), 2.64–2.73 (m, 2H, CH₂), 7.02 (d, $^3J_{H,P}$ = 25.07 Hz, 1H, C=CH), 7.65–7.75 (m, 12H, H_{Ph}), 7.84–7.88 (m, 3H, H_{Ph}). – ^{13}C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 13.42

(CH₃), 22.70 (CH₂), 29.64 (d, ²J_{C,P} = 9.7 Hz, CH₂), 31.98 (d, ³J_{C,P} = 1.3 Hz, CH₂), 53.33 (d, ⁴J_{C,P} = 2.9 Hz, C⁻), 117.19 (d, ¹J_{C,P} = 87.2 Hz, PC_{Ph}), 118.42 (C≡N), 121.78 (C≡N), 126.55 (d, ¹J_{C,P} = 68.8 Hz, PC=CH), 130.65 (d, ³J_{C,P} = 12.6 Hz, C_{PPh}), 134.52 (d, ²J_{C,P} = 10.0 Hz, C_{PPh}), 135.66 (d, ⁴J_{C,P} = 3.0 Hz, C_{PPh}), 151.64 (d, ²J_{C,P} = 7.4 Hz, C=C_H), 182.21 (d, ³J_{C,P} = 20.1 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 26.92 (⁺PPh₃, *E*); 26.09 (⁺PPh₃, *Z*).

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2195 (s), 2171 (s), 1611 (m), 1565 (s), 1438 (m), 1353 (m), 1107 (s), 995 (m), 723 (s), 691 (m), 526 (m). – HRMS (MALDI): *m/z* = 437.16654 [M + H]⁺; calcd. 437.17828. – Anal. for C₂₈H₂₅N₂OP (436.49 g/mol): calcd. C 77.05, H 5.77, N 6.42; found C 77.05, H 5.71, N 6.70.

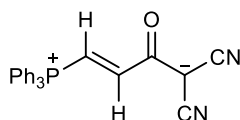
2.1.13. (*E*)-1,1-Dicyano-4-cyclopropyl-2-oxo-4-(triphenylphosphonio)but-3-en-1-ide ((*E*)-3n)



From triflamide **1** (R = cyclopropyl; 327 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and malononitrile (66 mg, 1.00 mmol), 2 hours. **E-3n** was obtained as a light yellow solid, which decomposed at 234 °C. Yield: 370 mg (0.88 mmol, 88%). The *Z* isomer was not observed. – ¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 0.74–0.83 (m, 4H, CH₂), 1.53–1.58 (m, 1H CH), 6.86 (dd, ³J_{H,P} = 24.04 Hz, ⁴J_{H,H} = 1.73 Hz, 1H, C=CH), 7.69–7.75 (m, 12H, H_{Ph}), 7.83–7.89 (m, 3H, H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 9.12 (d, ³J_{C,P} = 5.5 Hz, CH₂), 13.43 (d, ²J_{C,P} = 14.4 Hz, CH), 52.46 (d, ⁴J_{C,P} = 2.4 Hz, C⁻), 117.65 (d, ¹J_{C,P} = 87.0 Hz, PC_{Ph}), 118.19 (C≡N), 122.18 (C≡N), 125.57 (d, ¹J_{C,P} = 72.2 Hz, PC=CH), 130.54 (d, ³J_{C,P} = 12.6 Hz, C_{PPh}), 134.58 (d, ²J_{C,P} = 10.1 Hz, C_{PPh}), 135.58 (d, ⁴J_{C,P} = 3.0 Hz, C_{PPh}), 153.91 (d, ²J_{C,P} = 9.5 Hz, C=C_H), 183.49 (d, ³J_{C,P} = 19.2 Hz, CH-C=O). – ³¹P NMR (CDCl₃): δ [ppm] = 26.48.

IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2195 (s), 2171 (s), 1614 (w), 1560 (s), 1438 (m), 1353 (m), 1107 (s), 753 (m), 727 (m), 692 (m), 523 (m). – Anal. for C₂₇H₂₁N₂OP (420.45 g/mol): calcd. C 77.13, H 5.03, N 6.66; found C 77.22, H 4.88, N 6.80.

2.1.14. (*E*)-1,1-Dicyano-2-oxo-4-(triphenylphosphonio)but-3-en-1-ide ((*E*)-3o)



From *N*,3-Diphenyl-3-trimethylsilyl-*N*-triflyl-propiolamide (**1**, R = SiMe₃) (360 mg, 1.03 mmol), PPh₃ (262 mg, 1.00 mmol) and malononitrile (66 mg, 1.00 mmol); 2 hours at 0 °C. **E-3o** was obtained as an ocre solid (269 mg, 0.71 mmol, 71%), which decomposed at 220 °C. The *Z* isomer was not observed.

¹H NMR (CDCl₃, 500.13 MHz): δ [ppm] = 7.33 (dd, *J* = 20.27 and 16.27 Hz, 1H, C=CH), 7.55–7.65 (m, 6H, H_{Ph}, HC=C), 7.69–7.75 (m, 7H, H_{Ph}, HC=C), 7.84–7.87 (m, 3H, H_{Ph}). – ¹³C NMR (CDCl₃, 100.62 MHz): δ [ppm] = 56.19 (C⁻), 113.69 (d, ¹J_{C,P} = 87.9 Hz, PC=CH), 117.54 (d, ¹J_{C,P} = 91.3 Hz,

PC_{Ph}), 118.38 (C≡N), 120.40 (C≡N), 130.79 (d, $^3J_{C,P}$ = 13.1 Hz, C_{PPh}), 133.91 (d, $^2J_{C,P}$ = 10.7 Hz, C_{PPh}), 135.86 (d, $^4J_{C,P}$ = 3.1 Hz, C_{PPh}), 152.49 (d, $^2J_{C,P}$ = 3.9 Hz, C=CH), 177.07 (d, $^3J_{C,P}$ = 18.6 Hz, CH-CO).
 ^{31}P NMR (CDCl₃): δ [ppm] = 20.51.

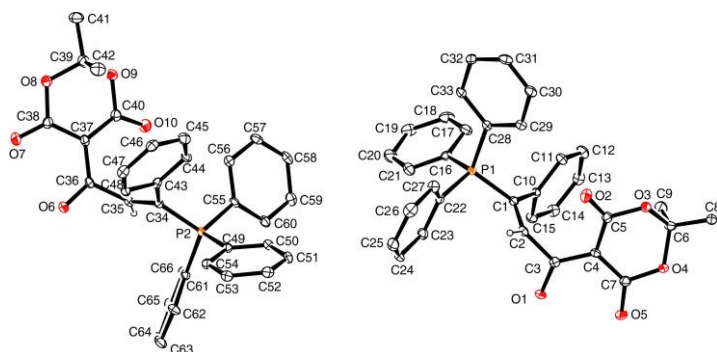
IR (KBr): $\tilde{\nu}$ [cm⁻¹] = 2198 (s), 2178 (s), 1613 (m), 1548 (s), 1438 (m), 1365 (m), 1110 (m), 726 (m). –

Anal. for C₂₄H₁₇N₂OP (380.39 g/mol): calcd. C 75.78, H 4.50, N 7.36; found C 75.77, H 4.52, N 7.26.

3. X-ray crystal structure determinations

CCDC-1937388 (*E*-**3a**), -1937389 (*E*-**3b**), -1937390 (*E*-**3c**), and -1937391 (*Z*-**3e**) contains the crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/getstructures.

3.1. Crystallographic data for (*E*)-**3a**



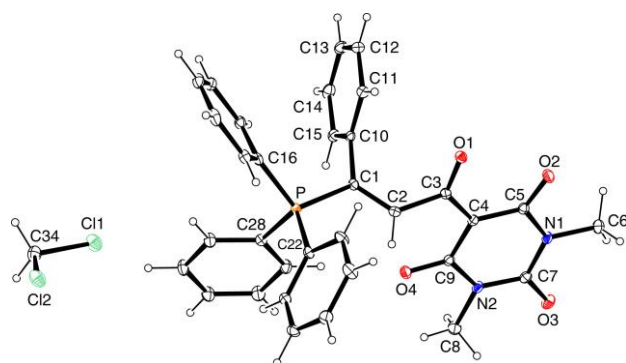
Data collection was performed on an Oxford Diffraction Rigaku instrument (SuperNova, Dual Source, Atlas CCD, Mo K_{α} radiation). Suitable crystals were obtained as pale yellow thin platelets by diffusion crystallization from CH_2Cl_2 /diethyl ether at 8 °C. Data collection at 150(1) K. Structure solution: SHELXS-97 [2]; refinement: SHELXL-2014/6 [3]; molecule plot: ORTEP-3 for Windows [4]. The triclinic unit cell contains two symmetry-independent molecules.

Table: Crystallographic data for (*E*)-**3a**

Identification code	AF291	
Empirical formula	$\text{C}_{33}\text{H}_{27}\text{O}_5\text{P}$	
Formula weight	534.51	
Temperature	149.95(10) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	<i>P</i> -1	
Unit cell dimensions	$a = 8.6737(4)$ Å	$\alpha = 92.180(3)^\circ$
	$b = 13.2993(4)$ Å	$\beta = 98.750(4)^\circ$
	$c = 23.5244(12)$ Å	$\gamma = 90.155(3)^\circ$
Volume	2680.0(2) Å ³	
<i>Z</i>	4	
Density (calculated)	1.325 Mg/m ³	
Absorption coefficient	0.145 mm ⁻¹	
<i>F</i> (000)	1120	
Crystal size	0.18 x 0.16 x 0.04 mm ³	
Theta range for data collection	2.839 to 25.681°.	
Index ranges	-10 ≤ <i>h</i> ≤ 8, -13 ≤ <i>k</i> ≤ 16, -24 ≤ <i>l</i> ≤ 28	
Reflections collected	26828	
Independent reflections	100.6283 [<i>R</i> (int) = 0.0597]	
Completeness to theta = 25.242°	99.8 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.84204	

Refinement method	Full-matrix least-squares on F^2
Data / restraints / parameters	100.6283 / 0 / 707
Goodness-of-fit on F^2	1.194
Final R indices [$I > 2\sigma(I)$]	$R1 = 0.1186$, $wR2 = 0.2714$
R indices (all data)	$R1 = 0.1393$, $wR2 = 0.2810$
Extinction coefficient	n/a
Largest diff. peak and hole	0.652 and -0.433 e.Å ⁻³

3.2. Crystallographic data for (*E*)-3b·CH₂Cl₂

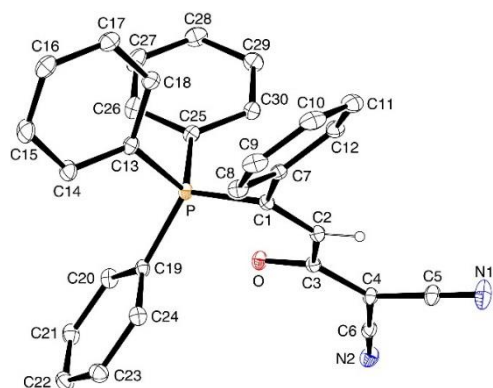


Data collection was performed on an Oxford Diffraction Rigaku instrument (SuperNova, Dual Source, Atlas CCD, Mo K α radiation). Colorless crystals were obtained by diffusion crystallization from CH₂Cl₂/*n*-pentane at 8 °C. A crystal piece cut from a larger columnar crystal was used. Data collection at 150(1) K. Structure solution: SHELXS [2]; refinement: SHELXL-2014 [3]; molecule plot: ORTEP-3 for Windows [4].

Table: Crystallographic data for (*E*)-3b × CH₂Cl₂.

Identification code	CF323	
Empirical formula	C ₃₄ H ₂₉ Cl ₂ N ₂ O ₄ P	
Formula weight	631.46	
Temperature	150.00(10) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	<i>P</i> -1	
Unit cell dimensions	<i>a</i> = 9.8481(7) Å	α = 111.210(7)°
	<i>b</i> = 11.3627(9) Å	β = 106.220(6)°
	<i>c</i> = 15.3689(11) Å	γ = 93.351(6)°
Volume	1514.8(2) Å ³	
<i>Z</i>	2	
Density (calculated)	1.384 Mg/m ³	
Absorption coefficient	0.310 mm ⁻¹	
<i>F</i> (000)	656	
Crystal size	0.22 x 0.16 x 0.15 mm ³	
Theta range for data collection	2.866 to 25.681°	
Index ranges	-11 ≤ <i>h</i> ≤ 12, -13 ≤ <i>k</i> ≤ 13, -18 ≤ <i>l</i> ≤ 17	
Reflections collected	11748	
Independent reflections	5732 [<i>R</i> (int) = 0.0278]	
Completeness to theta = 25.242°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.88639	
Refinement method	Full-matrix least-squares on <i>F</i> ²	
Data / restraints / parameters	5732 / 0 / 390	
Goodness-of-fit on <i>F</i> ²	1.069	
Final <i>R</i> indices [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> 1 = 0.0455, <i>wR</i> 2 = 0.1174	
<i>R</i> indices (all data)	<i>R</i> 1 = 0.0588, <i>wR</i> 2 = 0.1275	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.473 and -0.462 e.Å ⁻³	

3.3. Crystallographic data for (Z)-3e

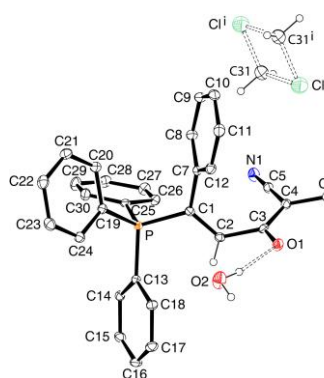


Data collection was performed on an Oxford Diffraction Rigaku instrument (SuperNova, Dual Source, Atlas CCD, Cu K α radiation). Crystals were obtained as yellow needles by diffusion crystallization from CH₂Cl₂/n-pentane at 8 °C. A crystal piece cut from a larger needle was used. Data collection at 150(1) K. Structure solution: SIR-92 [5]; refinement: SHELXL-2014/6 [3]; molecule plot: ORTEP-3 for Windows [4].

Table: Crystallographic data of (Z)-3e.

Identification code	CF325	
Empirical formula	C ₃₀ H ₂₁ N ₂ OP	
Formula weight	456.46	
Temperature	150.00(10) K	
Wavelength	1.54184 Å	
Crystal system	Monoclinic	
Space group	<i>I</i> 2/a	
Unit cell dimensions	<i>a</i> = 18.9715(3) Å	α = 90°
	<i>b</i> = 8.8404 (2) Å	β = 99.6230(14)°
	<i>c</i> = 28.5247(4) Å	γ = 90°
Volume	4716.72(13) Å ³	
<i>Z</i>	8	
Density (calculated)	1.286 Mg/m ³	
Absorption coefficient	1.228 mm ⁻¹	
<i>F</i> (000)	1904	
Crystal size	0.18 x 0.15 x 0.14 mm ³	
Theta range for data collection	4.728 to 70.060°	
Index ranges	-23 ≤ <i>h</i> ≤ 18, -10 ≤ <i>k</i> ≤ 10, -20 ≤ <i>l</i> ≤ 34	
Reflections collected	9147	
Independent reflections	4469 [<i>R</i> (int) = 0.0223]	
Completeness to theta = 67.684°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.92939	
Refinement method	Full-matrix least-squares on <i>F</i> ²	
Data / restraints / parameters	4469 / 0 / 307	
Goodness-of-fit on <i>F</i> ²	1.039	
Final <i>R</i> indices [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> 1 = 0.0338, <i>wR</i> 2 = 0.0858	
<i>R</i> indices (all data)	<i>R</i> 1 = 0.0383, <i>wR</i> 2 = 0.0899	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.287 and -0.285 e.Å ⁻³	

3.4. Crystallographic data for (*E*)-3e·H₂O·CH₂Cl₂



Data collection was performed on an Oxford Diffraction Rigaku instrument (SuperNova, Dual Source, Atlas CCD, Cu K α radiation). Crystals were obtained as colorless prisms by diffusion crystallization from CH₂Cl₂/n-pentane at 8 °C. A crystal piece cut from a larger needle was used. Data collection at 150(2) K. Structure solution: SIR-92 [5]; refinement: SHELXL-2014/6 [3]; molecule plot: ORTEP-3 for Windows [4]. Hydrogen atoms were in geometrically calculated positions and included in the refinement using the riding model. The positions of the water hydrogen atoms were taken from a difference Fourier electron map and refined. The dichloromethane solvate molecule is disordered around a crystallographic inversion center.

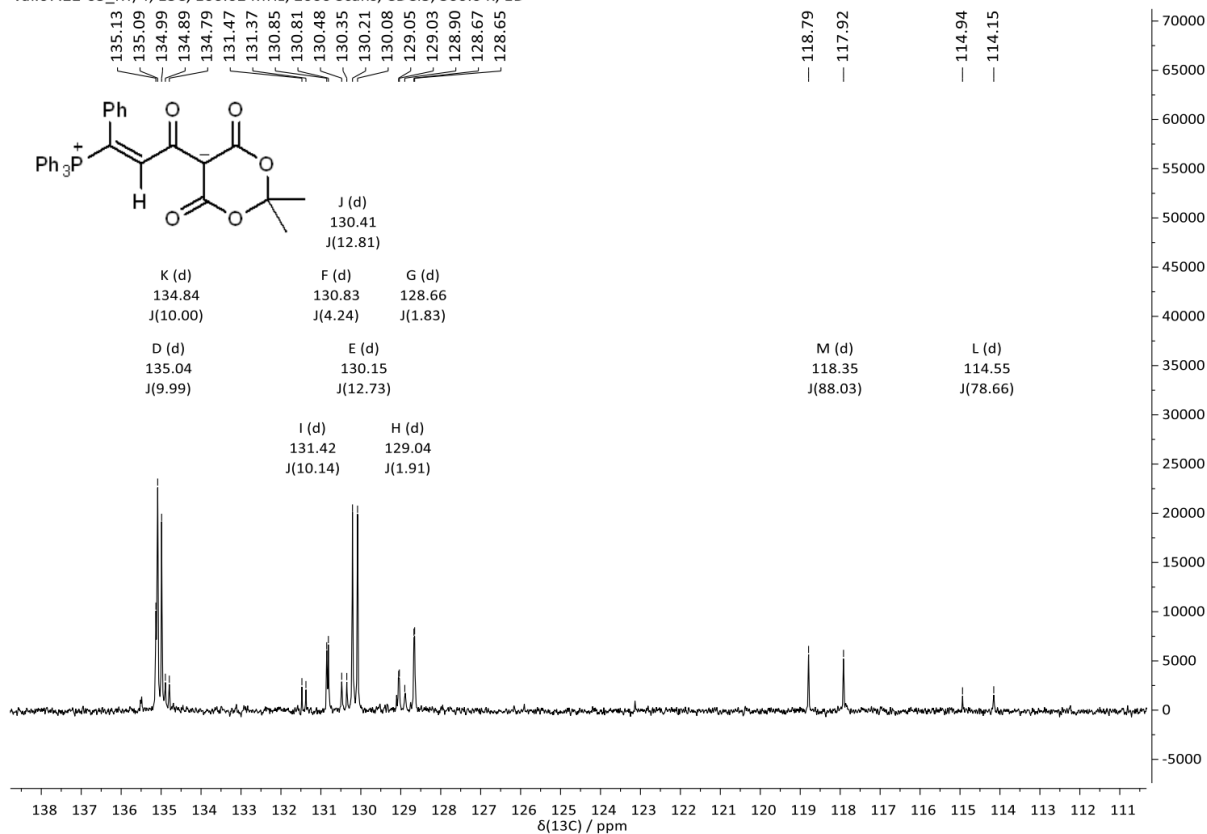
Table: Crystallographic data of (*E*)-3e.

Identification code	CF326	
Empirical formula	C _{30.50} H ₂₄ ClN ₂ O ₂ P	
Formula weight	516.94	
Temperature	150(2) K	
Wavelength	1.54184 Å	
Crystal system	Triclinic	
Space group	<i>P</i> -1	
Unit cell dimensions	<i>a</i> = 9.7424(5) Å	α = 99.933(4)°
	<i>b</i> = 12.5721(5) Å	β = 110.790(5)°
	<i>c</i> = 12.8181(7) Å	γ = 110.690(4)°
Volume	1290.99(12) Å ³	
<i>Z</i>	2	
Density (calculated)	1.330 Mg/m ³	
Absorption coefficient	2.143 mm ⁻¹	
<i>F</i> (000)	538	
Crystal size	0.18 x 0.15 x 0.14 mm ³	
Theta range for data collection	4.550 to 70.059°	
Index ranges	-11 ≤ <i>h</i> ≤ 10, -14 ≤ <i>k</i> ≤ 15, -15 ≤ <i>l</i> ≤ 15	
Reflections collected	8900	
Independent reflections	4881 [<i>R</i> (int) = 0.0214]	
Completeness to theta = 67.684°	99.9 %	
Refinement method	Full-matrix least-squares on <i>F</i> ²	
Data / restraints / parameters	4881 / 0 / 350	
Goodness-of-fit on <i>F</i> ²	1.046	
Final <i>R</i> indices [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> 1 = 0.0365, <i>wR</i> 2 = 0.0924	
<i>R</i> indices (all data)	<i>R</i> 1 = 0.0412, <i>wR</i> 2 = 0.0955	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.679 and -0.584 e.Å ⁻³	

3. References

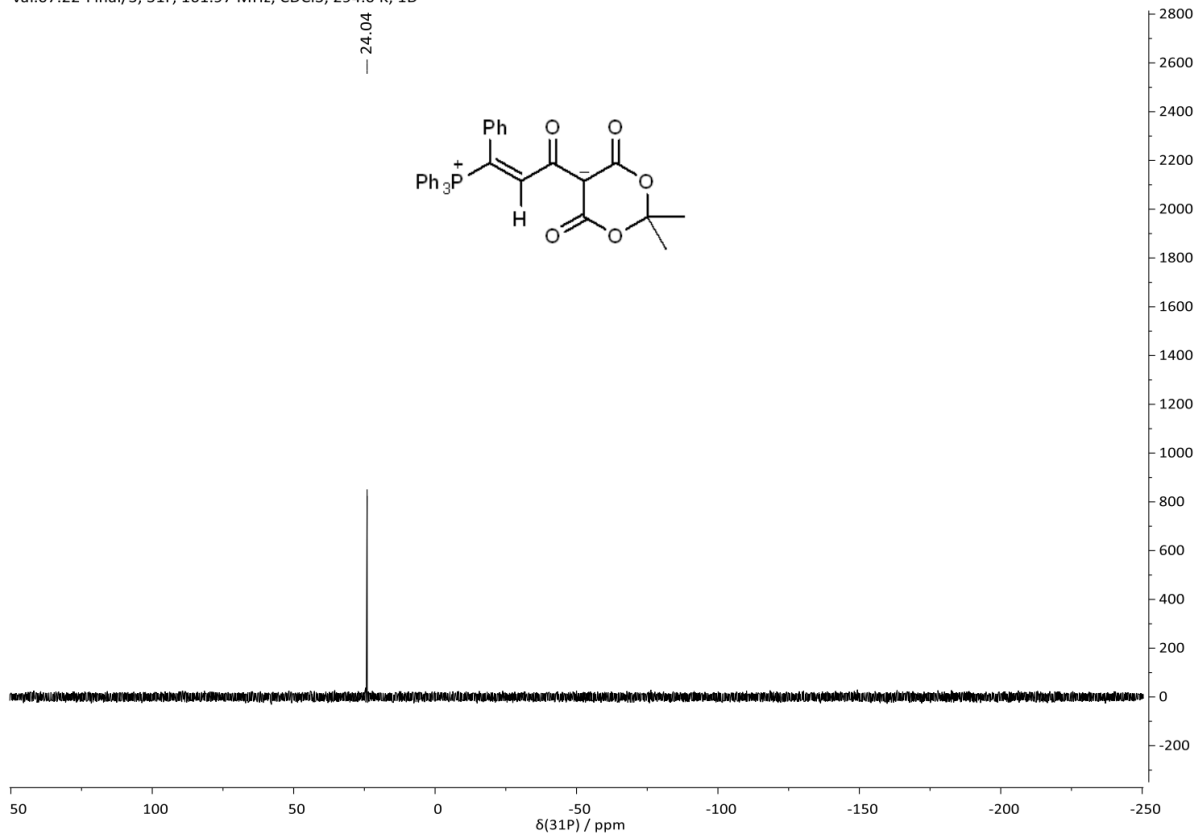
1. Fiore, V. A.; Maas, G. *Tetrahedron* **2019**, *75*, 3586–3595. DOI: 10.100.626/j.tet.2019.05.027.
2. SHELXS97: Sheldrick, G. M. *Acta Crystallogr.* **2008**, *A64*, 112–122.
3. SHELXL-2014: Sheldrick, G. M. Crystal Structure Refinement with SHELXL. *Acta Crystallogr.* **2015**, *C71*, 3–8.
4. ORTEP-3 for Windows: Farrugia, L. J. *J. Appl. Crystallogr.* **2012**, *45*, 849–854.
5. SIR97: Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G. L.; Giacovazzo, C.; Guagliardi, A.; Moliterni, A. G. G.; Polidori, G.; Spagna, R. *J. Appl. Cryst.* **1999**, *32*, 115–119.

