

## **Supporting Information**

for

Clustering and curation of electropherograms: an efficient method for analyzing large cohorts of capillary electrophoresis glycomic profiles for bioprocessing operations

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Beilstein J. Org. Chem. 2020, 16, 2087-2099. doi:10.3762/bjoc.16.176

Additional tables and figures

## **Supplementary information**

**Supplementary Table 1.** Statistics on the data used throughout the work.

	Technical replicate 1	Technical replicate 2	Technical replicate 3	Used for assessment
Biological replicates	12 days of: condition 1 condition 2 condition 3 condition 4 condition 5 condition 6 condition 7 condition 8 condition 9 condition 10 condition 11	12 days of: condition 1 condition 2 condition 3 condition 4 condition 5 condition 6 condition 7 condition 8 condition 9 condition 10 condition 11	12 days of: condition 1 condition 2 condition 3 condition 4 condition 5 condition 6 condition 7 condition 8 condition 9 condition 10 condition 11	(11 conditions x 12 days x 3 technical replicates) -5 sampling errors = 391 electropherograms

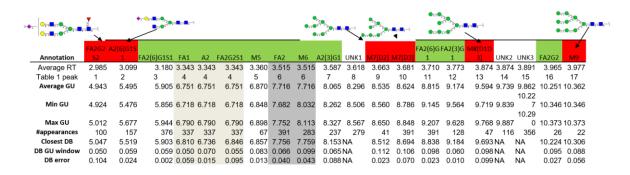
**Supplementary Table 2.** Glycans identified. SNFG is the Symbol nomenclature for graphical representations of glycans. Oxford are glycans represented using oxford nomenclature.

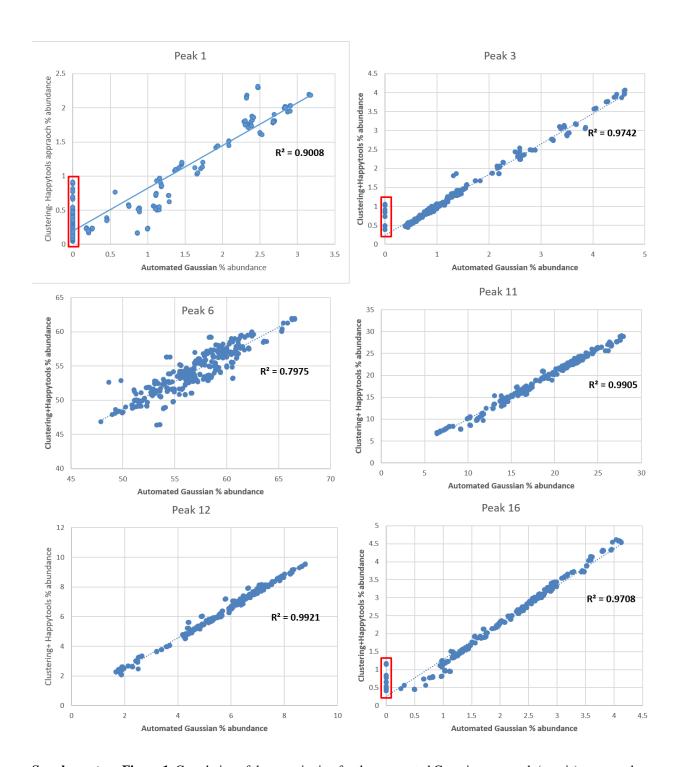
SNFG	Oxford	ATPS GU
	M3	4.925
	FA2G2S2	5.047
<b>→</b> • • • • • • • • • • • • • • • • • • •	A2[6]G1S1	5.519
	FA2(6)G1S1	5.903
	FA2(3)G1S1	6.081
<b>3 3 3 4 4 3 4 </b>	A2/FA1[6] FA2G2S1	6.736/6.81 0/6.846
	M5	6.857
	FA2/M6	7.756/7.75 9
	A2[3]G1	8.153
p + 1 p + 1	M7 D2	8.512
Q Q Q Q Q Q Q Q Q Q Q Q Q Q Q Q Q Q Q	M7 D2	8.694
β +	FA2[6]G1	8.838
p 2 0 0 p 4 1 p 1 p 1	FA2[3]G1	9.184
● - 2	M8 D1,D3	9.693
p 4 p 2 a p 4 p 4	FA2G2	10.224
	M9	10.306

**Supplementary Table 3.** The peak calibrated migration times (MT) and windows ( $\Delta$ RT) used to define peak boundaries in all 391 calibrated electropherograms. This table was input to HappyTools via its analysis input file. Using the peak definitions in this table, peak areas were integrated separately for the 6, 30 and 355 electropherograms for cluster 1, 2 and 3 respectively.

