



## Supporting Information

for

### **Antiviral therapy in shrimp through plant virus VLP containing VP28 dsRNA against WSSV**

Santiago Ramos-Carreño, Ivone Giffard-Mena, Jose N. Zamudio-Ocadiz, Alfredo Nuñez-Rivera, Ricardo Valencia-Yañez, Jaime Ruiz-Garcia, Maria Teresa Viana and Ruben D. Cadena-Nava

*Beilstein J. Org. Chem.* **2021**, *17*, 1360–1373. [doi:10.3762/bjoc.17.95](https://doi.org/10.3762/bjoc.17.95)

### **Tables of detailed experimental assays and methods to prepare the pellet feed containing VLP-dsRNAvp28**

**Table ST1.** Partial sequence of the VP28 gene of the main structural protein of WSSV isolated from Mexico (GenBank: EU931451.1).

---

ATGGATCTTTCTTTCACTCTTTTCGGTCGTGTGCGCCATCCTCGCCATCACTGCTGTGATTGCTGTATTTA  
 TTGTGATTTTTAGGTATCACAACACTGTGACCAAGACCATCGAAACCCACACAGACAATATCGAGACAAA  
 CATGGATGAAAACCTCCGCATTCCTGTGACTGCTGAGGTTGGATCAGGCTACTTCAAGATGACTGATGTG  
 TCCTTTGACAGCGACACCTTGGGCAAATCAAGATCCGCAATGGAAAGTCTGATGCACAGATGAAGGAAG  
 AAGATGCGGATCTTGTCACTACTCCCGTGGAGGGCCGAGCACTCGAAGTACTGTGGGGCAGAATCTCAC  
 CTTTGAGGGAACATTCAAGGTGTGGAACAACACATCAAGAAAGATCAACATCACTGGTATGCAGATGGTG  
 CCAAAGATTAACCCATCAAAGGCCTTTGTGCGGTAGCTCCAACACCTCCTCCTTACCCCCGTCTCTATTG  
 ATGAGGATGAAGTTGGCACCTTTGTGTGTGGTACCACCTTTGGCGACCAATTGCAGCTACCGCCGGTGG  
 AAA

---

**Table ST2.** Design of the challenge to evaluate the efficacy and optimal dose of dsRNA<sub>vp28</sub>.

<b>Group<sup>1</sup></b>	<b>Treatment scheme</b>	<b>No. of shrimp<sup>2</sup></b>
WSSV-Positive	+WSSV infected shrimp	4 × 5
3.0 µg WSSV-Negative	-WSSV free shrimp 3.0 µg dsRNA <sub>vp28</sub> IM	4 × 5
0.5 µg dsRNA <sub>vp28</sub>	+0.5 µg dsRNA <sub>vp28</sub> IM	4 × 5
1.0 µg dsRNA <sub>vp28</sub>	+1.0 µg dsRNA <sub>vp28</sub> IM	4 × 5
2.0 µg dsRNA <sub>vp28</sub>	+2.0 µg dsRNA <sub>vp28</sub> IM	4 × 5
3.0 µg dsRNA <sub>vp28</sub>	+3.0 µg dsRNA <sub>vp28</sub> IM	4 × 5

<sup>1</sup>Indicates the abbreviature for each treatment

(+) WSSV-Positive shrimp with a dose of 10<sup>-6</sup>; (-) WSSV-Negative or non-infected shrimp. <sup>2</sup>Indicates the number of replicates × number of shrimps in each replicate.

## **Methods to prepare the pellet feed containing VLP-dsRNAvp28.**

**Method I.** Pellets of similar size (Natural Force, VIMIFOS, México) were coated with 0.5 µg of VLP-dsRNAvp28 each. VLP-dsRNAvp28 were applied coating the pellets with a thin layer of fish oil before drying at room temperature [1]. The feed was dried at 28 °C for 15 min and stored at 4 °C. The pellets were offered twice a day (2.0 µg/shrimp/day), 2 pellets at 09:00 h. and two at 16:00 h.) for three continuous days. After 48 hour feeding period, the shrimp were challenged with the WSSV.