vaf.07.22-05_RT/4, ¹³C, 100.62 MHz, 2000 Scans, CDCl₃, 300.0 K, 1D

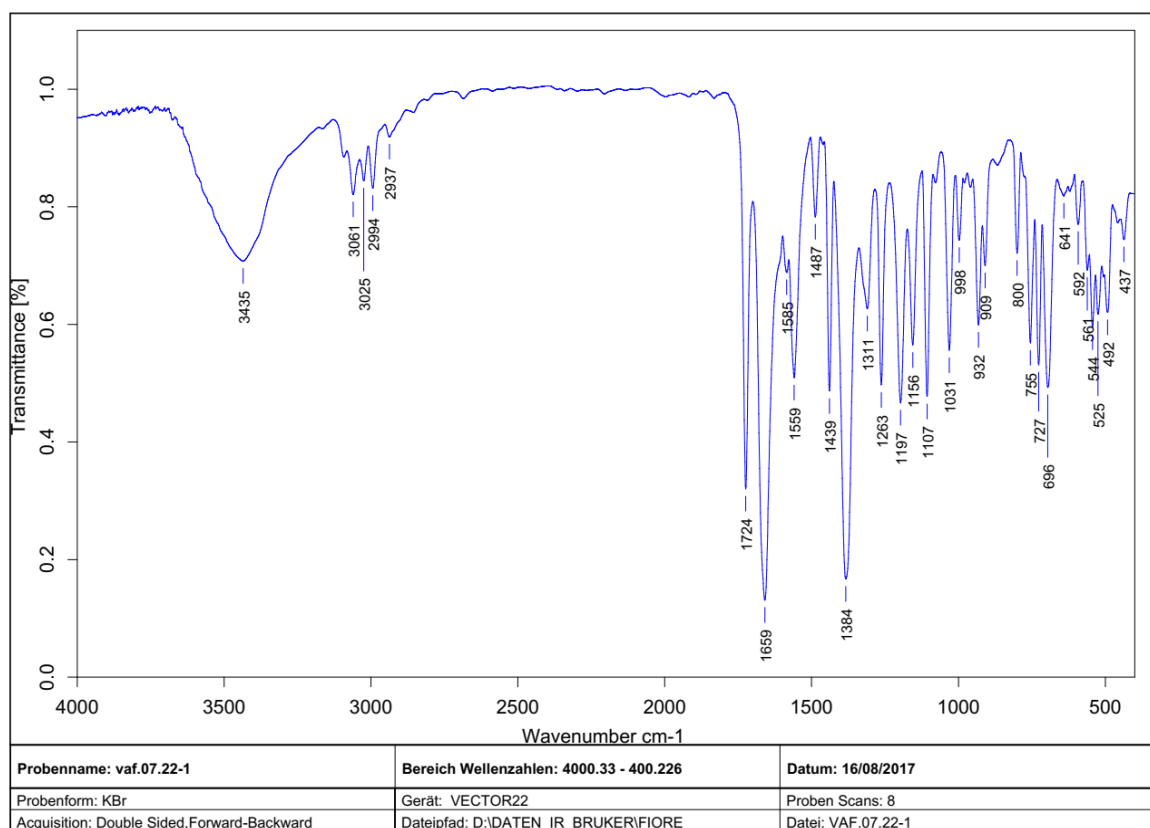


³¹P NMR

vaf.07.22-Final/3, ³¹P, 161.97 MHz, CDCl₃, 294.0 K, 1D

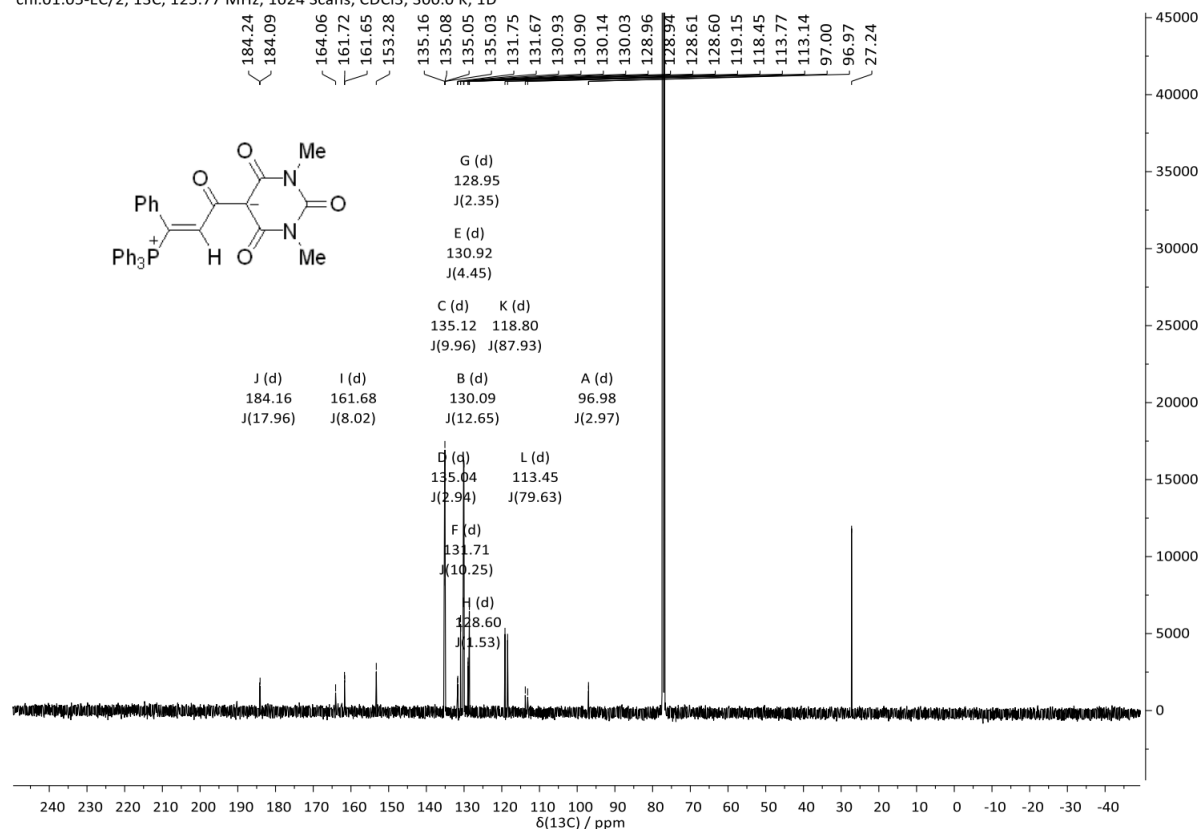


IR

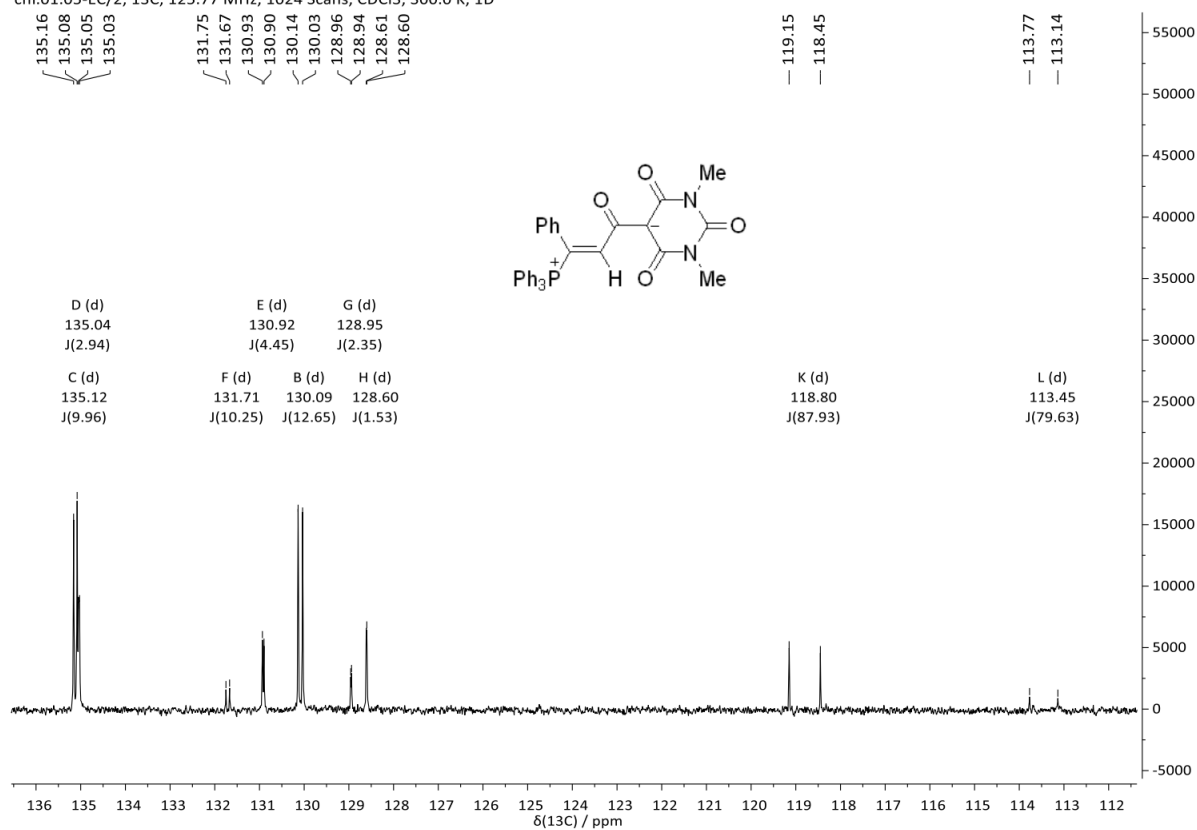


¹³C NMR

chf.01.05-EC/2, ¹³C, 125.77 MHz, 1024 Scans, CDCl₃, 300.0 K, 1D

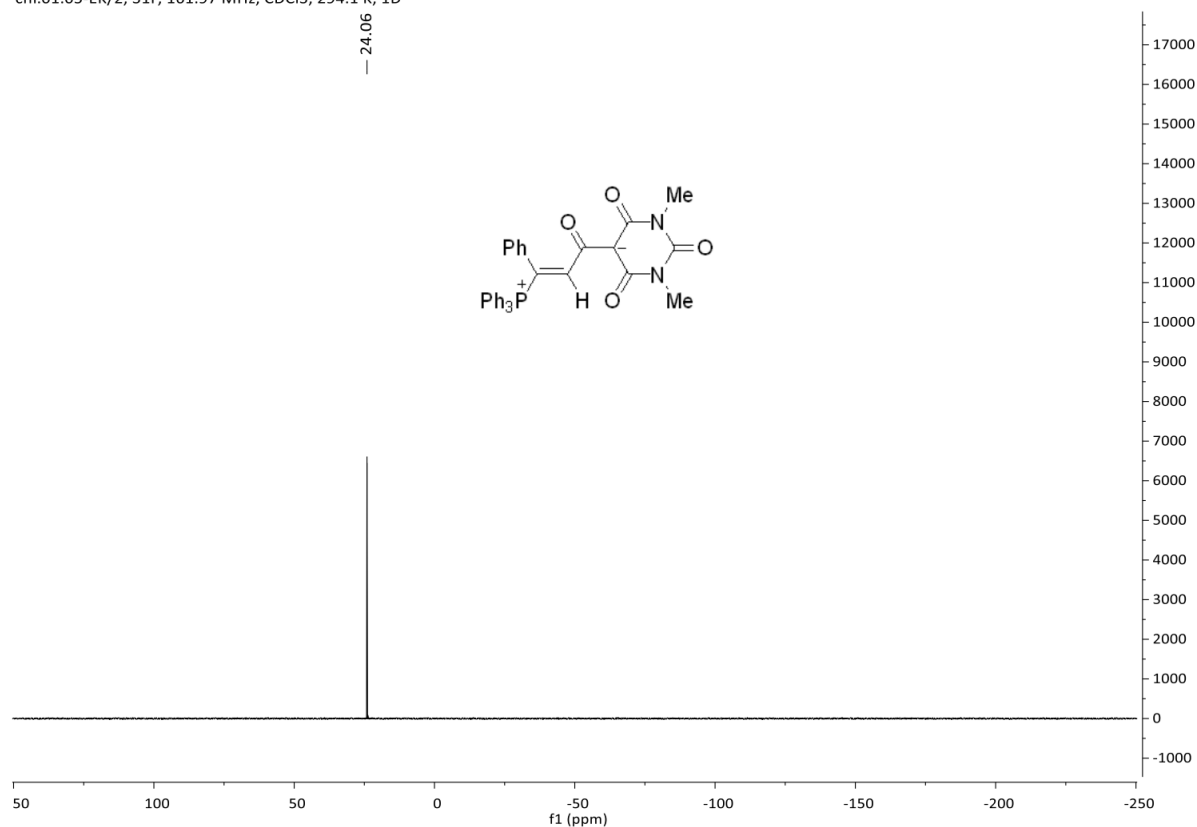


chf.01.05-EC/2, ¹³C, 125.77 MHz, 1024 Scans, CDCl₃, 300.0 K, 1D

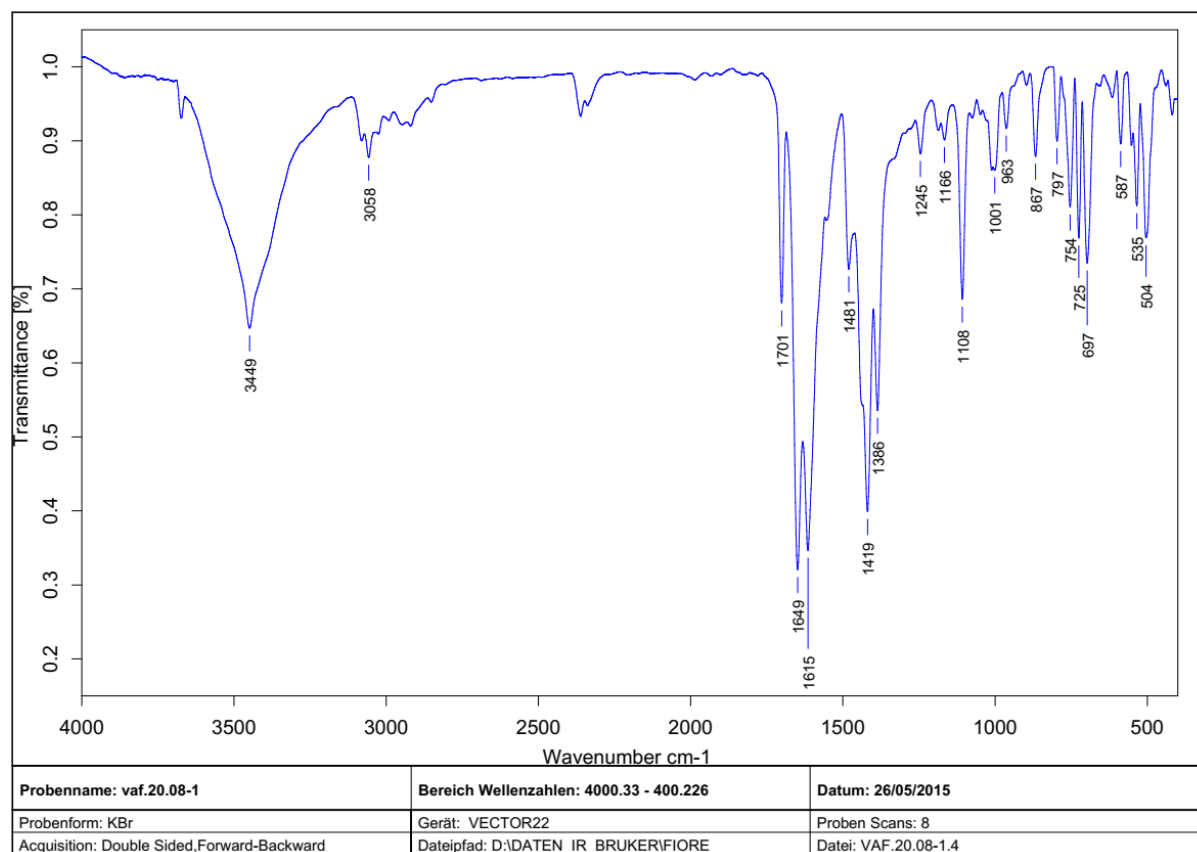


³¹P NMR

chf.01.05-EK/2, 31P, 161.97 MHz, CDCl₃, 294.1 K, 1D



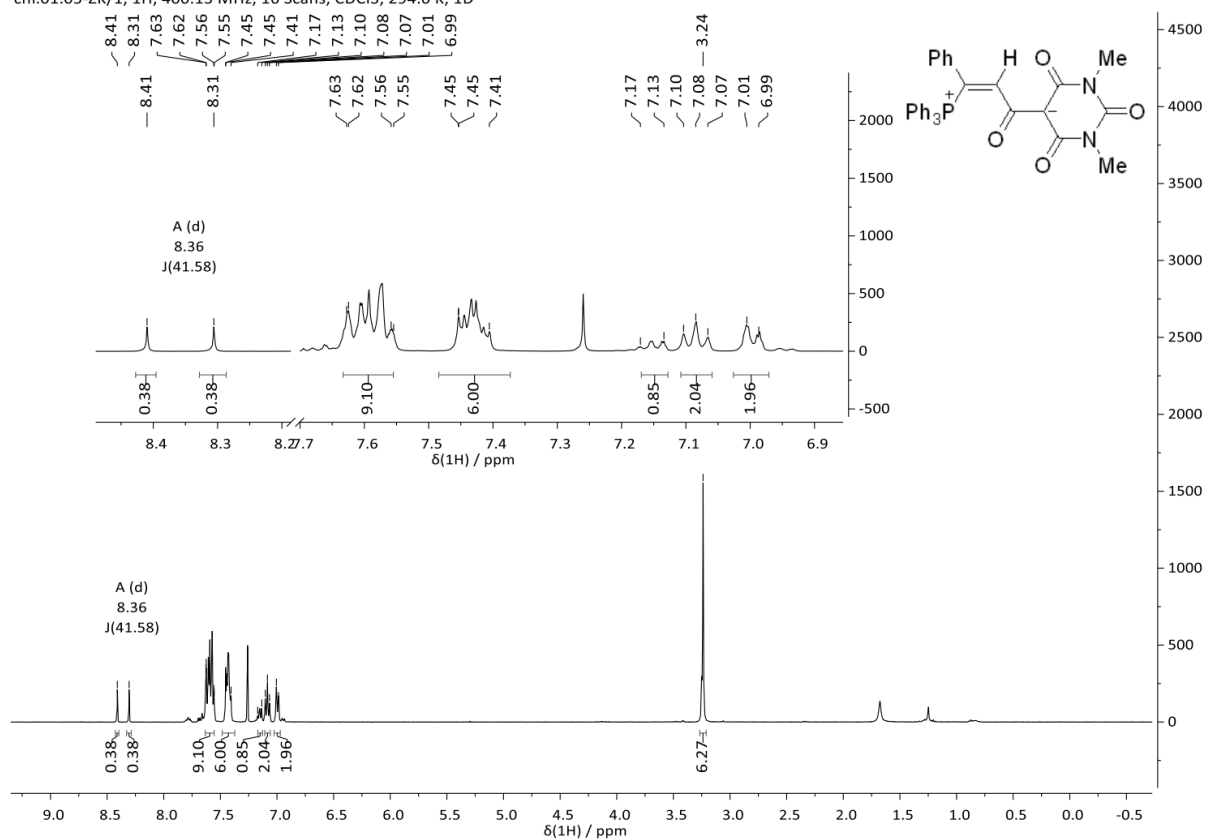
IR



(Z)-3b

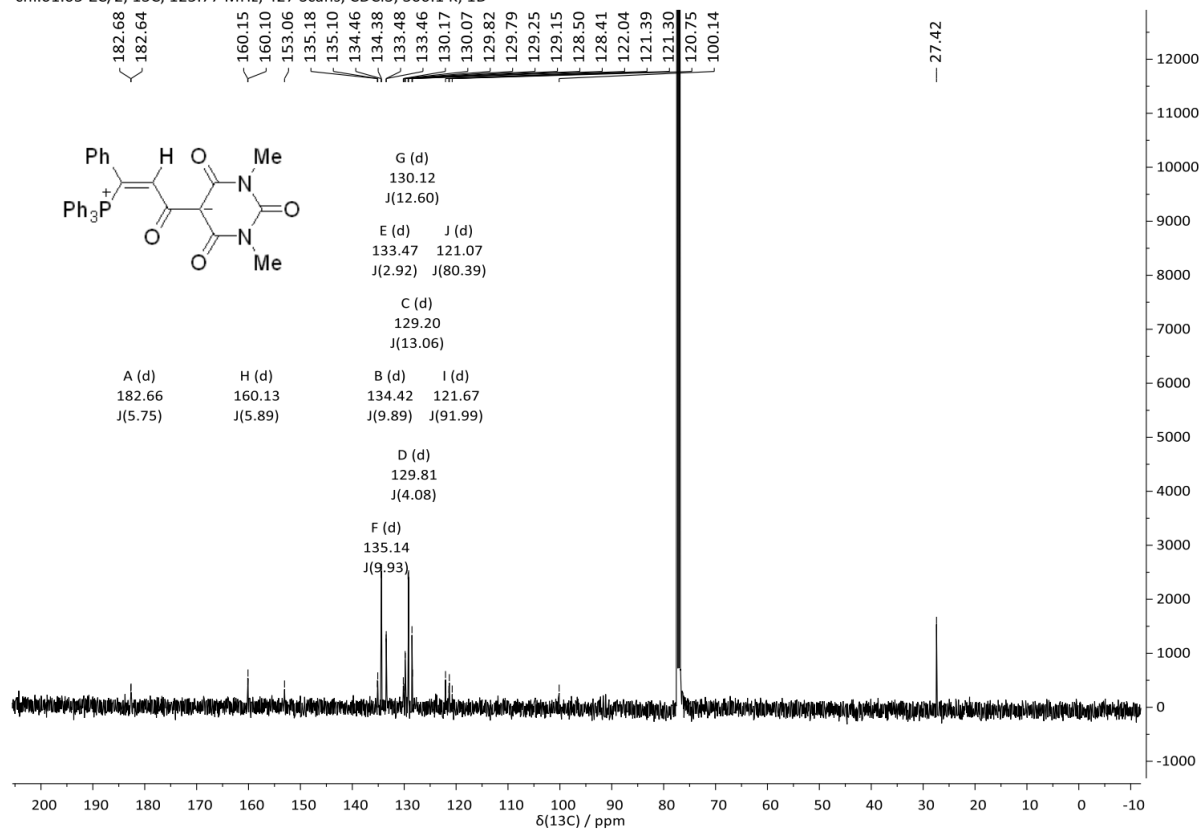
¹H NMR

chf.01.05-ZK/1, 1H, 400.13 MHz, 16 Scans, CDCl₃, 294.0 K, 1D

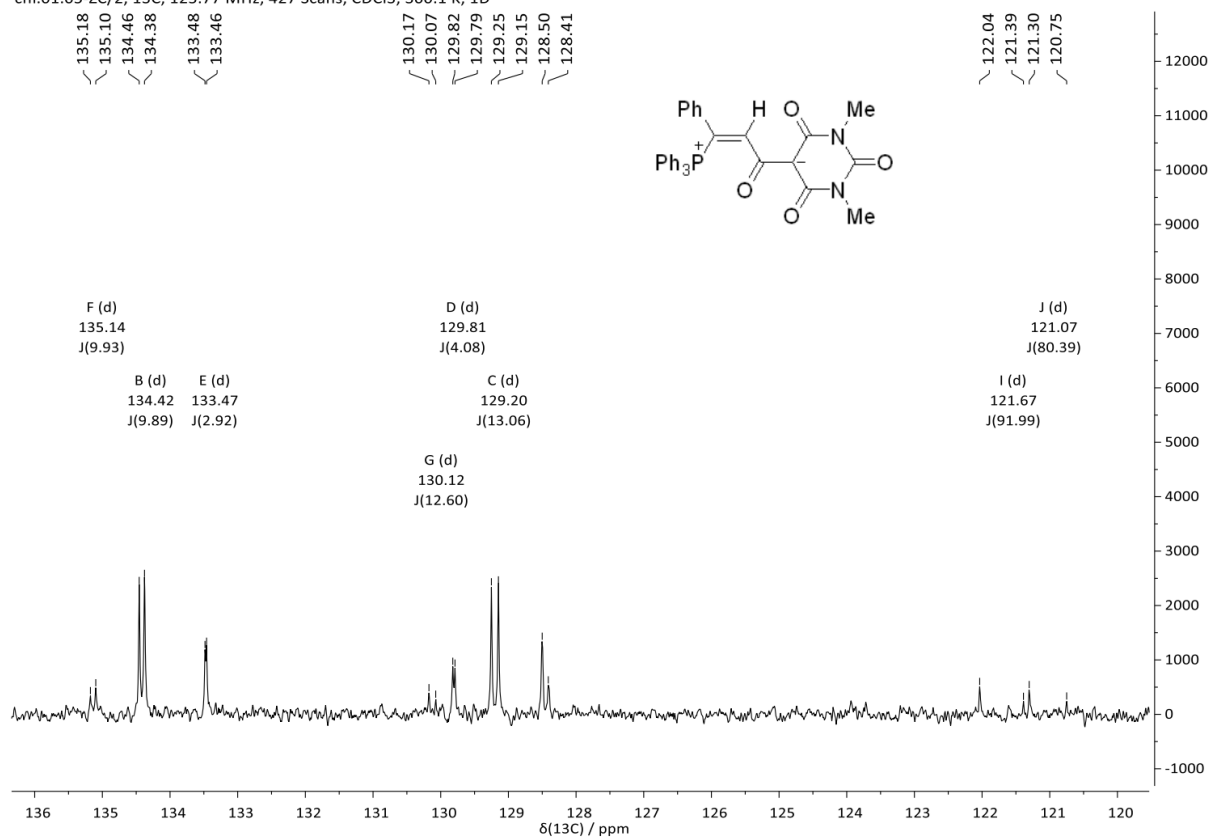


¹³C NMR

chf.01.05-ZC/2, 13C, 125.77 MHz, 427 Scans, CDCl₃, 300.1 K, 1D

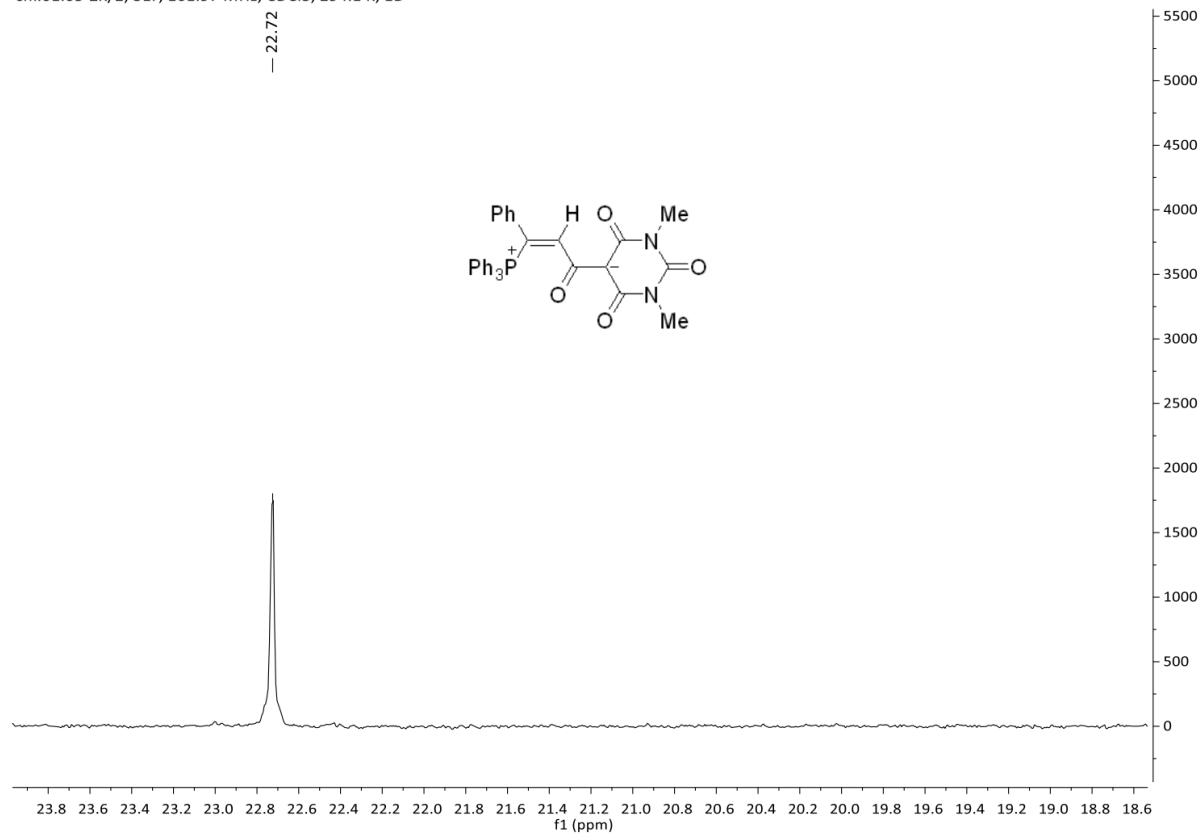


chf.01.05-ZC/2, ¹³C, 125.77 MHz, 427 Scans, CDCl₃, 300.1 K, 1D

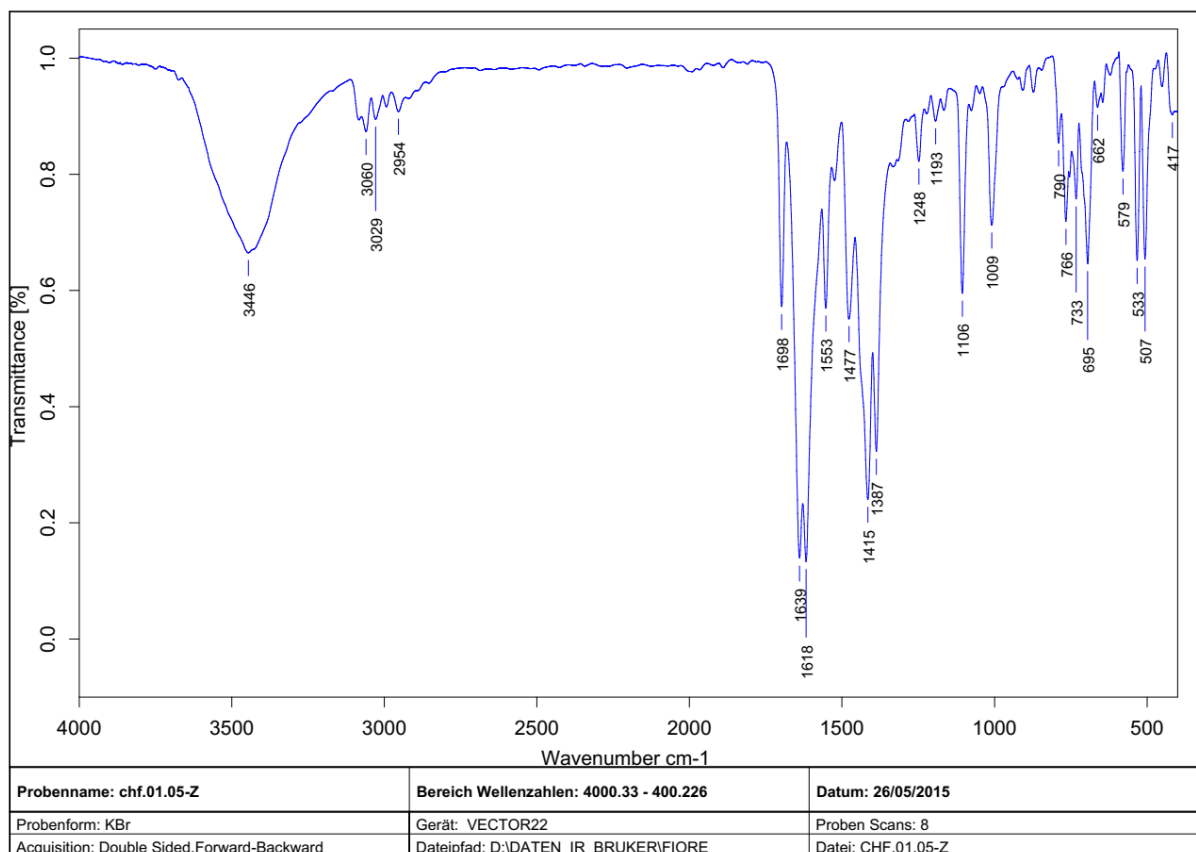


³¹P NMR

chf.01.05-ZK/2, ³¹P, 161.97 MHz, CDCl₃, 294.1 K, 1D



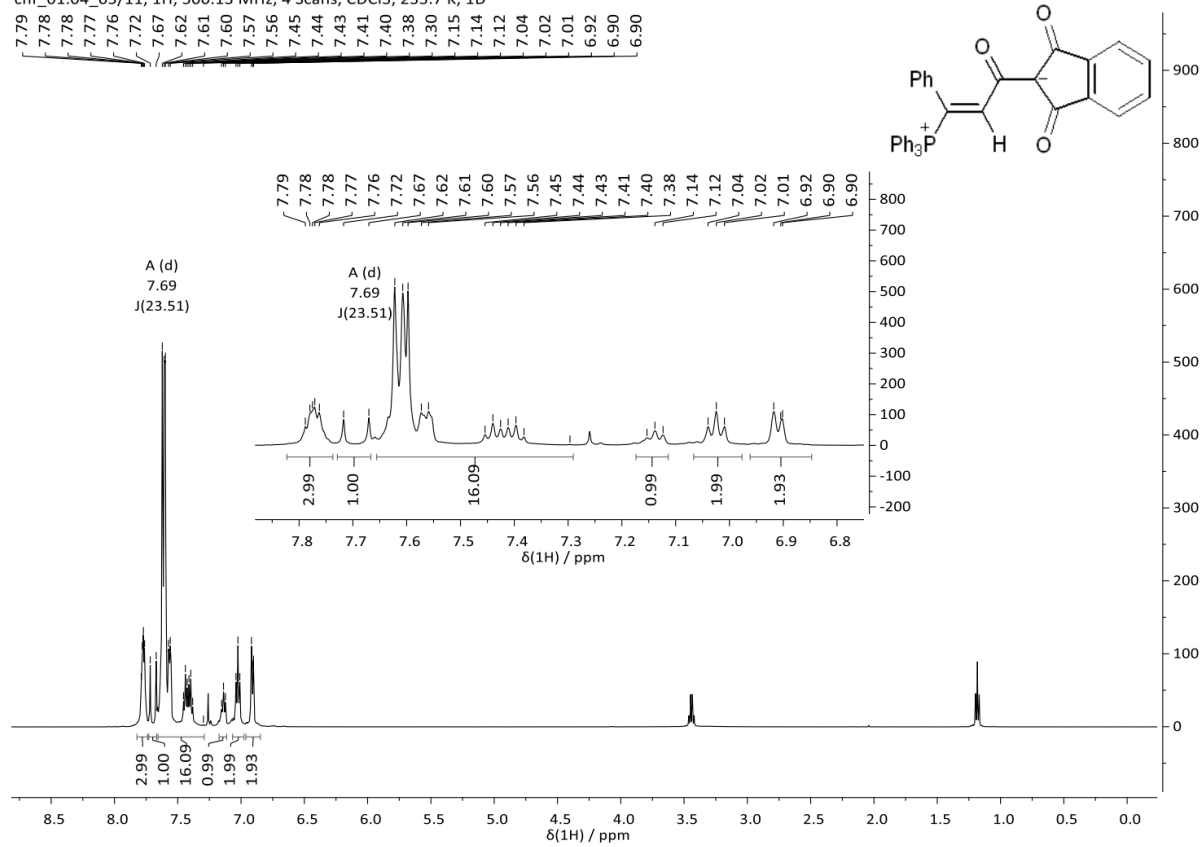
IR



4.3. Betaine (*E*)-3c

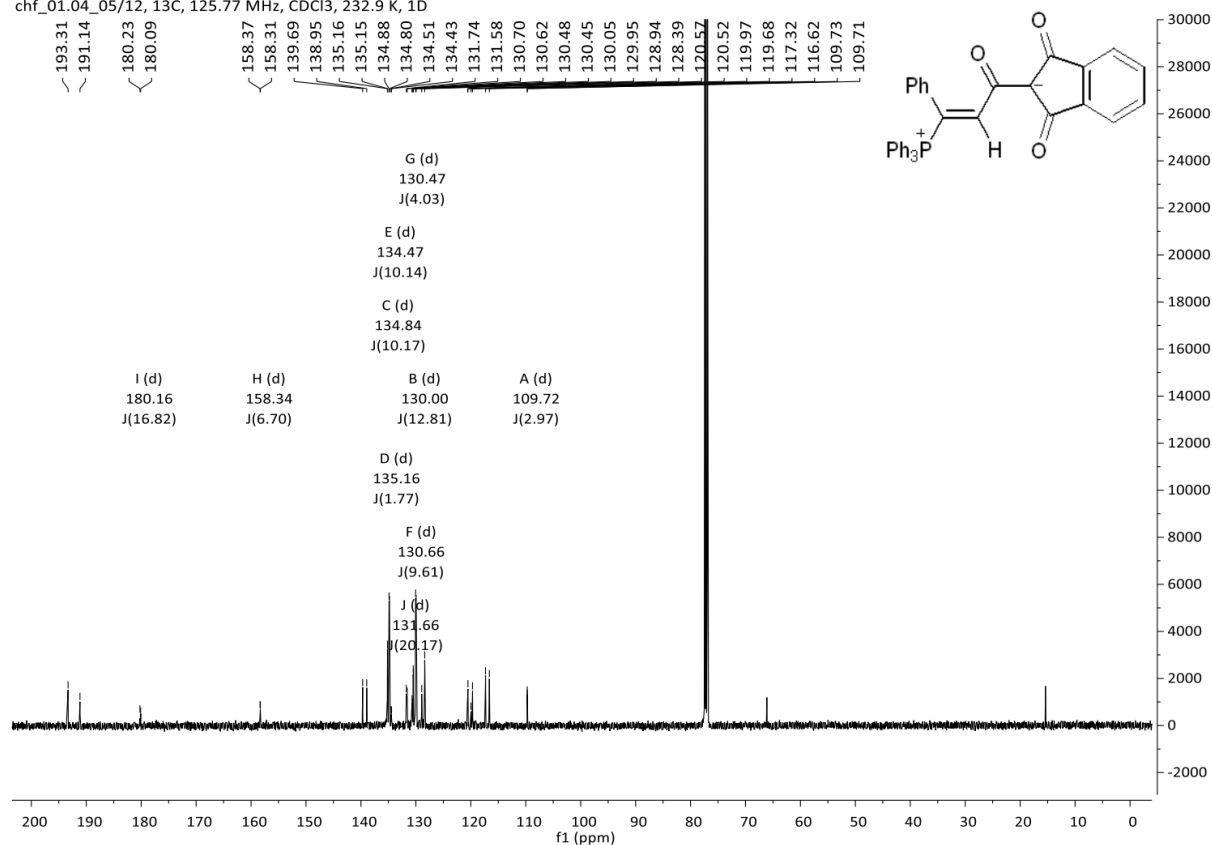
^1H NMR

chf_01.04_05/11, 1H, 500.13 MHz, 4 Scans, CDCl₃, 233.7 K, 1D

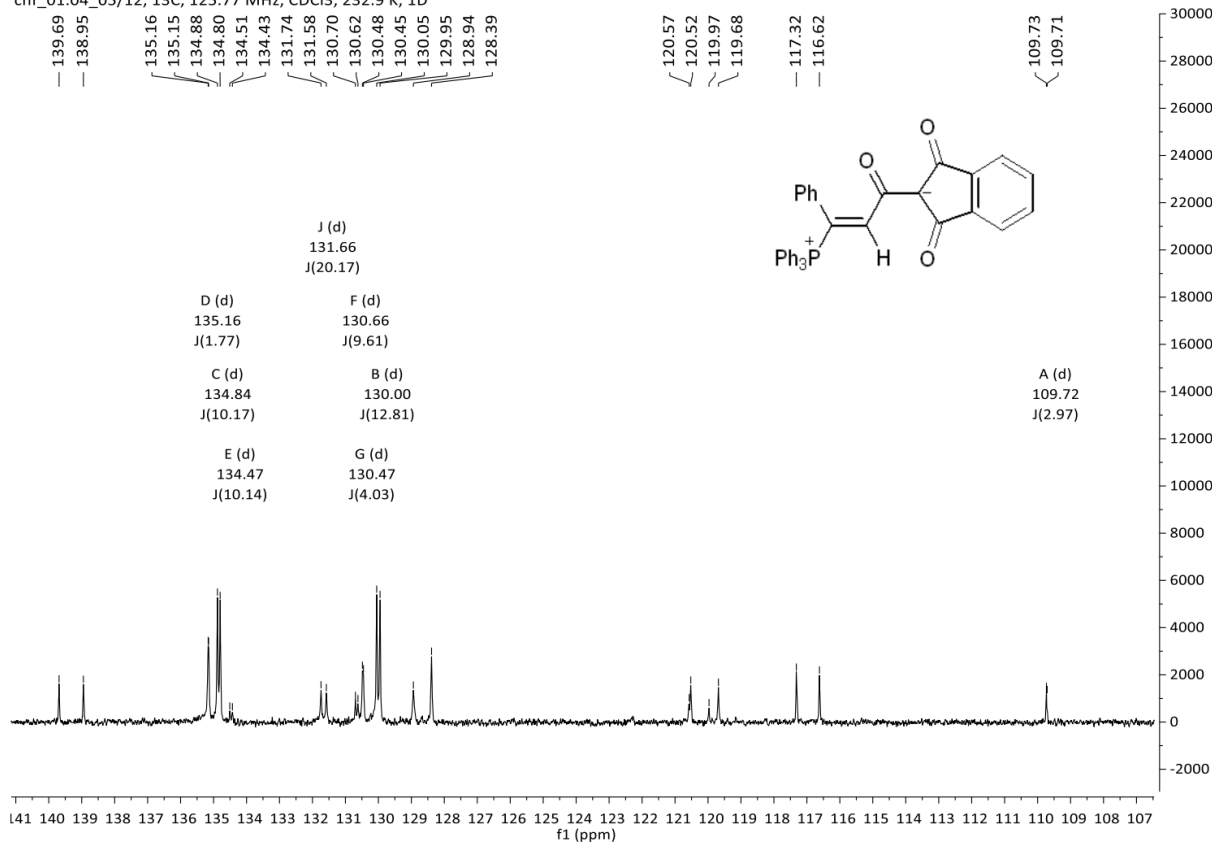


¹³C NMR

chf_01.04_05/12, ¹³C, 125.77 MHz, CDCl₃, 232.9 K, 1D

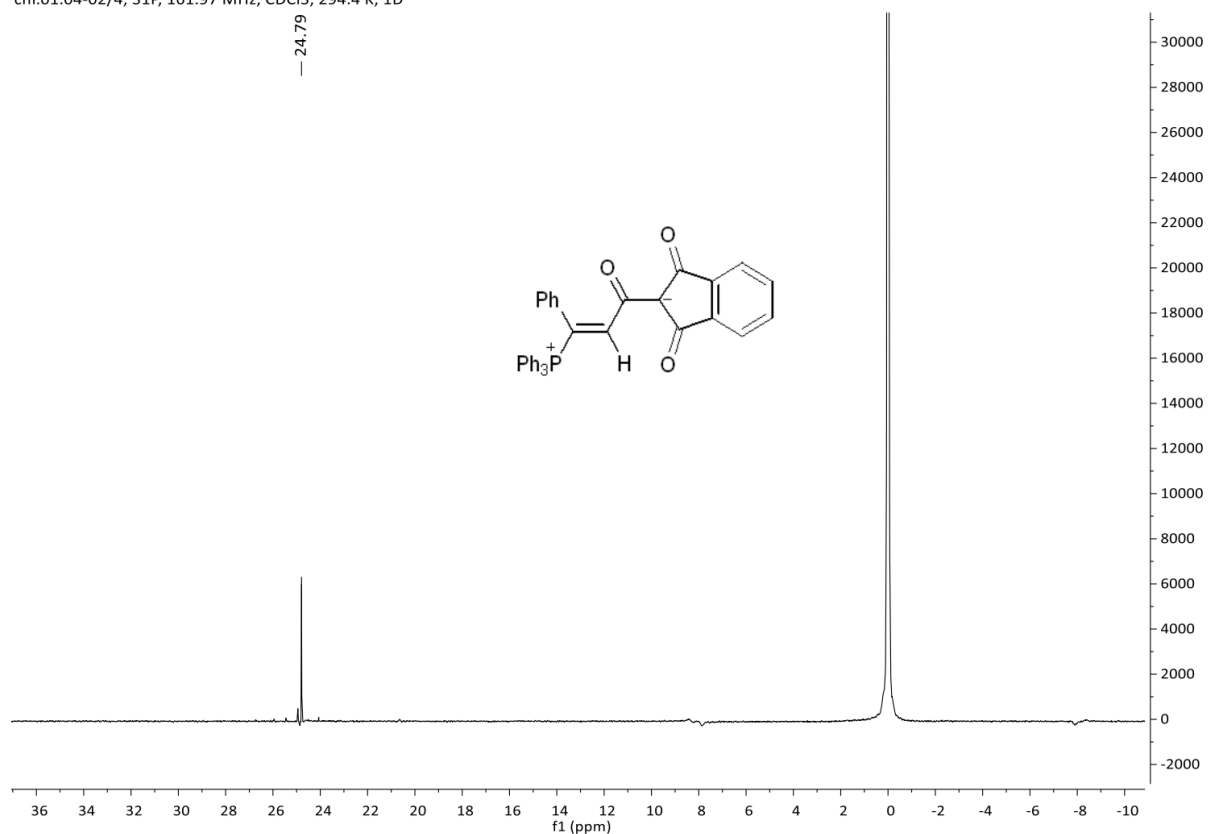


chf_01.04_05/12, ¹³C, 125.77 MHz, CDCl₃, 232.9 K, 1D

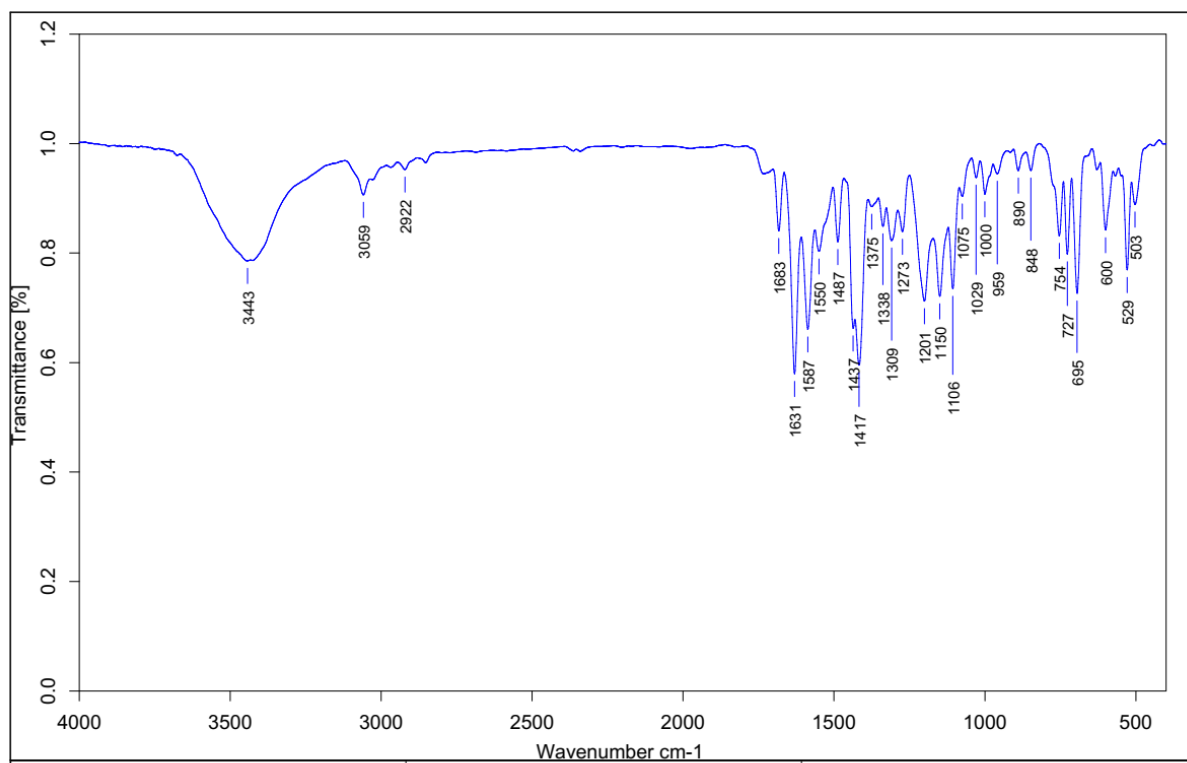


³¹P NMR

chf.01.04-02/4, 31P, 161.97 MHz, CDCl₃, 294.4 K, 1D



IR

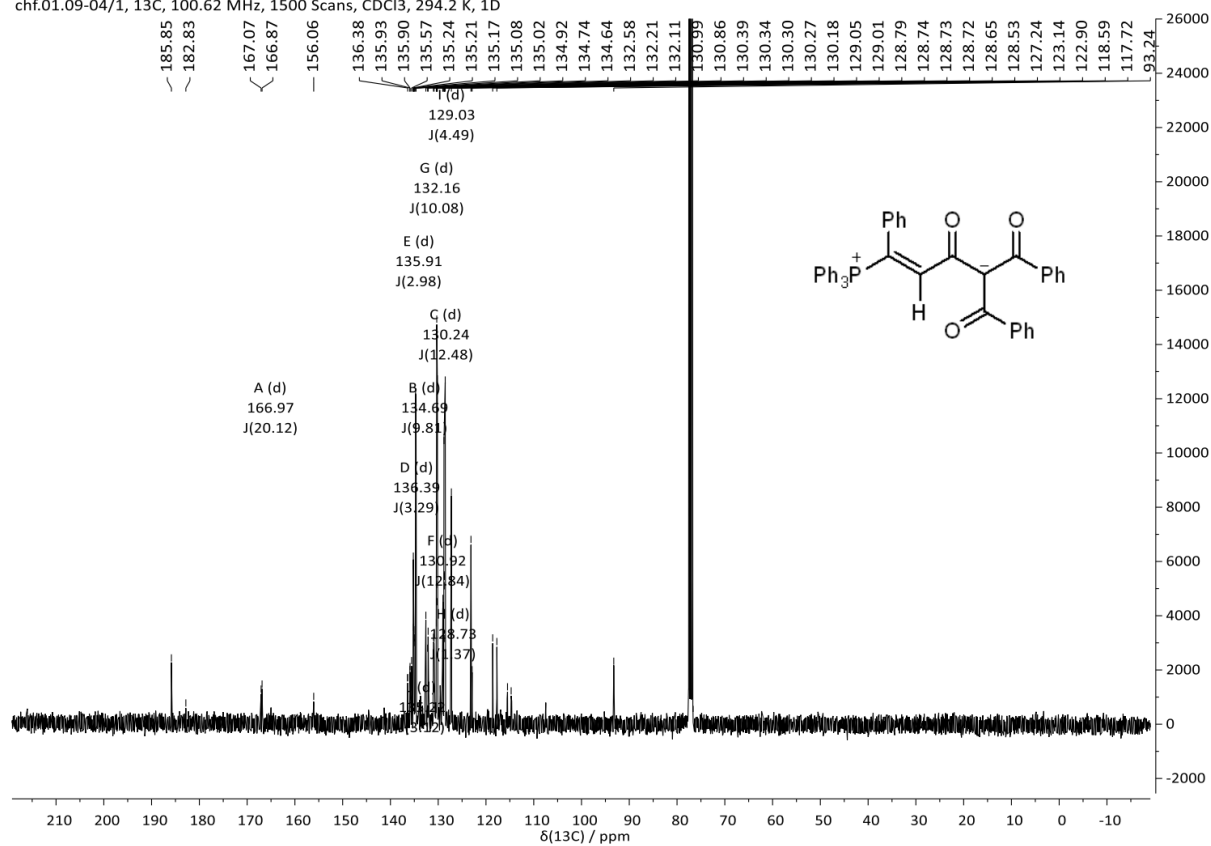


Probenname: chf.01.04-2	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 18/03/2015
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided,Forward-Backward	Dateipfad: D:\DATEN_IR_BRUKER\FIORE	Datei: CHF.01.04-2

