



Supporting Information

for

A *myo*-inositol dehydrogenase involved in aminocyclitol biosynthesis of hygromycin A

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Table S1: Characterized enzymes from PF01408.

Gene name	Uniprot ID	Function	Biological role	Reference
Inositol metabolism				
lolG	P26935	<i>Myo</i> -inositol 2-dehydrogenase	<i>Myo</i> -inositol catabolism	[1]
	Q9WYP5	<i>Myo</i> -inositol 2-dehydrogenase	<i>Myo</i> -inositol catabolism	[2]
	O68965	<i>Myo</i> -inositol 2-dehydrogenase	<i>Myo</i> -inositol catabolism	[3]
	Q8NTY7	<i>Myo</i> -inositol 2-dehydrogenase	<i>Myo</i> -inositol catabolism	[4]
	P40332	<i>Scyllo</i> -inositol 2-dehydrogenase	<i>Scyllo</i> -inositol catabolism	[5]
	Q8NNS7	<i>Scyllo</i> -inositol 2-dehydrogenase	<i>Scyllo</i> -inositol catabolism	[4]
lolU	O05265	<i>Scyllo</i> -inositol 2-dehydrogenase	<i>Scyllo</i> -inositol catabolism	[6]
lolW	O32223	<i>Scyllo</i> -inositol 2-dehydrogenase	<i>Scyllo</i> -inositol catabolism	[6]
	Q8NTY4	<i>Scyllo</i> -inositol 2-dehydrogenase	<i>Scyllo</i> -inositol catabolism	[4]
Natural product biosynthesis				
KijD10	B3TMR8	dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose 3-reductase	L-Digitoxose biosynthesis	[7]
SpnN	Q9ALN5	dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose 3-reductase	Forosamine biosynthesis	[8]
OleW	Q9RR32	dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose 3-reductase	L-Oleandrose biosynthesis	[9]
IgdH	F0M433	Levoglucozan dehydrogenase	Levoglucozan biosynthesis	[10]
NtdC	O07564	Glucose-6-phosphate 3-dehydrogenase	Kanosamine biosynthesis	[11]
KanD2	Q6L737	Glucose-6-phosphate 3-dehydrogenase	Kanosamine biosynthesis	[12]
RifL	Q7BUE1	UDP-glucose dehydrogenase	Kanosamine biosynthesis	[13]
Irp3	W9BA38	Thiazolinyil imine reductase	Yersiniabactin biosynthesis	[14]
Carbohydrate metabolism				
NagA	A4Q8F7	α -N-Acetylgalactosaminidase	Blood group antigen degradation	[1-5][15]
	A4Q8G1	α -N-Acetylgalactosaminidase	Blood group antigen degradation	[15]
	Q8ECL7	α -N-Acetylgalactosaminidase	Blood group antigen degradation	[15]
AraA	Q53TZ2	L-arabinose 1-dehydrogenase	L-Arabinose degradation	[16]
Gal	P11886	Galactose 1-dehydrogenase	Galactose metabolism	[17]

ApsD	B1G894	Apiose dehydrogenase	D-Apiose catabolism	[18]
	B9JK80	Apiose dehydrogenase	D-Apiose catabolism	[18]
Xdh	Q8GAK6	D-Xylose 1-dehydrogenase	D-Xylose degradation	[19]
	Q5UY95	D-Xylose 1-dehydrogenase	D-Xylose catabolism	[24–26][20]
PLH35	T2KNC8	Oxidoreductase P35	Ulvan degradation	[21]
Gfo	A0A143PP41	Aldose-Aldose Oxidoreductase	Unknown	[22]
YjhC	P39353	2,7-anhydro-Neu5Ac dehydrogenase	Sialic acid metabolism	[23]
YcjS	P77503	Glucoside 3-dehydrogenase	Unknown carbohydrate catabolism	[24]
LPS biosynthesis				
WbpB	G3XD23	UDP-N-acetyl-2-amino-2-deoxy-D-glucuronate dehydrogenase	LPS biosynthesis.	[25–27]
WlbA	I6UWH1	UDP-N-acetyl-2-amino-2-deoxy-D-glucuronate dehydrogenase	LPS biosynthesis	[28]
GnnA	A0A543Q2K9	UDP-N-acetylglucosamine 3-dehydrogenase	LPS biosynthesis	[29]
Other functions				
PhtC	Q05184	Phthalate cis-4,5-dihydrodiol dehydrogenase	Cis-3,4-dihydrodiol phthalate catabolism	[30]
DgpA	A0A3Q9WWX8	C and O glycoside isomerase	Puerarin catabolism	[31]
LigC	Q9KWL3	4-carboxy-2-hydroxymuconate-6-semialdehyde dehydrogenase	Protocatechuate catabolism	[32]
Ddh	Q5L9Q6	<i>m</i> -Diaminopimelate dehydrogenase	L-Lysine biosynthesis	[33]

Table S2: Comparison of the hygromycin A biosynthetic cluster [34] and unknown hygromycin A-like cluster.