**Method II.** The pellets were pulverized using a sterile porcelain mortar until a fine powder was obtained and mixed with 3% (w/v) NutriKelp® binder and fish oil in hot water (90 °C). After the blend was cooled to ca. 35 °C, the solution of VLP-dsRNAvp28 and 5% fish oil was added. The proportion recipe for the preparation of the pellets is shown in Table ST3. Afterward, the mixture was pelleted pressing through a 50 mL syringe without a needle (Terumo® 50 mL) to make long pellets (≈ 10–12 cm) and dried in an incubator (VWR Incubator F Air 2.3 CF, USA) at 28 °C for 40 min. The pellets were cut into pieces of ≈ 2 to 2.5 mm long. Feed prepared with VLPs was stored at 4 °C for subsequent doses. The pellets obtained (10 g) were weighed and divided by the number of organisms to be treated ( $n = 20$ ). The ration for each shrimp (0.5 g) was divided into six portions, to offer twice a day. The feed was administered at 2 µg of dsRNAi/shrimp daily, as described in the Method I section, giving a total of 6 µg of dsRNA/shrimp over a period of three days. In both cases the organisms were starved for 24 hours [2] before giving them the VLP-dsRNAvp28 treatment.

**Pellets with VLP-dsRNAvp28 coated with industrial grade fish oil.** Industrial grade fish oil was used to coat the pellets with VLPs following the Method I and II. The treatments were performed using shrimp ( $10.0 \text{ g} \pm 1.1 \text{ g}$ ) acclimated for 15 days at a salinity of 16 ppt. The experimental conditions are detailed in Table ST4 (Experiment 1).

**Pellet with VLP-dsRNAvp28 coated with salmon fish oil.** The salmon fish oil (Carlson Labs, Wild Norwegian Cod Liver Oil) was used to coat the pellets with VLPs following the Method I and II. Seven treatments were performed, each with four replicates with four shrimp ( $14.5 \text{ g} \pm 2.1 \text{ g}$ ) each ( $n = 16$ ) in 16 ppt saline water. The details are described in Table ST4, Experiment 2.

**Pellet with VLP-dsRNAvp28 prepared with commercial binders.**

**a). Dry Oil®.** The pellet feeds were prepared as described in Method I with slight changes. Dry Oil® (DO, Innova-Codemet SA de CV, Mexico) was used as a binder to coat pellets with VLPs, following the manufacturer's instructions. The pellets were directly submerged in a solution of Dry Oil in TN buffer at 30 °C containing the VLP-dsRNAvp28. The Table ST4 describe in details of this treatment, Experiment 3. Due to the characteristics of this product, it was only used to coat the pellets.

**b). NutriKelp®.** Pellets with VLPs using NutriKelp® (NK, NutriKelp Algas & Bioderivados Marinos, Mexico) as a binder. The procedure to make this food is described in “Method II.” Experimental details are in Table ST4 (Experiment 3). The Nutrikelp has an alginate base; it was only used to remake the pellet and not to coat them.

**Table ST3.** Recipe to prepare pellets mixed with VLPdsRNA $v_p28$  to feed shrimps.

Ingredient	Amount	To observe
Powdered pellet	8.0 g/ Xg <sup>a</sup>	The formation of fine grains
Natural grenetine hydrated		3% total in the mix
Fish oil	0.5 mL	
VLPdsRNA $v_p28$ * (0.12 $\mu\text{g}/\mu\text{L}$ )	1.0 mL	1X PBS <sup>b</sup>
Pellet mixed with VLPdsRNA $v_p28$	$\approx 10$ g	Make pellet

<sup>a</sup>The amount used may vary depending on the shrimp weight, to administrate twice a day, approximately 3% daily biomass for 3.3 g shrimp.