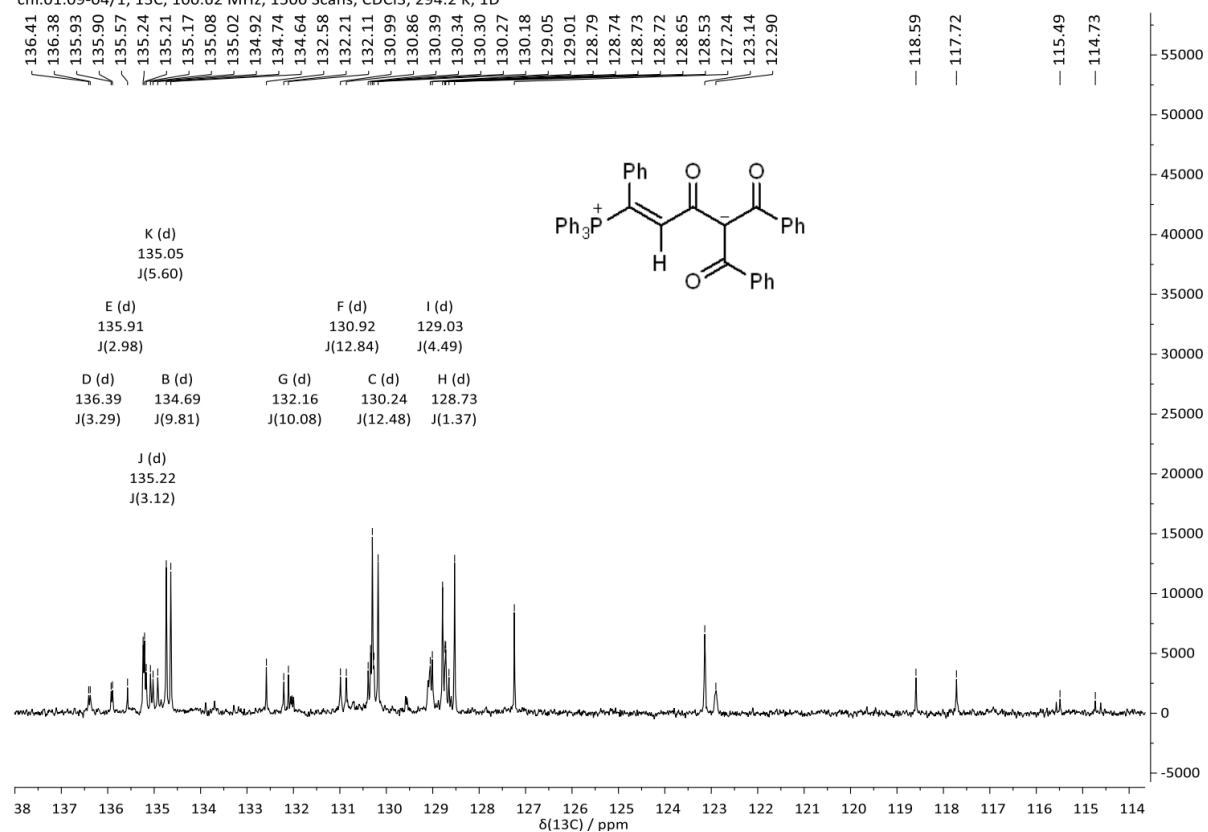
4.4. Betaine (*E*)-3d

¹³C NMR

chf.01.09-04/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

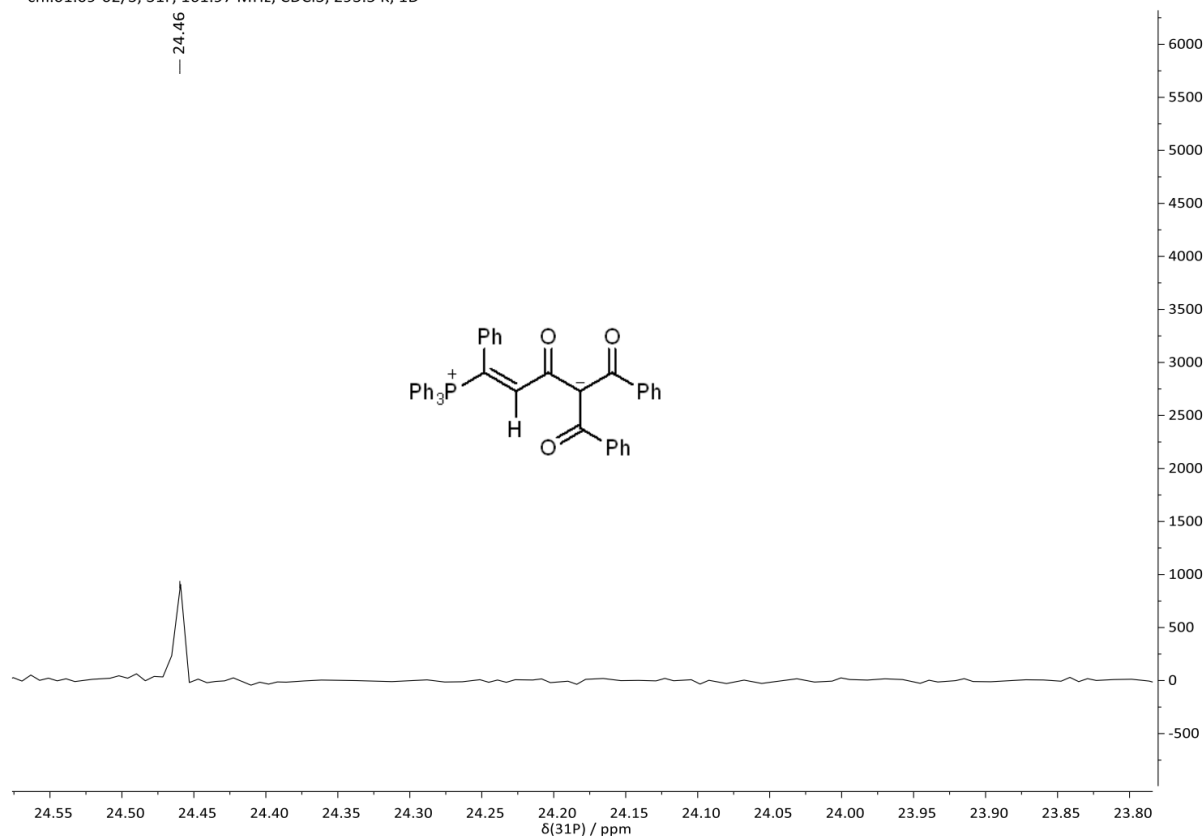


chf.01.09-04/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

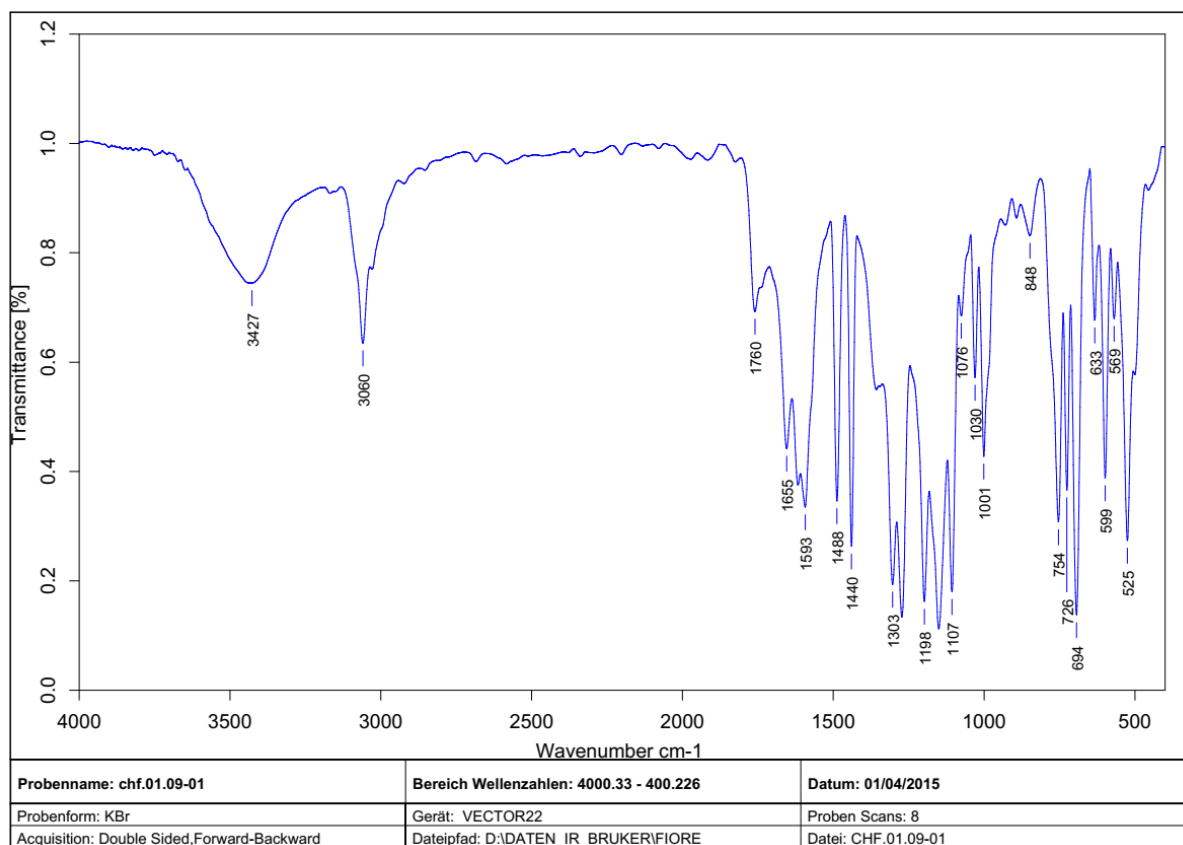


³¹P NMR

chf.01.09-02/3, 31P, 161.97 MHz, CDCl₃, 293.5 K, 1D



IR

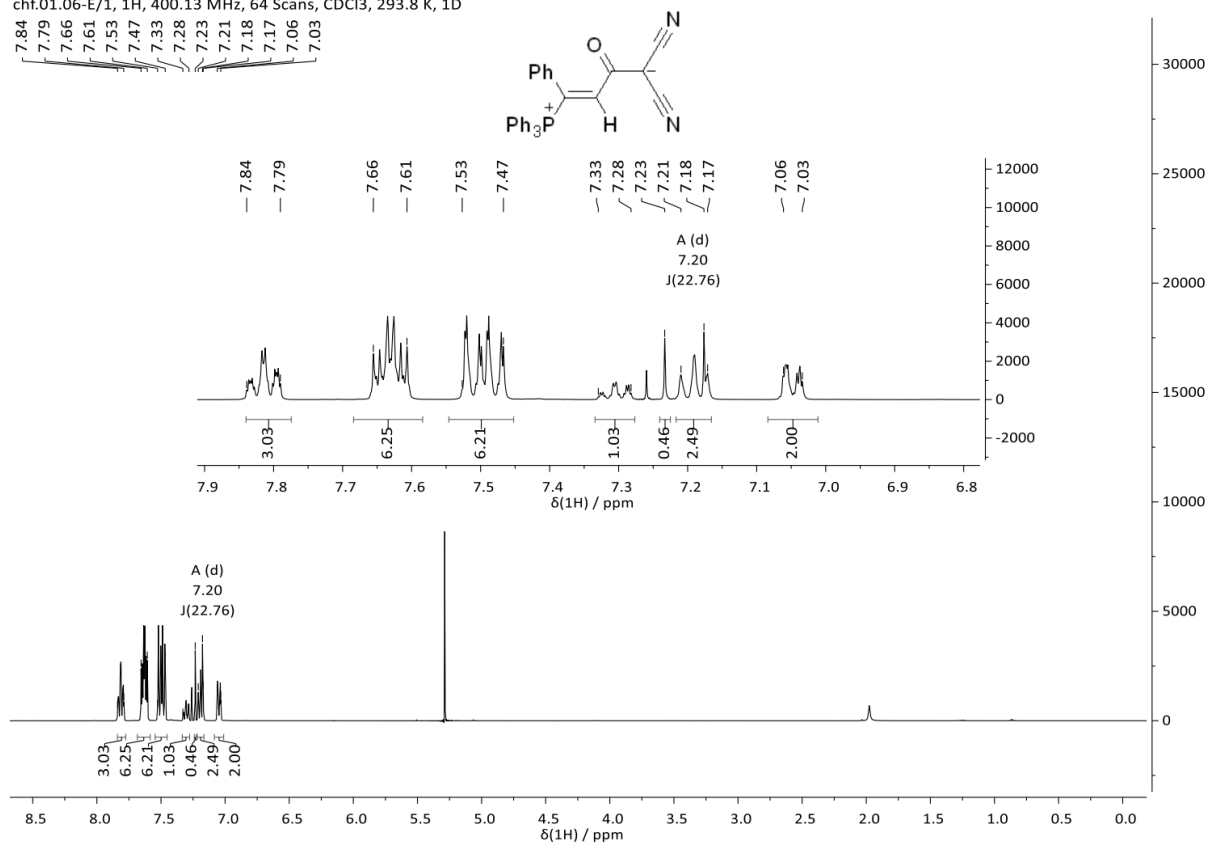


4.5. Betaines (E)- and (Z)-3e

(E)-3e

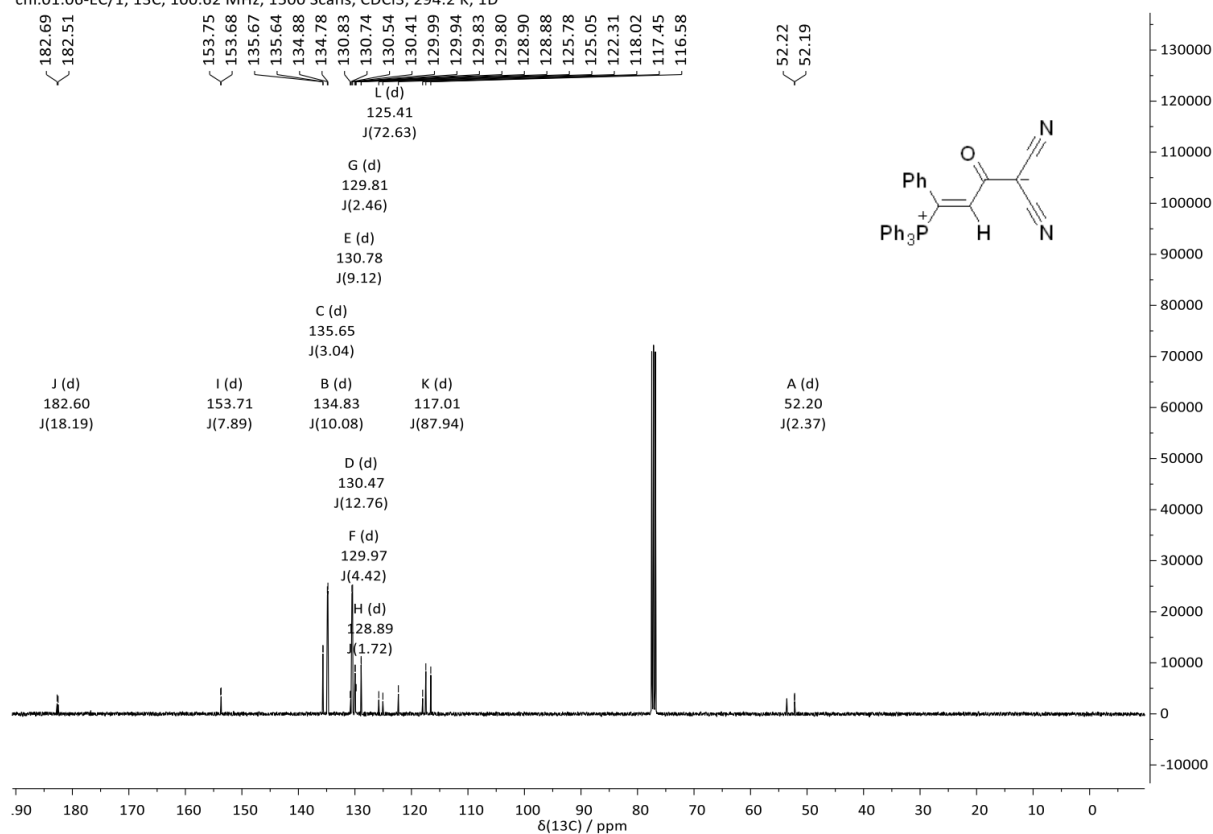
¹H NMR

chf.01.06-E/1, 1H, 400.13 MHz, 64 Scans, CDCl₃, 293.8 K, 1D

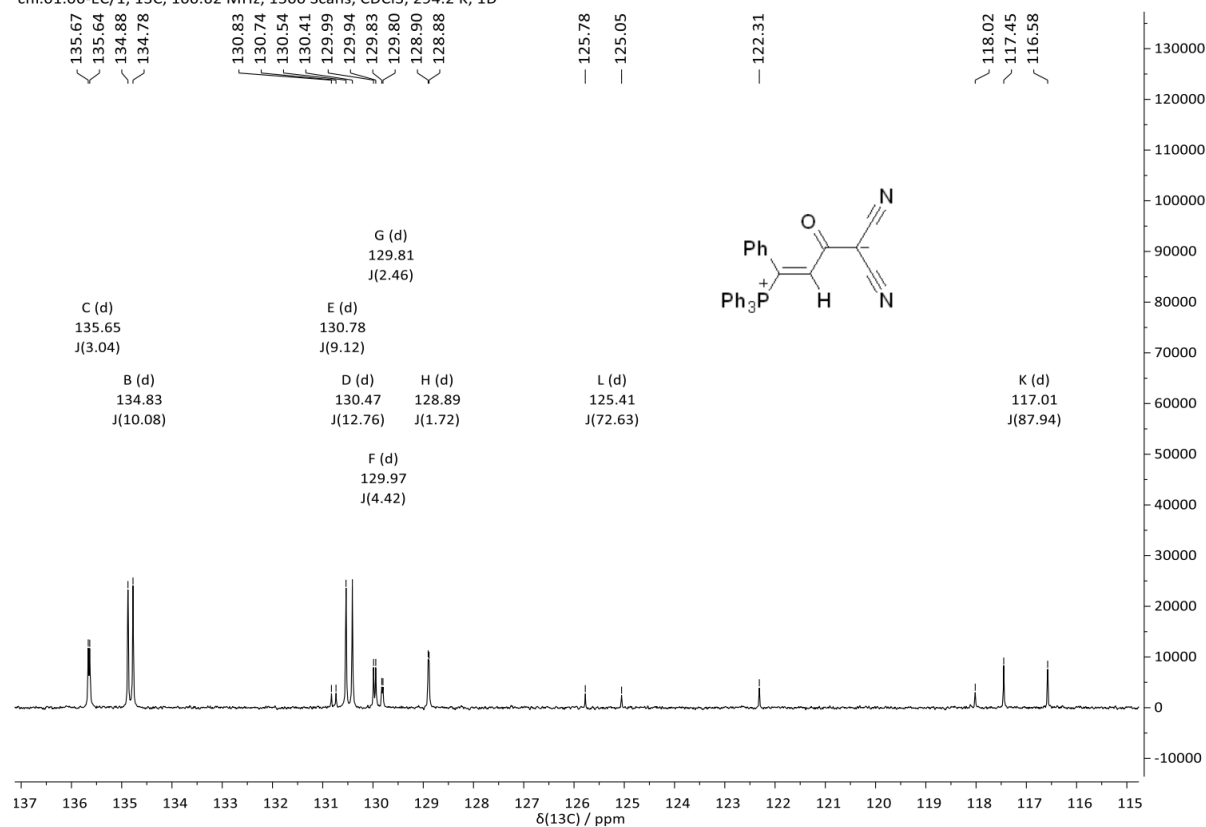


¹³C NMR

chf.01.06-EC/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

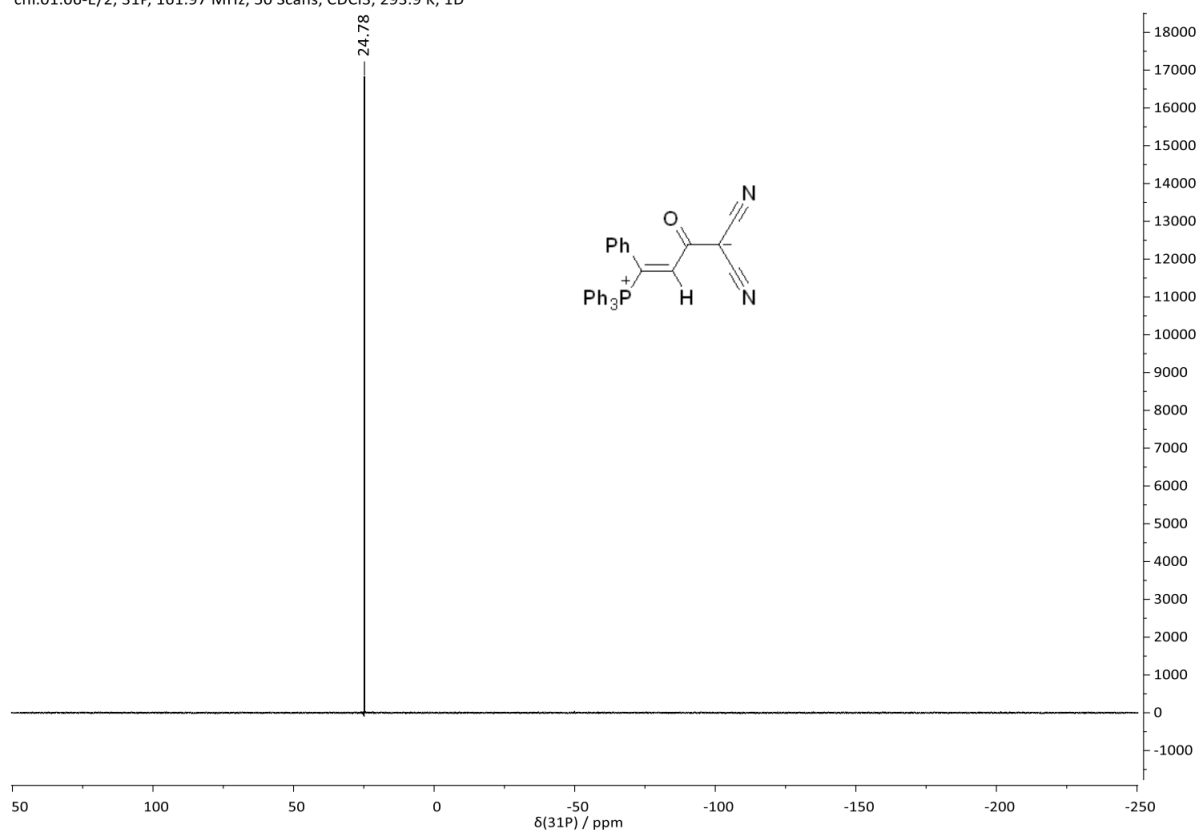


chf.01.06-EC/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

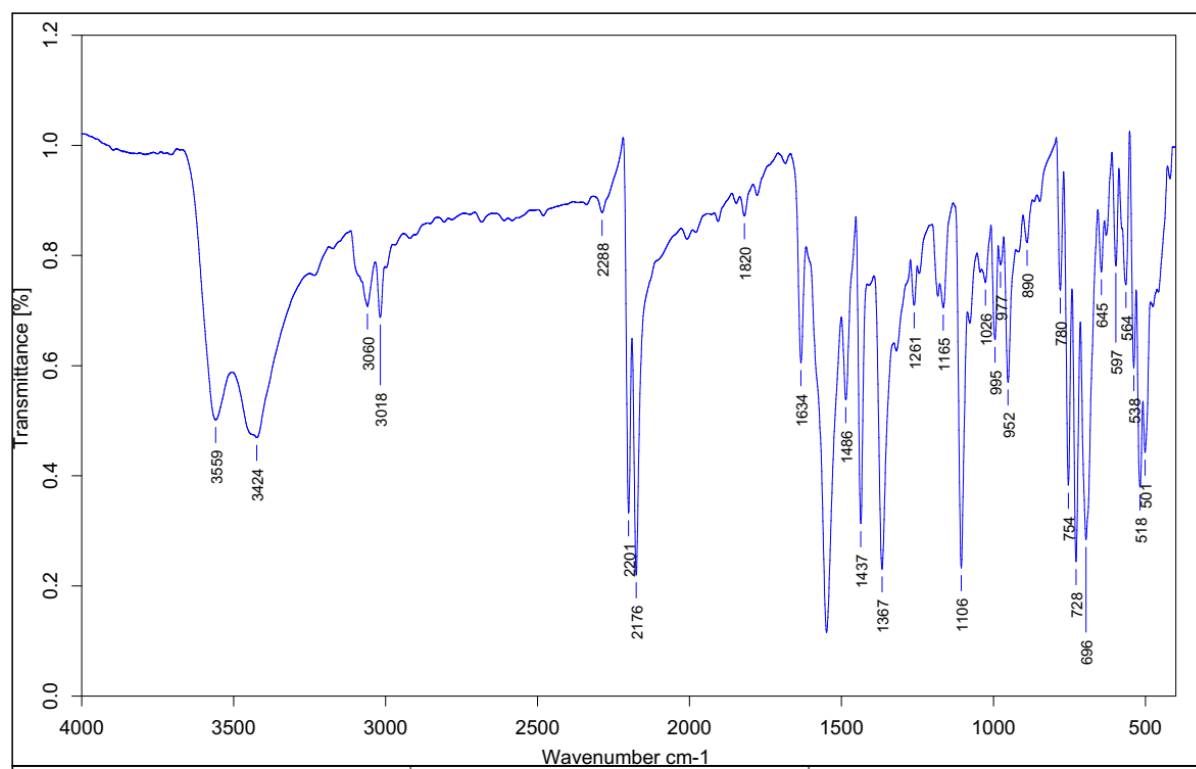


³¹P NMR

chf.01.06-E/2, 31P, 161.97 MHz, 50 Scans, CDCl₃, 293.9 K, 1D



IR

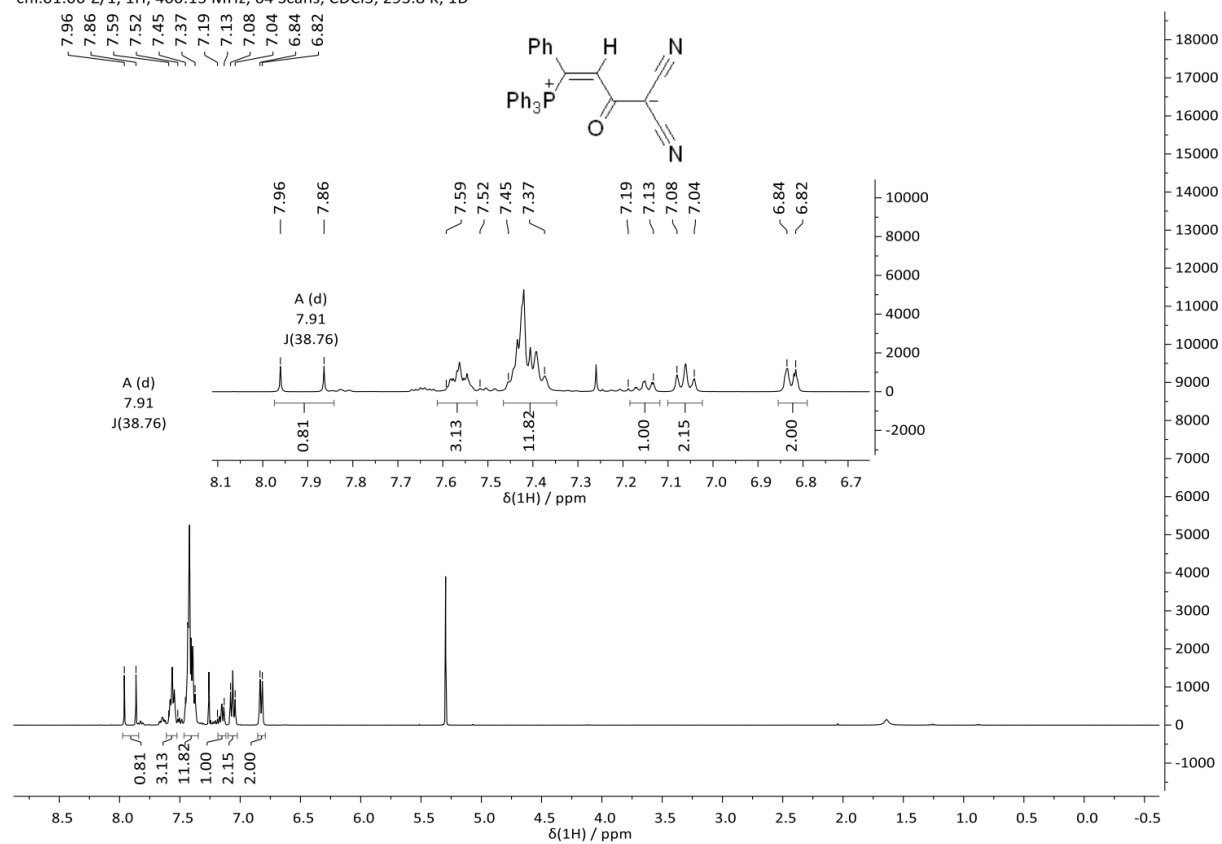


Probenname: chf.01.06-E	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 26/05/2015
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided, Forward-Backward	Dateipfad: D:\DATEN_IR_BRUKER\FIORE	Datei: CHF.01.06-E