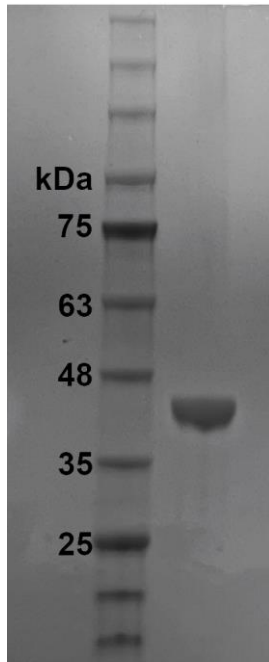
Protein	Annotated function	HygA biosynthetic role	% ID to unknown cluster
Hyg1	AfsR regulatory protein	Regulation	---
Hyg2	<i>p</i> -Hydroxybenzoate hydroxylase	Cinnamic acid	---
Hyg3	Regulatory protein	Regulation	---
Hyg4	Chorismate lyase	Cinnamic acid	---
Hyg5	Mannose dehydratase	Furanose	---
Hyg6	Methyltransferase	Aminocyclitol	56
Hyg7	Unknown	Aminocyclitol	69
Hyg8	Aminotransferase	Aminocyclitol	65
Hyg9	ACP	Cinnamic acid	31
Hyg10	b-ketoacyl synthase I	Cinnamic acid	50
Hyg11	Unknown	Unknown	---
Hyg12	CoA-ligase	Cinnamic acid	37
Hyg13	ACP	Cinnamic acid	
Hyg14	3-Hydroxyacyl ACP dehydratase	Cinnamic acid	34
Hyg15	3-Hydroxyacyl ACP reductase	Cinnamic acid	44
Hyg16	Glycosyltransferase	Furanose attachment	45
Hyg17	<i>Myo</i> -inositol dehydrogenase	Aminocyclitol	50
Hyg18	<i>Myo</i> -inositol-1-phosphate synthase	Aminocyclitol	---
Hyg19	Transmembrane protein	Unknown	---
Hyg20	Transglucosylase	Furanose	57
Hyg21	Phosphotransferase	Resistance	37
Hyg22	Acyltransferase	Cinnamic acid	---
Hyg23	Fucose synthase	Furanose	32
Hyg24	Unknown	Unknown	
Hyg25	<i>Myo</i> -inositol-1-phosphatase	Aminocyclitol	---
Hyg26	Short chain dehydrogenase	Furanose	---
Hyg27	DAHPS synthase	Cinnamic acid	---
Hyg28	ABC transporter	Unknown	---
Hyg29	Methyltransferase	Unknown	---

Table S3: Genomic neighborhood analysis of PF01408 sequences.

PFAM	Function	# Sequences with PFAM neighbors	# SSN clusters that contain PFAM
	NP biosynthetic enzymes		
PF00109	Ketoacyl synthase	584	45
PF00550	Acyl carrier protein (ACP) domain	340	71
PF00975	Thioesterase (TE) domain*	1,193	22
	Aminotransferases		
PF01041	DegT/DnrJ/EryCq/StrS aminotransferase	15,139	340
PF00202	Aminotransferase – class III	1,236	149
PF00155	Aminotransferase – class I and II	1,318	136
	Putative resistance proteins		
PF04655	Aminoglycoside antibiotic resistance kinase	22	4
PF02537	CrcB-like protein, Camphor Resistance	81	5
PF00903/ PF13660	Glyoxalase/Bleomycin resistance protein/Dioxygenase superfamily	1354	160
PF13536	Putative multidrug resistance efflux protein	2	2
PF05099	Tellurite resistance protein TerB	8	5

*TE domains only on their own. Some other TE domains are also found with ACP domains.

Figure S1: SDS-PAGE gel of purified Hyg17.



Expected MW
36 kDa

Figure S2: pH Activity profile for reactions with 10 mM *myo*-inositol, 10 mM NAD⁺, 1 μM Hyg17, 100 mM HEPEs, 100 mM Tris, 100 mM CHES, and 100 mM CAPS, 50 mM NaCl, pH 9.5–11.

