Supporting Information

for

A two step synthesis of a key unit B precursor of

cryptophycins by asymmetric hydrogenation

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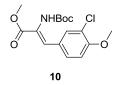
Full experimental procedures and detailed analytical data for the synthesis of 10 and 4 including chiral HPLC spectra.

General information

CH₂Cl₂ was dried over CaH₂. Dry methanol was obtained by carefully adding magnesium shavings and a few iodine crystals to an excess of methanol (Caution!) and subsequent heating to reflux. TLC plates were stained by dipping into ammonium heptamolybdate/cerium(IV) sulfate solution [10.0 g (NH₄)₆Mo₇O₂₄ · 4 H₂O, 0.4 g Ce(SO₄)₂, 5.4 mL conc. H₂SO₄, 180 mL H₂O], followed by subsequent heating. All NMR spectra were recorded at 298 K in CDCl₃. TMS was used for internal calibration (¹H NMR and ¹³C NMR: 0.00 ppm). IR spectra were obtained on an instrument containing an ATR accessory. The enantiomeric excess (*ee*) of the asymmetric hydrogenation reaction utilizing [(COD)Rh-(*R*,*R*)-Et-DuPhos]BF₄ and its enantiomer for comparison was determined by chiral HPLC (Chiralpak AD, hexane/2-propanol 9:1, 1 mL/min, 254 nm).

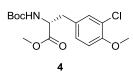
Experimental

(*Z*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonylamino)acrylate (10):



According to a slightly modified literature procedure [1] 3-chloro-4methoxybenzaldehyde (0.449 g, 2.63 mmol) and *rac*-Boc- α -phosphonoglycine trimethyl ester (**9**) (1.00 g, 3.36 mmol) were dissolved in dry CH₂Cl₂ (26 mL) at 0 °C. 1,1,3,3-Tetramethylguanidine (0.496 mL, 0.455 g, 3.95 mmol) was added over a 15 min period. After 4 h at 0 °C the solution was stirred overnight at rt. CH₂Cl₂ (100 mL) was added and the organic phase washed with 10% tartaric acid (40 mL), then brine (20 mL) and dried over solid MgSO₄. The solvent was removed under reduced pressure. Purification by column chromatography (hexane/EtOAc: 3:1) gave olefin **10** (0.761 g, 84%) as a colorless solid. R_f (hexane/EtOAc: 7:3) = 0.47; ¹H **NMR** (300 MHz, CDCl₃): δ [ppm] = 1.43 (s, 9H, NHCO₂C(CH₃)₃), 3.85 (s, 3H, CO₂CH₃), 3.92 (s, 3H, C_{ar}OCH₃), 6.24 (bs, 1H, NHBoc), 6.90 (d, *J* = 8.6 Hz, 1H, C⁵H), 7.22 (s, 1H, CO₂C=CH), 7.40 (dd, *J* = 1.5 Hz, 8.5 Hz, 1H, C⁶H) 7.64 (d, *J* = 2.1 Hz, 1H, C²H). ¹³C **NMR** (151 MHz, CDCl₃) δ [ppm] = 28.2 (NHCO₂C(CH₃)₃), 52.7 (CO₂CH₃), 56.2 (C_{ar}OCH₃), 81.1 (C(CH₃)₃), 111.5 (C⁵H), 122.5 (CO₂C=CH), 127.7 (C¹ and C^{'3}Cl), 129.1 (CO₂C=CH), 130.3 (C⁶H), 131.3 (C²H), 155.5 (NHCO₂C(CH₃)₃) and C^{4'}OCH₃), 166.1 (CO₂CH₃). **ESI-MS:** *m/z* 363.9 [M+Na]⁺. **HR-ESI:** calculated for C₁₆H₂₀N₁O₅ClNa [M+Na]⁺ 364.09222, found 364.09215. **IR** (neat, cm⁻¹): 3211, 3103, 1697, 1504, 804. **Elemental analysis:** calculated (%) for C₁₆H₂₀ClNO₅: C 56.32, H 5.90, N 4.10; found: C 56.31, H 6.04, N 4.07.