(Z)-3e

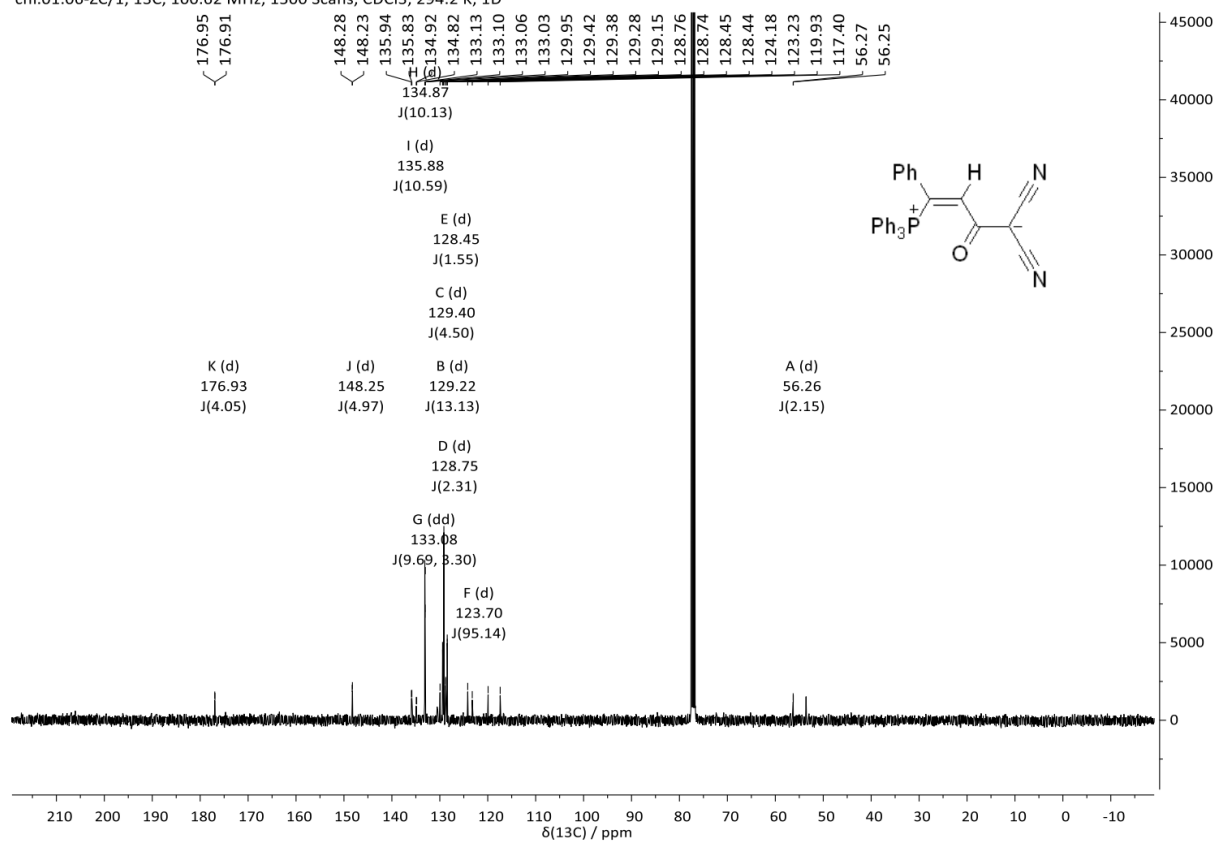
¹H NMR

chf.01.06-Z/1, 1H, 400.13 MHz, 64 Scans, CDCl₃, 293.8 K, 1D

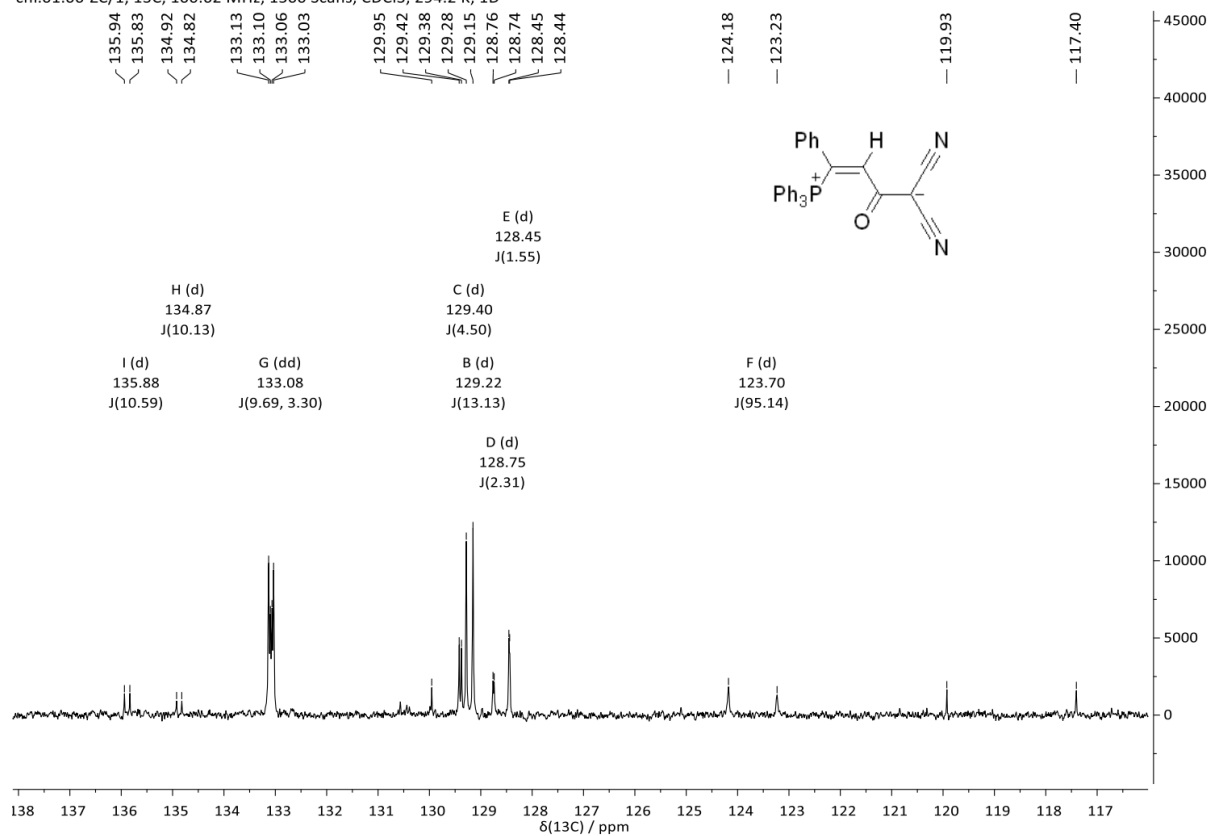


¹³C NMR

chf.01.06-ZC/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

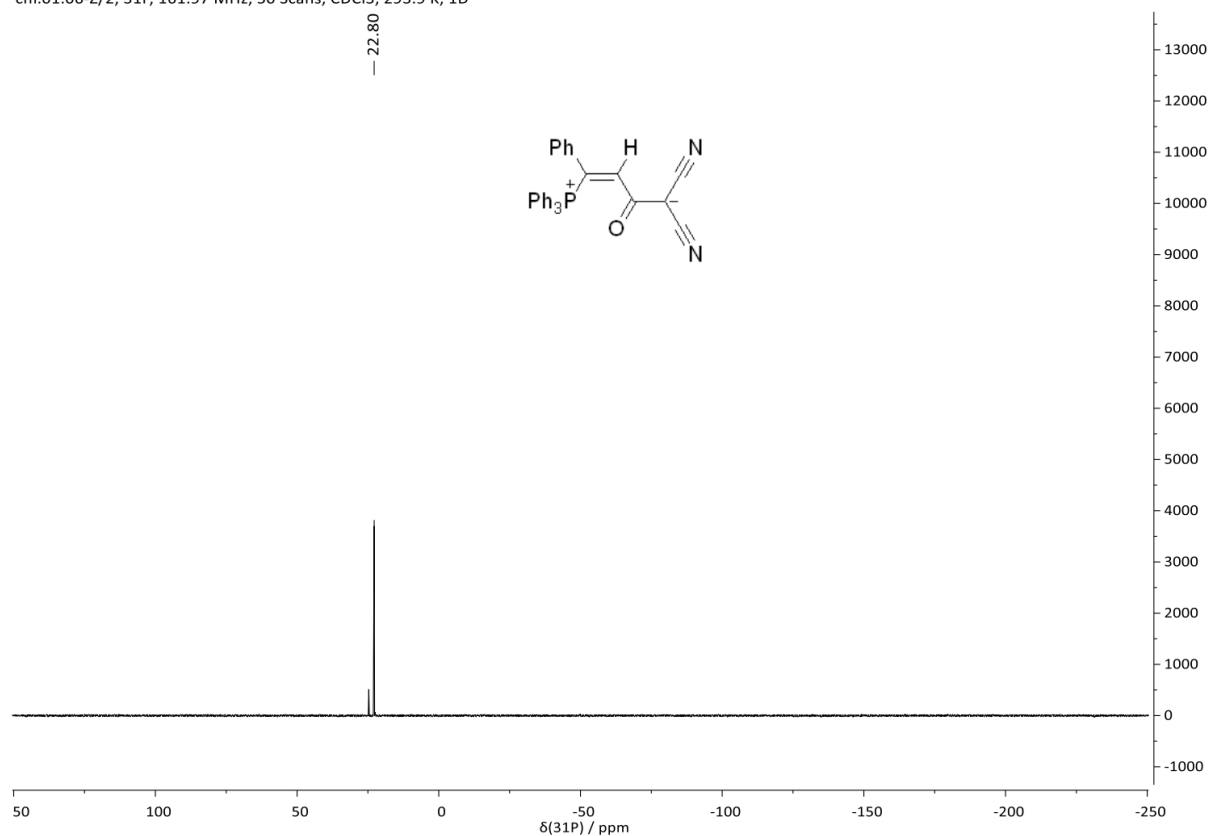


chf.01.06-ZC/1, ¹³C, 100.62 MHz, 1500 Scans, CDCl₃, 294.2 K, 1D

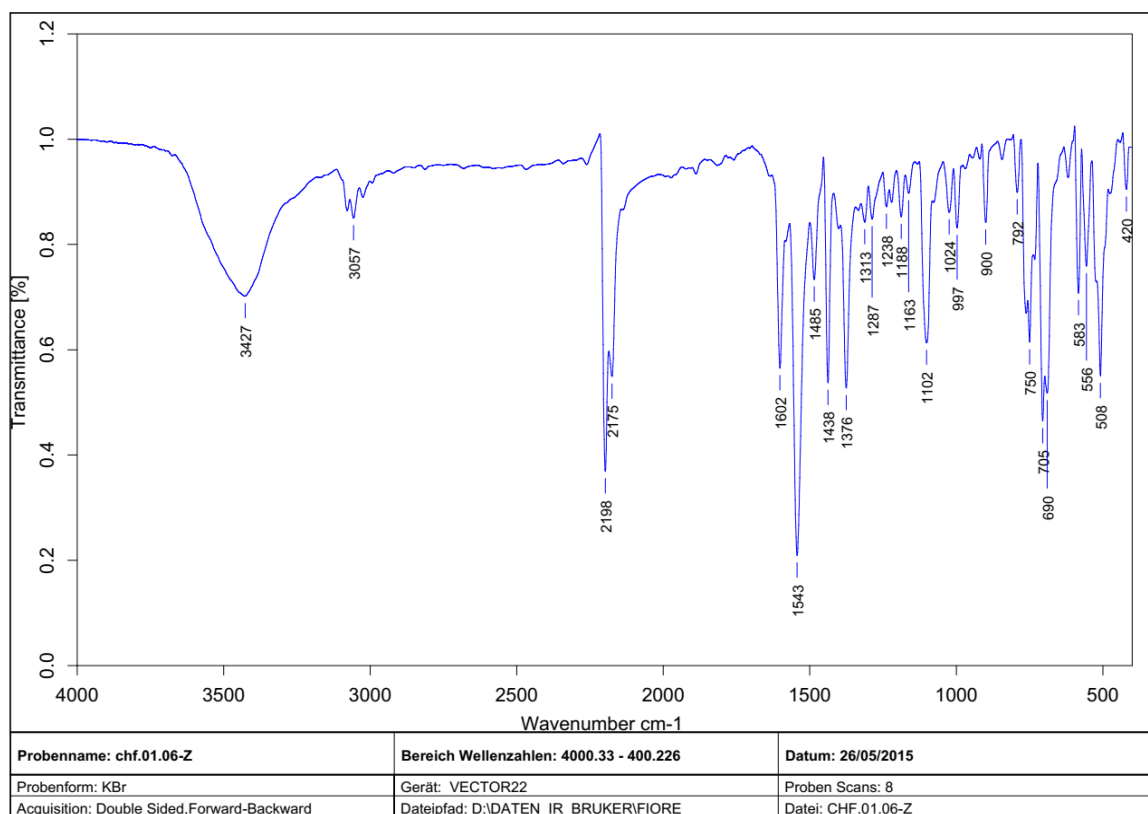


³¹P NMR

chf.01.06-Z/2, 31P, 161.97 MHz, 50 Scans, CDCl₃, 293.9 K, 1D



IR

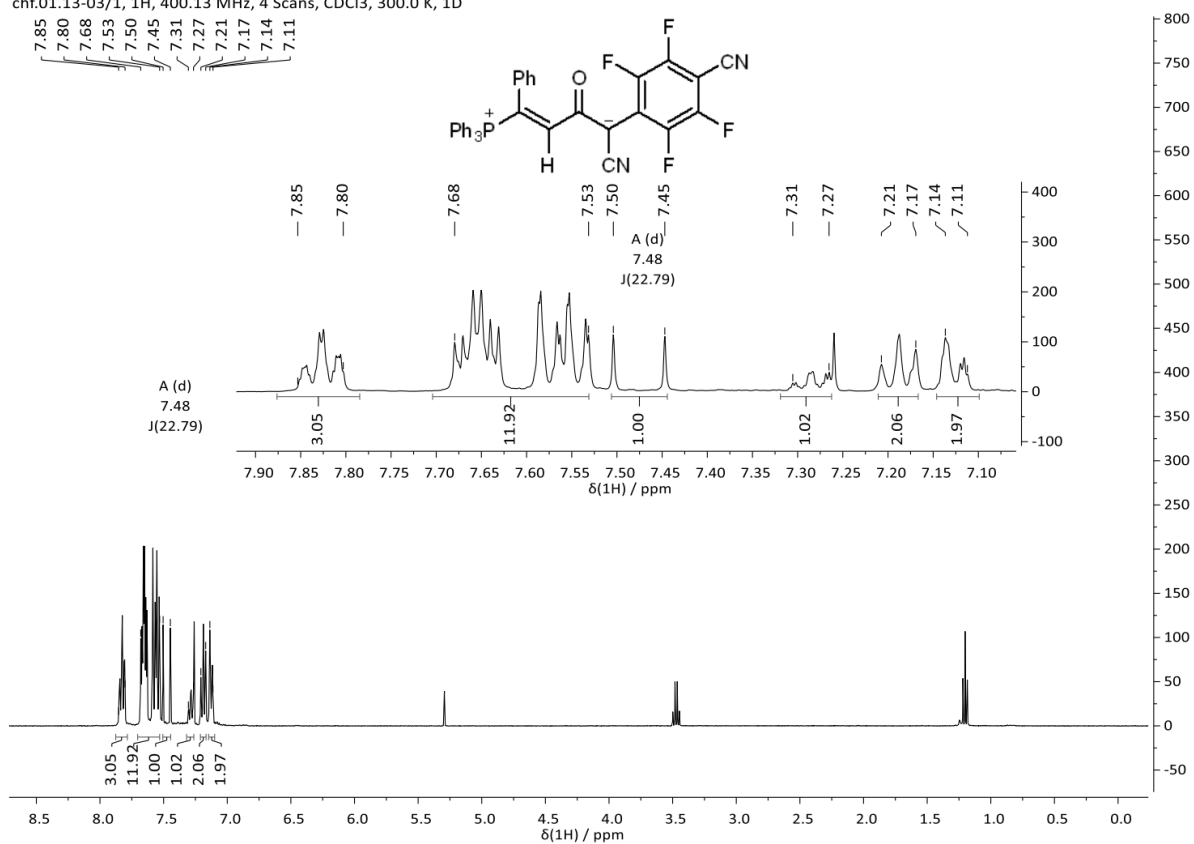


4.6. Betaines (*E*)- and (*Z*)-3f

(*E*)-3f

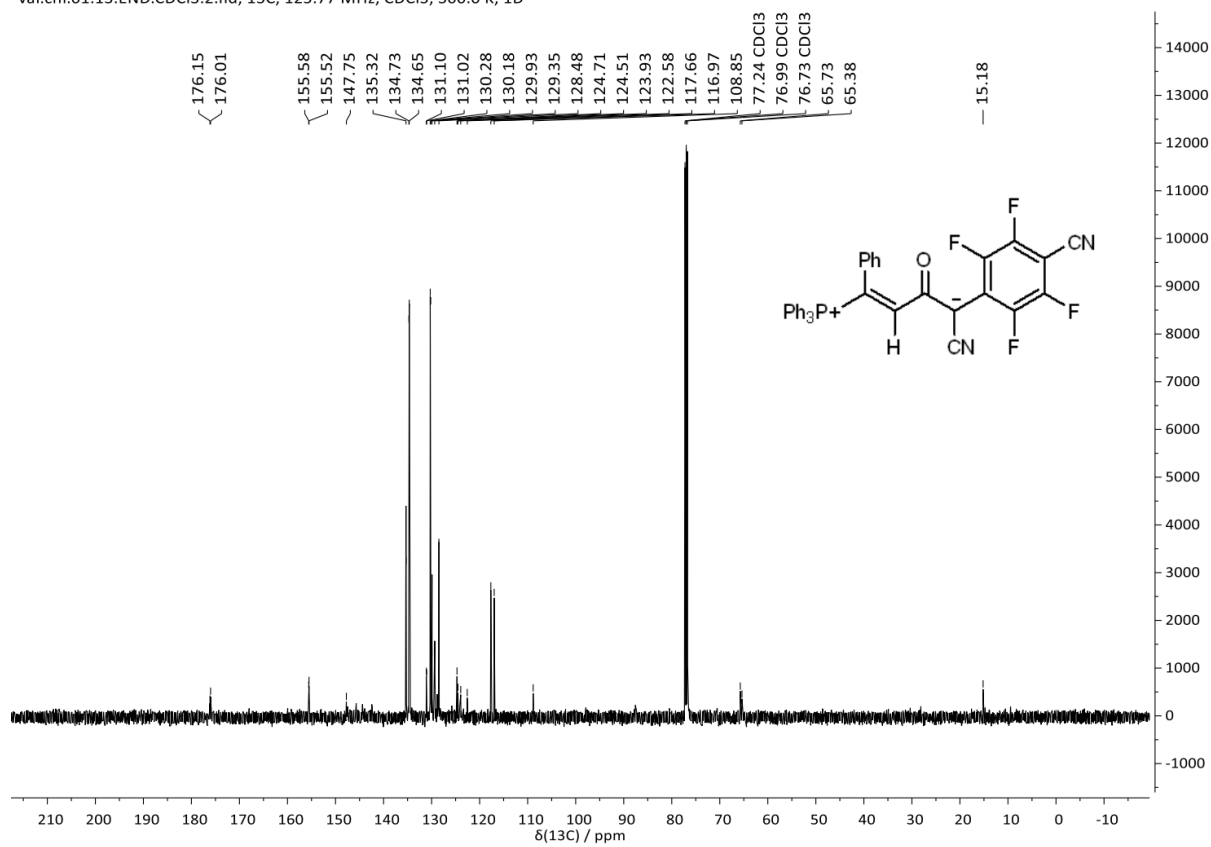
¹H NMR

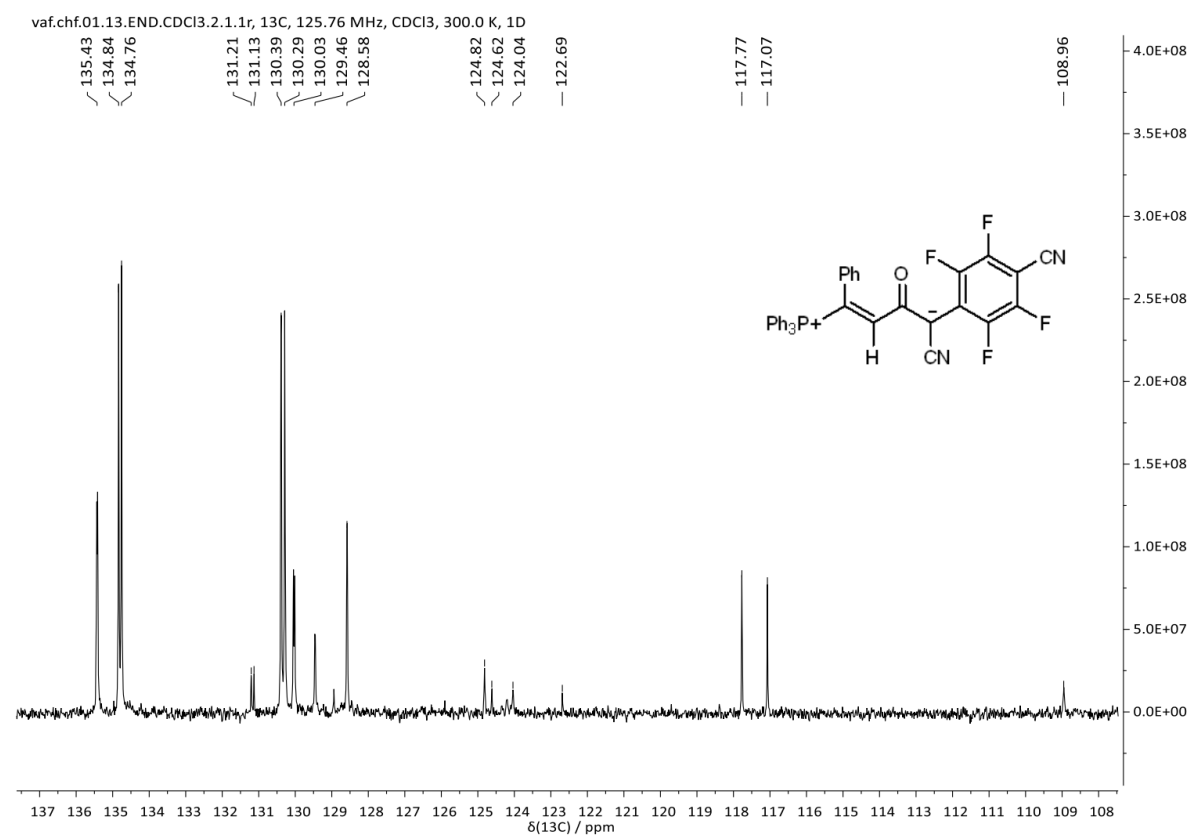
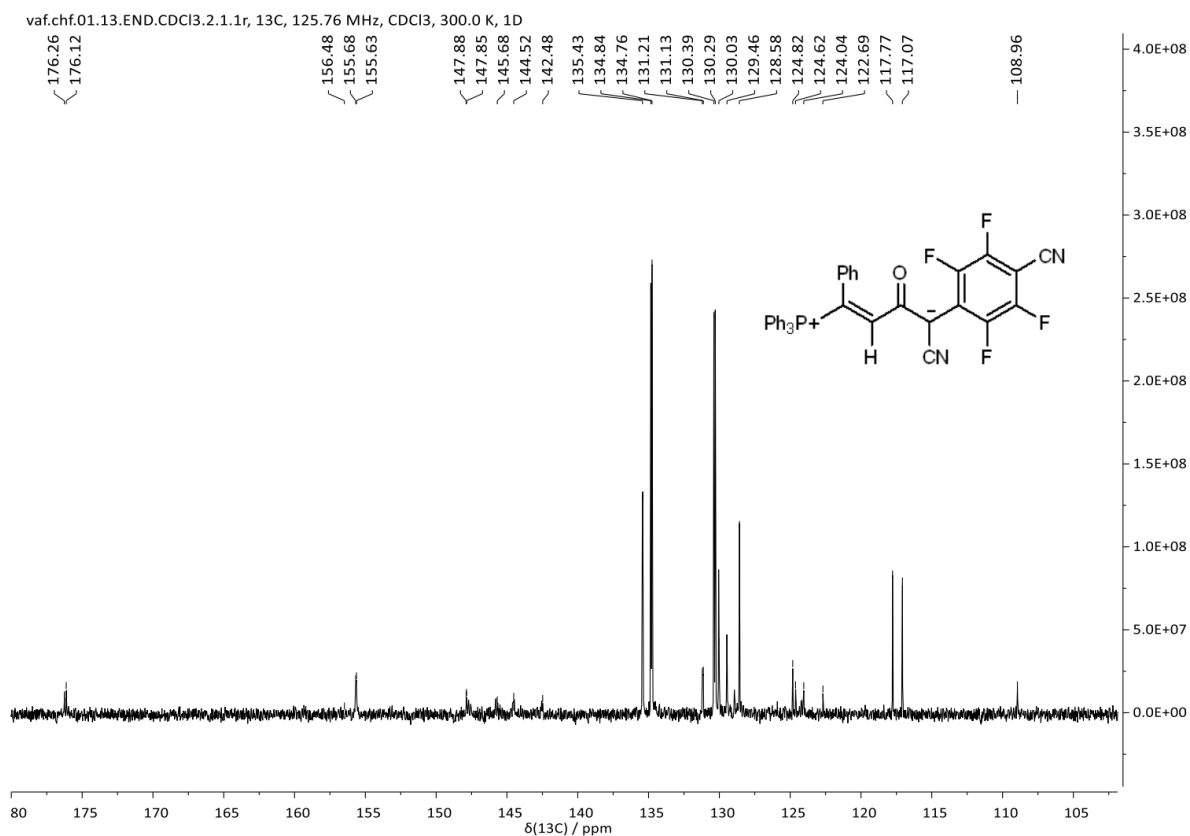
chf.01.13-03/1, 1H, 400.13 MHz, 4 Scans, CDCl₃, 300.0 K, 1D



¹³C NMR

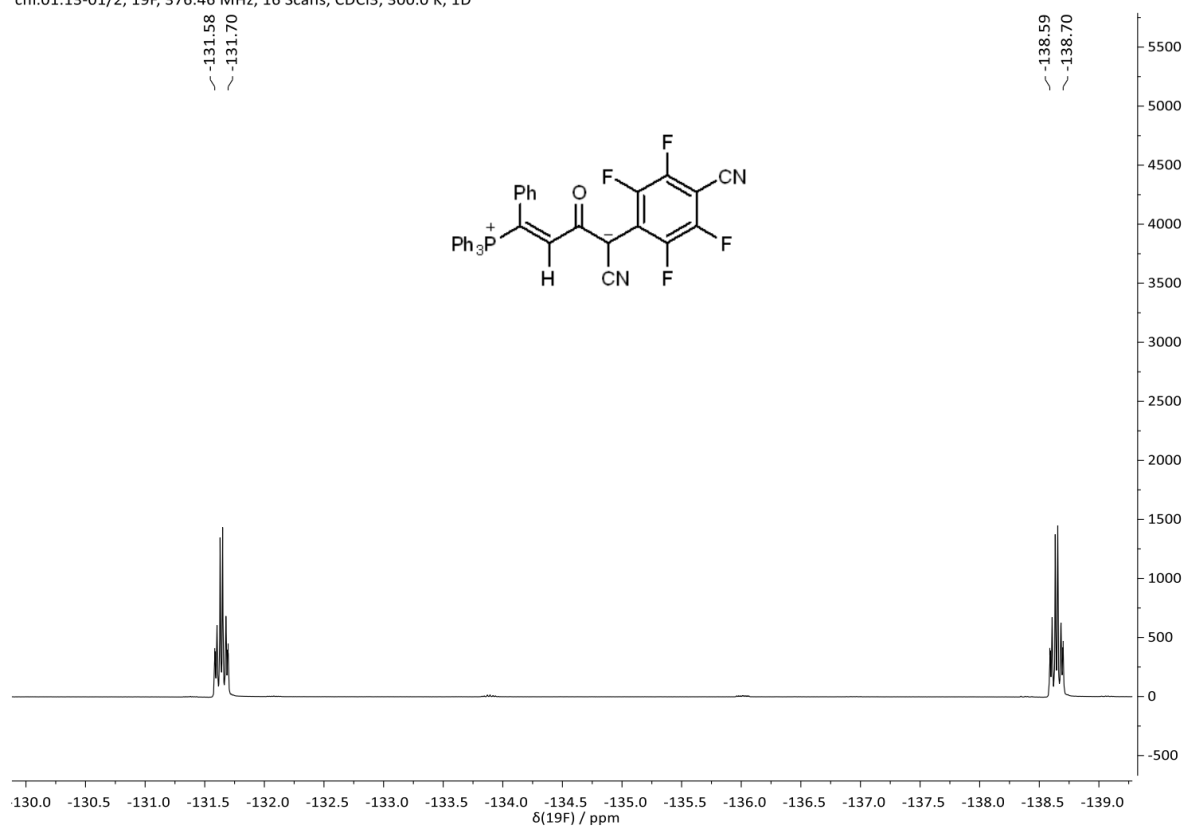
vaf.chf.01.13.END.CDCI3.2.fid, 13C, 125.77 MHz, CDCl₃, 300.0 K, 1D





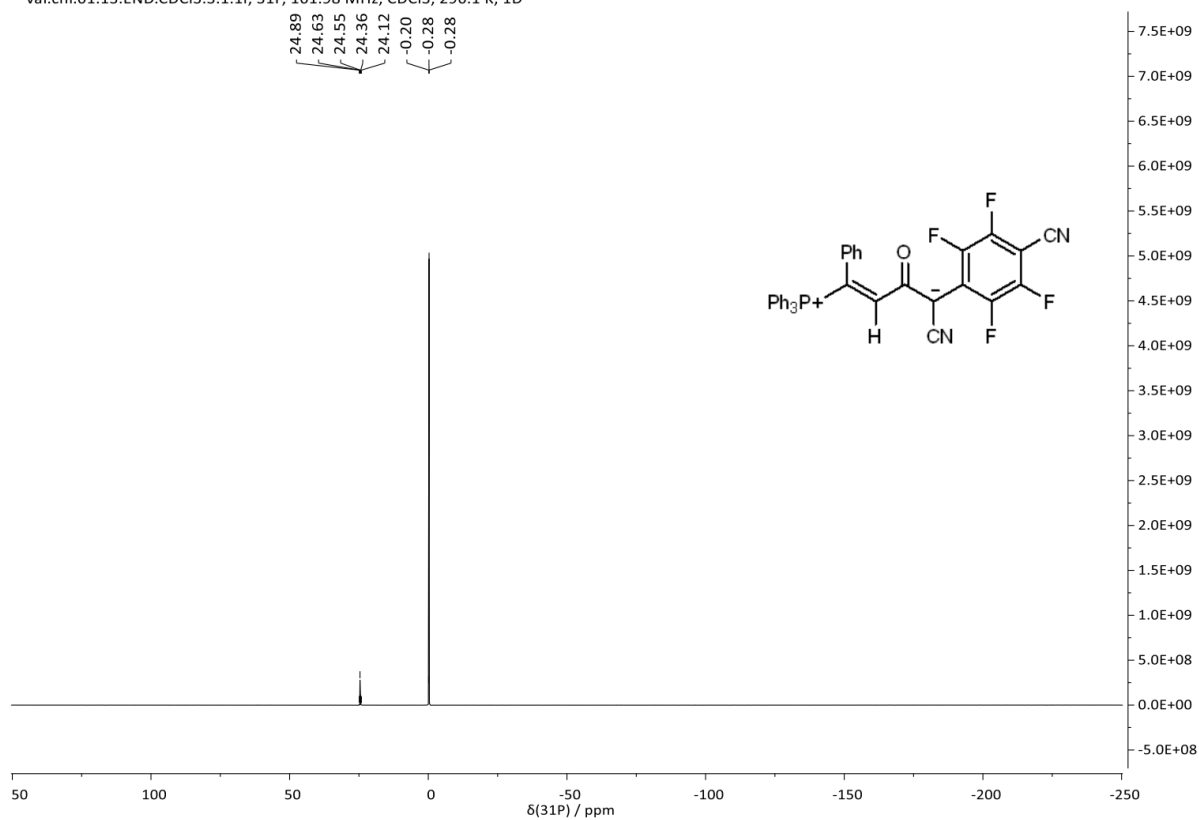
¹⁹F NMR

chf.01.13-01/2, 19F, 376.46 MHz, 16 Scans, CDCl₃, 300.0 K, 1D

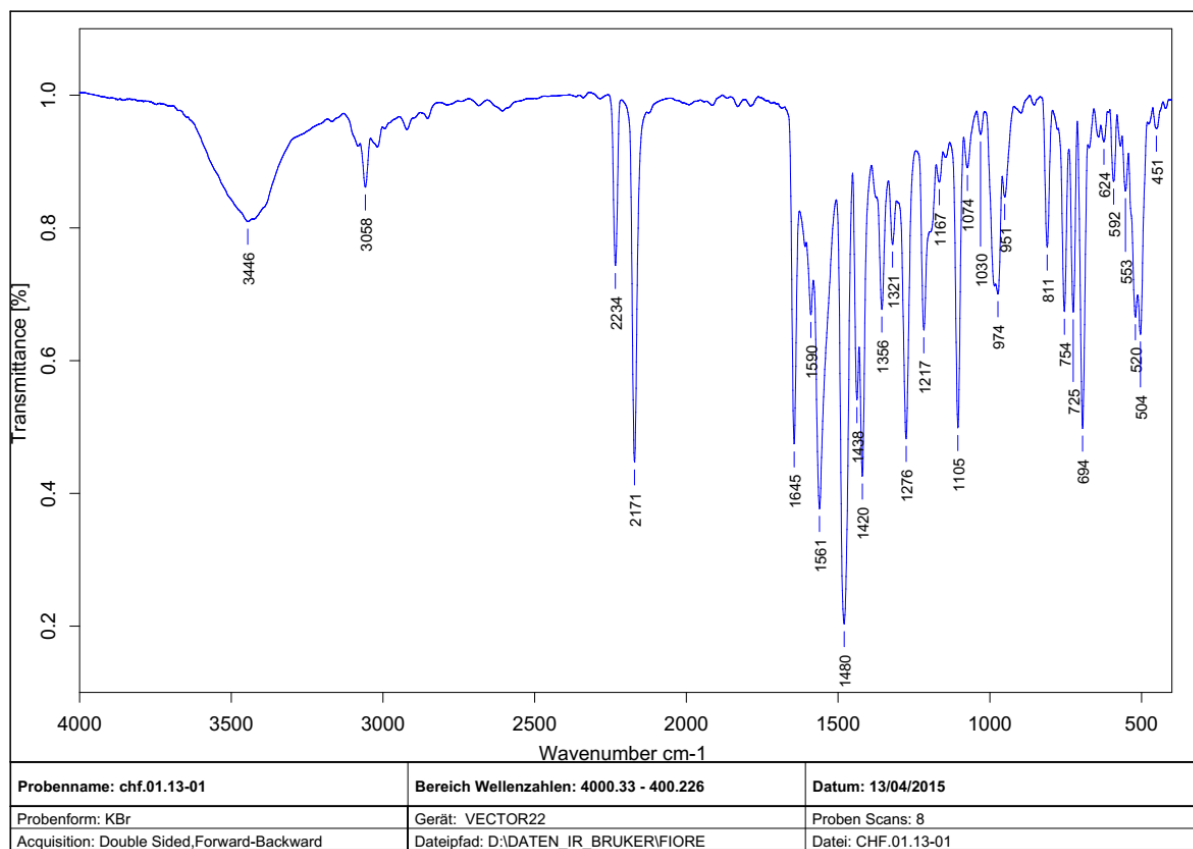


³¹P NMR

vaf.chf.01.13.END.CDCl₃.3.1.1r, 31P, 161.98 MHz, CDCl₃, 296.1 K, 1D



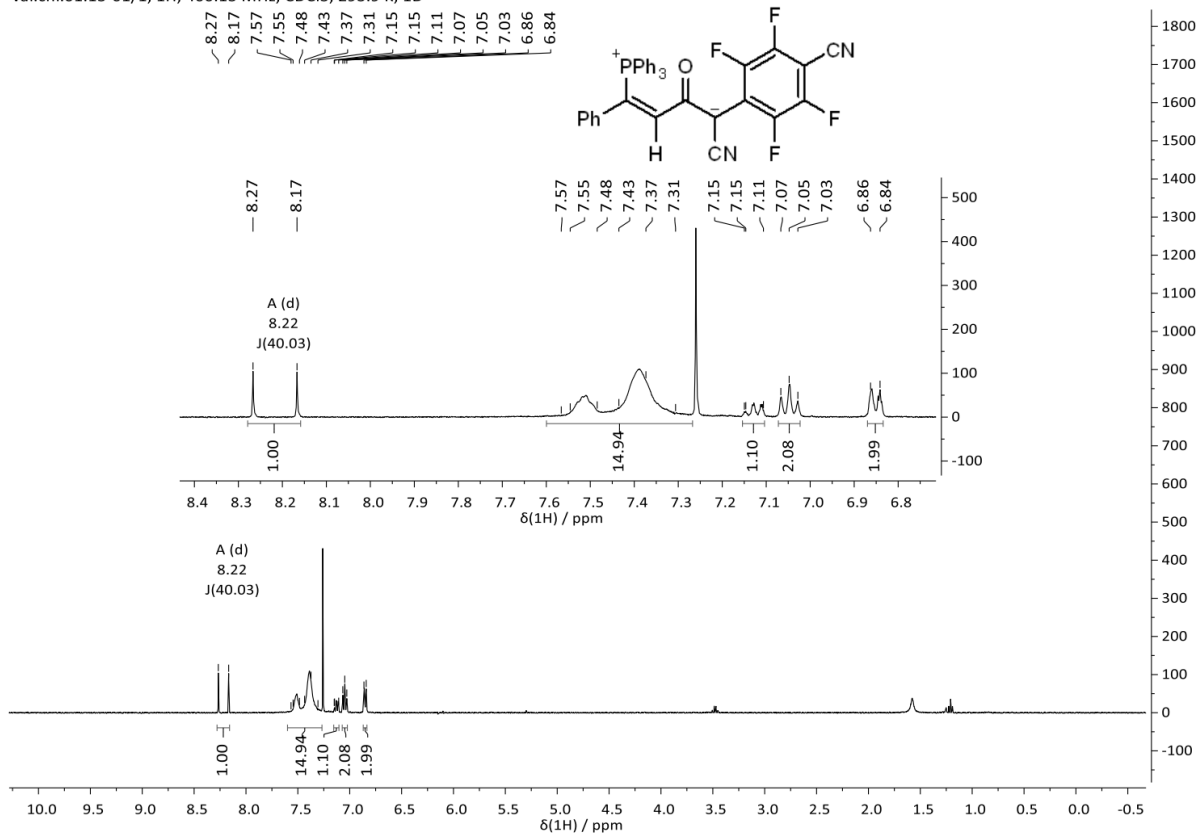
IR



(Z)-3f

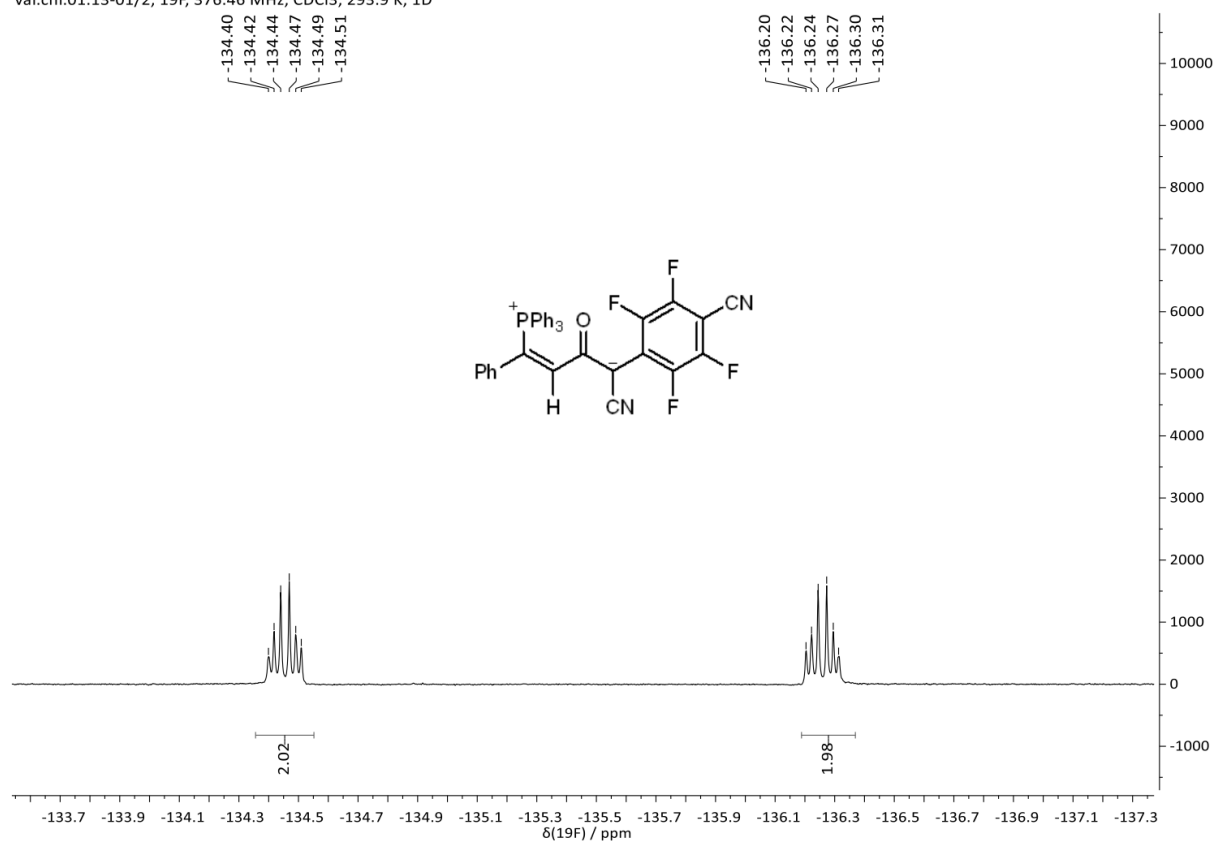
¹H NMR

vaf.chf.01.13-01/1, 1H, 400.13 MHz, CDCl₃, 293.9 K, 1D



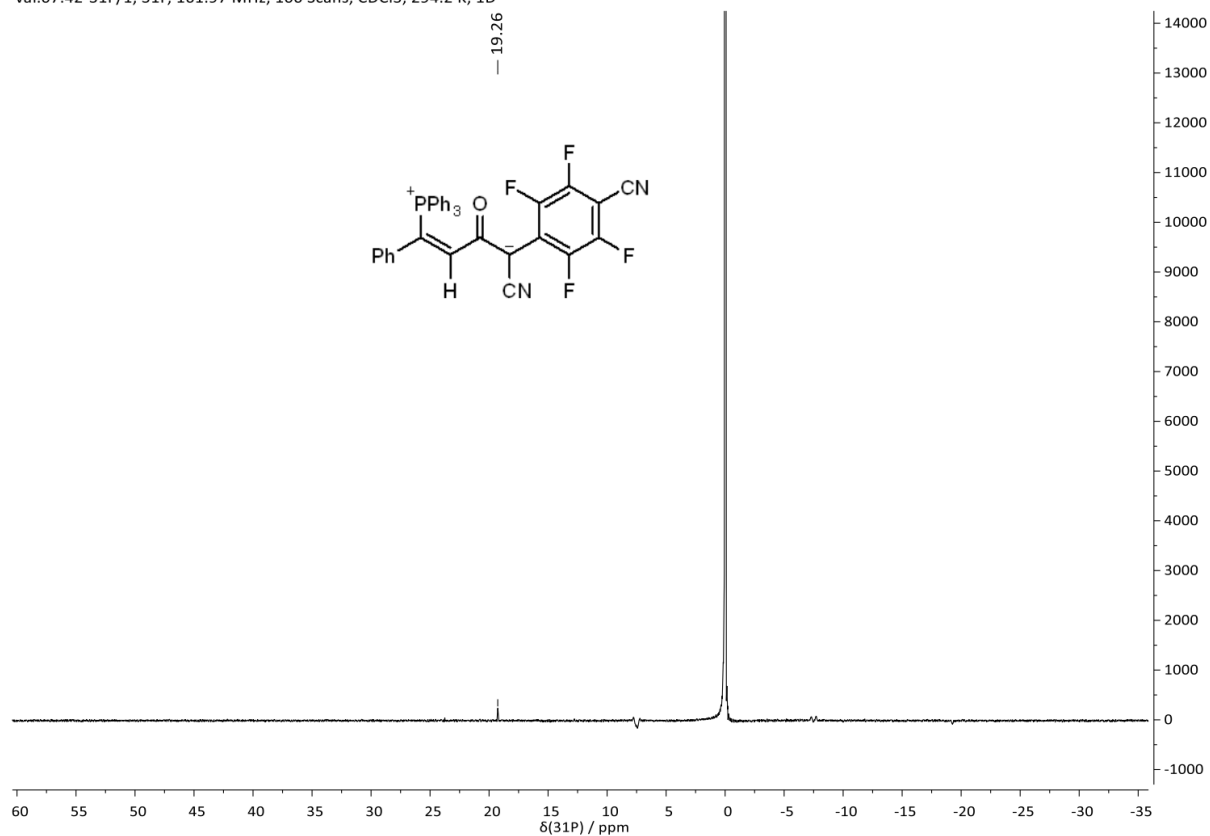
¹⁹F NMR

vaf.chf.01.13-01/2, 19F, 376.46 MHz, CDCl₃, 293.9 K, 1D



³¹P NMR

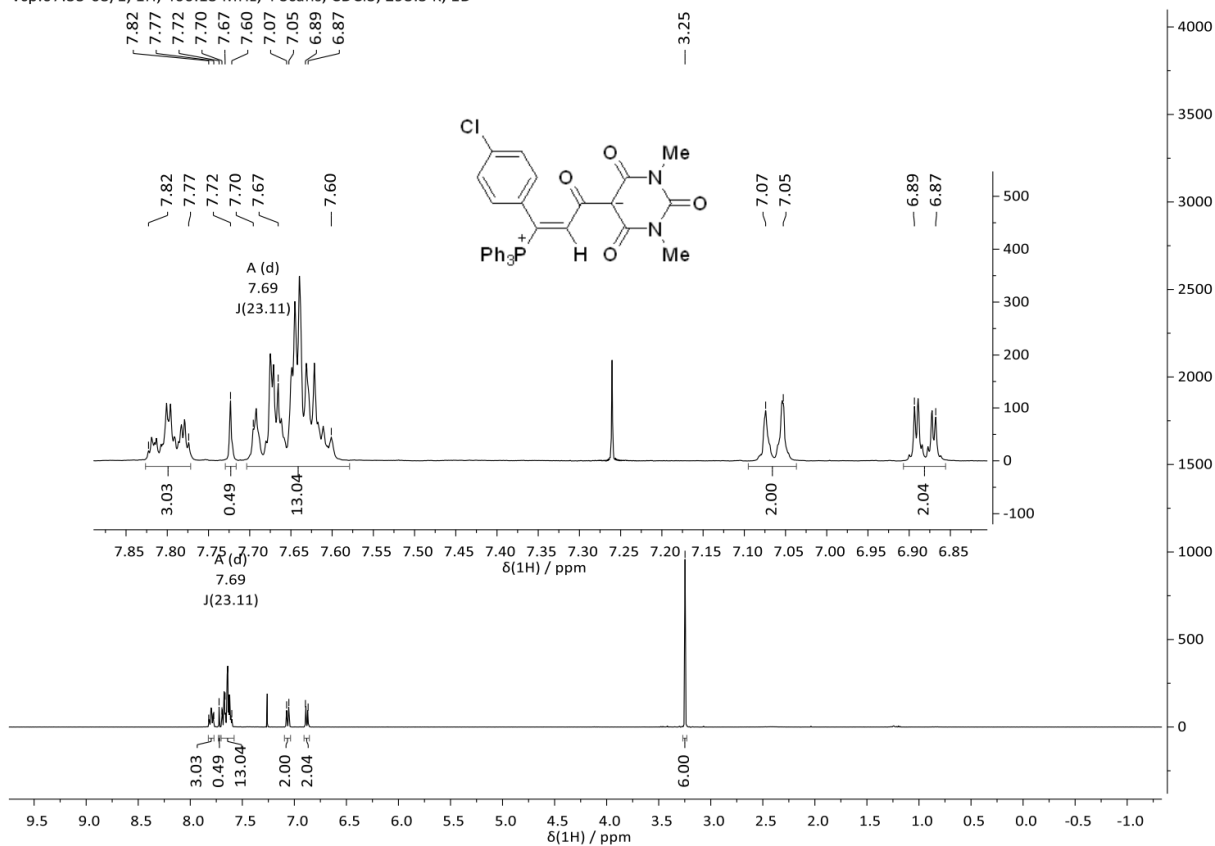
vaf.07.42-31P/1, 31P, 161.97 MHz, 100 Scans, CDCl₃, 294.2 K, 1D



