Supporting Information

for

Asymmetric desymmetrization of *meso*-diols by C_2 -symmetric chiral 4-pyrrolidinopyridines

Hartmut Schedel, Keizo Kan, Yoshihiro Ueda, Kenji Mishiro, Keisuke Yoshida, Takumi Furuta, and Takeo Kawabata*

Address: Institute for Chemical Research, Kyoto University, Uji, Kyoto 611-0011, Japan Email: Takeo Kawabata - kawabata@scl.kyoto-u.ac.jp

* Corresponding author

Experimental details and characterization data of new compounds, copies of 1H NMR and ^{13}C NMR

Content:

S2
S2
S3
S3
S3
S4
S4
S5
S5
S6
S6
S7

(2S,5S)-2,5-Bis[N-methyl-N-hexylamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (15)	S7
(2S,5S)-Dihexyl 1-(pyridin-4-yl)pyrrolidine-2,5-dicarboxylate (16)	S8
4-Pyridinyl-L-proline 1'-hexyloxyamide (17)	S8
Typical procedure for asymmetric desymmetrization	S9
1,3-Cyclohexanediol isobutyrate (20)	S9
Acylative kinetic resolution of racemic-6 with catalyst 12b	S9
Determination of the absolute configuration of 2,3-butanediol isobutyrate (23a)	S9
2,3-Butanediol diisobutyrate (24a)	S10
Conformation search of cat 11 and meso-1,2-cyclohexanediol	S11
References	S11
¹ H and ¹³ C NMR spectra	S12

Some of the results shown are already mentioned in the patent, JP2005132746.

General

NMR spectra were obtained with a Varian Gemini 200 (¹H NMR: 200 MHz, ¹³C NMR: 50 MHz) or JEOL JMN 400 spectrometer (¹H NMR: 400 MHz, ¹³C NMR: 100 MHz), chemical shifts being given in ppm units (tetramethylsilane or the solvent residual signal for CD₃OD as internal standard, indicating 0 and 3.30, respectively). IR spectra were recorded with a JASCO FT/IR-300 spectrometer. Specific rotation was measured with a Horiba SEPA-200 automatic digital polarimeter. MS spectra were recorded with a JEOL JMS-DX300 mass spectrometer or JEOL JMS-700 mass spectrometer. TLC analysis and preparative TLC were performed on commercial glass plates bearing a 0.25 mm layer and 0.5 mm layer of Merck Kiesel-gel 60 F₂₅₄, respectively. Silica gel chromatography was carried with Wakogel C-200, Fuji Silysia BW-1277H, or Nacalai Tesque Silica gel 60 (150–325 mesh). Dry solvents (THF, ether, hexane, dichloromethane, and toluene; <50 ppm water contents) were purchased from Kanto Chemical CO., Inc. and used without further treatment.

List of abbreviations

AcOEt ethyl acetate

DCM dichloromethane

DMAP 4-dimethylaminopyridine

DMF dimethylformamide

EDCI 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride

HOBt 1-hydroxybenzotriazole NMM *N*-methylmorpholine

THF tetrahydrofuran

General procedure for the preparation of C_2 -symmetric catalysts

To a solution of (2S,5S)-1-(pyridin-4-yl)pyrrolidine-2,5-dicarboxylic acid [1] (130 mg, 0.55 mmol) and the corresponding amino acid or amino alcohol (1.6 mmol, 2.9 equiv) in DMF or DCM (10 mL) were added NMM (148 μ l, 1.6 mmol, 2.9 equiv), EDCI (312 mg, 1.6 mmol, 2.9 equiv) and HOBt (222 mg, 1,6 mmol, 2.9 equiv). After stirring for 24 h, the mixture was evaporated and diluted with EtOAc (200 mL). The organic layer was washed with sat. aq. NaHCO₃, brine, dried over Na₂SO₄, filtered and evaporated in vacuo. The residue was purified by SiO₂ column chromatography or preparative TLC on silica gel, or recrystallization to give the corresponding C_2 -symmetric catalyst.