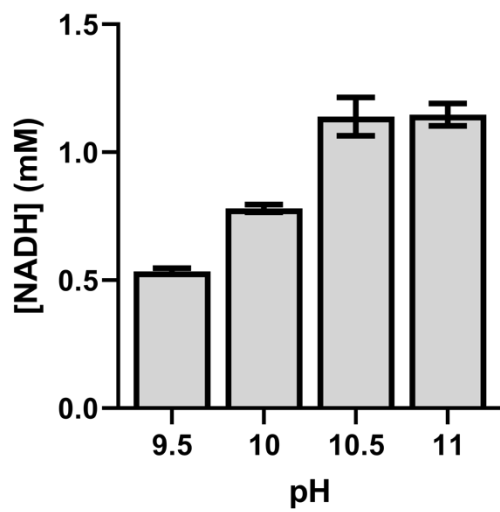
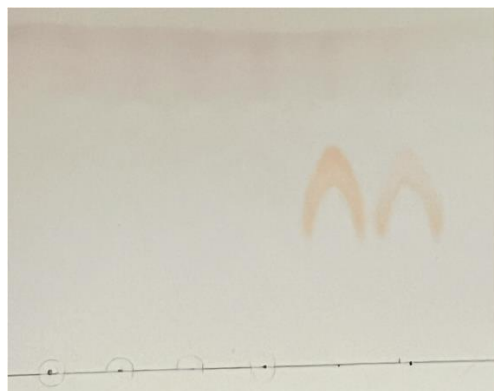


Figure S3: TLC analysis for Hyg17 and BslDH reactions with *myo*-inositol and NAD⁺. *p*-Anisaldehyde was used to stain ketones light pink.



-	+	-	+	+	+	NAD ⁺
-	-	+	+	+	+	<i>myo</i> -inositol
-	-	-	-	+	-	Hyg17
-	-	-	-	-	+	BslDH

References:

1. Ramaley, R.; Fujita, Y.; Freese, E. *J Biol Chem*, **1979**, *254*, 7684–7690
2. Rodionova, I. A.; Leyn, S. A.; Burkart, M. D.; Boucher, N.; Noll, K. M.; Osterman, A. L.; Rodionov, D. A. *Environ Microbiol* **2013**, *15*, 2254–2266.
3. Galbraith, M. P.; Feng, S. F.; Borneman, J.; Triplett, E. W.; De Bruijn, F. J.; Rossbachl, S. *Microbiology*, **1998**, *144*, 2915–2924.
4. Ramp, P.; Pflieger, C.; Dittrich, J.; Mack, C.; Gohlke, H.; Bott, M. *Microbiol Spectr* **2022**, *10*, e01950-22.
5. Morinaga, T.; Ashida, H.; Yoshida, K. *Microbiology*, **2010**, *156*, 1538–1546.
6. Kang, D.-M.; Tanaka, K.; Takenaka, S.; Ishikawa, S.; Yoshida, K. *Bioscience, Biotechnology, and Biochemistry*, **2017**, *81*, 1026–1032.
7. Kubiak, R. L.; Holden, H. M. *Biochemistry*, **2011**, *50*, 5905–5917.
8. Waldron, C.; Matsushima, P.; Rosteck, P. R.; Broughton, M. C.; Turner, J.; Madduri, K.; Crawford, K. P.; Merlo, D. J.; Baltz, R. H. *Chemistry & Biology*, **2001**, *8*, 487–499.
9. Aguirrezabalaga, I.; Olano, C.; Allende, N.; Rodriguez, L.; Braña, A. F.; Méndez, C.; Salas, J. A. *Antimicrob Agents Chemother*, **2000**, *44*, 1266–1275.
10. Sugiura, M.; Nakahara, M.; Yamada, C.; Arakawa, T.; Kitaoka, M.; Fushinobu, S. *Journal of Biological Chemistry*, **2018**, *293*, 17375–17386.
11. Vetter, N. D.; Langill, D. M.; Anjum, S.; Boisvert-Martel, J.; Jagdhane, R. C.; Omene, E.; Zheng, H.; Van Straaten, K. E.; Asiamah, I.; Krol, E. S.; Sanders, D. A. R.; Palmer, D. R. J. *J. Am. Chem. Soc.*, **2013**, *135*, 5970–5973.
12. Kudo, F.; Kitayama, Y.; Miyanaga, A.; Hirayama, A.; Eguchi, T. *Biochemistry*, **2020**, *59*, 1470–1473.
13. Yu, T.-W.; Müller, R.; Müller, M.; Zhang, X.; Draeger, G.; Kim, C.-G.; Leistner, E.; Floss, H. G. *Journal of Biological Chemistry*, **2001**, *276*, 12546–12555.
14. Meneely, K. M.; Lamb, A. L. *Biochemistry*, **2012**, *51*, 9002–9013.
15. Liu, Q. P.; Sulzenbacher, G.; Yuan, H.; Bennett, E. P.; Pietz, G.; Saunders, K.; Spence, J.; Nudelman, E.; Levery, S. B.; White, T.; Neveu, J. M.; Lane, W. S.; Bourne, Y.; Olsson, M. L.; Henrissat, B.; Clausen, H. *Nat Biotechnol*, **2007**, *25*, 454–464.
16. Watanabe, S.; Kodak, T.; Makino, K. *Journal of Biological Chemistry*, **2006**, *281*, 2612–2623.
17. Blachnitzky, E.; Wengenmayer, F.; Kurz, G. *European Journal of Biochemistry*, **1974**, *47*, 235–250.
18. Carter, M. S.; Zhang, X.; Huang, H.; Bouvier, J. T.; Francisco, B. S.; Vetting, M. W.; Al-Obaidi, N.; Bonanno, J. B.; Ghosh, A.; Zallot, R. G.; Andersen, H. M.; Almo, S. C.; Gerlt, J. A. *Nat Chem Biol*, **2018**, *14*, 696–705.
19. Mihasan, M.; Stefan, M.; Hritcu, L.; Artenie, V.; Brandsch, R. *Research in Microbiology*, **2013**, *164*, 22–30.
20. Johnsen, U.; Schönheit, P. *J Bacteriol*, **2004**, *186*, 6198–6207.
21. Baumgen, M.; Dutschei, T.; Bartosik, D.; Suster, C.; Reisky, L.; Gerlach, N.; Stanetty, C.; Mihovilovic, M. D.; Schweder, T.; Hehemann, J.-H.; Bornscheuer, U. T. *Journal of Biological Chemistry*, **2021**, 101210.