<sup>b</sup>Make pellet with 50 mL syringe without needle, and dry at 28–29 °C  $\times$  40 min, then cut in pieces 2 to 2.5 mm long.

**Table ST4.** Oral feeding challenges with VLPdsRNA $v_p28$  in *Penaeus vannamei*.

Experiment	Group*	Treatment scheme	No. of shrimp in group <sup>a</sup>
<b>Exp. 1.</b>	WSSV-Positive-E1	+WSSV infected shrimp	4 $\times$ 5
	WSSV-Negative-E1	-WSSV free shrimp	4 $\times$ 5
	VLP28-IM-E1	+Control VLPdsRNA $v_p28$ IM	4 $\times$ 5
	ApVLP28-mix-E1	+VLPdsRNA $v_p28$ /Mixpell	4 $\times$ 5
	ApVLP28-coat-E1	+VLPdsRNA $v_p28$ /coatpell	4 $\times$ 5
	VLP28-oral cav-E1	+VLPdsRNA $v_p28$ /oral cavity	4 $\times$ 5
	dsRNA28-200 $\mu\text{g}$ -IM	+Naked dsRNA $v_p28$ /200 $\mu\text{g}$ IM	2 $\times$ 6
	dsRNA28- 6 $\mu\text{g}$ IM-E1	+Naked dsRNA $v_p28$ -IM	4 $\times$ 5
<b>Exp. 2.</b>	WSSV-Positive-E2	+ WSSV infected shrimp	4 $\times$ 4
	WSSV-Negative-E2	-WSSV free shrimp	4 $\times$ 4

	VLP28-IM-E2	+Control VLPdsRNA $v_p28$ IM	4 × 4
	ApsVLP28-mix-E2	+VLPdsRNA $v_p28$ /Mixpell	4 × 4
	ApsVLP28-coat-E2	+VLPdsRNA $v_p28$ /coatpell	4 × 4
	VLP28-Oral cav-E2	+VLPdsRNA $v_p28$ /oral cavity	4 × 4
	dsRNA28-Oral cav-E2	+Naked dsRNA $v_p28$ /Oral cavity	4 × 4
	WSSV-PositiveE3	+WSSV infected shrimp	1 × 10
	WSSV-Negative-E3	-WSSV free shrimp	2 × 15
<b>Exp. 3.</b>	VLP28-IM-E3	+Control VLPdsRNA $v_p28$ IM	3 × 10
	NKVLP28-mix-E3	+VLPdsRNA $v_p28$ /Mixpell	3 × 10
	DOVLP28-coat-E3	+VLPdsRNA $v_p28$ /coatpell	3 × 10

\*=Treatment key. E = Experiment number.

"+"=WSSV-Positive (dose  $10^{-6}$ ); "-" WSSV-Negative, Mixpell = mixed pellet, coatpell = coated pellet.

Shrimps treated with VLPdsRNA $v_p28$  or naked dsRNA $v_p28$  received a dose of 6  $\mu$ g dsRNA, unless otherwise specified. <sup>a</sup>Indicates the number of replicates × number of shrimps in each replicate.

**Exp. 1.** Pellets with VLPdsRNA $v_p28$  prepared with fish oil (Ap) (industrial grade)

**Exp. 2.** Pellet with VLPdsRNA $v_p28$  coated with salmon fish oil (Aps)

**Exp. 3.** Pellet with VLPdsRNA $v_p28$  prepared with commercial binders (DO/NK= (Dry Oil<sup>®</sup> and NutriKelp<sup>®</sup>, respectively).

## References

1. Jha, R. K.; Xu, Z. R.; Bai, S. J.; Sun, J. Y.; Li, W. F.; Shen, J. *Fish Shellfish Immunol.* **2007**, *22*, 295–307. doi:10.1016/j.fsi.2006.04.006
2. Ning, J. F.; Zhu, W.; Xu, J. P.; Zheng, C. Y.; Meng, X. L. *Vaccine* **2009**, *27*, 1127–1135. doi:10.1016/j.vaccine.2008.11.075