Preparation of (2S,5S)-1-(pyridin-4-yl)pyrrolidine-2,5-dicarboxylic acid was previously reported [1].

(2S,5S)-2,5-Bis[(2S)-3-(1H-indol-3-yl)-1-(methyloxy)-1-oxopropan-2-ylamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (8a)

According to the general procedure, 8a was prepared with (S)-tryptophane methyl ester hydrochloride in DMF and purified by SiO_2 column chromatography (AcOEt/MeOH = 5/1) as a white powder in 46% yield.

M.p. 223–225 °C. [α]_D²⁰ = –26 (c 0.5, DMSO). ¹H NMR (200 MHz, DMSO) δ 1.65–1.85 (m, 2H), 2.20 (br s, 2H), 3.05 (dd, J = 10.6, 14.6 Hz, 1H), 3.24 (dd, J = 4.2, 14.6 Hz, 1H), 4.22 (d, J = 7.8 Hz, 1H), 4.20–4.60 (m, 1H), 5.78 (d, J = 6.0 Hz, 1H), 6.90-7.16 (m, 2H), 7.21 (s, 1H), 7.38 (d, J = 8.0 Hz, H), 7.55 (d, J = 7.2 Hz, H), 7.74 (d, J = 5.0 Hz, 1H), 8.68 (d, J = 8.4 Hz, 1H). ¹³C NMR (50 MHz, DMSO) δ 24.3, 26.6, 49.6, 50.4, 58.6, 104.9, 107.3, 109.0, 115.5, 116.0, 118.5, 121.3, 124.5, 133.7, 146.2, 147.5, 169.4, 169.7. IR (KBr) 3255, 1755, 1735, 1655, 1600 cm⁻¹. MS (FAB) m/z (rel intensity) 659 (MNa⁺), 637 (MH⁺). HRMS (FAB) calcd for $C_{35}H_{37}N_6O_6$ (MH): 637.2775, found, 637.2764. Anal calcd for $C_{35}H_{36}N_6O_6$: C, 66.03; H, 5.70; N, 13.20. Found C, 65.91; H, 5.79; N, 13.00.

(2S,5S)-2,5-Bis[(2R)-3-(1H-indol-3-yl)-1-(methyloxy)-1-oxopropan-2-ylamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (8b)

According to the general procedure, **8b** was prepared with (R)-tryptophane methyl ester hydrochloride in DMF and purified by SiO₂ column chromatography (AcOEt/MeOH = 5/1)

in 34% yield as a crystalline powder.

M.p. 176–178 °C. [α]_D²⁰ = –58 (c 0.5, DMSO). ¹H NMR (200 MHz, CD₃OD) δ 1.40–1.70 (m, 2H), 2.03 (br s, 2H), 3.12 (dd, J = 9.4, 14.6 Hz, 2H), 3.20–3.40 (m, 1H), 3.70 (s, 6H), 4.29 (d, J = 7.8 Hz, 2H), 4.75 (dd, J = 4.8, 9.2, 2H), 6.28 (d, J = 6.8 Hz, 2H), 6.80-7.20 (m, 8H), 7.33 (d, J = 8.0 Hz, 2H), 7.45 (d, J = 7.4 Hz, 2H), 7.93 (d, J = 6.8 Hz, 2H). ¹³C NMR (50 MHz, CD₃OD) δ 26.2, 28.4, 51.0, 52.5, 61.6, 107.9, 108.7, 110.5, 117.1, 118.0, 120.6, 122.4, 126.6, 136.1, 144.9, 151.7, 171.7, 172.0. IR (KBr) 3380, 3260, 1740, 1665, 1600 cm⁻¹. MS (FAB) m/z (rel intensity) 637 (MH⁺). HRMS (FAB) calcd for C₃₅H₃₇O₆N₄ (MH): 637.2775, found, 637.2748.