4.7. Betaines ((E)-3g)

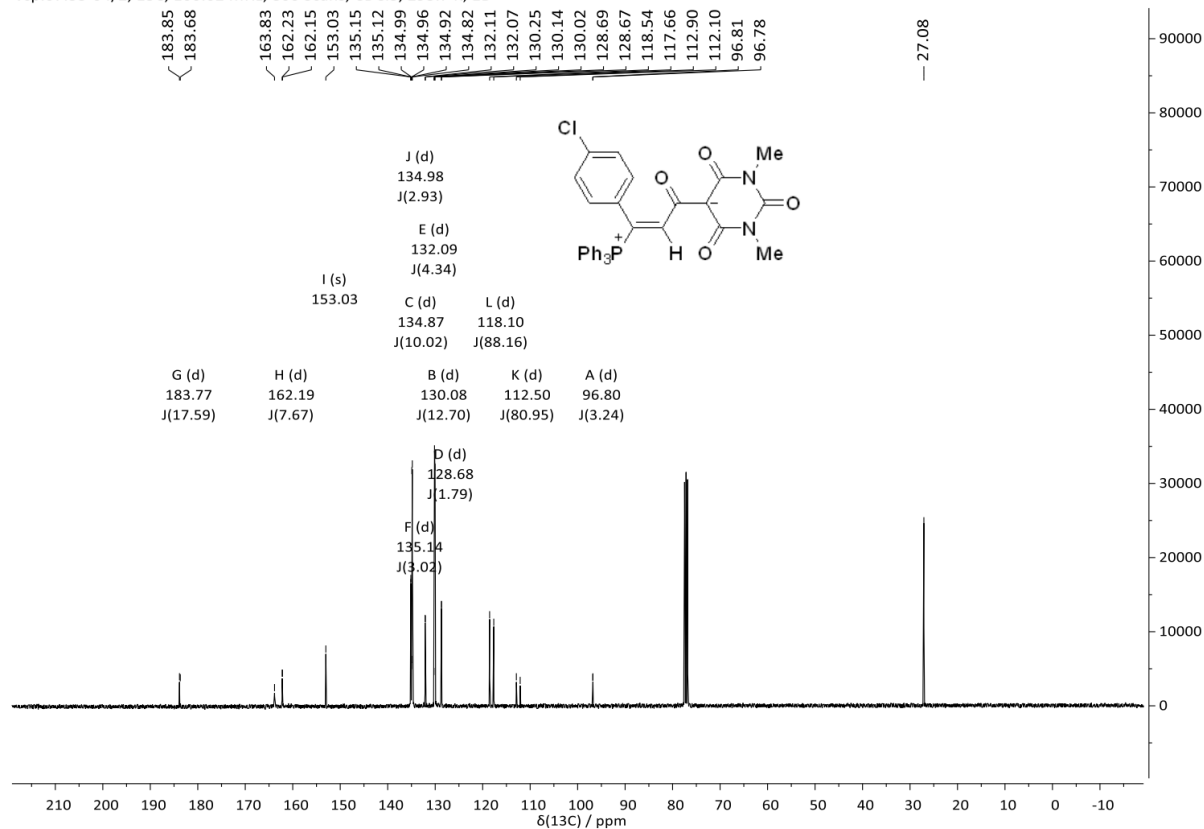
¹H NMR

vcp.07.33-03/1, 1H, 400.13 MHz, 4 Scans, CDCl₃, 293.5 K, 1D



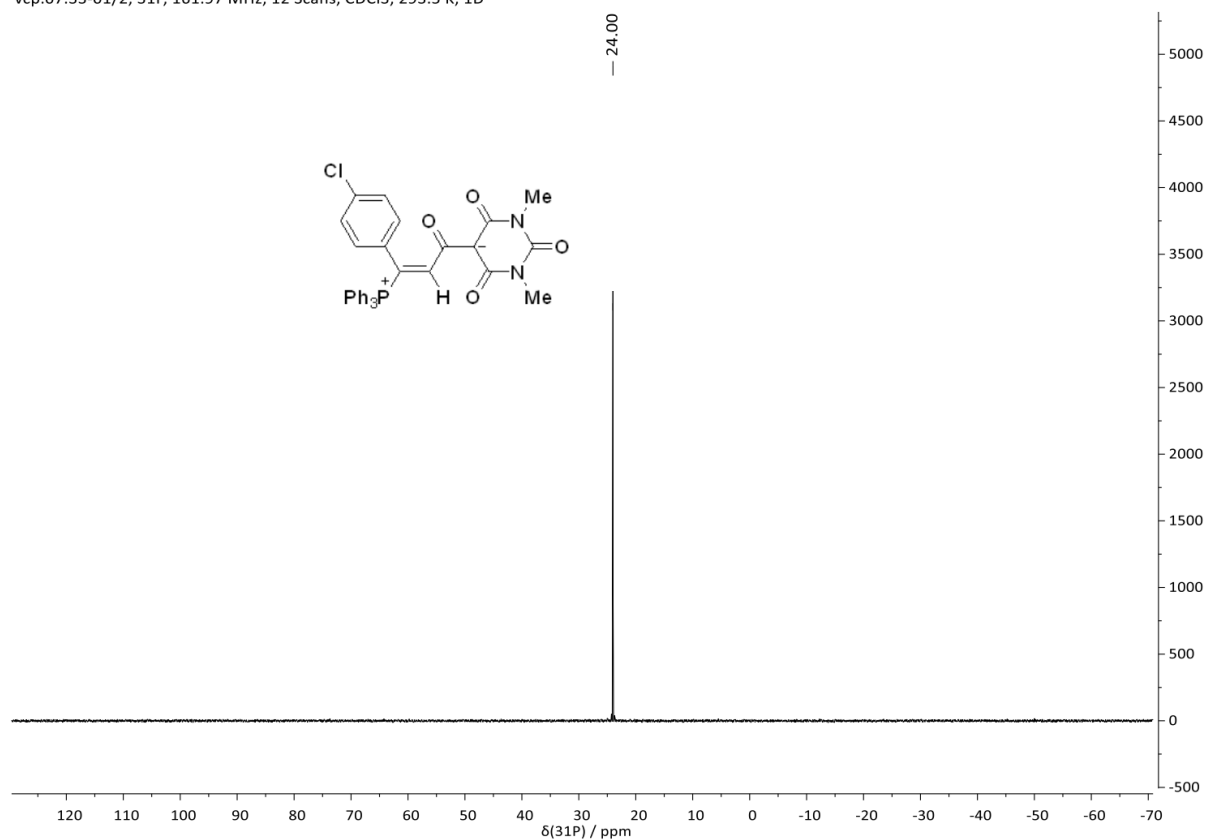
¹³C NMR

vcp.07.33-04/2, 13C, 100.62 MHz, 800 Scans, CDCl₃, 293.7 K, 1D

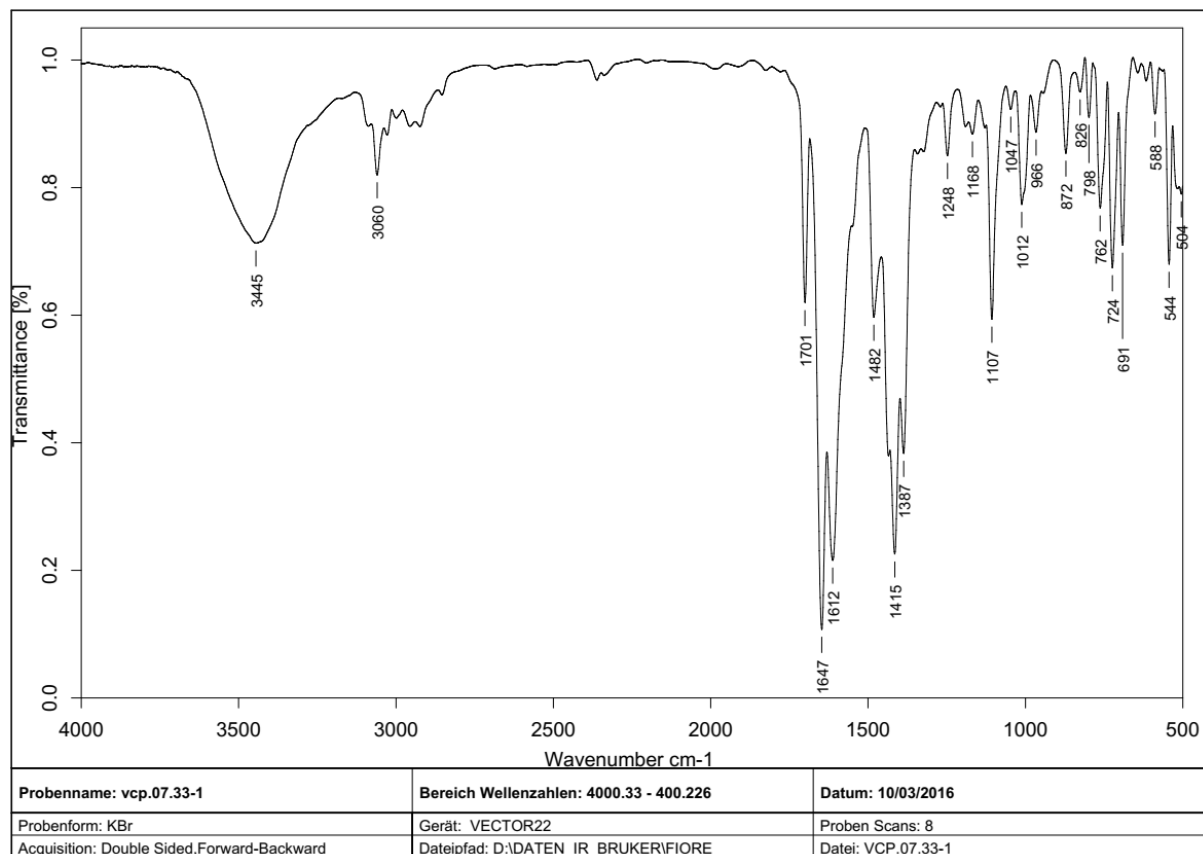


³¹P NMR

vcp.07.33-01/2, 31P, 161.97 MHz, 12 Scans, CDCl₃, 293.5 K, 1D



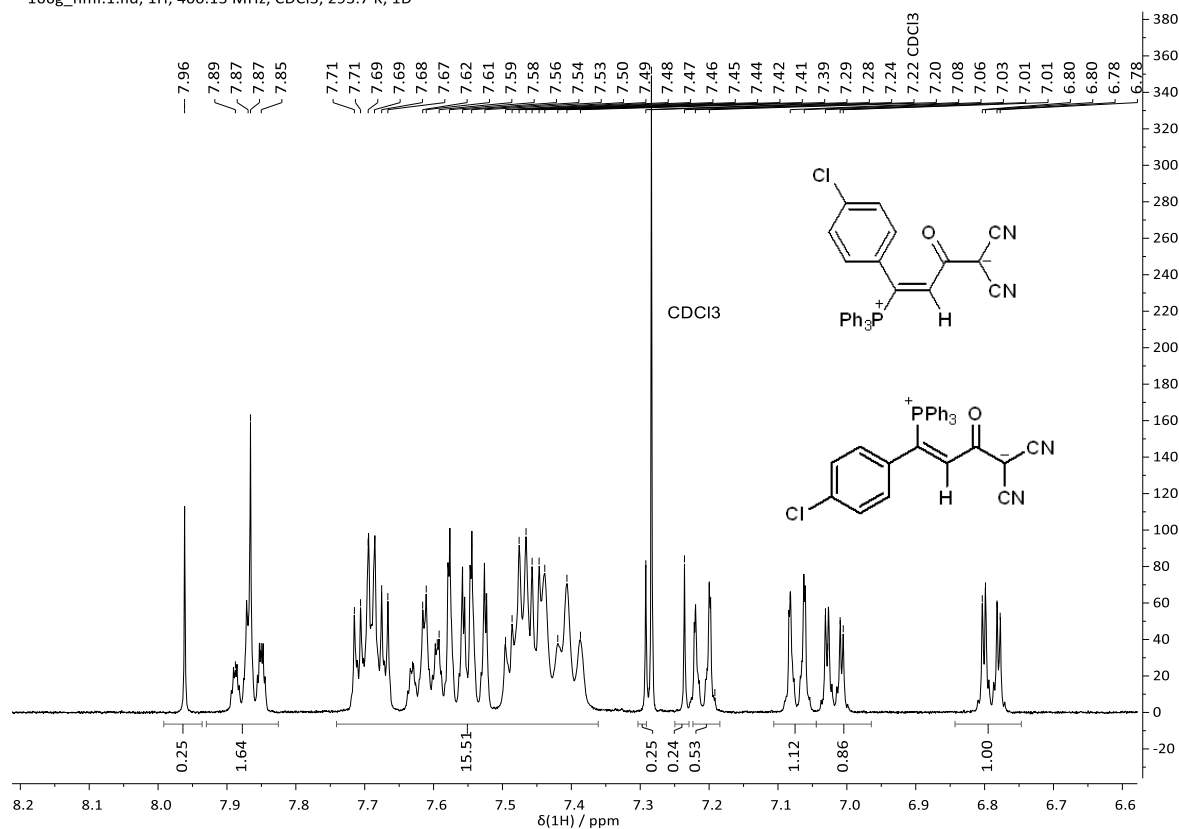
IR



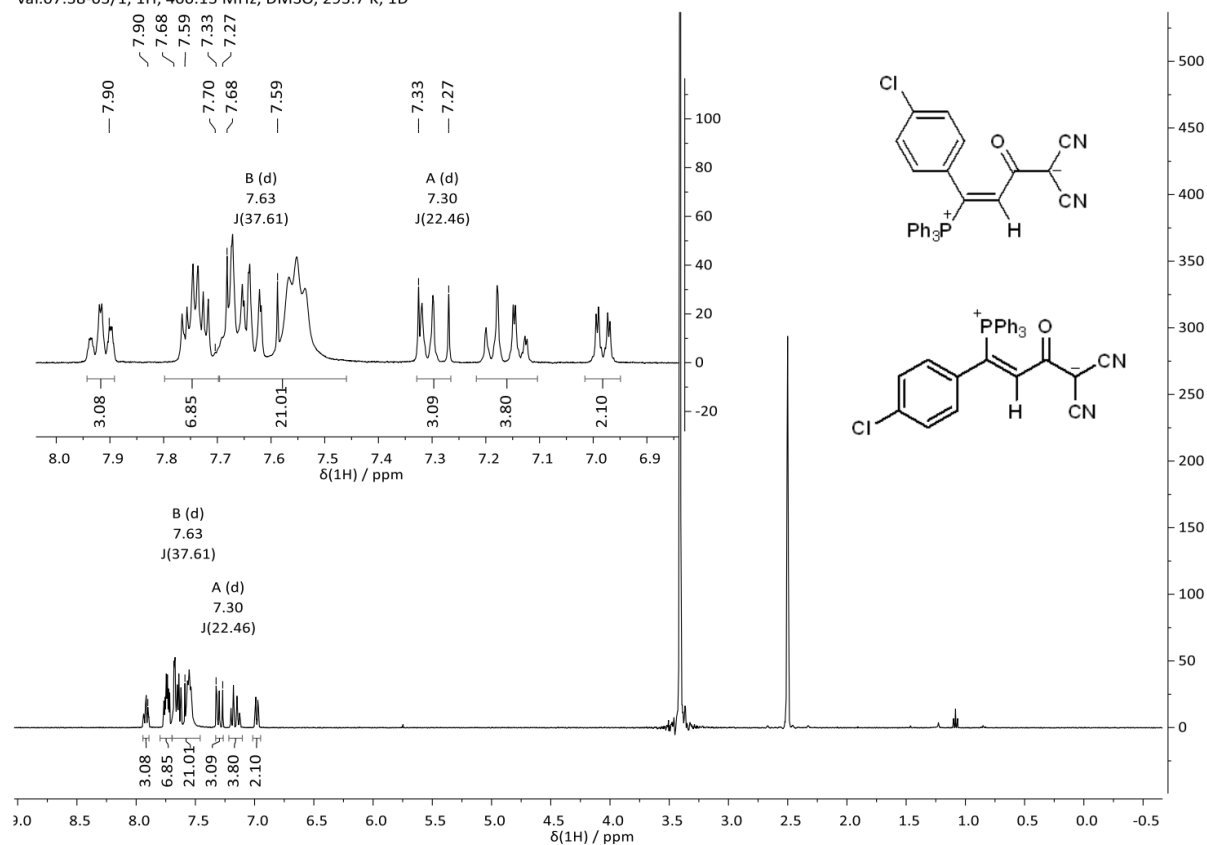
4.8 Betaines (E)- and (Z)-3h

^1H NMR

166g_nmr.1.fid, 1H, 400.13 MHz, CDCl_3 , 293.7 K, 1D

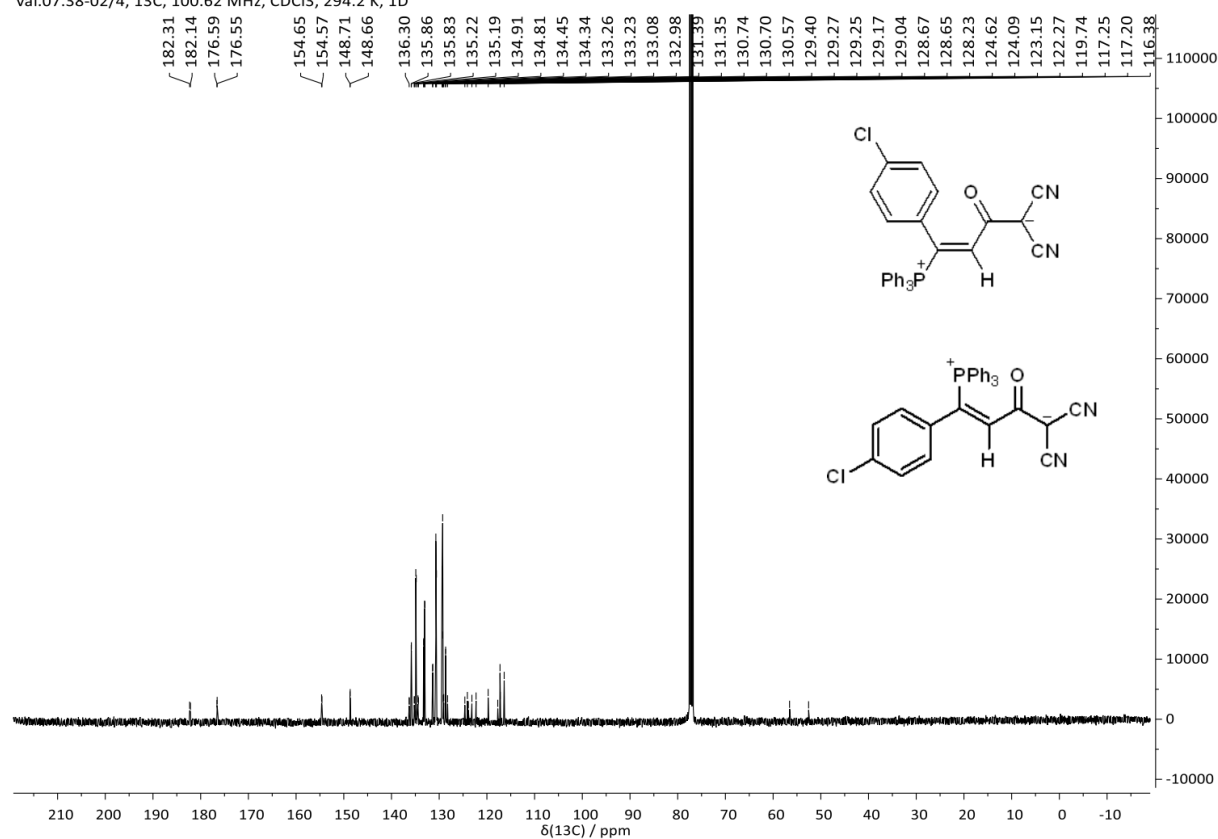


vaf.07.38-03/1, 1H, 400.13 MHz, DMSO , 293.7 K, 1D

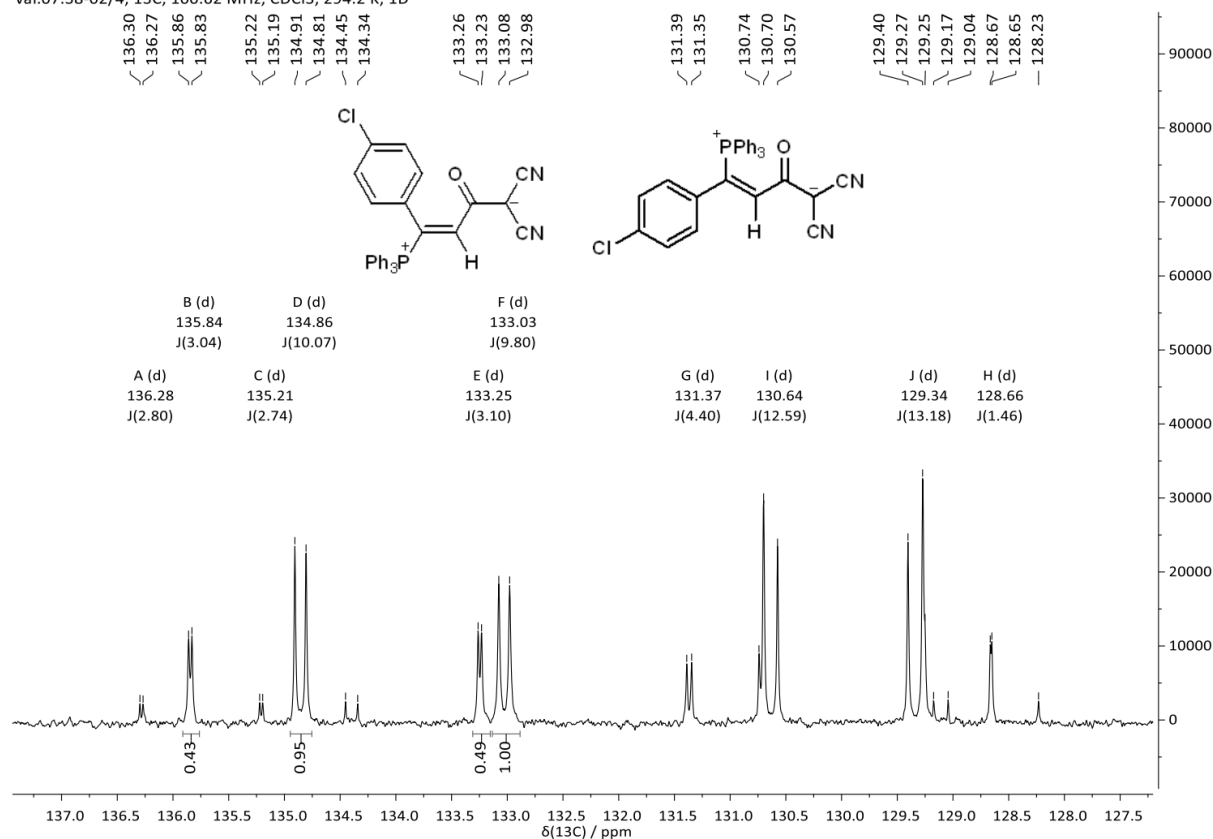


¹³C NMR

vaf.07.38-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D

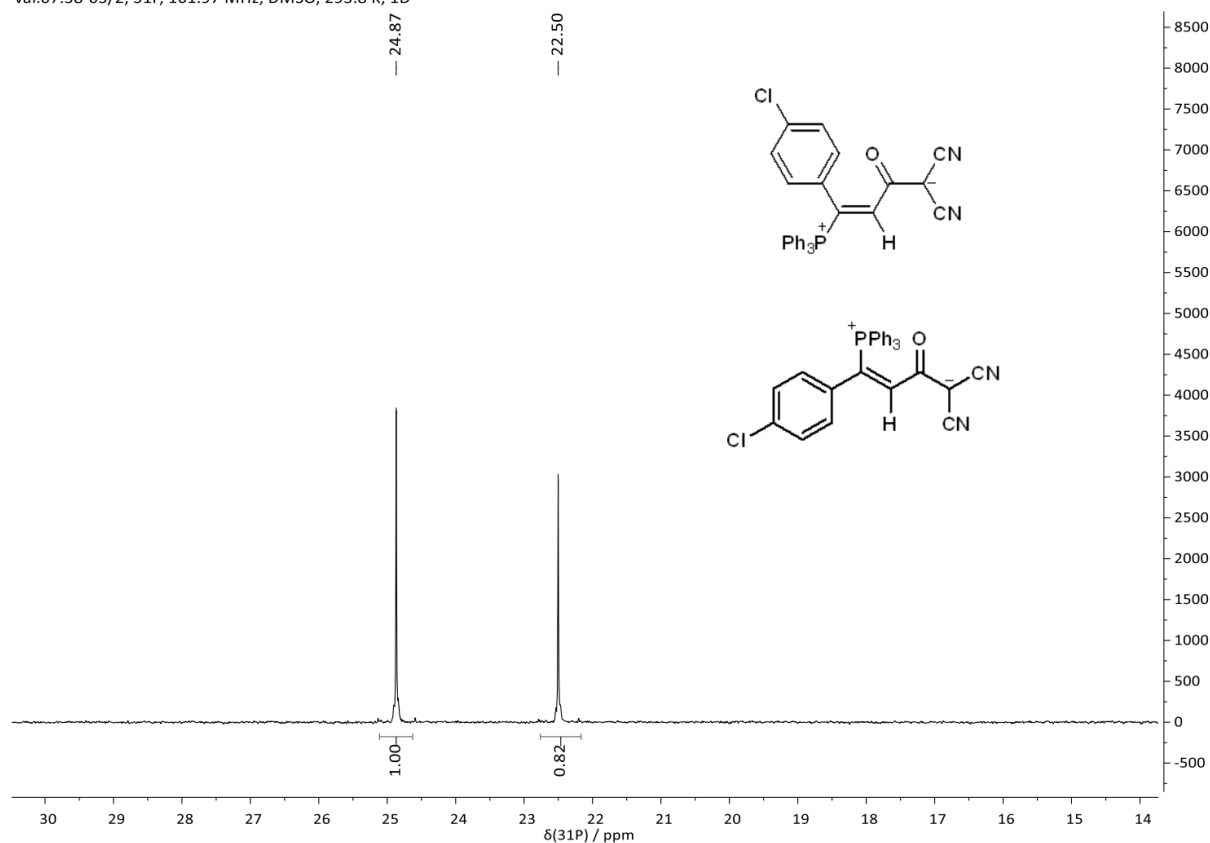


vaf.07.38-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D

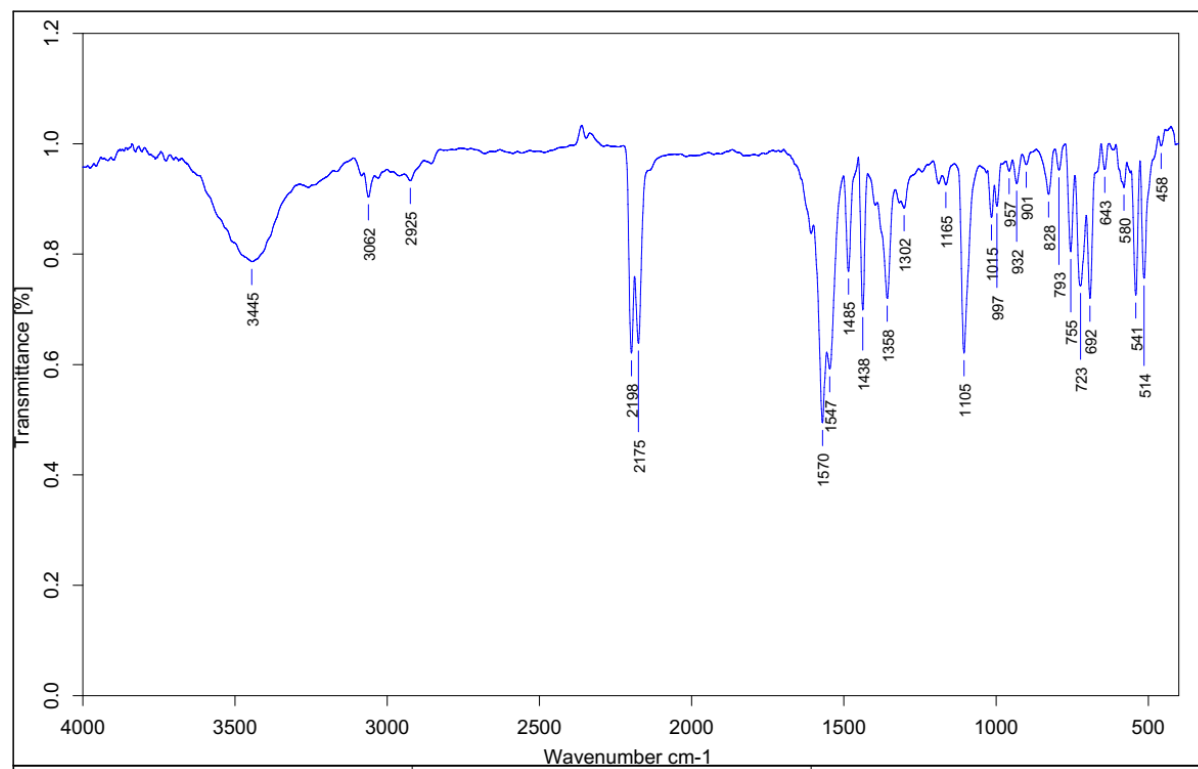


³¹P NMR

vaf.07.38-03/2, 31P, 161.97 MHz, DMSO, 293.8 K, 1D



IR

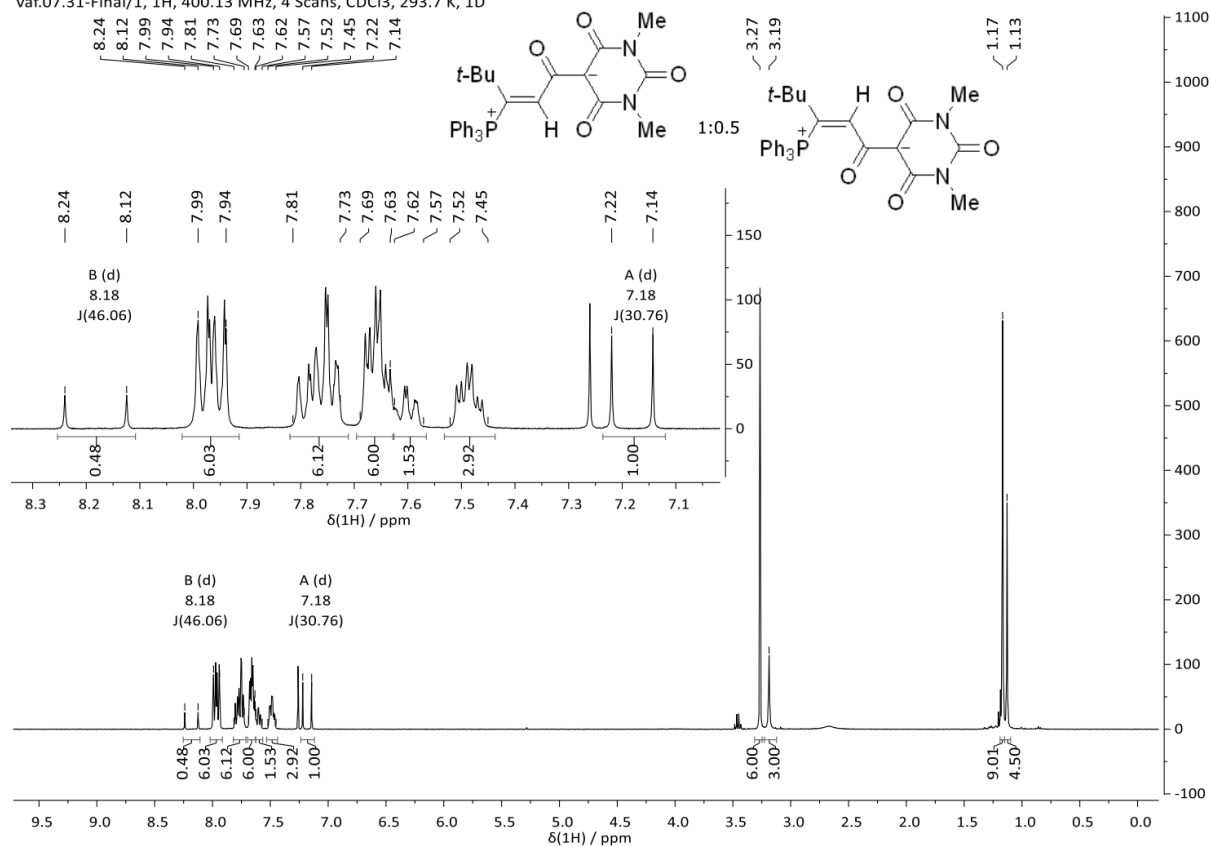


Probenname: vaf.07.38-1	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 17/03/2017
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided, Forward-Backward	Dateipfad: D:\DATEN_IR_BRUKER\FIORE	Datei: VAF.07.38-1