(*R*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonylamino)propanoate (4):

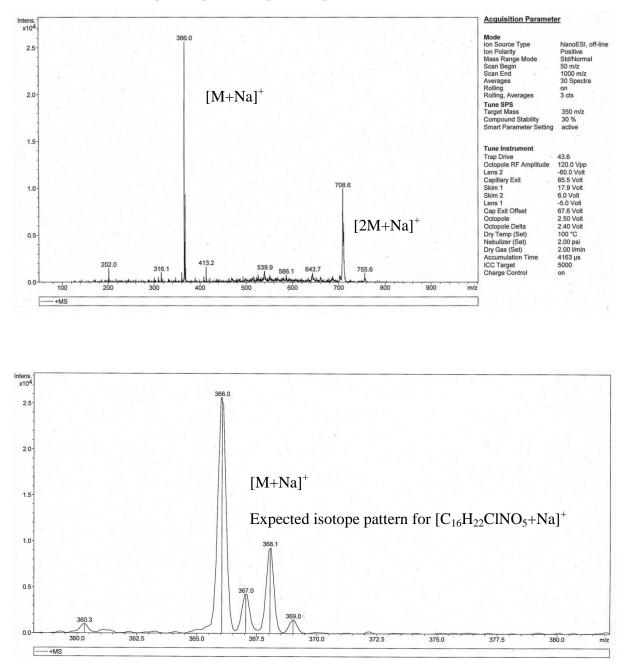


According to a slightly modified literature procedure [1] dry and degassed methanol (16 mL) was added to olefin **10** (0.410 g, 1.20 mmol) and [(COD)Rh-(R,R)-Et-DuPhos]BF₄ (0.015 g, 0.023 mmol, 1.9 mol %) in a hydrogenation flask. After three purging cycles the initial suspension was stirred under a hydrogen atmosphere (3–6 bar) for 21.5 h in a hydrogenation apparatus. The solvent was removed in vacuo and the catalyst separated from the already highly pure product by column chromatography (hexane/EtOAc: 3:1) to yield ester **4** (0.399 g, 97%, 98% *ee*) as a

colorless oil. R_f (hexane/EtOAc: 7:3) = 0.63; HPLC (Chiralpak AD, hexane/2-propanol 9:1, 1 mL/min, 254 nm): t_R = 9.8 min (4), 11.2 min (*ent-4*). $[\alpha]_D^{24} = -49$ (c = 0.78 in CHCl₃; Lit. $[\alpha]_D^{24} = -45$ [2]). ¹H NMR (300 MHz, CDCl₃): δ [ppm] = 1.43 (s, 9H, NHCO₂C(C<u>H</u>₃)₃), 2.96 (dd, J = 5.9, 14.1 Hz, 1H, C<u>H</u>^AH^B), 3.06 (dd, J = 5.6 Hz, 14.0 Hz, 1H, CH^A<u>H</u>^B), 3.73 (s, 3H, CO₂C<u>H</u>₃), 3.88 (s, 3H, C_{ar}OC<u>H</u>₃), 4.52 (m, 1H, C^a<u>H</u>), 4.99 (d, J = 7.5 Hz, 1H, N<u>H</u>), 6.85 (d, J = 8.4 Hz, 1H, C⁵<u>H</u>), 6.99 (dd, J = 2.1 Hz, 8.4 Hz, 1H, C⁶<u>H</u>), 7.13 (d, J = 1.6 Hz, 1H, C²<u>H</u>). ¹³C NMR (151 MHz, CDCl₃) δ [ppm] = 28.3 (NHCO₂C(<u>C</u>H₃)₃), 37.2 (<u>C</u>H^AH^B), 52.3 (CO₂<u>C</u>H₃), 54.4 (<u>C</u>^aH), 56.1 (C⁴O<u>C</u>H₃), 80.1 (<u>C</u>(CH₃)₃), 112.1 (<u>C</u>⁵<u>H</u>), 122.3 (<u>C</u>³Cl), 128.5 (<u>C</u>⁶<u>H</u>), 129.1 (<u>C</u>¹), 131.1 (<u>C</u>²<u>H</u>), 154.1 (<u>C</u>⁴OCH₃), 155.0 (NH<u>C</u>O₂C(CH₃)₃), 172.1 (<u>C</u>O₂CH₃).