(2S,5S)-2,5-Bis[tryptamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (9)

According to the general procedure, **9** was prepared with tryptamin hydrochloride in DMF and purified by SiO_2 column chromatography (AcOEt/MeOH = 5/1) in 40% yield as a crystalline powder.

M.p. 140–145 °C. $[\alpha]_D^{20} = -36$ (c 0.5 MeOH). ¹H NMR (200 MHz, CD₃OD) δ 1.70–1.90 (m, 2H), 2.10–2.40 (m, 2H), 2.91 (t, J = 7.6 Hz, 4H), 3.49 (dd, J = 7.0 Hz, 4H), 4.31 (d, J = 7.6 Hz, 2H), 6.16 (d, J = 6.6 Hz, 2H), 6.90–7.20 (m, 6H), 7.30–7.40 (m, 2H), 7.42–7.60 (m, 2H), 7.85 (d, 6.6, 2H). ¹³C NMR (50 MHz, CD₃OD) δ 24.1, 28.7, 39.2, 61.8, 107.6, 110.4, 111.0, 117.4, 117.8, 120.5, 121.7, 126.9, 136.2, 147.3, 150.7, 172.4. IR (KBr) 3240, 1660, 1600, 1518 cm⁻¹. MS (FAB) m/z (rel intensity) 521 (MH⁺). HRMS (FAB) calcd for C₃₁H₃₂O₂N₆ (MH): 521.2665, found, 521.2657.

(2S,5S)-2,5-Bis[(2S)-3-(1-(methyloxy)-1-oxoethan-2-ylamino)carbonyl]-1-(pyridin-4-yl) pyrrolidine (10)

According to the general procedure, **10** was prepared with glycine methyl ester hydrochloride in 16% yield as a white powder.

M.p. 222–225 °C. $[\alpha]_D^{20} = -76$ (c 0.25, CH₃OH). ¹H NMR (200 MHz, CD₃OD) δ 2.10–2.30 (m, 2H), 2.42–2.33 (m, 2H), 3.73 (s, 6H), 3.96 (d, J = 6.2 Hz, 4H), 4.59 (d, J = 8.2 Hz, 2H), 6.56 (d, J = 6.4 Hz, 2H), 8.14 (d, J = 6.4 Hz, 2H). ¹³C NMR (50 MHz, CD₃OD) δ 28.7, 39.9, 50.7, 61.7, 108.1, 147.2, 150.8, 169.5, 173.4. IR (KBr) 3245, 1742, 1642, 1600, 1542, 1519 cm⁻¹. MS (FAB) m/z (rel intensity) 401 (MNa⁺, 12), 379 (MH⁺, 100), 262 (30). HRMS (FAB) calcd for C₁₇H₂₃O₆N₄ (MH): 379.1618, found, 379.1598.

(2S,5S)-2,5-Bis[hexylamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (11)

According to the general procedure, **11** was prepared with *n*-hexylamine in DCM, purified by repeated preparative TLC (SiO₂, EtOAc/MeOH = 3:1 and CHCl₃/MeOH = 5:1) and recrystallization from EtOAc-CH₂Cl₂ to afford **11** in 54% yield as a colorless powder.

M.p. 222–223 °C. [α]_D²⁰ = -86 (c 0.5, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ 0.85 (t, J = 6.8 Hz, 6H), 1.10–1.35 (m, 12H), 1.35–1.50 (m, 4H), 2.10–2.30 (m, 2H), 2.30–2.60 (m, 2H), 3.10–3.40 (m, 4H), 4.42 (d, J = 7.6 Hz, 2H), 6.30–6.60 (m, 4H), 8.13 (dd, J = 1.4, 5.0 Hz, 2H). ¹³C NMR (50 MHz, CDCl₃) δ 14.1, 22.7, 26.7, 29.6, 30.1, 31.6, 39.7, 62.4, 108.3, 149.0, 150.5, 172.3. IR (KBr) 3288, 2928, 1651, 1596, 1552, 1510 cm⁻¹. MS (FAB) m/z (rel intensity) 402 (M, 3), 274 (100), 145 (60). HRMS (FAB) calcd for $C_{23}H_{38}O_2N_4$ (M⁺) 402.2995, found, 402.2974. Anal calcd for $C_{23}H_{38}N_2O_4$: C, 68.62; H, 9.51; N, 13.92. Found C, 68.35; H, 9.40; N, 13.93.