4.9. Betaines (*E*)- and (*Z*)-3i (mixture)

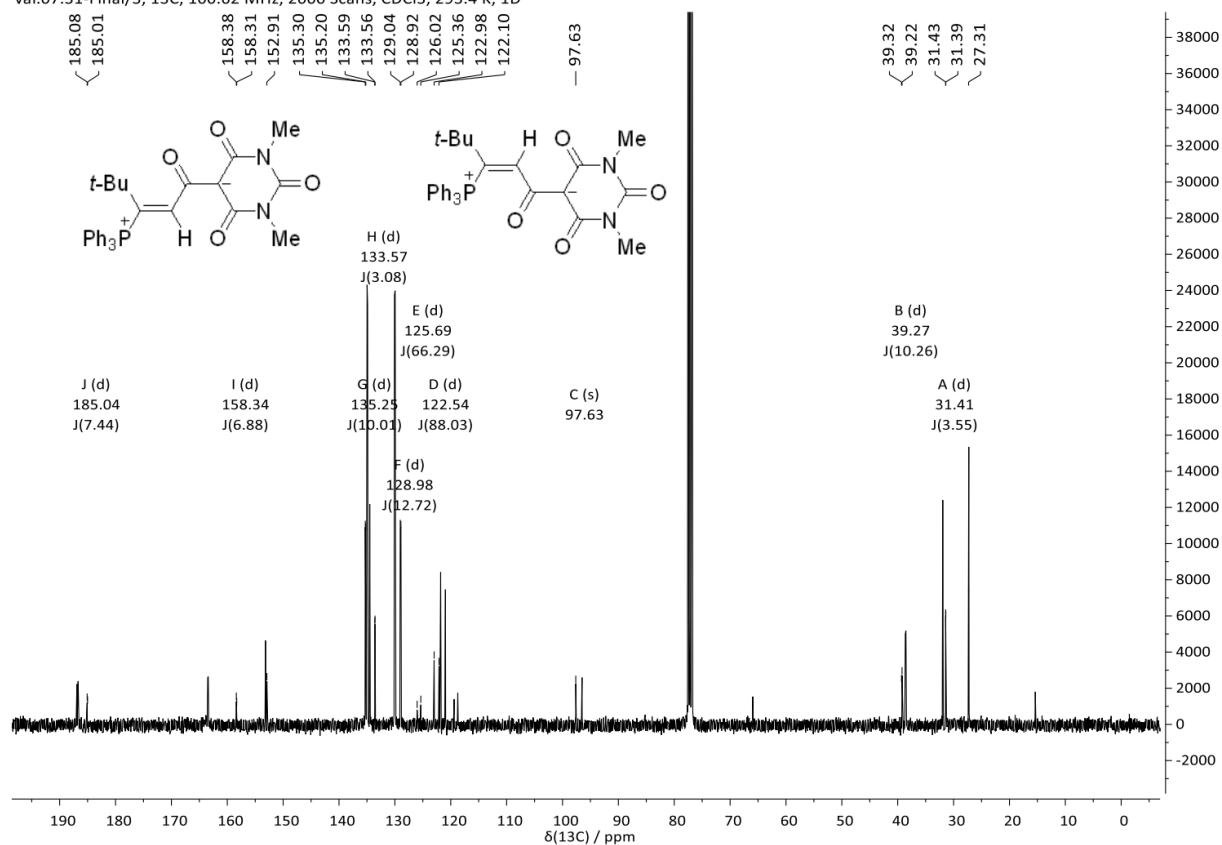
¹H NMR

vaf.07.31-Final/1, 1H, 400.13 MHz, 4 Scans, CDCl₃, 293.7 K, 1D

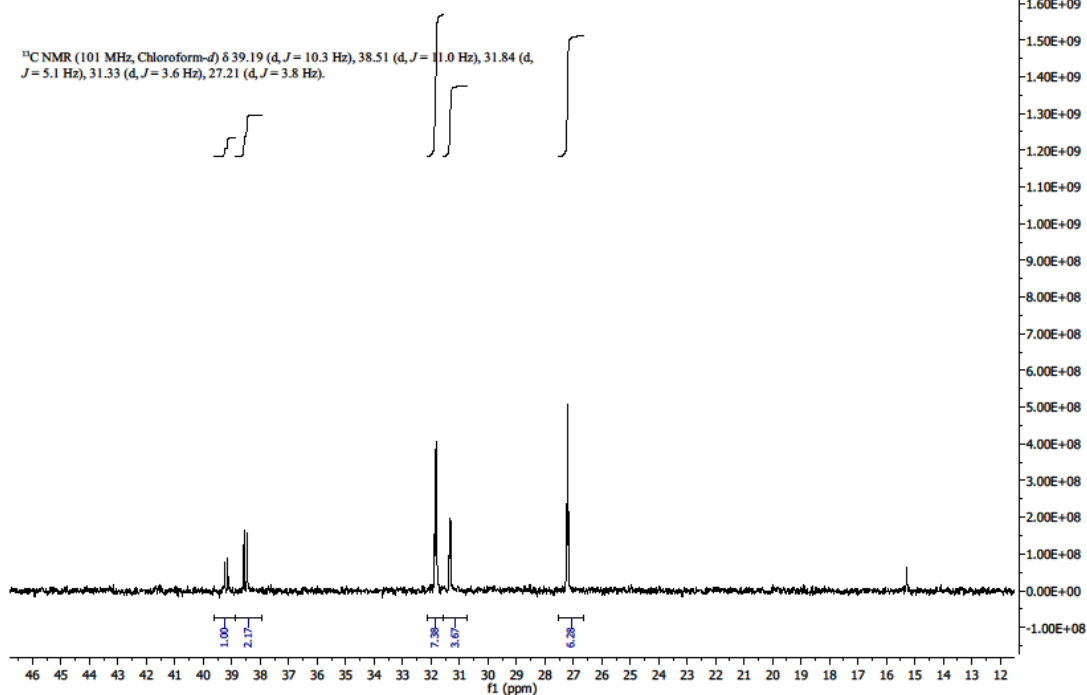


¹³C NMR

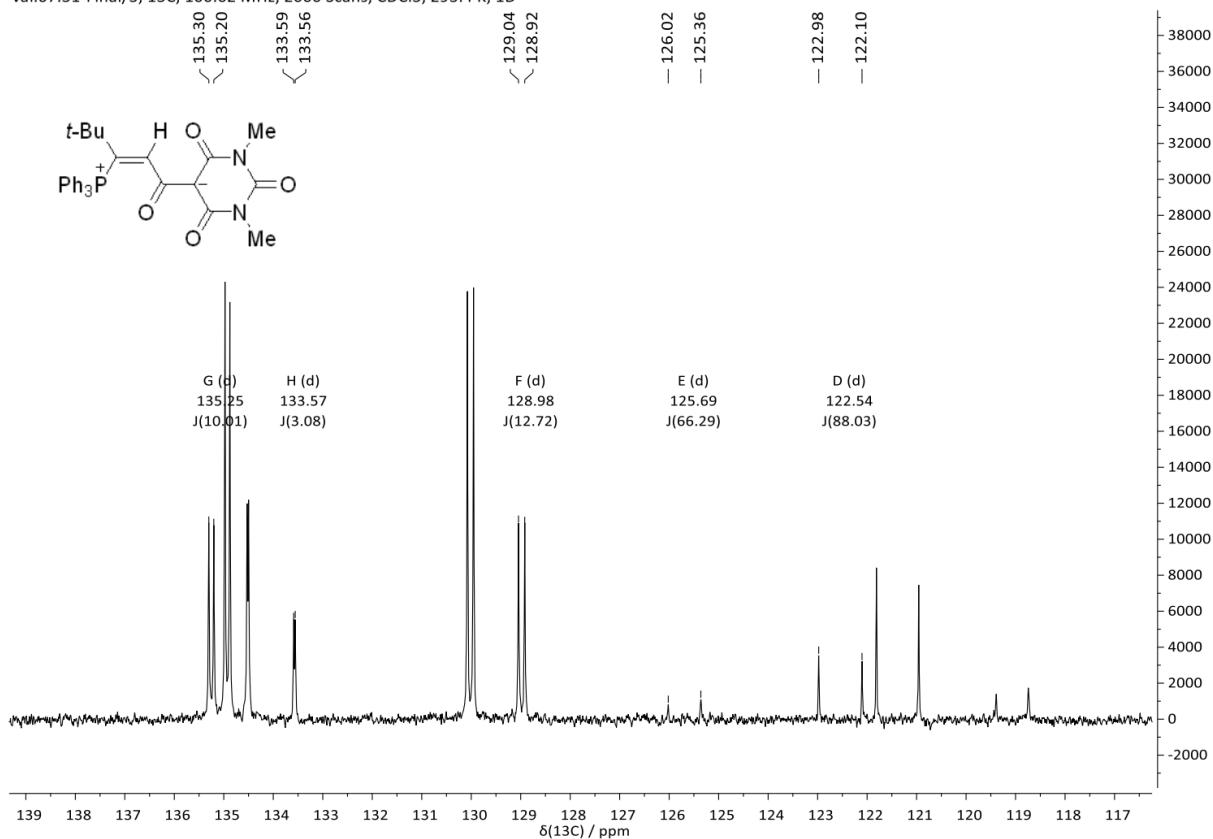
vaf.07.31-Final/3, 13C, 100.62 MHz, 2000 Scans, CDCl₃, 293.4 K, 1D



166j_nmr.3.1.1r
C13CPD CDCl3 /opt/topspin akmaas 57

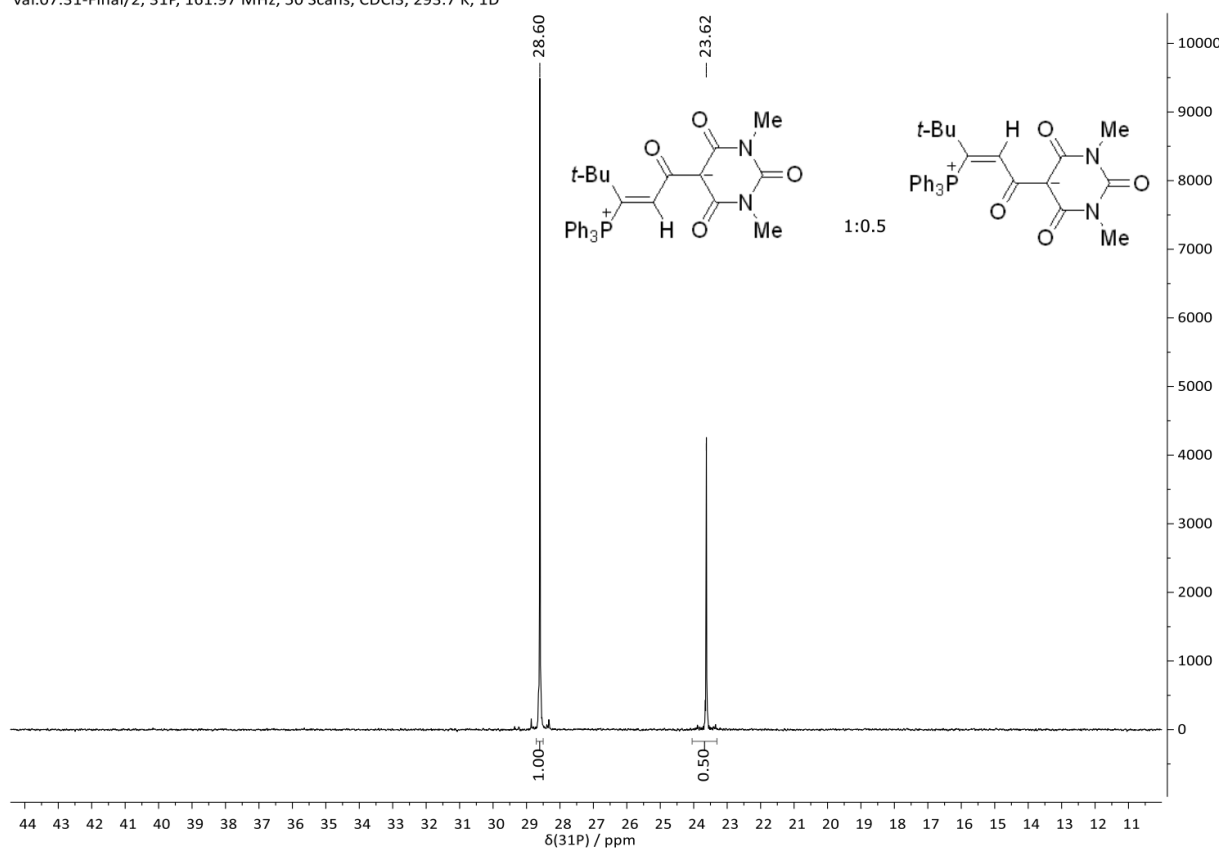


vaf.07.31-Final/3, ^{13}C , 100.62 MHz, 2000 Scans, CDCl_3 , 293.4 K, 1D

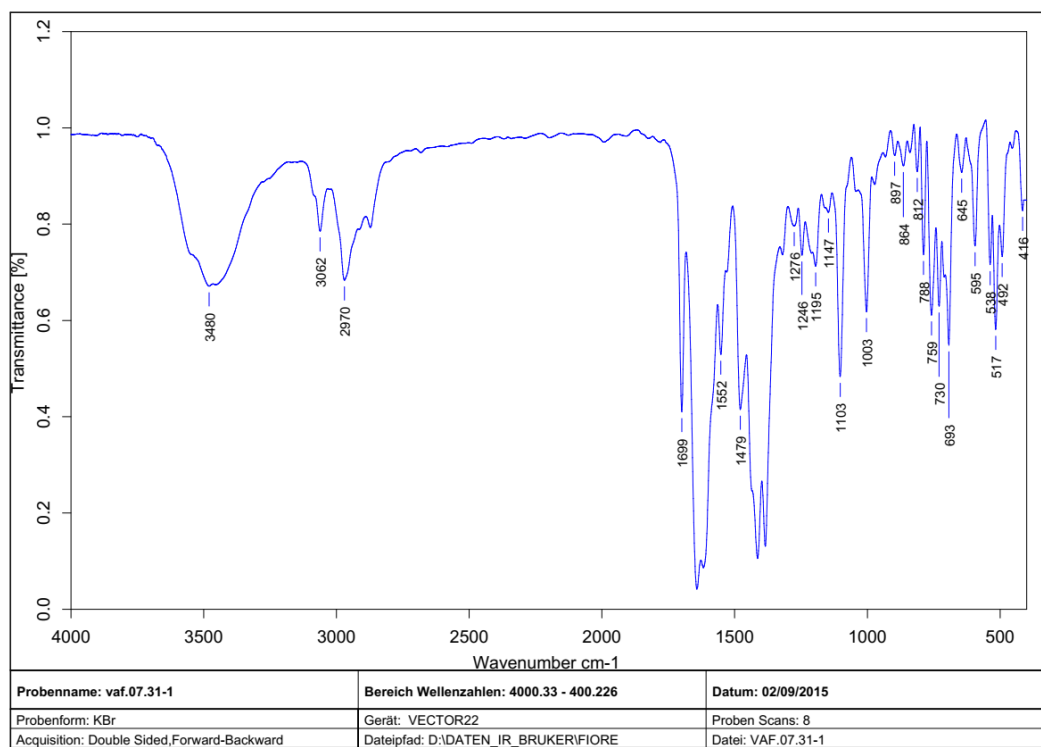


³¹P NMR

vaf.07.31-Final/2, 31P, 161.97 MHz, 50 Scans, CDCl₃, 293.7 K, 1D



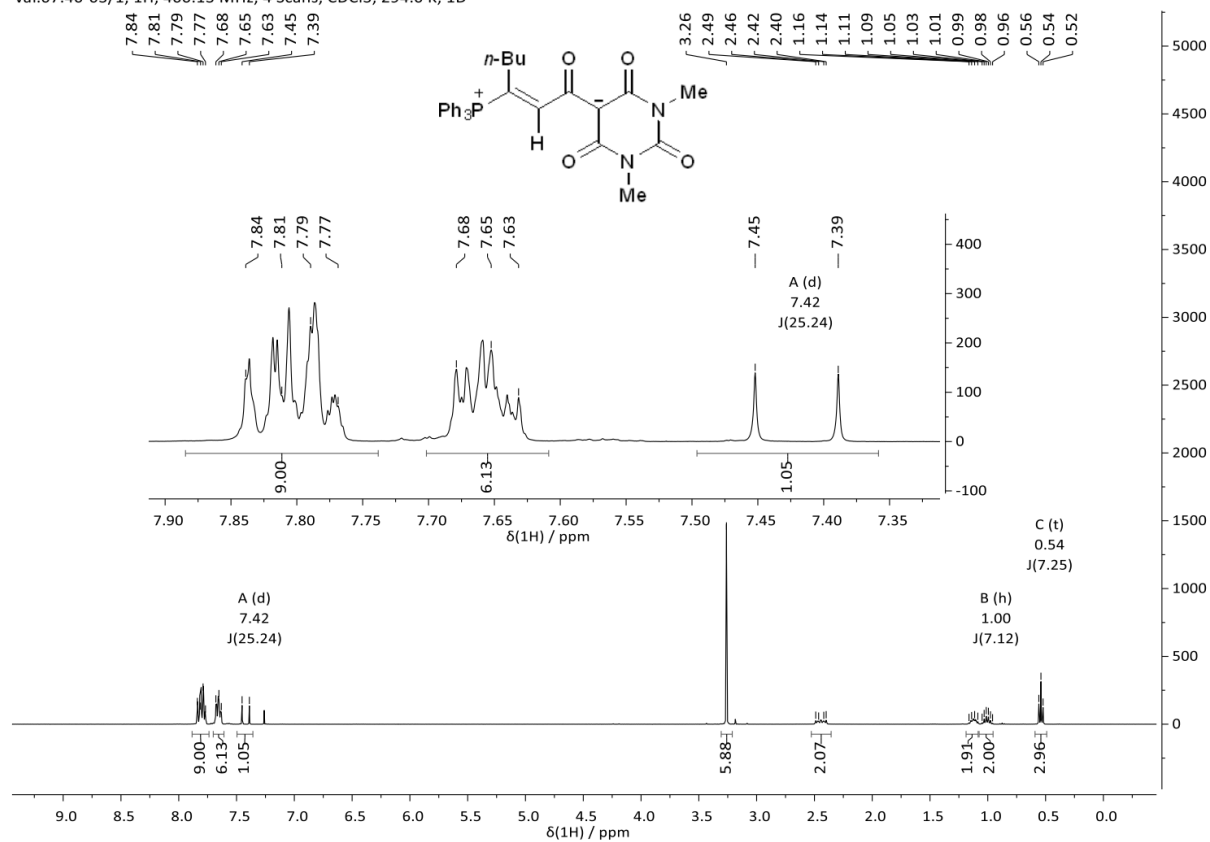
IR



4.10. Betaine (*E*)-3j

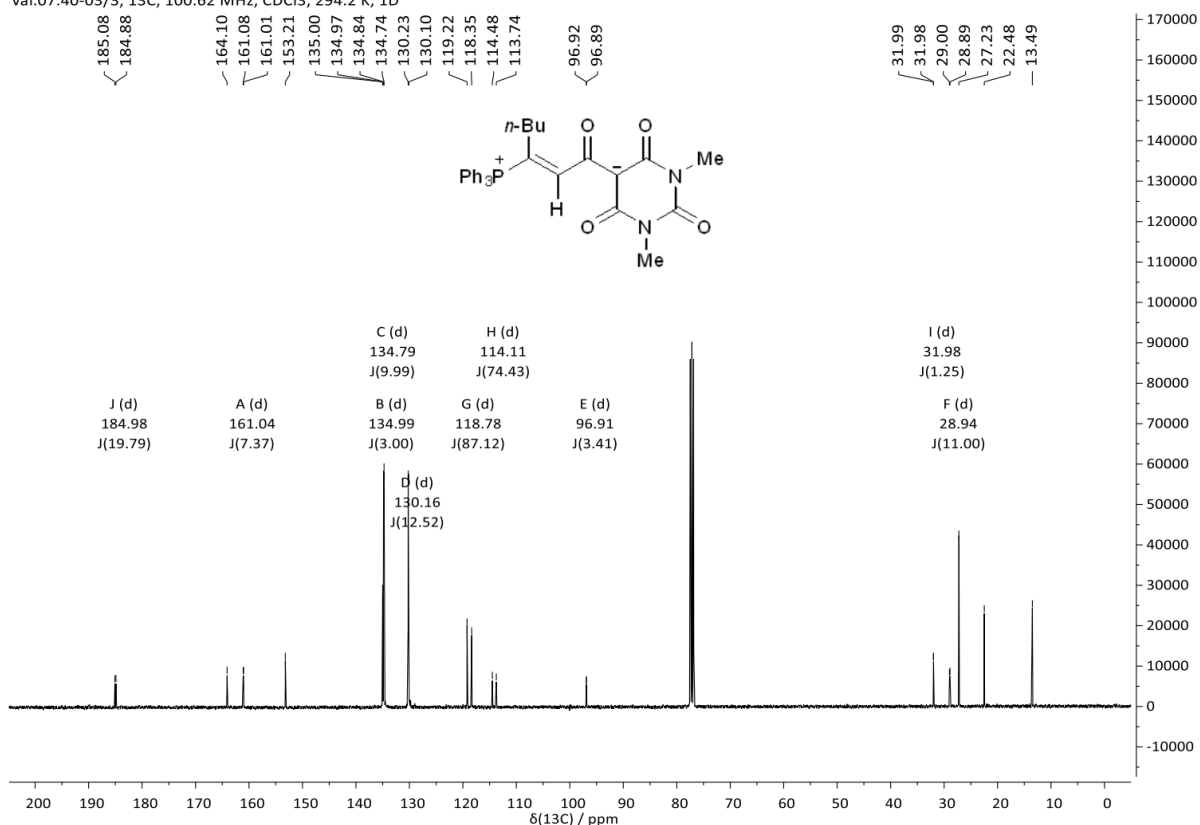
¹H NMR

vaf.07.40-03/1, 1H, 400.13 MHz, 4 Scans, CDCl₃, 294.0 K, 1D

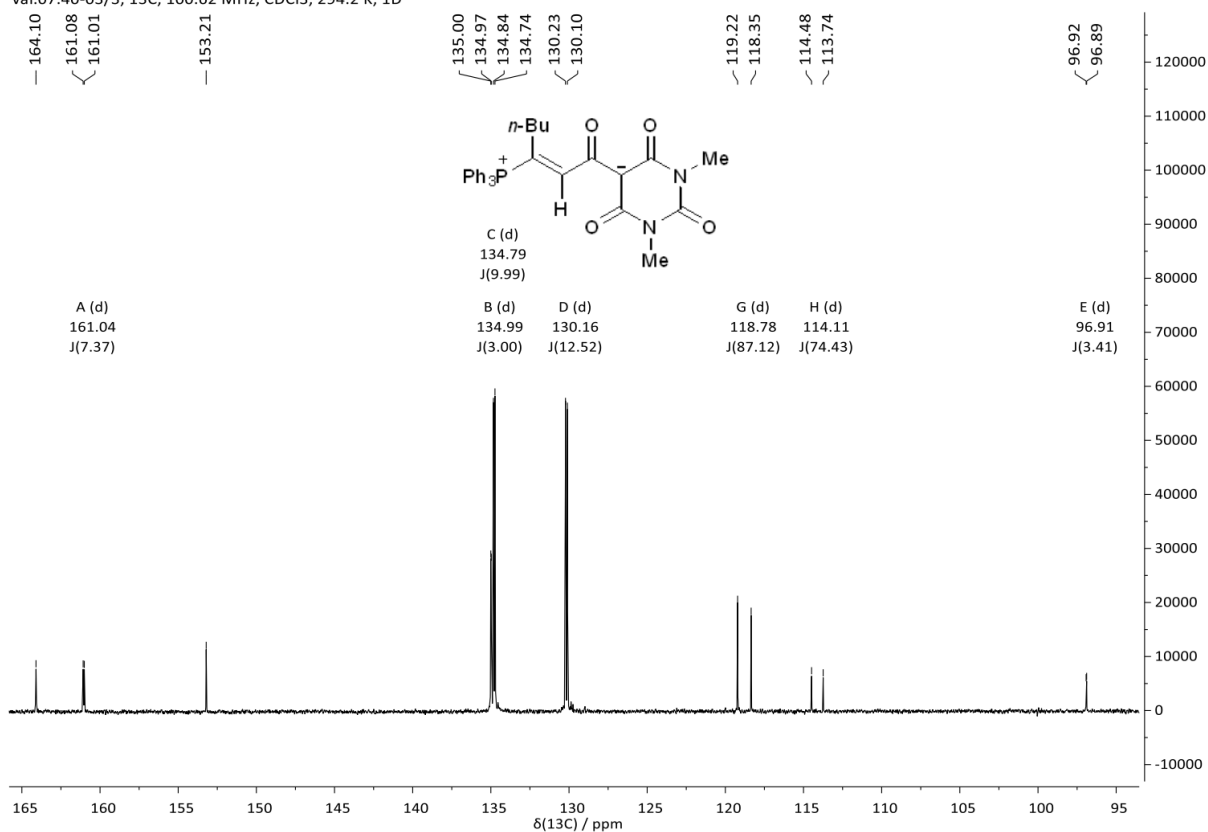


¹³C NMR

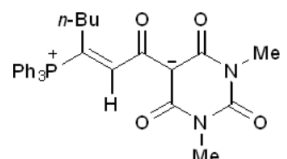
vaf.07.40-03/3, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D



vaf.07.40-03/3, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D



vaf.07.40-31P/1, 31P, 161.97 MHz, 100 Scans, CDCl₃, 294.4 K, 1D

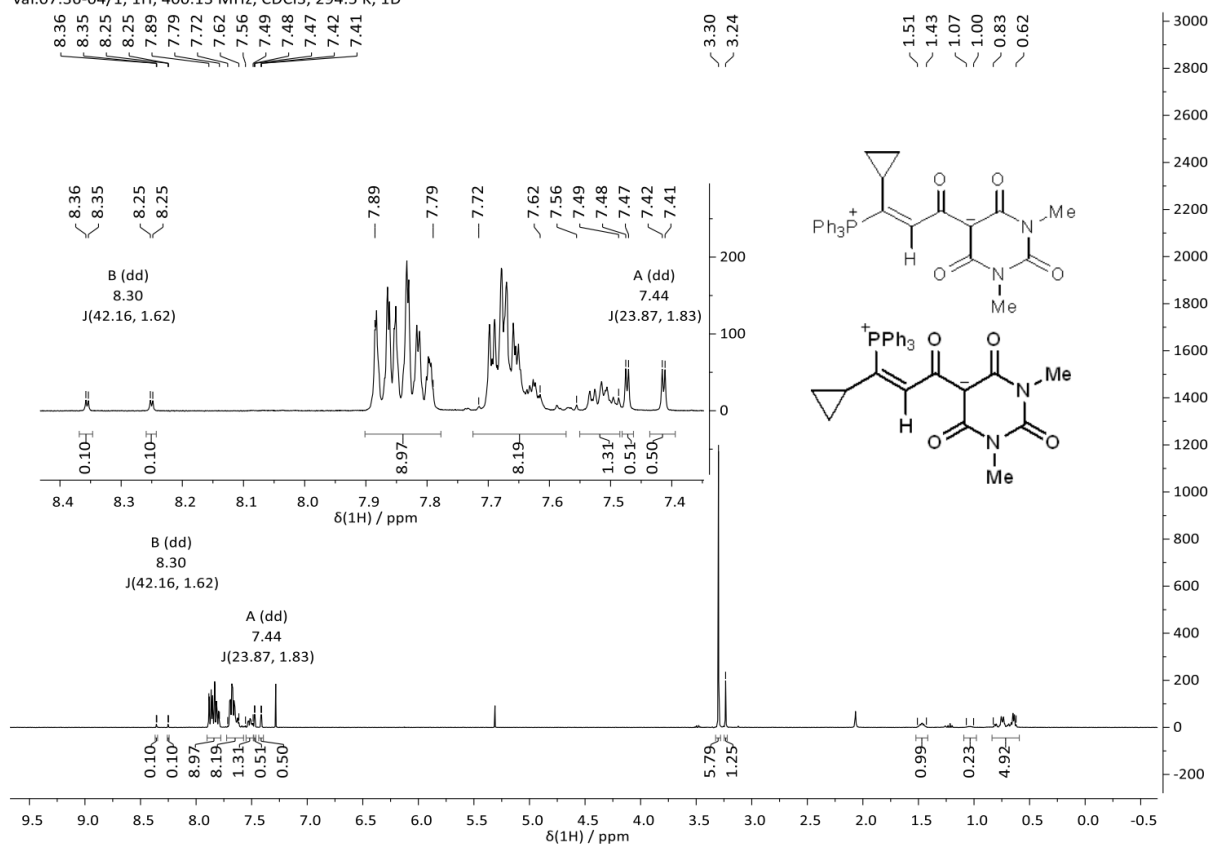


Probenname: vaf.07.40-1	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 21/03/2017
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided, Forward-Backward	Dateipfad: D:\DATEN IR BRUKER\FIORE	Datei: VAF.07.40-1.0