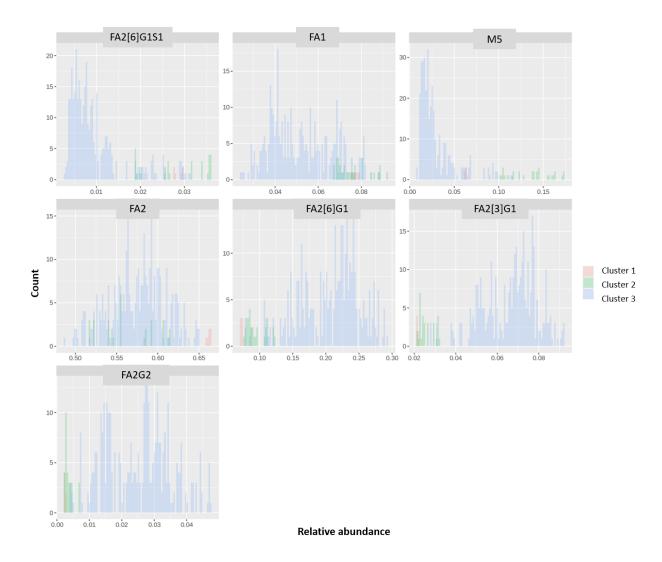
Cluster 1			Cluster 2		Cluster 3	
Peak Time Window		Time Window		Time Window		
1	2.9390	0.0230	3.0825	0.0145	2.9943	0.0205
	2.9390	0.0230	3.0623	0.0143		0.0203
2	3.0475	0.0165	3.1640	0.0110	3.1090	0.0180
3	3.1225	0.0185	3.2260	0.0170	3.1889	0.0330
4	3.2790	0.0160	3.3550	0.0120	3.3451	0.0150
5	3.3100	0.0130	3.3815	0.0145	3.3660	0.0140
6	3.4560	0.0330	3.5190	0.0290	3.5190	0.0300
7	3.5120	0.0160	3.5645	0.0145	3.5750	0.0150
8	3.5570	0.0160	3.6130	0.0130	3.6170	0.0190
9	3.5915	0.0115	3.6465	0.0075	3.6580	0.0100
10	3.6140	0.0110	3.6625	0.0095	3.6760	0.0110
11	3.6560	0.0320	3.6990	0.0180	3.7100	0.0200
12	3.7130	0.0200	3.7620	0.0190	3.7745	0.0185
13	3.7835	0.0125	3.8285	0.0125	3.8725	0.0135
14	3.8100	0.0130	3.8585	0.0165	3.8985	0.0145
15	3.8650	0.0160	3.9090	0.0140	3.9275	0.0145
16	3.8985	0.0135	3.9440	0.0120	3.9665	0.0195
17	3.9255	0.0125	3.9690	0.0130	4.1110	0.0170

Supplementary Table 4. Identified glycans and GU database matching statistics. Red columns were not identified in the UPLC-MS innovator (green columns identified in UPLC-MS). Average RT: raw migration time, before calibration with HappyTools, averaged over all occurrences ('#appearances' row). Supplementary Table 2 peak: the ordering of peaks and similar migration times allowed us to associate peaks in Table 1 with glycans. Average, min and max GU: the average, minimum and maximum of all of observed GU '#appearances' values. Closest DB: the closest GU value in the Sciex CE APTS database. DB GU window: tolerance of GU value matches (correspond to horizontal error bars in B). DB error: |Average GU - Closest DB|. Co-elutions are shaded the same color.

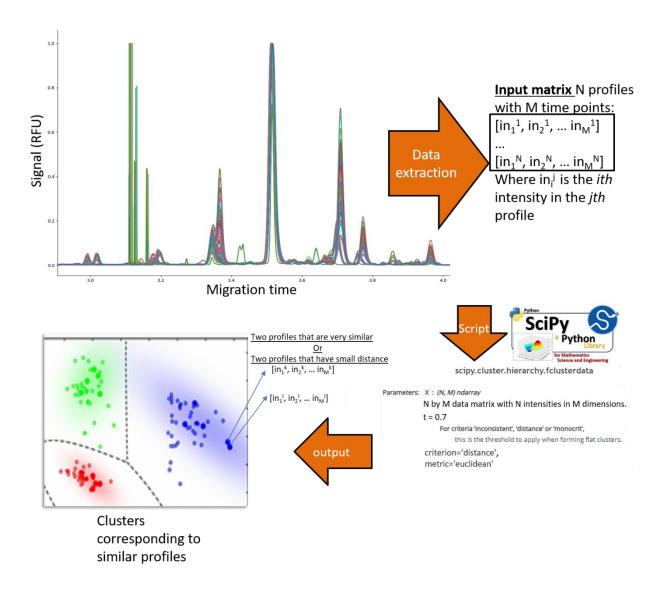




**Supplementary Figure 1.** Correlation of the quantitation for the automated Gaussian approach (x-axis) compared to our clustering+HappyTools quantitation (y-axis) for the same glycan peaks. Correlations only shown for the major peaks (at least one peak in the 391 electropherograms > 3%). The red rectangles show cases where the Gaussian automated approach missed peaks. The correlations for peaks 4 and 5 are shown in Figure 2 G, H and I.



**Supplementary Figure 2.** Distribution of glycan abundances in the three clusters for all glycans >1% average abundance. Cluster 3 had the largest number of observations. Cluster 1 had higher abundance of FA2[6]G1S1, FA1, M5 and lower abundance of FA2[6]G1, FA2[3]G1, and FA2G2. Cluster 2 had higher abundance of FA2[6]G1S1, FA1, FA2, M5 and lower abundance of FA2[6]G1, FA2[3]G1, and FA2G2.



**Supplementary Figure 3.** The clustering algorithm in more detail. Python packages were used for clustering using the hierarchical Euclidean distance function.