(2S,5S)-2,5-Bis[(2S)-3-(2-naphthylmethyl)-1-(methyloxy)-1-oxopropan-2-ylamino]carb onyl]-1-(pyridin-4-yl)pyrrolidine (12a)

According to the general procedure, **12a** was prepared with (*S*)-naphtylalanine methyl ester hydrochloride in DMF in 41% yield as a white powder.

M.p. 225–228 °C. $[\alpha]_D^{20} = +39$ (c 0.25, MeOH). ¹H NMR (200 MHz, DMSO) δ 1.50–1.65 (m, 2H), 2.09 (br s, 2H), 3.05 (t, J = 13.0 Hz, 2H), 3.20–3.50 (m, 2H), 3,63 (s, 6H), 4.10 (d, J = 7.2 Hz, 2H), 4.50–4.65 (m, 2H), 5.53 (d, J = 6.2 Hz, 2H), 7.10 (d, J = 5.8 Hz, 2H), 7.38-7.60 (m, 6H), 7.70–8.00 (m, 8H), 8.66 (d, J = 8.4 Hz, 2H). ¹³C NMR (50 MHz, DMSO) δ 29.6, 37.1, 52.9, 53.7, 61.9, 108.0, 126.34, 126.8, 128.1, 128.3, 128.4, 132.6, 133.6, 135.9, 149.2, 150.4, 172.4, 172.5. IR (KBr) 3255, 1750, 1650, 1600 cm⁻¹. MS (FAB) m/z (rel intensity) 659 (MH⁺). HRMS (FAB) calcd for $C_{39}H_{39}O_6N_4$ (MH): 659.2869, found, 659.2859.

(2S,5S)-2,5-Bis[(2R)-3-(2-naphthylmethyl)-1-(methyloxy)-1-oxopropan-2-ylamino]carb onyl]-1-(pyridin-4-yl)pyrrolidine (12b)

According to the general procedure, 12b was prepared with (R)-naphtylalanine methyl ester hydrochloride in DMF and purified by SiO_2 column chromatography (SiO_2 , AcOEt /MeOH = 4/1) in 53% yield as an amorphous powder.

M.p. 191–193 °C. $[\alpha]_D^{20} = -90$ (c 0.5, MeOH). ¹H NMR (200 MHz, CD₃OD) δ 1.20–1.50 (m, 2 H), 1.87 (br s, 2 H), 3.05 (dd, J = 10.8, 13.9 Hz, 2 H), 3.41 (dd, J = 4.4, 13.9 Hz, 2H), 3.73 (s, 6H), 4.26 (d, J = 7.6 Hz, 2H), 4.83 (dd, J = 4.6, 10.8 Hz, 2H), 6.18 (d, J = 6.6 Hz, 2H), 7.25 (dd, J = 1.6, 8.4 Hz, 2H), 7.40–7.60 (m, 6H), 7.60–7.90 (m, 8H). ¹³C NMR (100 MHz, CD₃OD) δ 28.4, 36.2, 51.1, 52.6, 61.4, 107.7, 124.9, 125.3, 126.2, 126.6, 126.7, 126.8, 127.2, 131.89, 132.9, 133.8, 146.7, 150.7, 171.3, 172.4. IR (KBr) 3280, 1740, 1657, 1600 cm⁻¹. MS m/z (FAB) (rel intensity) 659 (MH⁺). HRMS (FAB) calcd for C₃₉H₃₉O₆N₄ (MH): 659.2869, found, 659.2859.