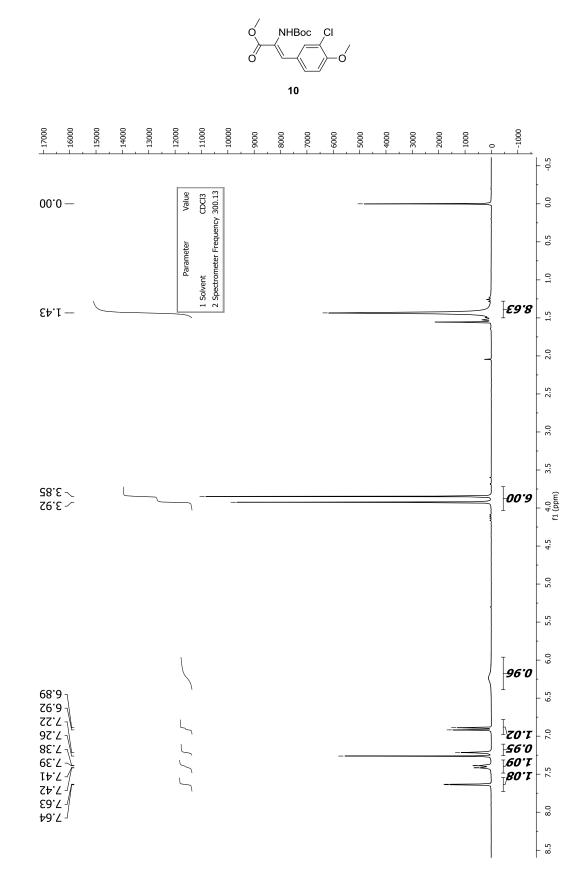
The ¹H and ¹³C NMR data are in complete agreement with previously published data, where **4** was synthesized by the chlorination and methylation of D-tyrosine [3]. However, the ¹³C NMR shift value of the aromatic carbon C³Cl (122.3 ppm), where the chloro substituent is attached to, disagrees with specifications from two other references: 135.6 ppm [2] and 132.4 ppm [4]. As we have obtained compound **4** according to a completely different route, we are confident that our data is correct.

ESI-MS: *m/z* 366.0 [M+Na]⁺; 708.6 [2M+Na]⁺.

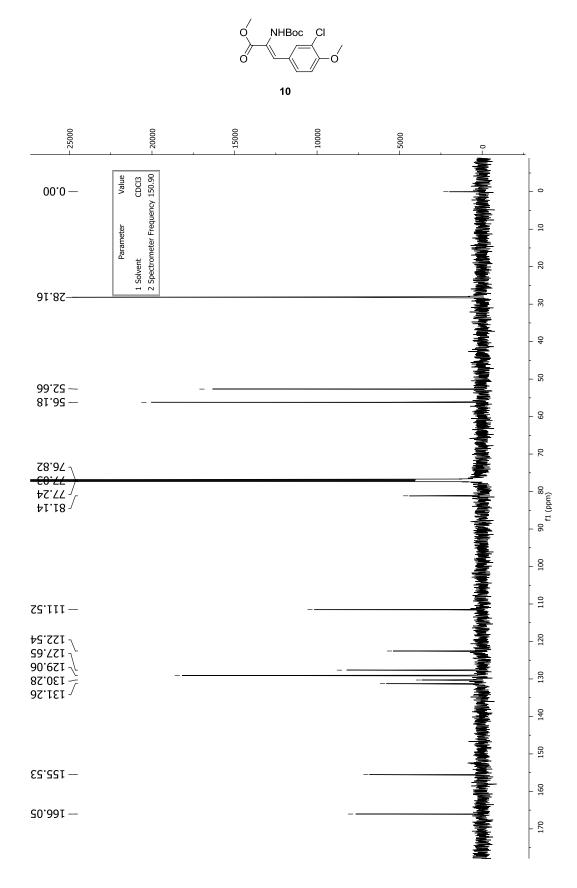


Elemental analysis: calculated (%) for C₁₆H₂₂CINO₅: C 55.90, H 6.45, N 4.07; found: C 55.74, H 6.31, N 4.00.

¹H NMR: (*Z*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonylamino) acrylate (10)

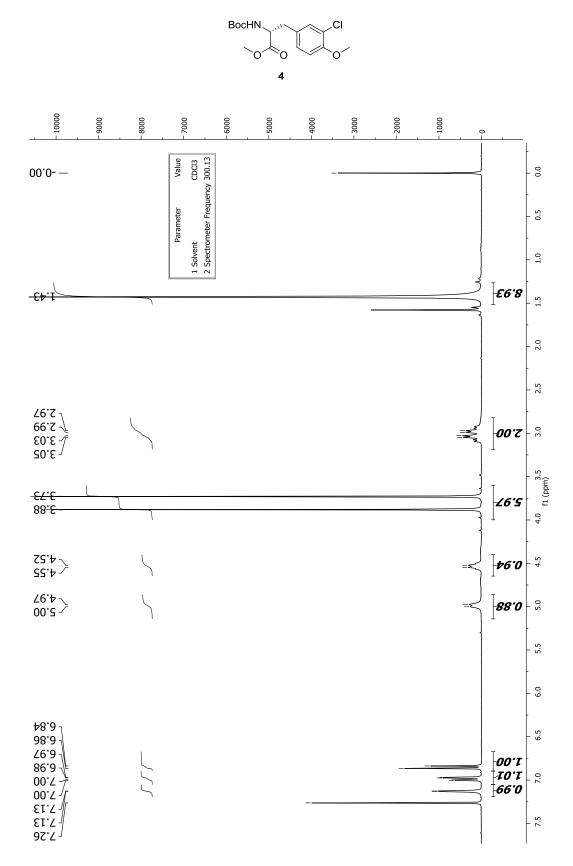


¹³C NMR: (*Z*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonylamino) acrylate (10)