4.11. Betaines (E)- and (Z)-3k (mixture of diastereoisomers)

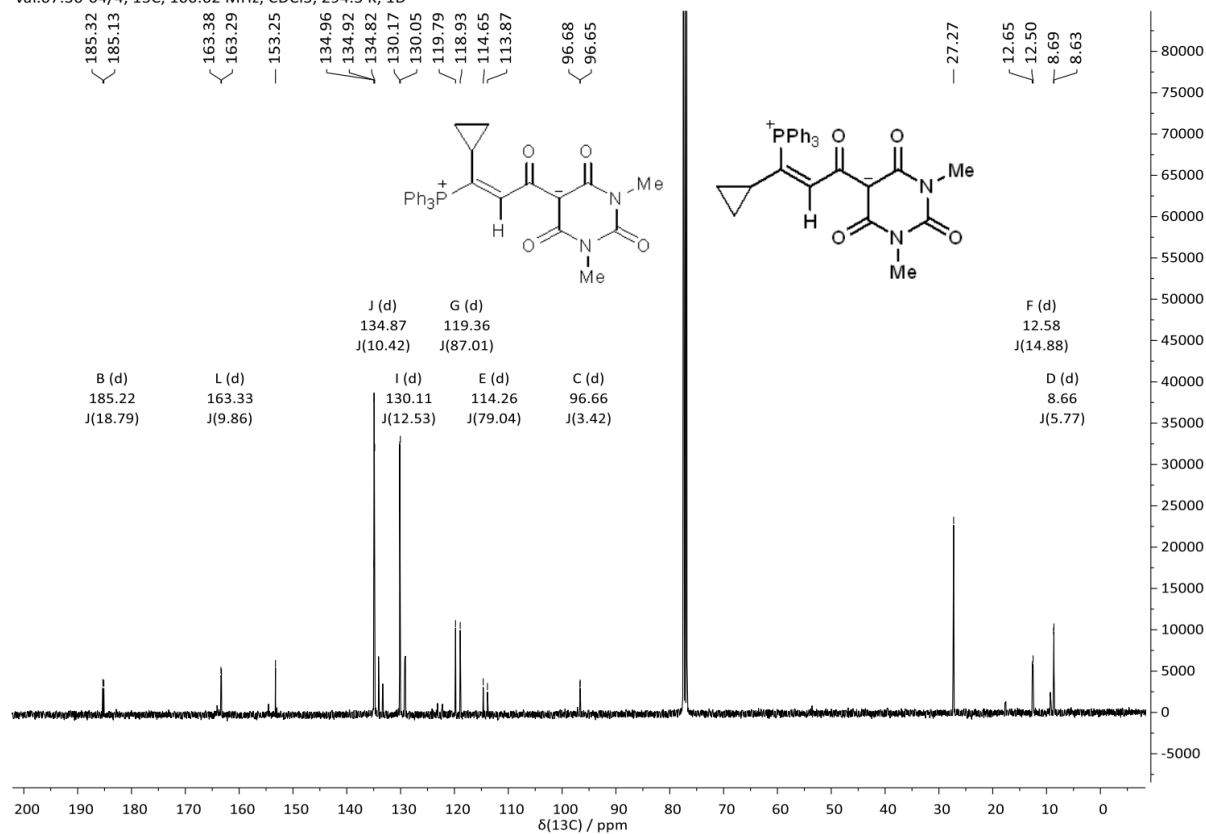
^1H NMR

vaf.07.36-04/1, ^1H , 400.13 MHz, CDCl_3 , 294.5 K, 1D

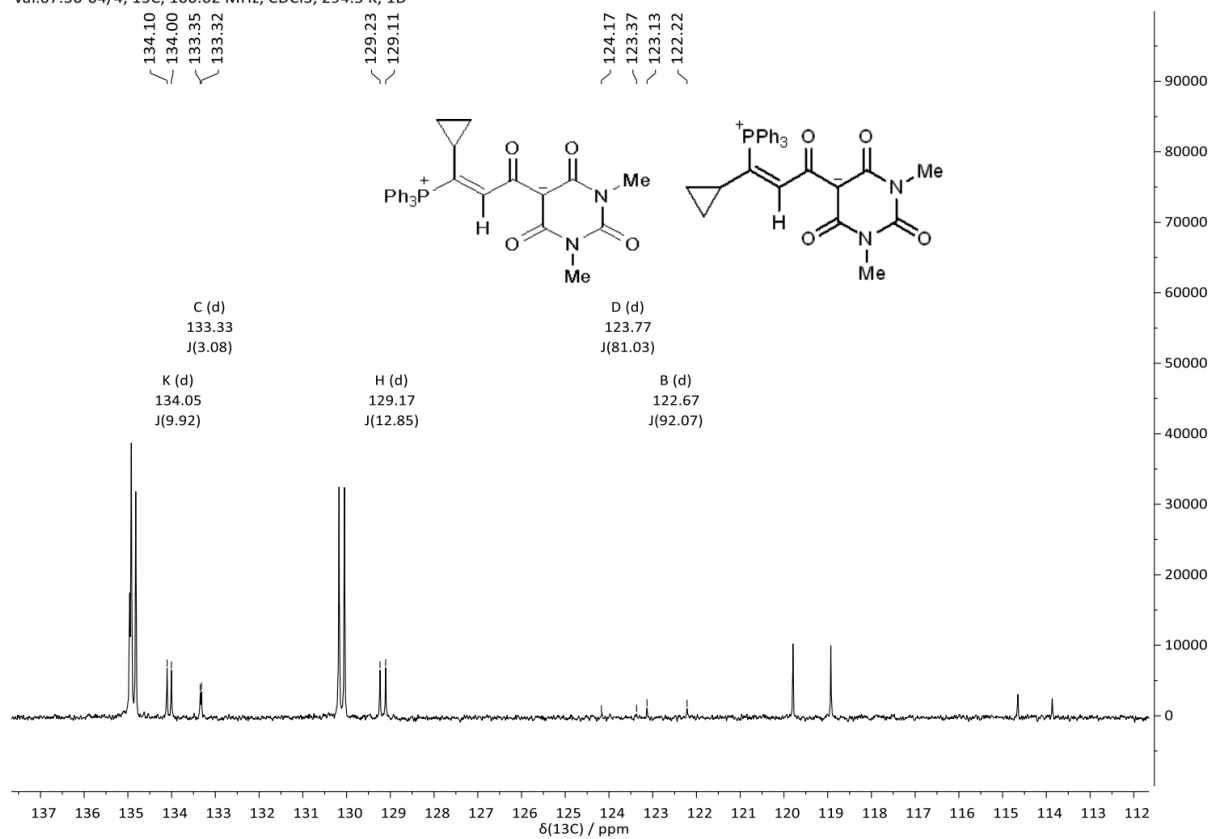


^{13}C NMR

vaf.07.36-04/4, ^{13}C , 100.62 MHz, CDCl_3 , 294.5 K, 1D

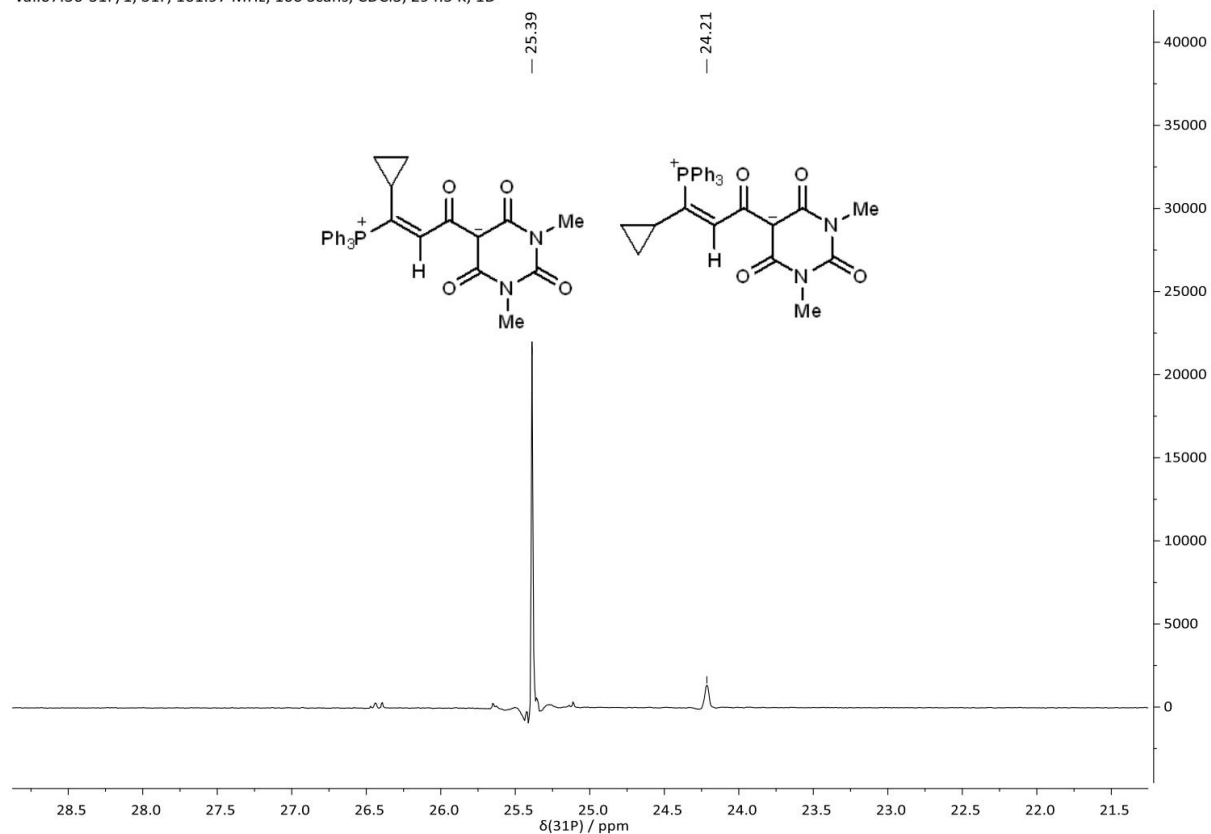


vaf.07.36-04/4, ¹³C, 100.62 MHz, CDCl₃, 294.5 K, 1D

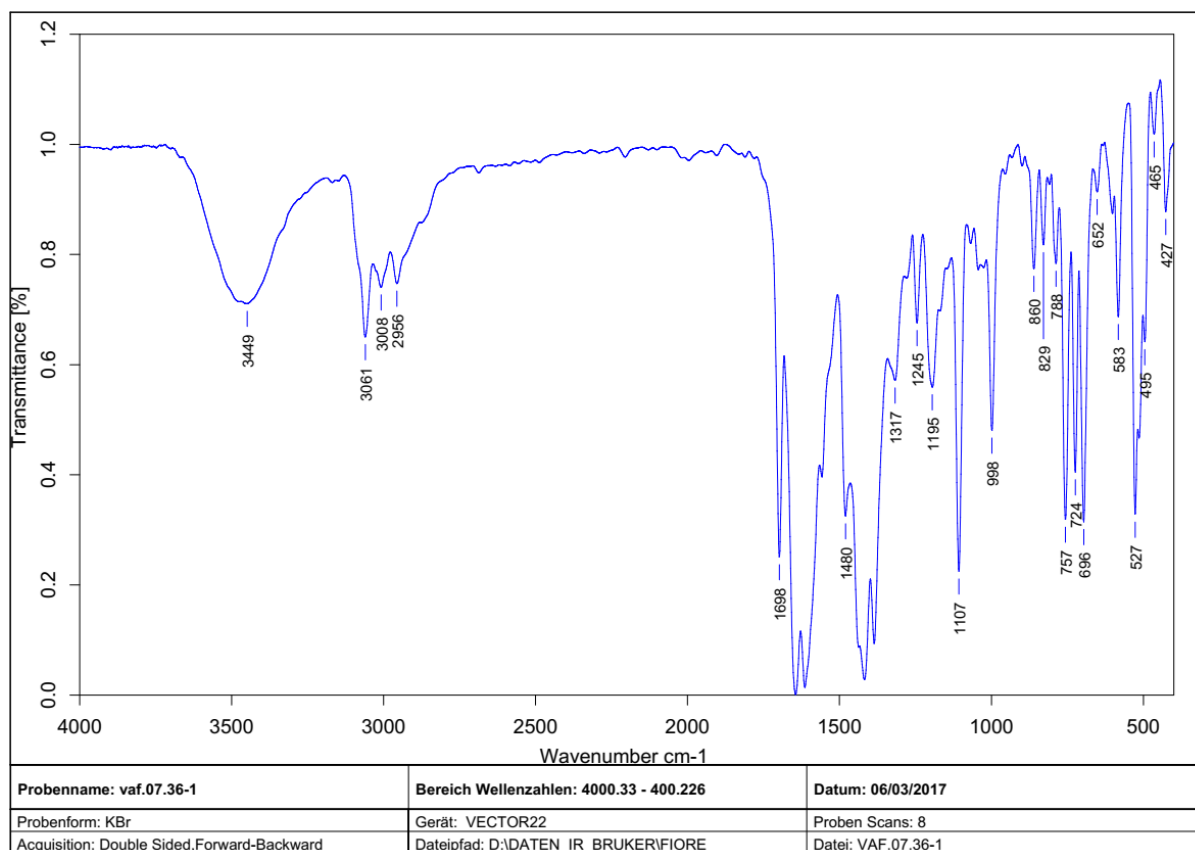


³¹P NMR

vaf.07.36-31P/1, ³¹P, 161.97 MHz, 100 Scans, CDCl₃, 294.5 K, 1D



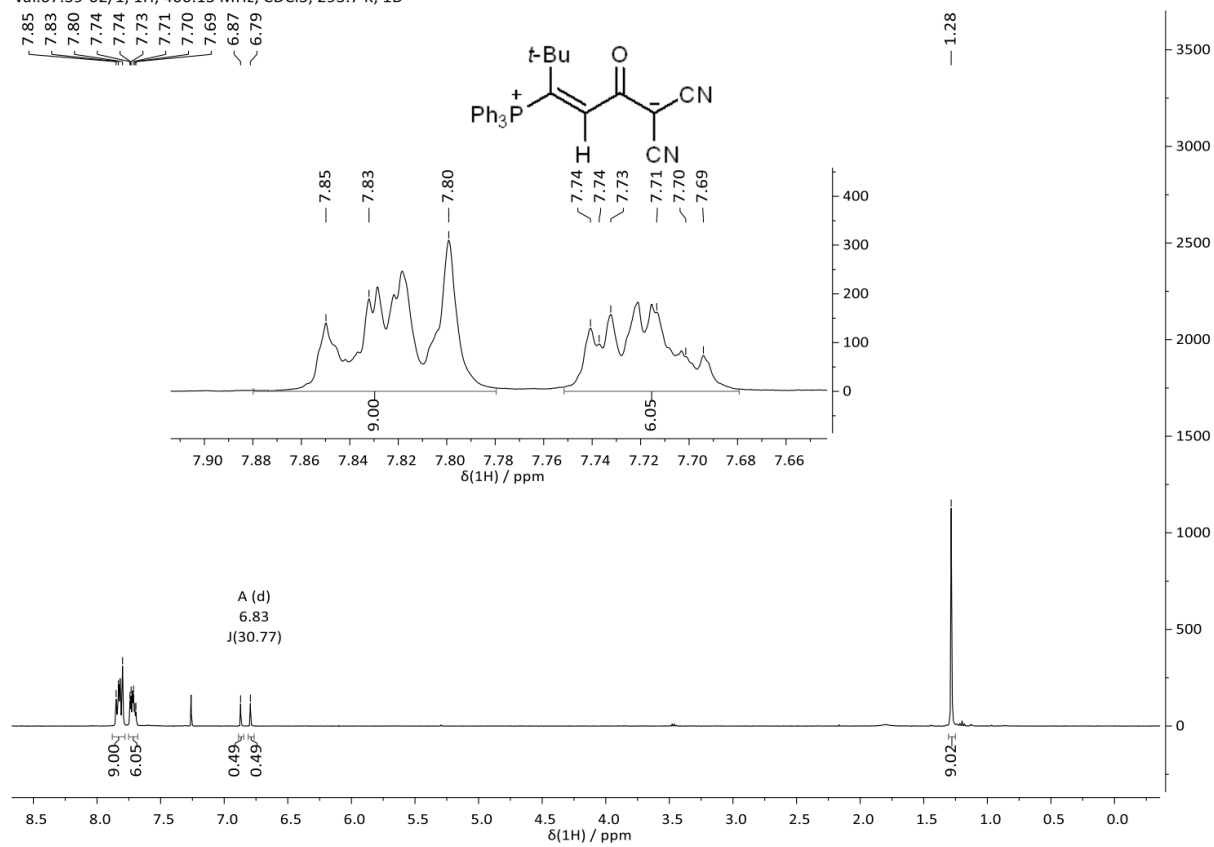
IR



4.12. Betaine (*E*)-3l

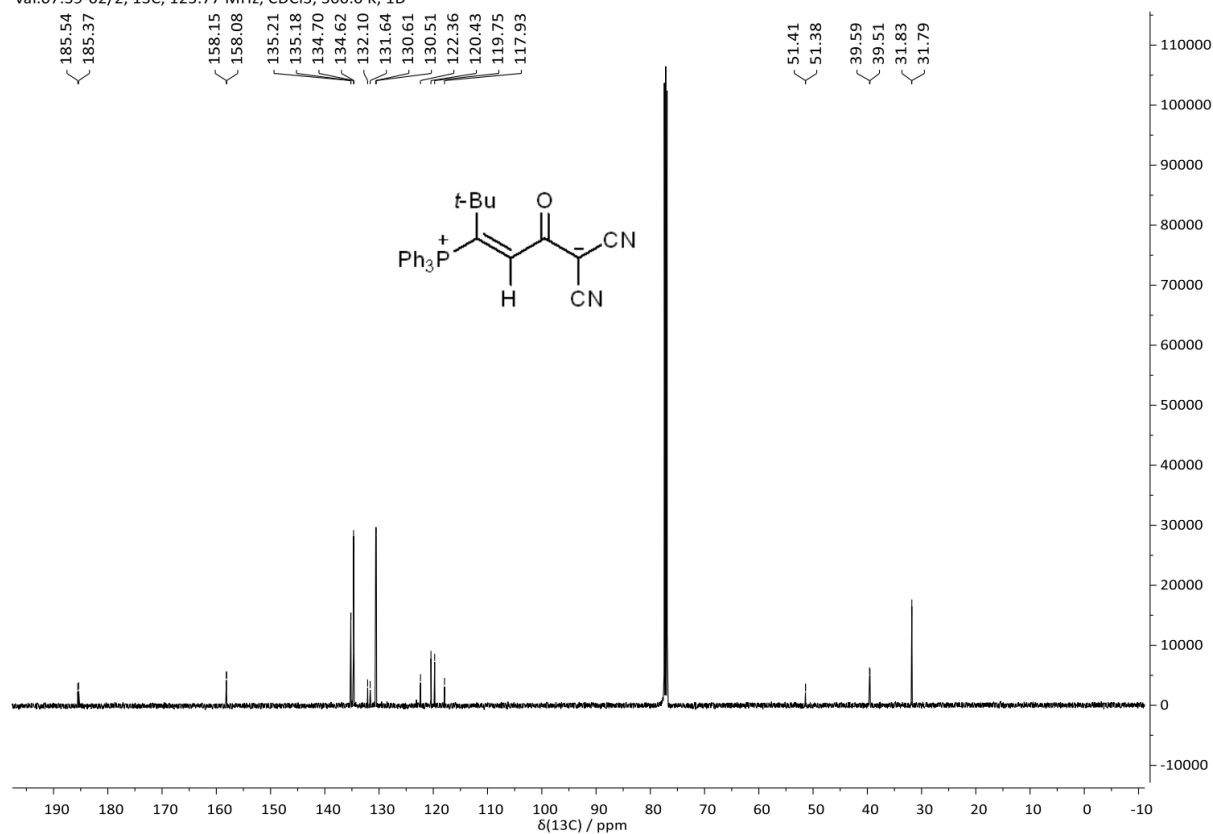
^1H NMR

vaf.07.39-02/1, ^1H , 400.13 MHz, CDCl_3 , 293.7 K, 1D

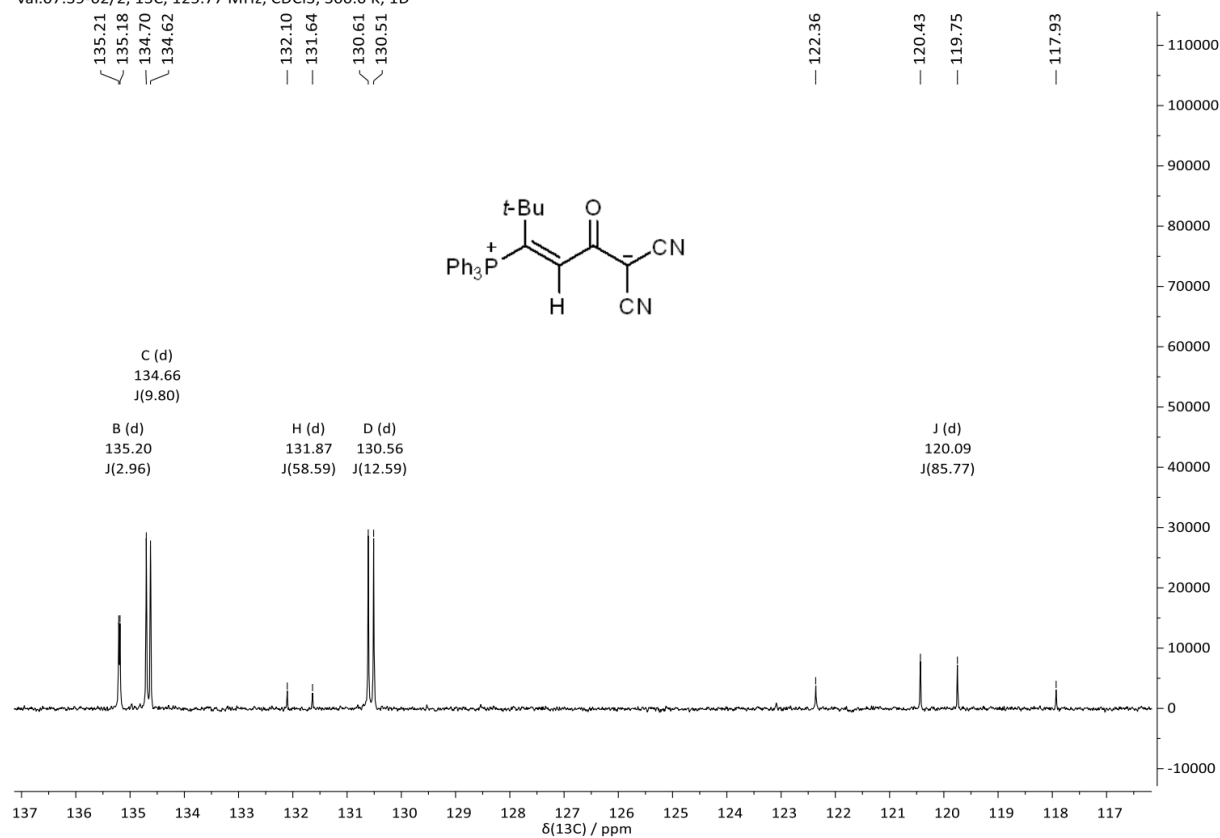


¹³C NMR

vaf.07.39-02/2, ¹³C, 125.77 MHz, CDCl₃, 300.0 K, 1D

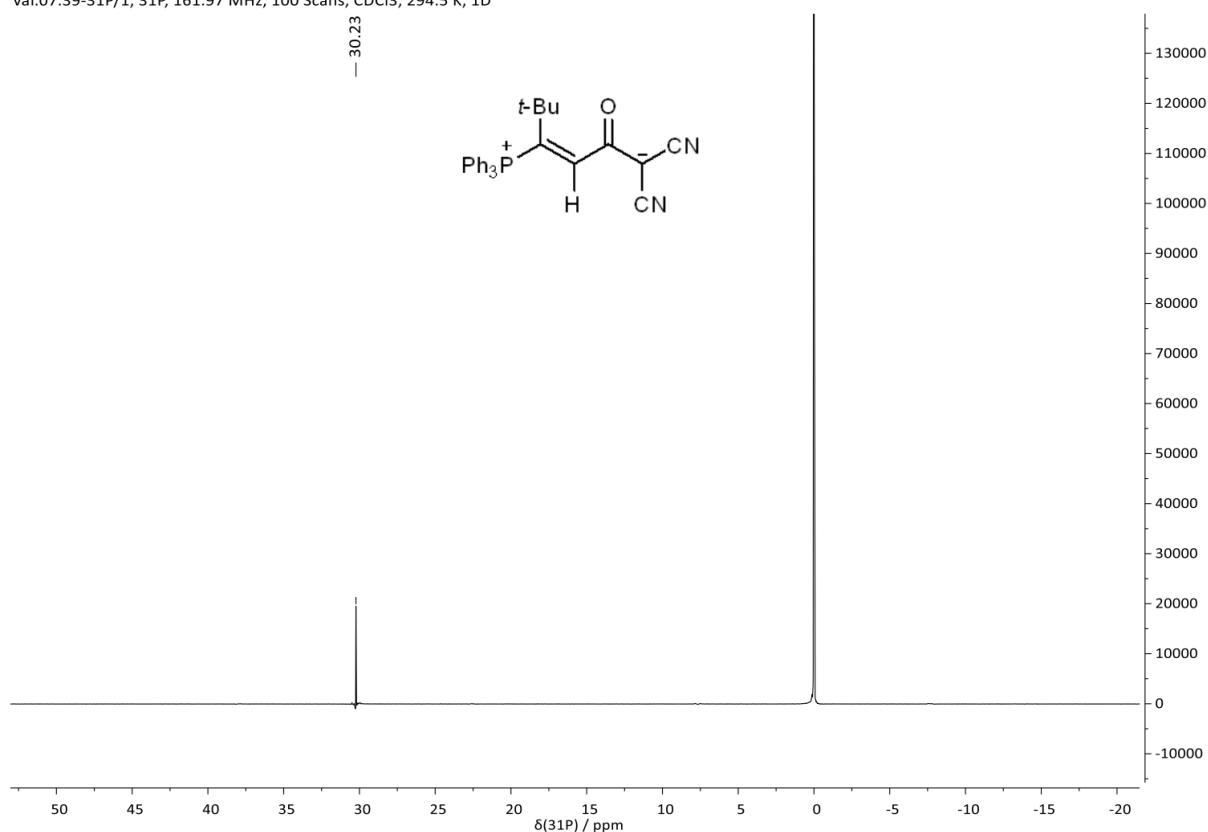


vaf.07.39-02/2, ¹³C, 125.77 MHz, CDCl₃, 300.0 K, 1D

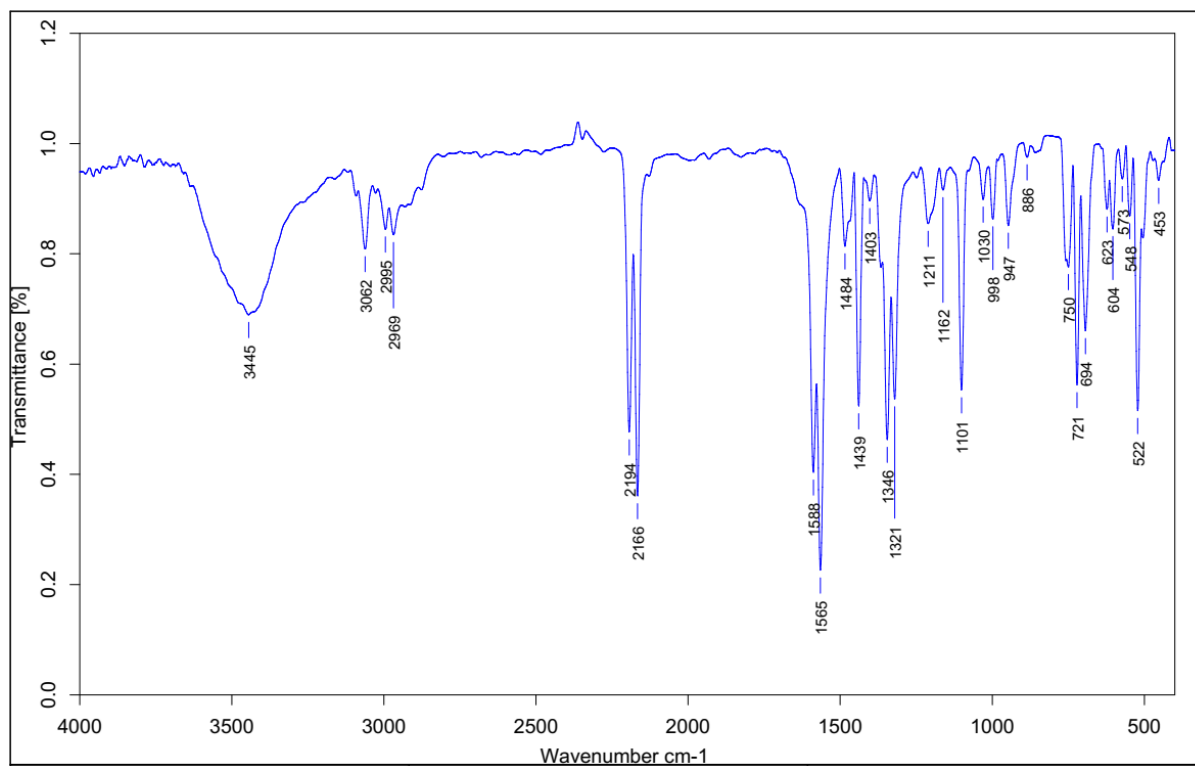


³¹P NMR

vaf.07.39-31P/1, 31P, 161.97 MHz, 100 Scans, CDCl₃, 294.5 K, 1D



IR

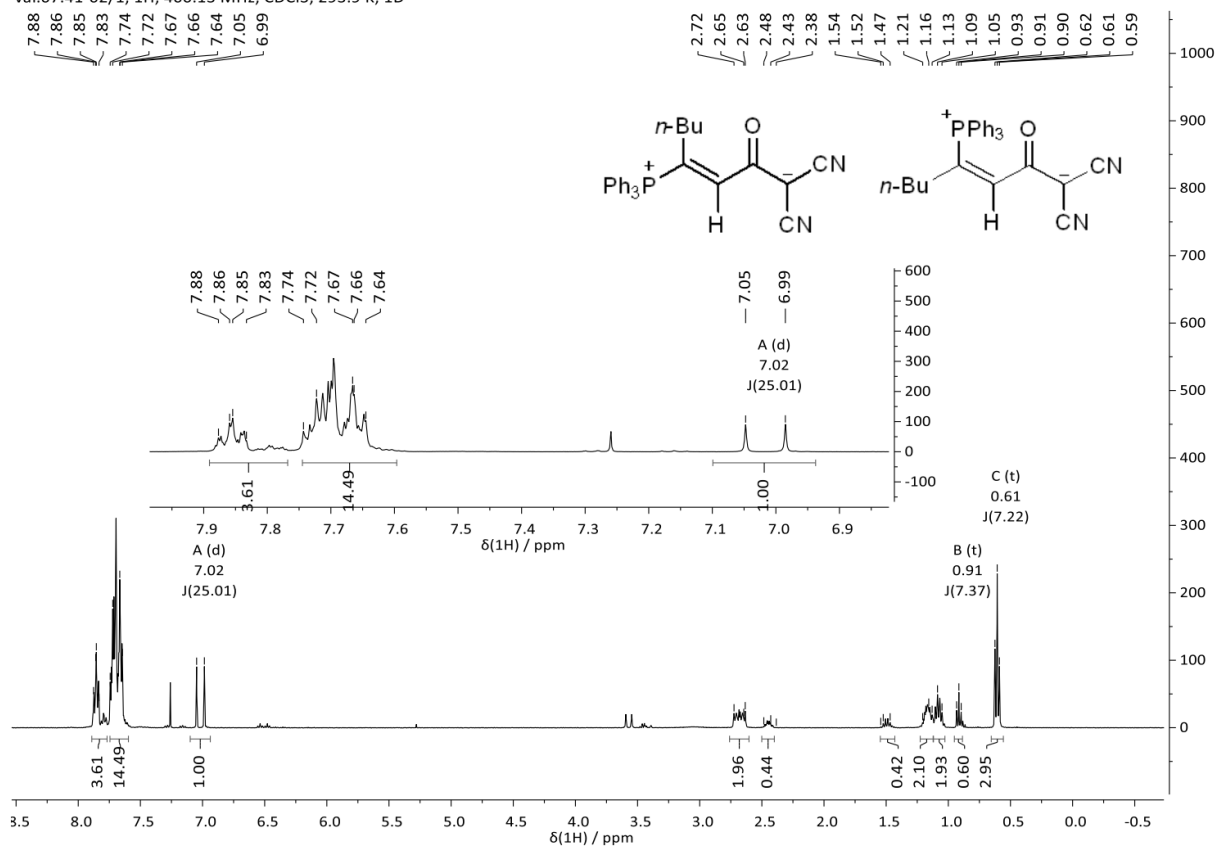


Probenname: vaf.07.39-1	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 17/03/2017
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided,Forward-Backward	Dateipfad: D:\DATEN_IR_BRUKER\FIORE	Datei: VAF.07.39-1

4.13. Betaines (E)- and (Z)-3m

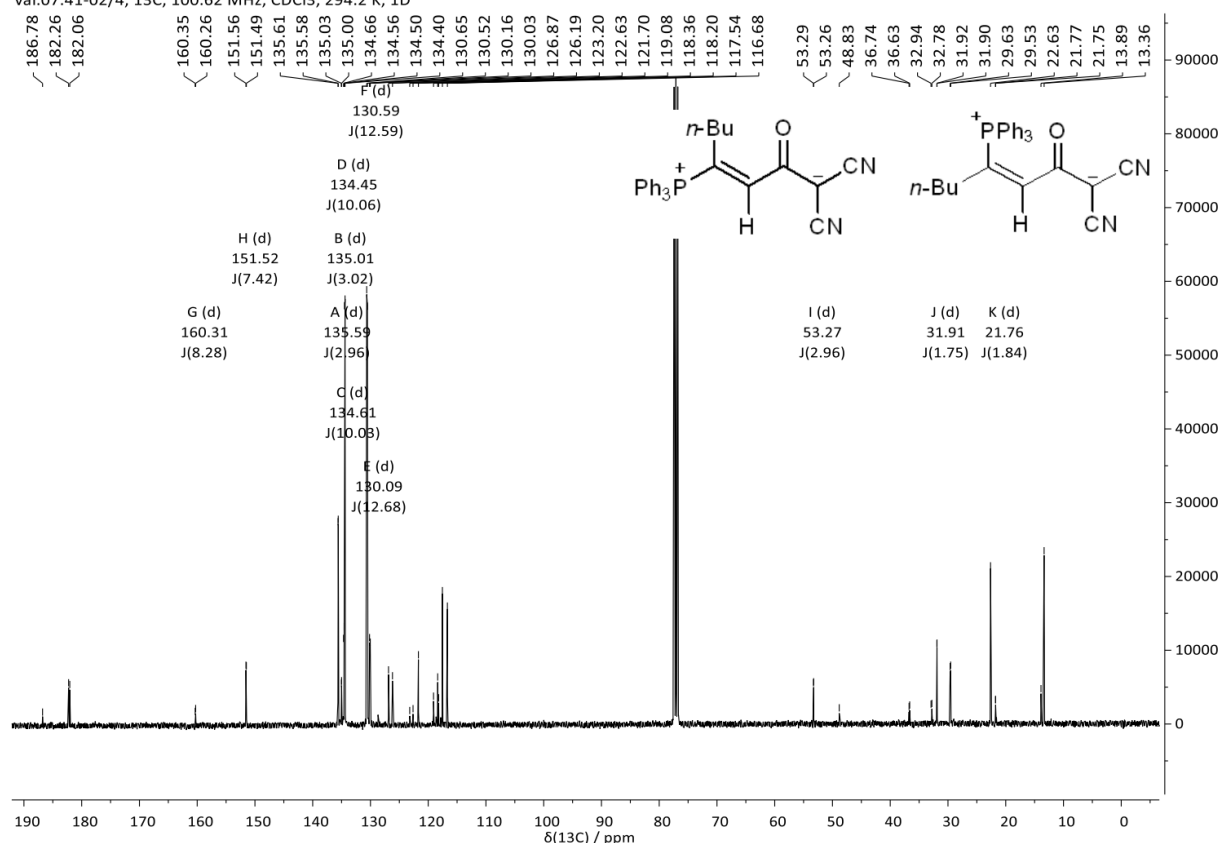
^1H NMR

vaf.07.41-02/1, 1H, 400.13 MHz, CDCl_3 , 293.9 K, 1D

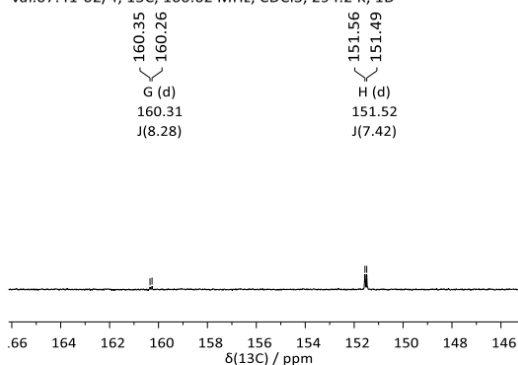


¹³C NMR

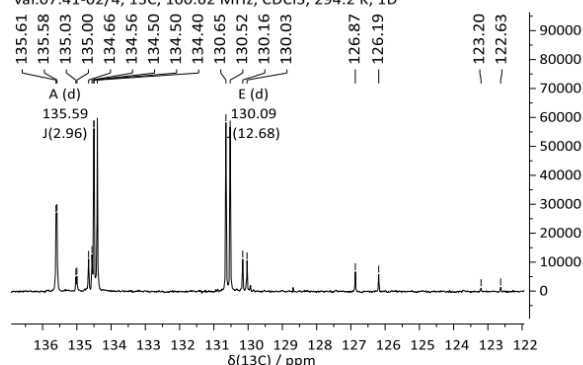
vaf.07.41-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D



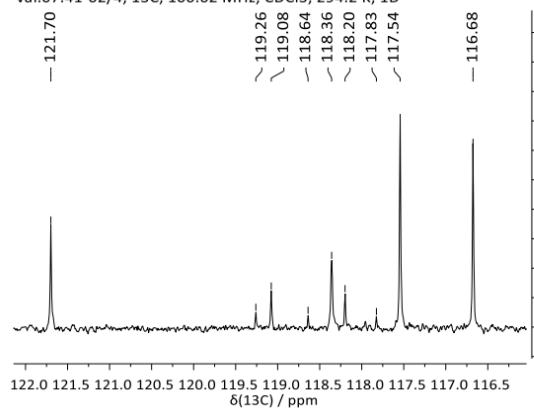
vaf.07.41-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D



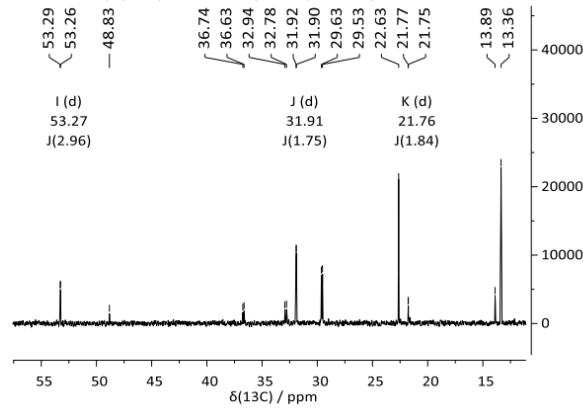
vaf.07.41-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D