(2S,5S)-2,5-Bis[(2S)-3-(isopropyl)-1-(methyloxy)-1-oxopropan-2-ylamino]-carbonyl]-1-(pyridin-4-yl)pyrrolidine (13)

According to the general procedure, 13 was prepared with (S)-valine methyl ester hydrochloride in CH_2Cl_2 with preparative TLC (SiO₂, AcOEt /MeOH = 3/1) in 29% yield as a white powder.

M.p. 203–205 °C. $[\alpha]_D^{21} = -117$ (c 0.5, MeOH). ¹H NMR (200 MHz, CD₃OD) δ 0.99–1.10 (m, 12H), 2.07–2.35 (m, 4H), 2.40–2.60 (m, 2H), 3.73 (s, 6H), 4.37 (d, J = 5.6 Hz, 2H), 4.66 (d, J = 7.6 Hz, 2H), 6.47 (d, J = 6.4 Hz, 2H), 8.09 (d, J = 6.4 Hz, 2H). ¹³C NMR (50 MHz, CD₃OD) δ 16.7, 17.8, 28.7, 29.7, 50.6, 57.4, 61.2, 107.5, 147.3, 151.0, 171.3, 173.1. IR (KBr) 3255, 2880, 1740, 1655, 1600 cm⁻¹. MS (FAB) m/z (rel intensity) 462 (M, 7), 304 (55), 71 (100). HRMS (FAB) Calcd for C₂₃H₃₄O₆N₄ (M): 462.2478, found, 462.2490.

(2S,5S)-2,5-Bis[(2S)-3-(isobutyl)-1-(methyloxy)-1-oxopropan-2-ylamino]-carbonyl]-1-(pyridin-4-yl)pyrrolidine (14)

According to the general procedure, **14** was prepared with (*S*)-leucine methyl ester hydrochloride in CH_2Cl_2 with SiO_2 column chromatography ($CHCl_3/MeOH = 6/1$) and successive preparative TLC (SiO_2 , $CHCl_3/MeOH = 6/1$) in 71% yield as a white powder. M.p. 187–189 °C. [α]_D²⁰ = -133 (c 0.5, CH_3OH). ¹H NMR (200 MHz, CD_3OD) δ 0.93 (d, J = 5.8 Hz, 6H), 1.02 (d, J = 6.2 Hz, 6H), 1.50–1.90 (m, 4H), 2.10–2.30 (m, 2H), 2.35–2.60 (m, 2H), 3.70 (s, 6H), 4.20–4.60 (m, 4H), 6.46 (d, J = 5.5 Hz, 2H), 8.10 (d, J = 5.5 Hz, 2H). ¹³C NMR (50 MHz, CD_3OD) δ 19.6, 21.6, 24.4, 28.6, 39.1, 50.2, 50.8, 61.4, 107.7, 147.1, 151.0, 172.2, 172.93. IR (KBr) 2880, 1745, 1660, 1600, 1545, 1518 cm⁻¹. MS (FAB) m/z (rel intensity) 490 (M, 4), 318 (100), 145 (40). HRMS (FAB) calcd for $C_{25}H_{38}O_6N_4$ (M): 490.2791, found, 490.2796.

(2S,5S)-2,5-Bis[N-methyl-N-hexylamino]carbonyl]-1-(pyridin-4-yl)pyrrolidine (15)

According to the general procedure, **15** was prepared with *N*-methyl hexylamine in CH_2Cl_2 without addition of NMM and purified by preparative TLC (SiO_2 , $CHCl_3/MeOH = 5/1$) in 42% yield as a white solid.