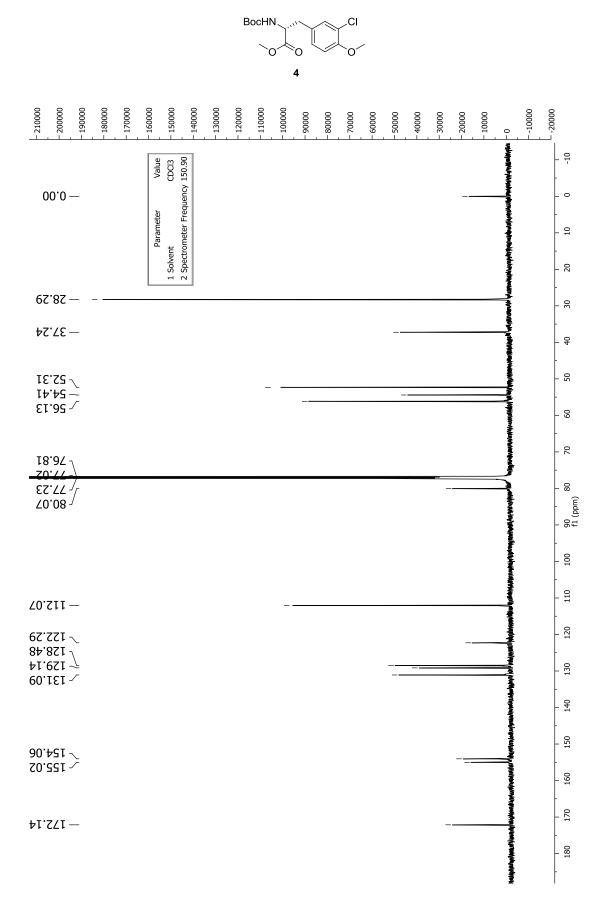
¹H NMR: (*R*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonyl-

amino)propanoate (4)



¹³C NMR: (*R*)-Methyl 3-(3-chloro-4-methoxyphenyl)-2-(*tert*-butoxycarbonyl-

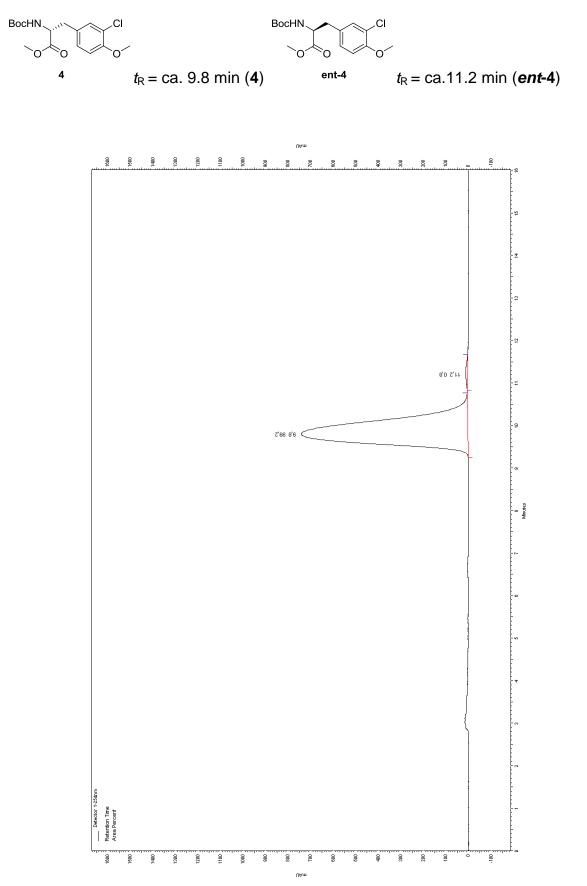
amino)propanoate (4)