vaf.07.41-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D

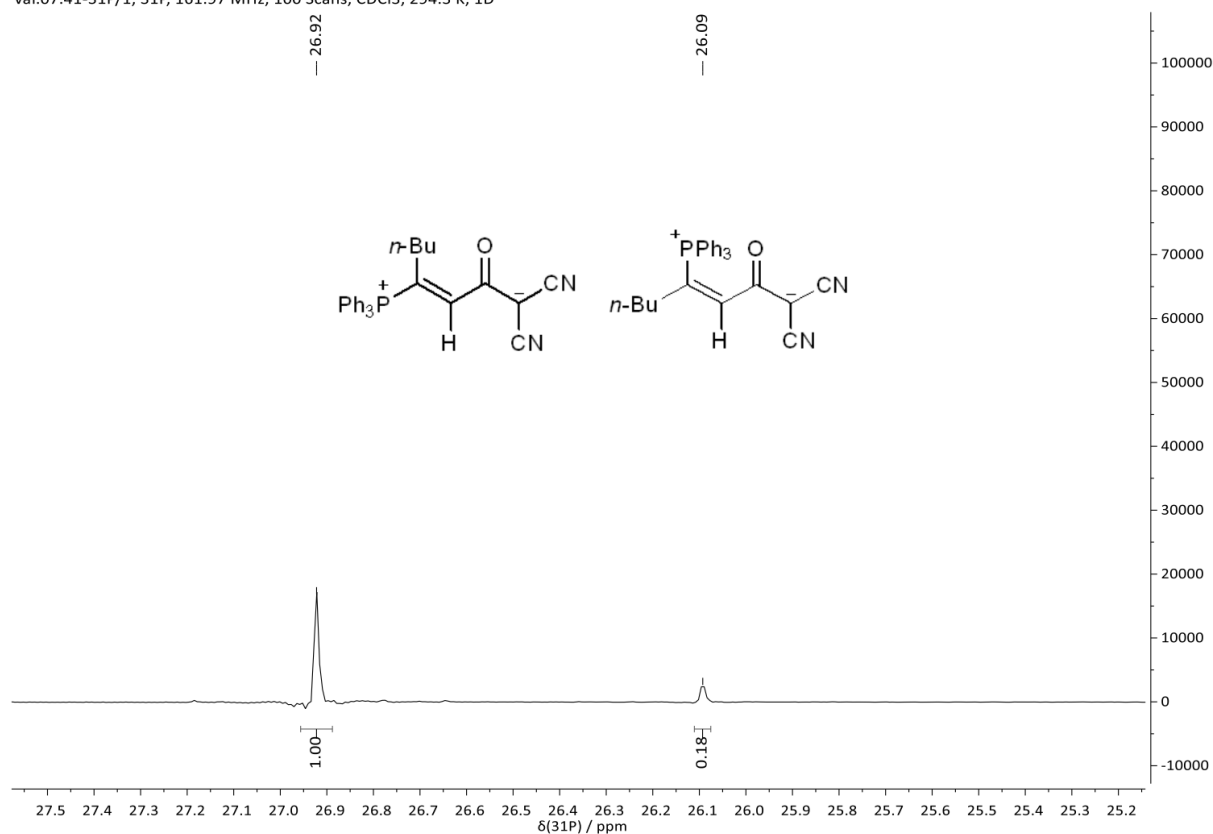


vaf.07.41-02/4, ¹³C, 100.62 MHz, CDCl₃, 294.2 K, 1D

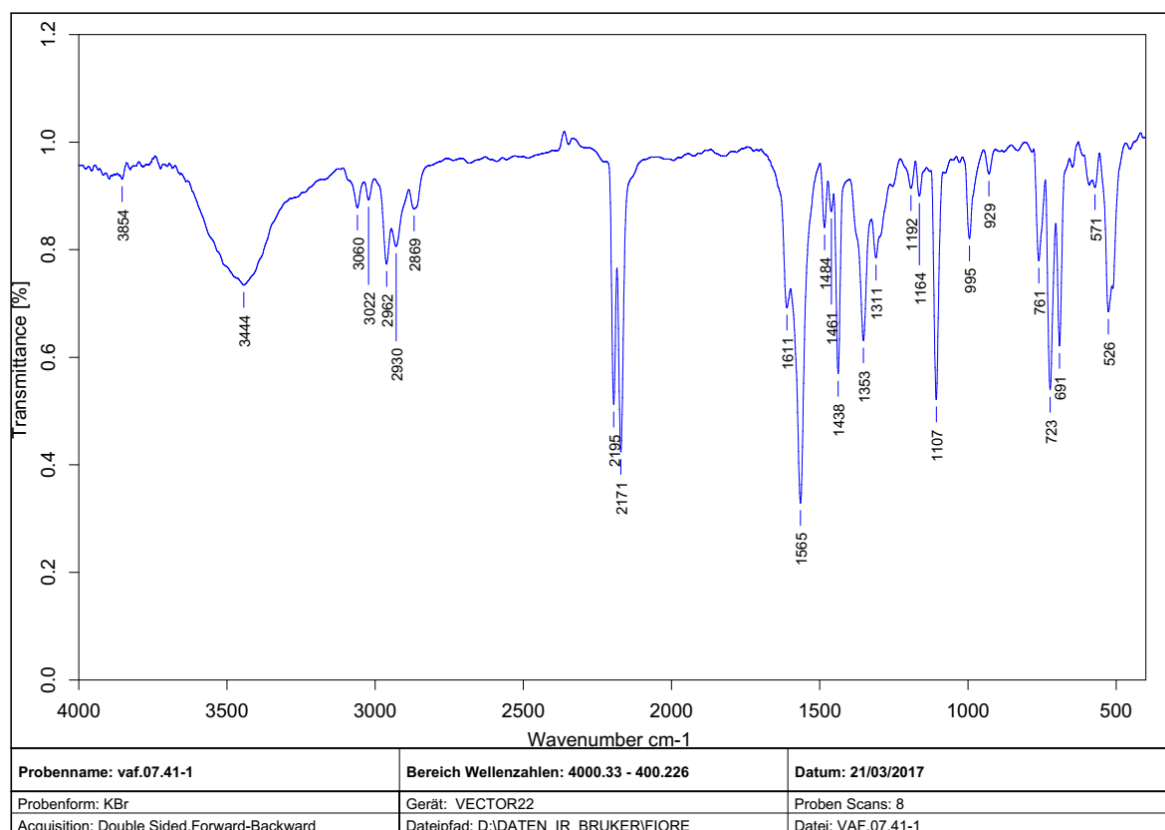


³¹P NMR

vaf.07.41-31P/1, 31P, 161.97 MHz, 100 Scans, CDCl₃, 294.3 K, 1D



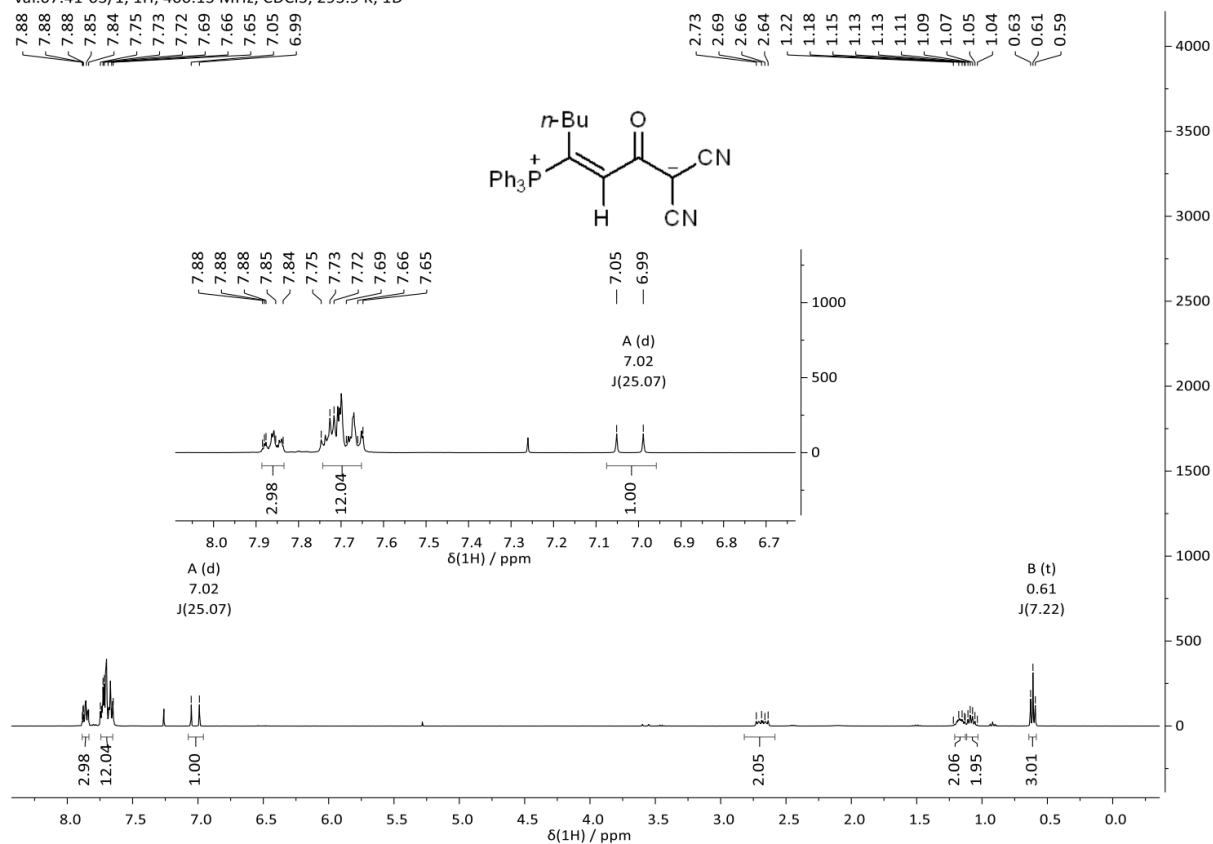
IR



(E)-3m

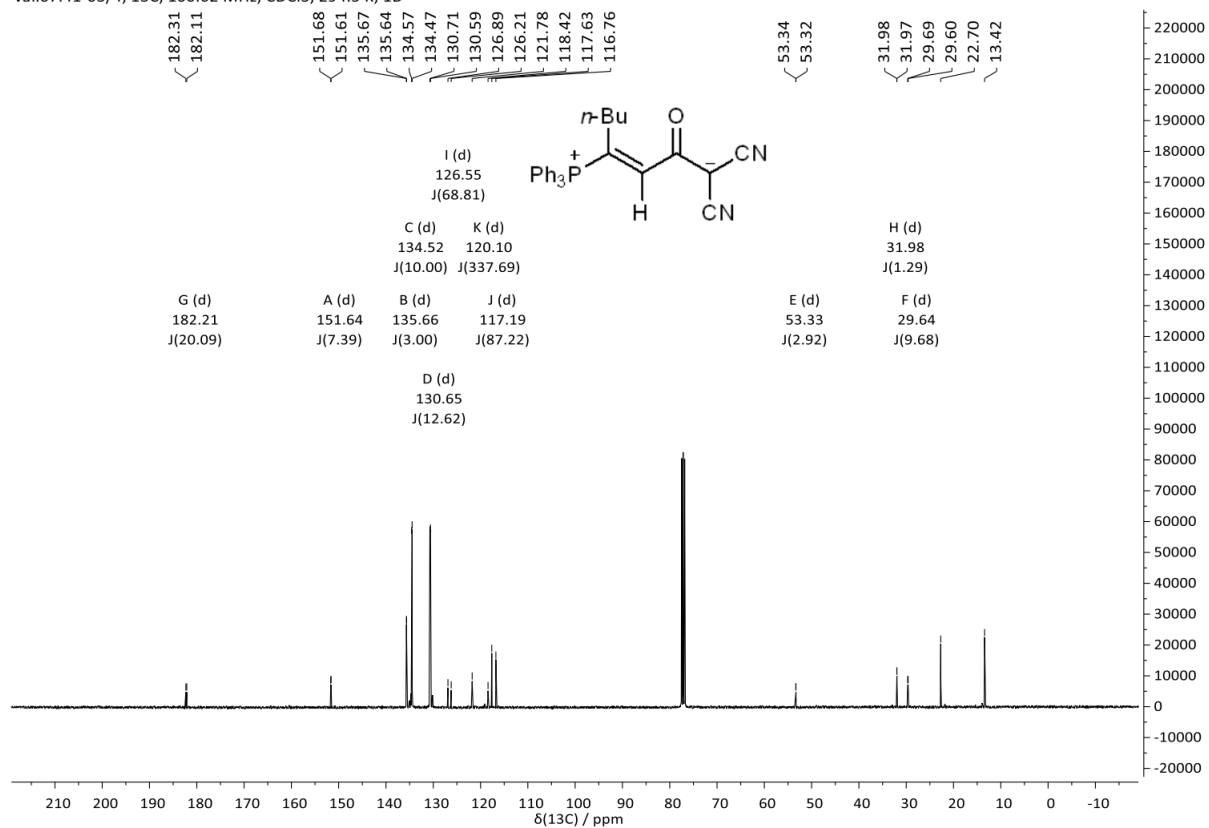
¹H NMR

vaf.07.41-03/1, ¹H, 400.13 MHz, CDCl₃, 293.9 K, 1D

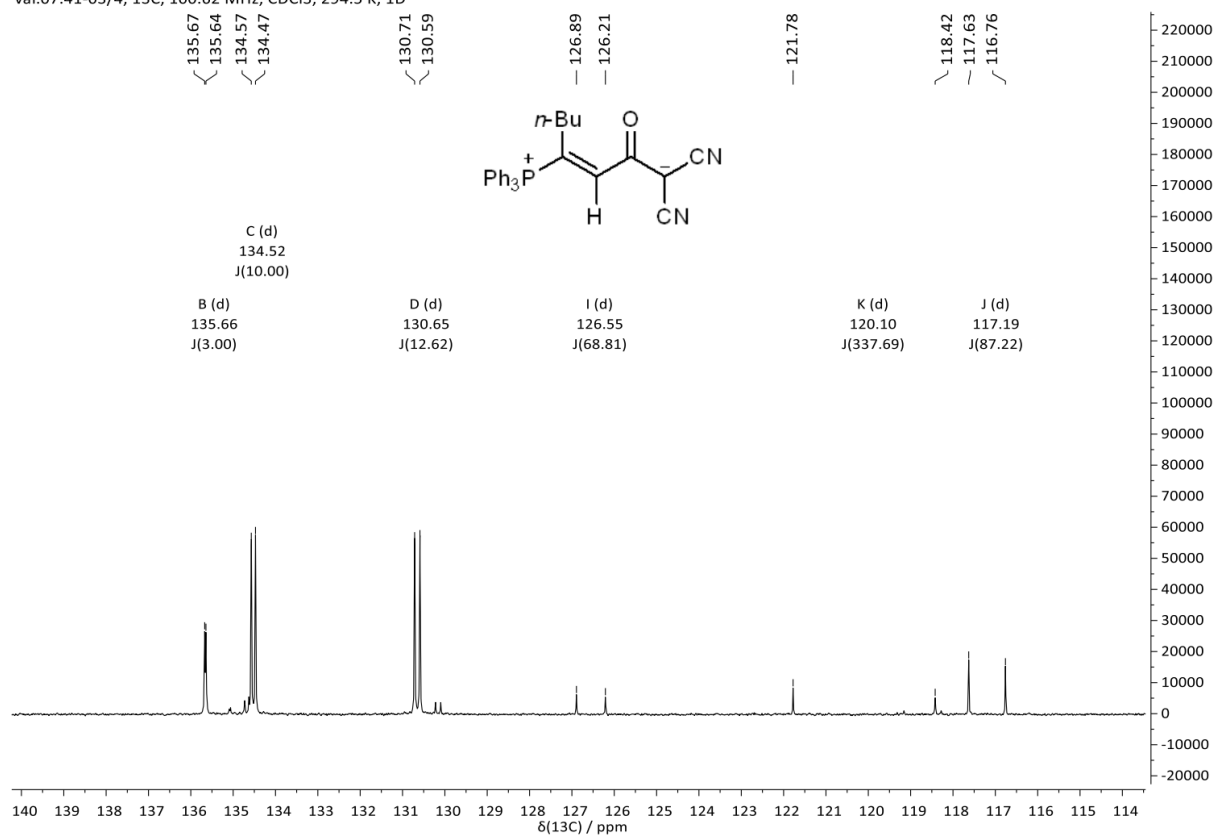


¹³C NMR

vaf.07.41-03/4, ¹³C, 100.62 MHz, CDCl₃, 294.5 K, 1D



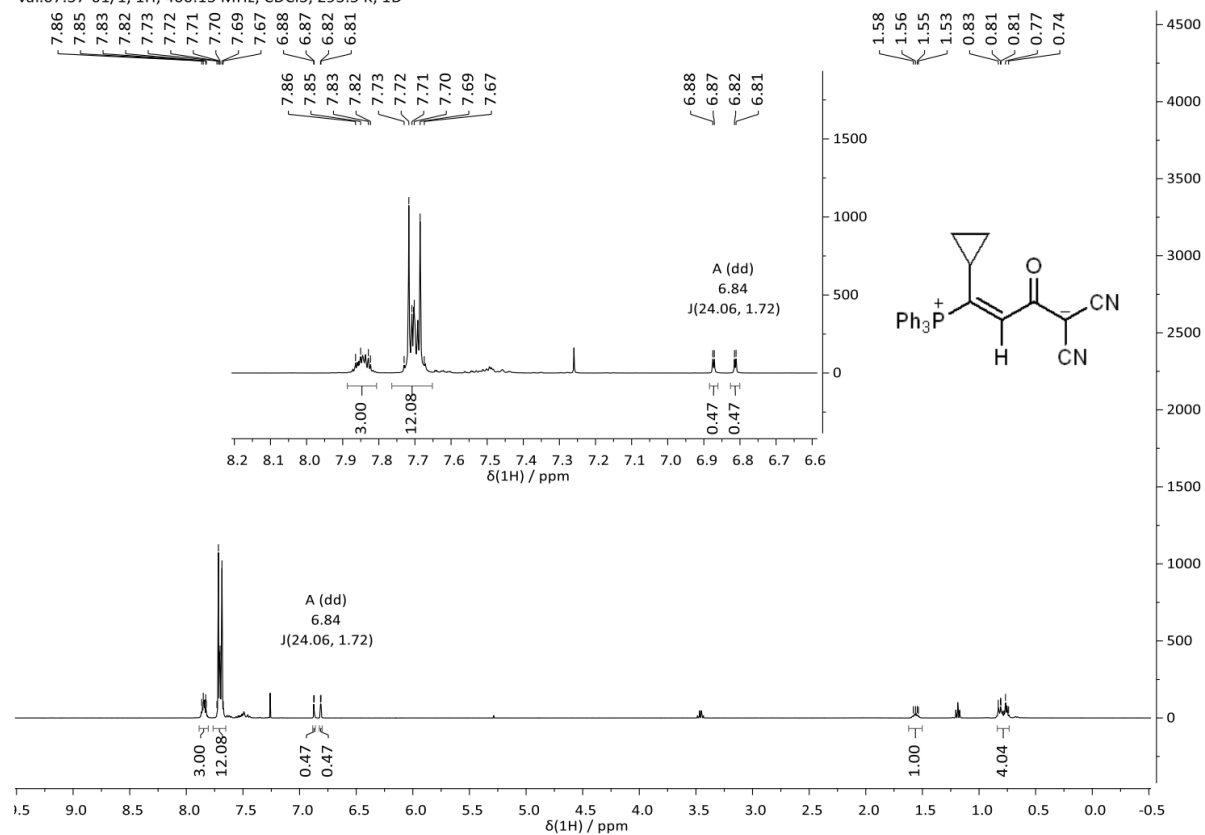
vaf.07.41-03/4, 13C, 100.62 MHz, CDCl3, 294.5 K, 1D



4.14. Betaine (*E*)-3n

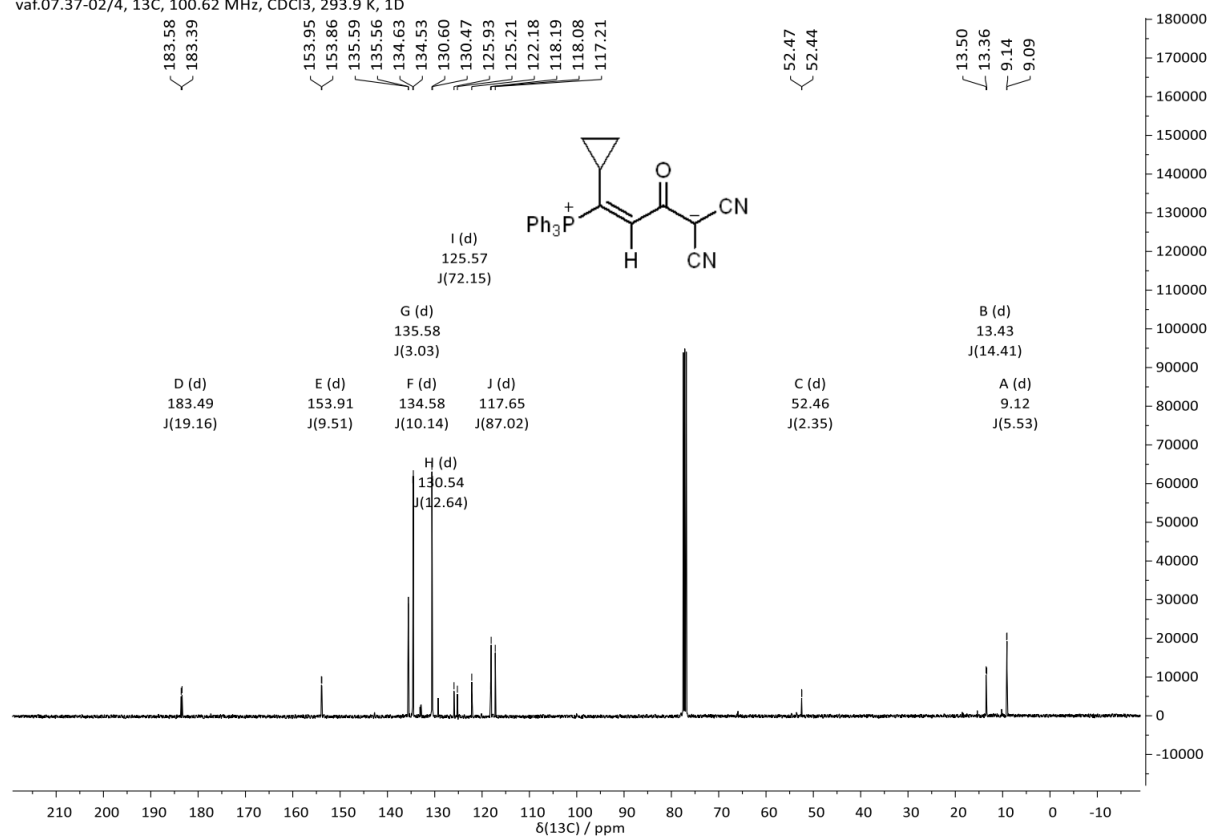
¹H NMR

vaf.07.37-01/1, 1H, 400.13 MHz, CDCl₃, 293.5 K, 1D

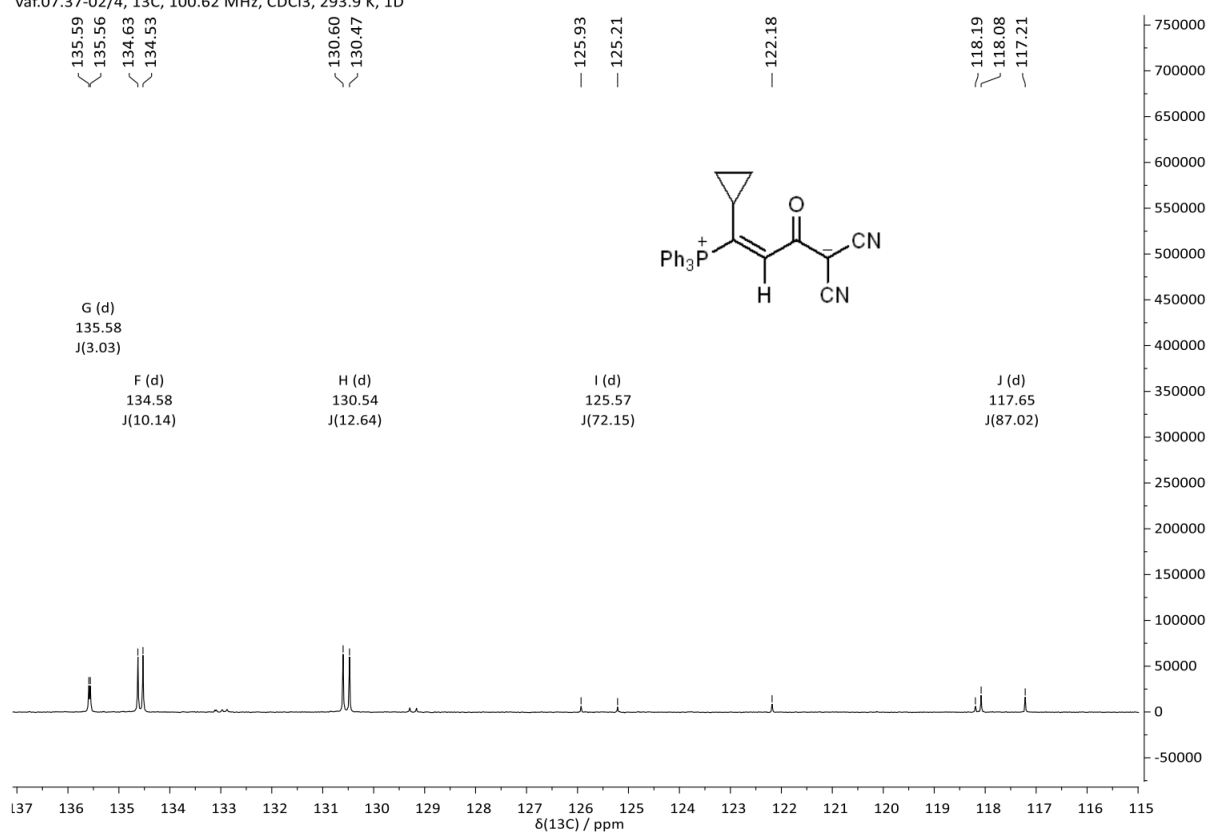


¹³C NMR

vaf.07.37-02/4, ¹³C, 100.62 MHz, CDCl₃, 293.9 K, 1D

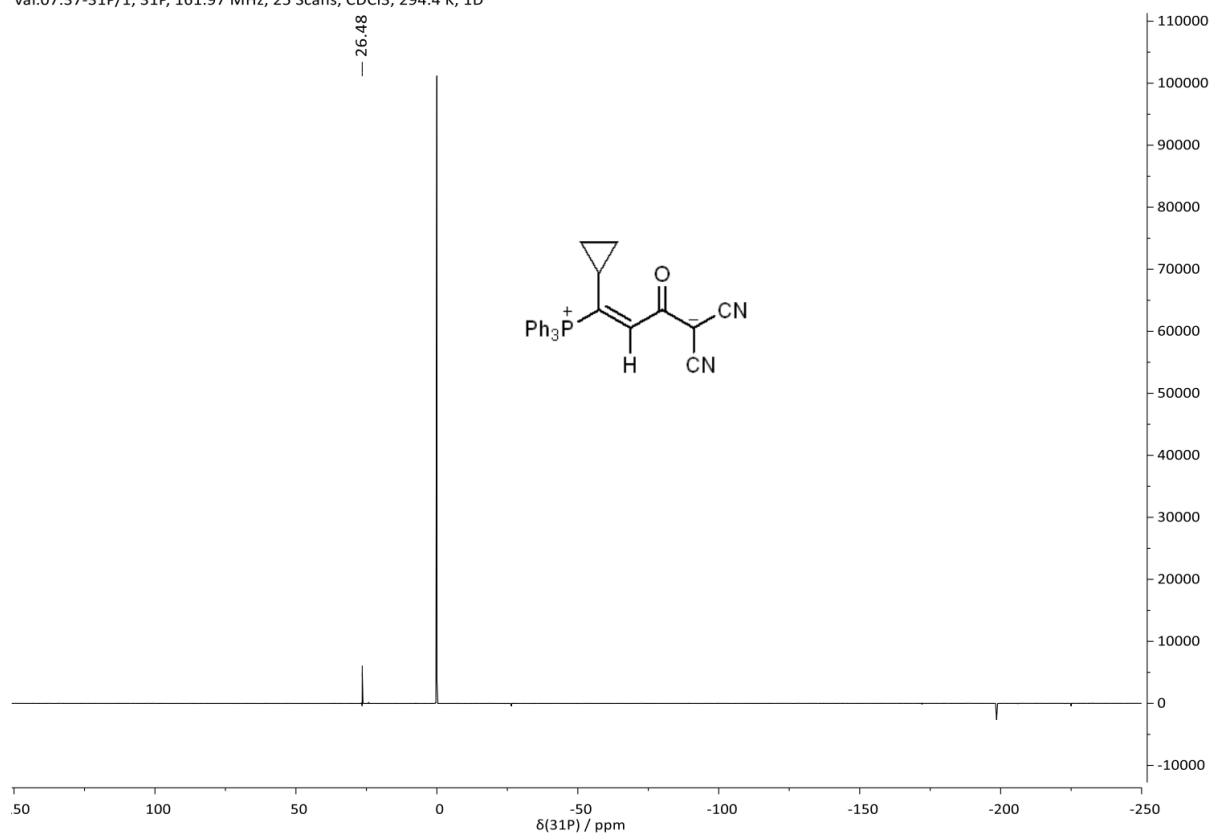


vaf.07.37-02/4, ¹³C, 100.62 MHz, CDCl₃, 293.9 K, 1D

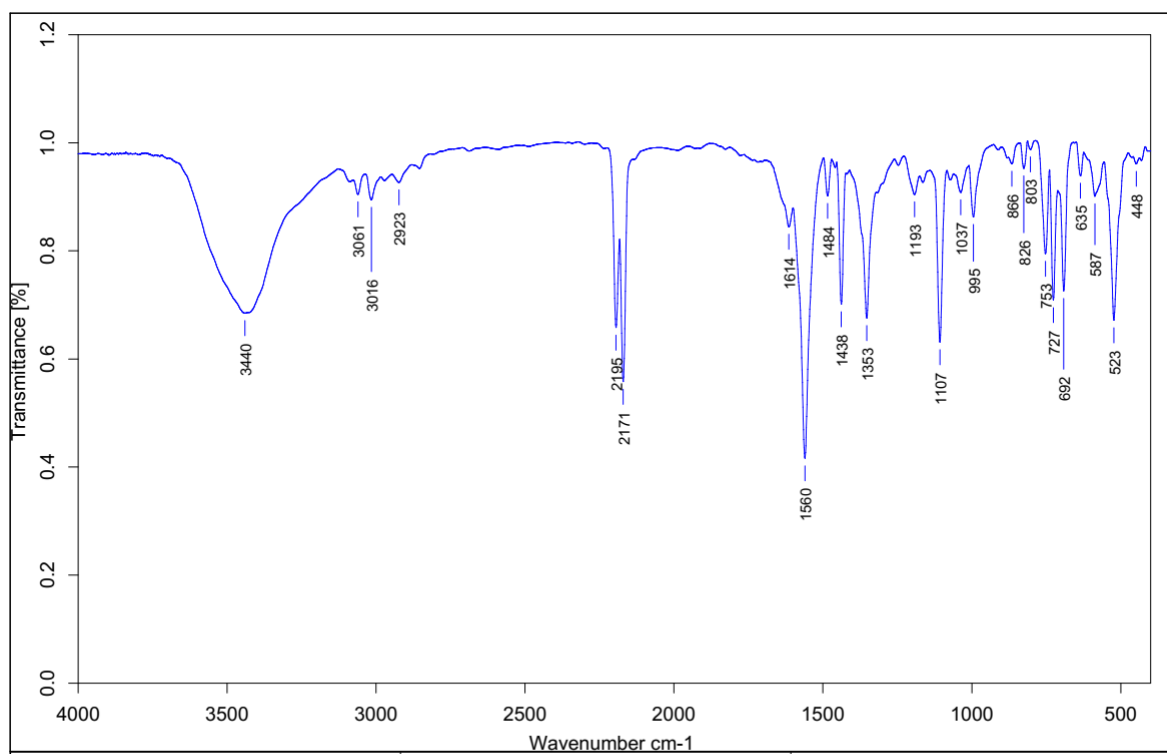


³¹P NMR

vaf.07.37-31P/1, 31P, 161.97 MHz, 25 Scans, CDCl₃, 294.4 K, 1D



IR

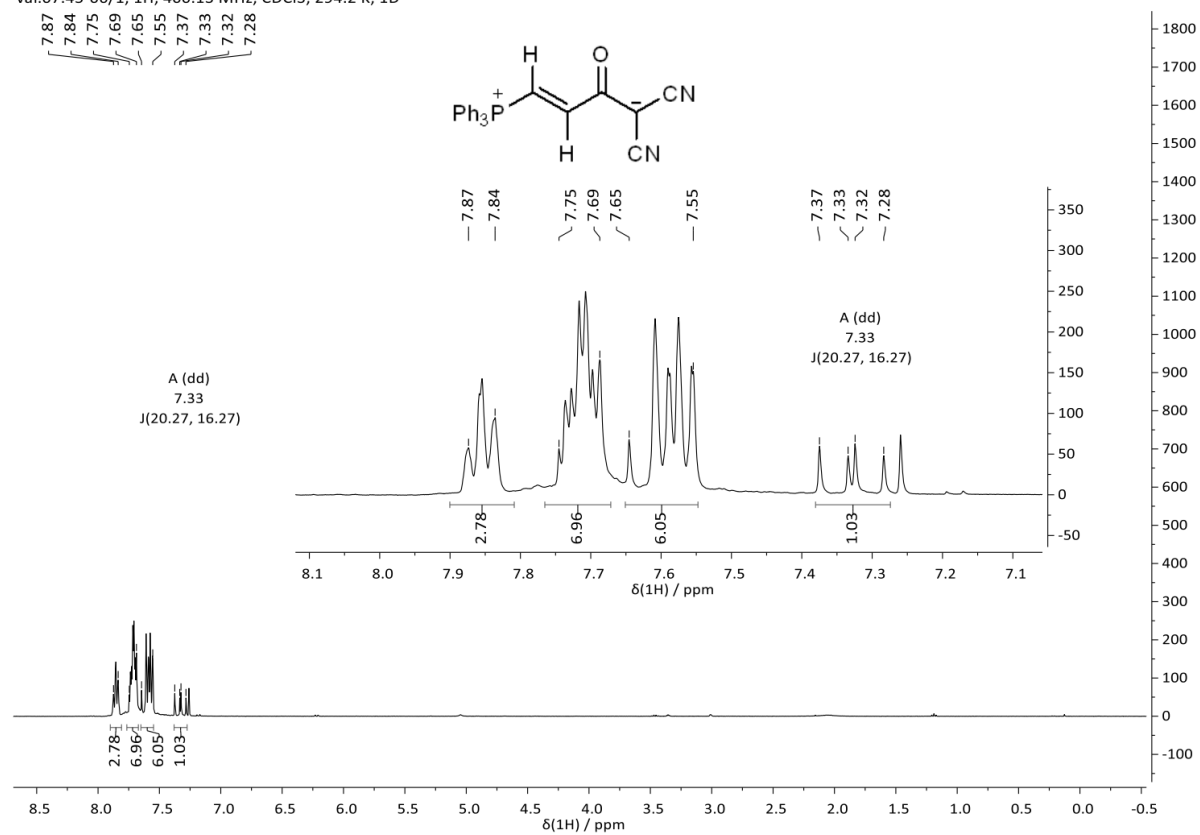


Probenname: vaf.07.37-1	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 08/03/2017
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided,Forward-Backward	Dateipfad: D:\DATEN IR_BRUKER\FIORE	Datei: VAF.07.37-1

4.15. Betaine (*E*)-3o

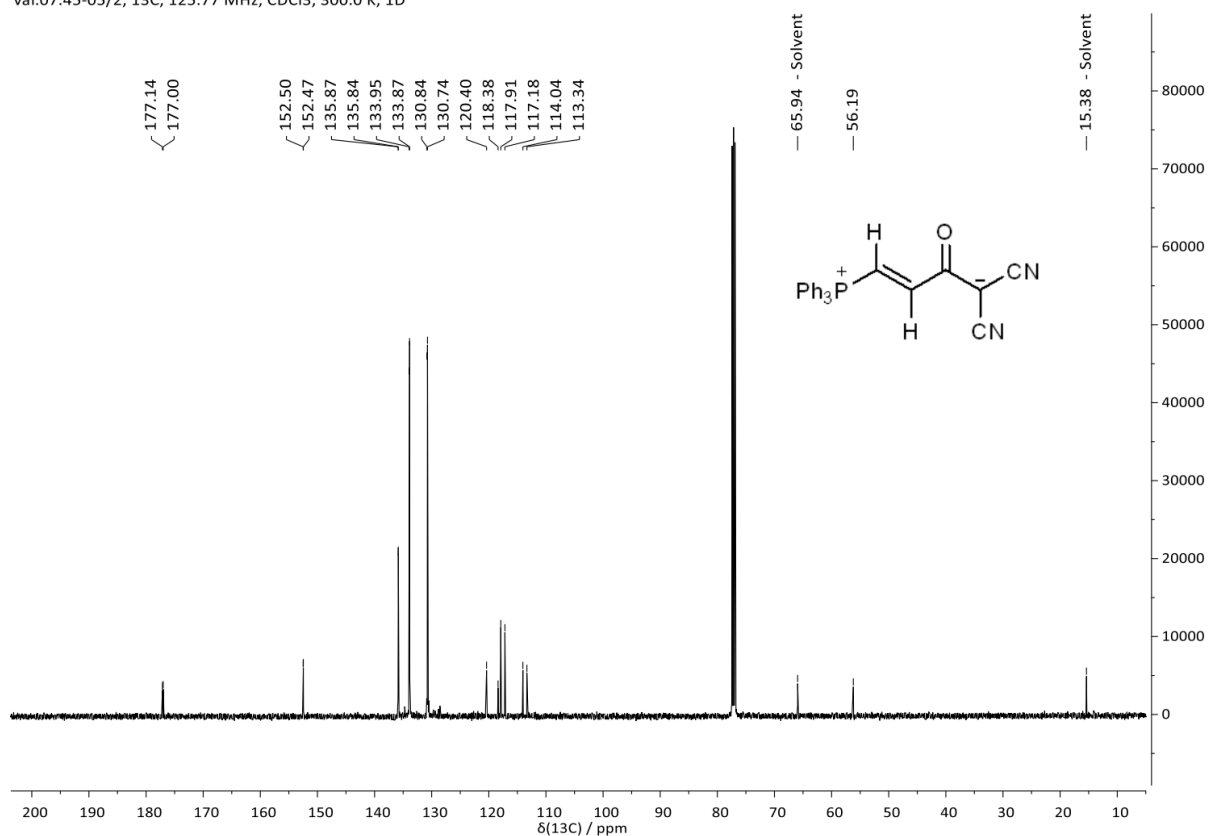
¹H NMR

vaf.07.45-06/1, 1H, 400.13 MHz, CDCl₃, 294.2 K, 1D

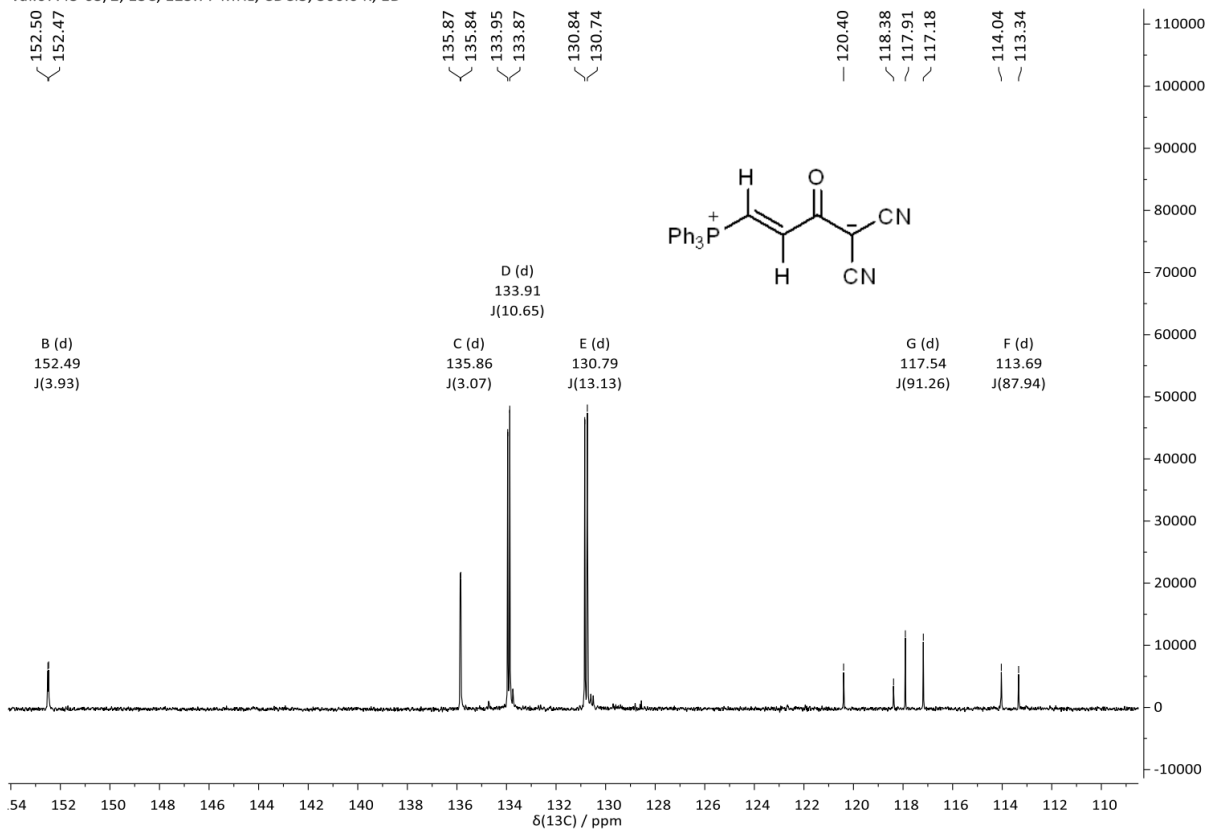


¹³C NMR

vaf.07.45-05/2, ¹³C, 125.77 MHz, CDCl₃, 300.0 K, 1D

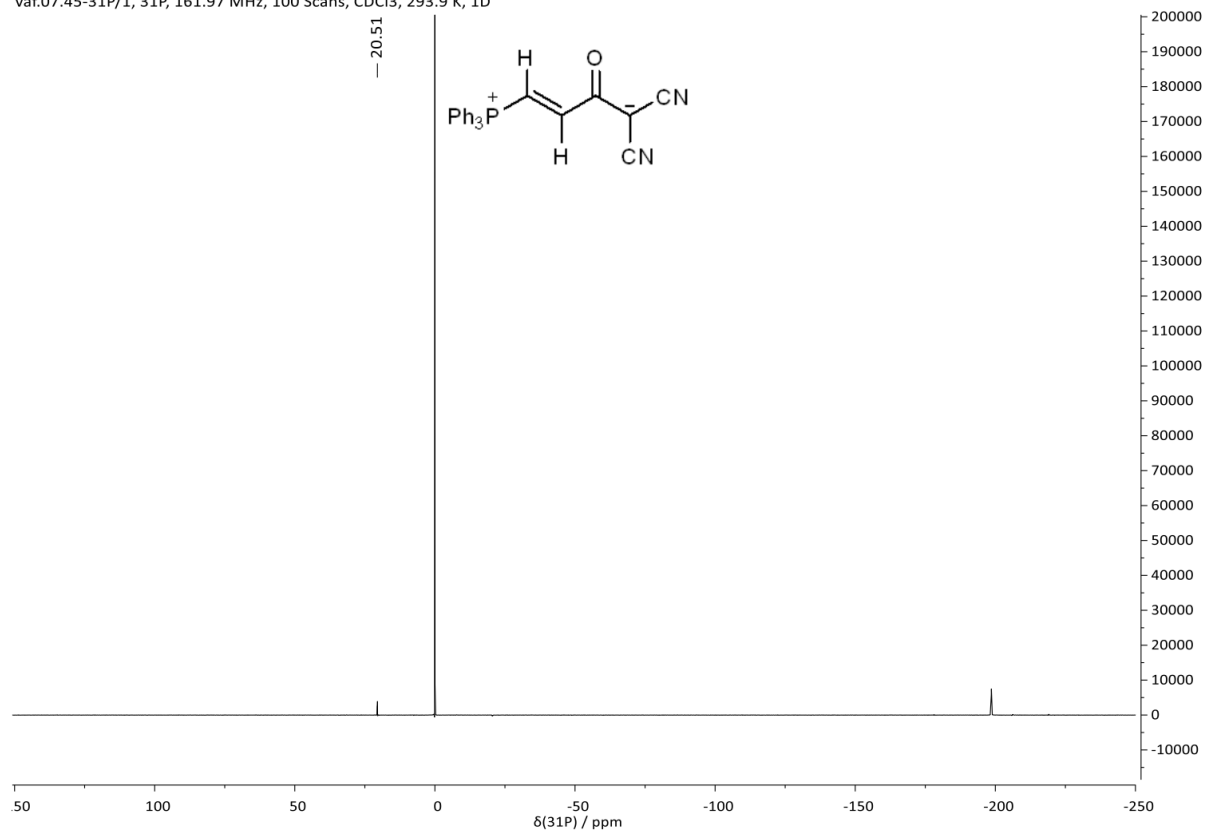


vaf.07.45-05/2, ¹³C, 125.77 MHz, CDCl₃, 300.0 K, 1D

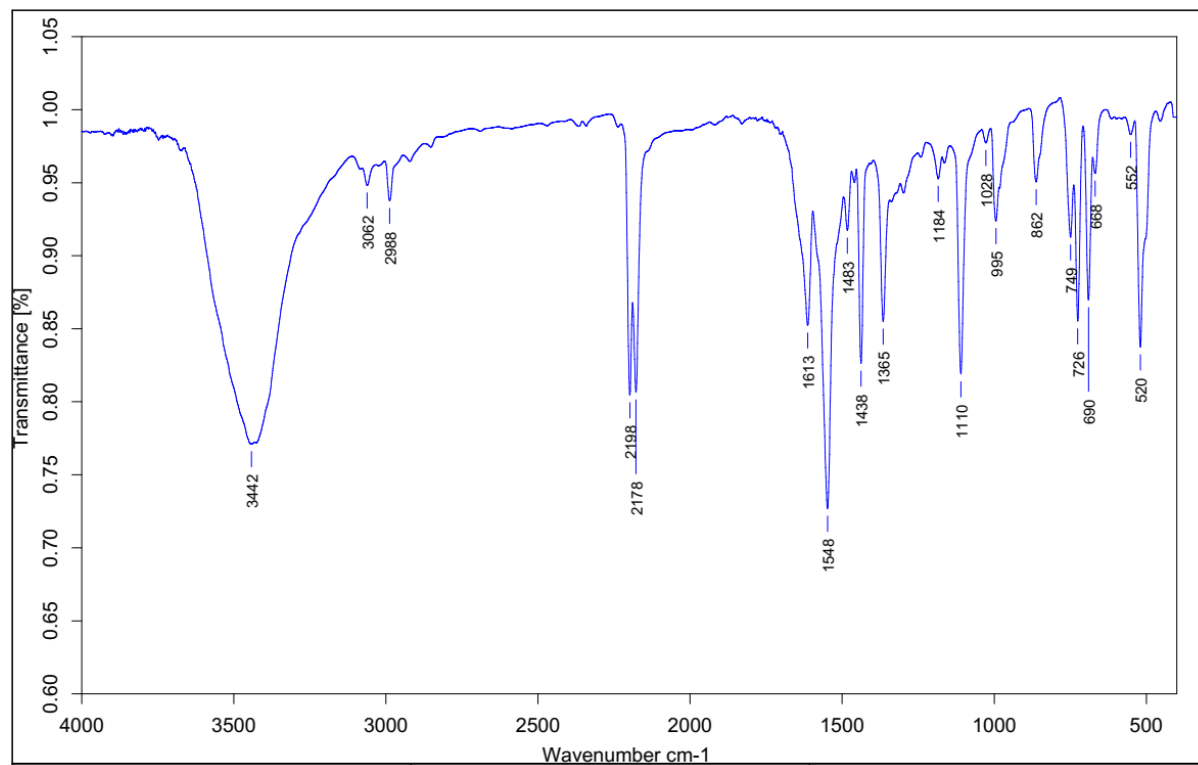


³¹P NMR

vaf.07.45-31P/1, 31P, 161.97 MHz, 100 Scans, CDCl₃, 293.9 K, 1D



IR



Probenname: vaf.07.45-1	Bereich Wellenzahlen: 4000.33 - 400.226	Datum: 11/05/2017
Probenform: KBr	Gerät: VECTOR22	Proben Scans: 8
Acquisition: Double Sided, Forward-Backward	Dateipfad: D:\DATEN_IR_BRUKER\FIORE	Datei: VAF.07.45-1