22. Taberman, H.; Andberg, M.; Koivula, A.; Hakulinen, N.; Penttilä, M.; Rouvinen, J.; Parkkinen, T. *Biochemical Journal*, **2015**, *472*, 297–307.
23. Horne, C. R.; Kind, L.; Davies, J. S.; Dobson, R. C. J. *Proteins* **2020**, *88*, 654–668.
24. Mukherjee, K.; Huddleston, J. P.; Narindoshvili, T.; Nemmara, V. V.; Raushel, F. M. *Biochemistry*, **2019**, *58*, 1388–1399.
25. Westman, E. L.; Preston, A.; Field, R. A.; Lam, J. S. *J Bacteriol*, **2008**, *190*, 6060–6069.
26. Westman, E. L.; McNally, D. J.; Charchoglyan, A.; Brewer, D.; Field, R. A.; Lam, J. S. *Journal of Biological Chemistry*, **2009**, *284*, 11854–11862.
27. Larkin, A.; Imperiali, B. *Biochemistry*, **2009**, *48*, 5446–5455. doi:10.1021/bi900186u
28. Thoden, J. B.; Holden, H. M. *Biochemistry*, **2010**, *49*, 7939–7948.
29. Manissorn, J.; Sitthiyotha, T.; Montalban, J. R. E.; Chunsrivirod, S.; Thongnuek, P.; Wangkanont, K. *ACS Chem. Biol.*, **2020**, *15*, 3235–3243.
30. Mahto, J. K.; Sharma, M.; Neetu, N.; Kayastha, A.; Aggarwal, S.; Kumar, P. *Archives of Biochemistry and Biophysics*, **2022**, *727*, 109314.
31. He, P.; Wang, S.; Li, S.; Liu, S.; Zhou, S.; Wang, J.; Tao, J.; Wang, D.; Wang, R.; Ma, W. *Acta Pharmaceutica Sinica B*, **2023**, *13*, 246–255.
32. Masai, E.; Momose, K.; Hara, H.; Nishikawa, S.; Katayama, Y.; Fukuda, M. *J Bacteriol*, **2000**, *182*, 6651–6658.
33. Hudson, A. O.; Klartag, A.; Gilvarg, C.; Dobson, R. C. J.; Marques, F. G.; Leustek, T. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics*, **2011**, *1814*, 1162–1168.
34. Palaniappan, N.; Ayers, S.; Gupta, S.; Habib, E.-S.; Reynolds, K. A. *Chemistry & Biology*, **2006**, *13*, 753–764.