S9

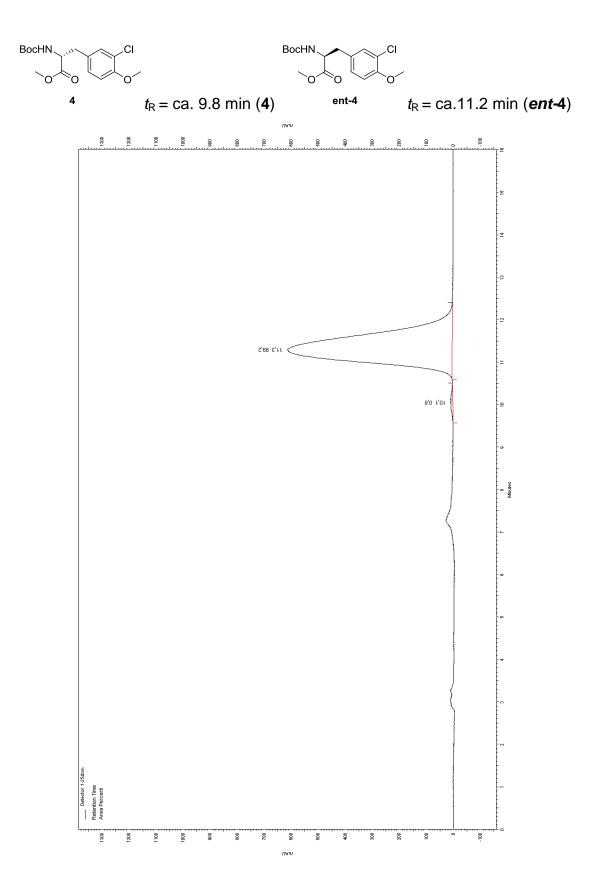
Chiral HPLC Run: Synthesis of (R)-methyl 3-(3-chloro-4-methoxyphenyl)-2-(tert-

butoxycarbonylamino)propanoate (4)

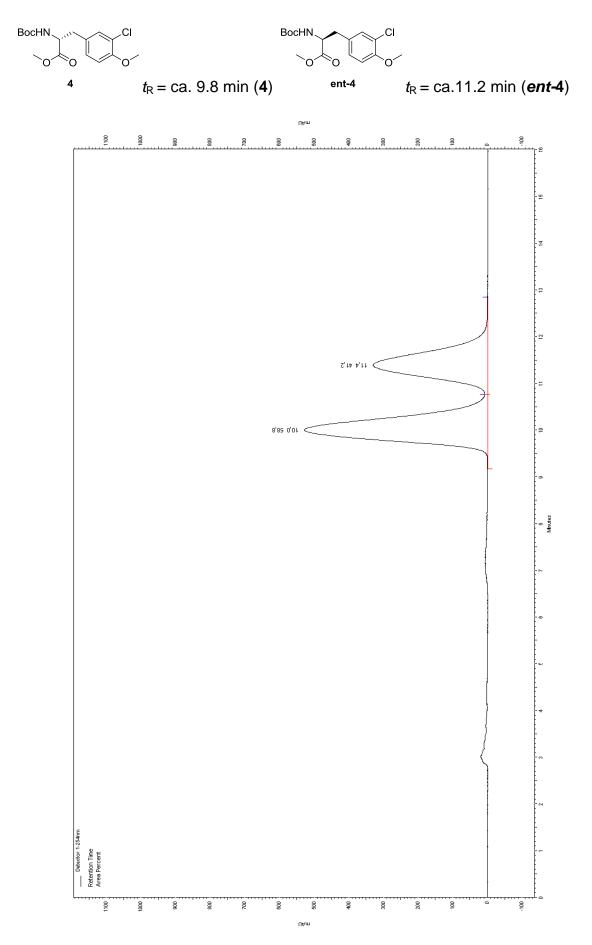


Chiral HPLC Run: Synthesis of (S)-methyl 3-(3-chloro-4-methoxyphenyl)-2-(tert-

butoxycarbonylamino)propanoate (ent-4)



Chiral HPLC Run: Mixture of 4 and ent-4



Literature

- [1] Bower, J. F.; Szeto, P.; Gallagher, T. *Chem. Comm.* **2005**, 5793-5795.
- Barrow, R. A.; Hemscheidt, T.; Liang, J.; Paik, S.; Moore, R. E.; Tius, M.
 A. J. Am. Chem. Soc. 1995, 117, 2479-2490
- [3] Nahrwold, M. β²-Aminosäuren als Bausteine funktionalisierter Cryptophycin-Analoga, Ph.D. Thesis, Bielefeld University, Germany,
 2009. http://bieson.ub.uni-bielefeld.de/volltexte/2010/1673/ (accessed Dec 19, 2010).
- [4] McCubbin, J. A.; Maddess, M. L.; Lautens, M. Org. Lett. 2006, 8, 2993-2996.