Decomposition. [α]_D²⁰ = -28 (c 0.5, CHCl₃). ¹H NMR (400 MHz, DMSO, 150 °C) δ 0.90 (t, J = 6.8 Hz, 6H), 1.20–1.40 (m, 12H), 1.45–1.65 (m, 4H), 1.80–1.95 (m, 2H), 2.40–3.15 (m, 2H), 3.04 (m, 6H), 3.38 (t, J = 6.8 Hz, 4H), 4.81 (d, J = 7.8 Hz, 2H), 6.12 (d, J = 6.2 Hz, 2H), 8.02 (d, J = 6.2 Hz, 2H). ¹³C NMR (50 MHz, CDCl₃) δ 14.1, 22.7, 26.6, 26.8, 27.2, 28.5, 28.8, 29.2, 31.6, 33.9, 35.0, 48.4, 49.8, 59.2, 107.7, 149.6, 150.4, 171.1, 171.4. IR (KBr) 2931, 1650, 1603, 1513 cm⁻¹. MS (FAB) m/z (rel intensity) 430 (M, 4), 402 (2), 288 (92), 145 (100). HRMS (FAB) calcd for $C_{25}H_{42}O_2N_4$ (M): 430.3307, found, 430.3311.

(2S,5S)-Dihexyl 1-(pyridin-4-yl)pyrrolidine-2,5-dicarboxylate (16)

According to the general procedure, **16** was prepared with n-hexanol in CH_2Cl_2 without addition of NMM and purified by preparative TLC (SiO_2 , $Et_2O/MeOH = 10/1$) in 38 % yield as a colorless oil.

[α]_D²⁰ = -77 (c 0.25, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ 0.88 (t, J = 6.6 Hz, 6H), 1.00–1.43 (m, 12 H), 1.43–1.70 (m, 4H), 2.10–2.30 (m, 2H), 2.40–2.65 (m, 2H), 4.00–4.20 (m, 4H), 4.48 (d, J = 8.2 Hz, 2H), 6.33 (dd, J = 1.6, 4.8 Hz, 2H), 8.24 (dd, J = 1.4, 4.8 Hz, 2H). ¹³C NMR (50 MHz, CDCl₃) δ 14.1, 22.6, 25.6, 28.7, 29.1, 31.4, 60.9, 65.7, 107.8, 150.0, 150.4, 172.7. IR (film) 2956, 2858, 1743, 1595, 1510 cm⁻¹. MS (FAB) m/z (rel intensity) 404 (M, 7), 275 (100), 191 (12), 145 (30). HRMS (FAB) calcd for C₂₃H₃₆O₄N₂ (M): 404.2675, found, 404.2673.

4-Pyridinyl-L-proline 1'-hexylamide (17)

To a solution of *n*-hexylamine (87 μ L, 0.66 mmol), *N*-(4-pyridyl)-L-proline hydrochloride (100 mg, 0.44 mmol) in DCM (10 mL) were added NMM (48 μ l, 0.44 mmol), EDCI (125 mg, 0.66 mmol) and HOBt (89 mg, 0.66 mmol). After stirring for 24 h, the mixture was evaporated and diluted with EtOAc (200 mL). The organic layer was washed with sat. aq. NaHCO₃ (10 mL), brine (10 mL), dried over Na₂SO₄, filtered and evaporated in vacuo. The residue was purified by preparative TLC (SiO₂, CHCl₃/MeOH = 5/1) to give **17** (41 mg, 34%) as colorless needles.

M.p. 45 °C. $[\alpha]_D^{20}$ –110 (c 0.5, CHCl₃). ¹H NMR (200 MHz, CDCl₃) δ 0.85 (t, J = 6.4 Hz, 3 H), 1.10–1.35 (m, 6 H), 1.42 (t, J = 6.6 Hz, 2 H), 1.90–2.20 (m, 2 H), 2.20–2.50 (m, 2 H), 3.17–3.40 (m, 3 H), 3.50–3.80 (m, 1 H), 4.10 (t, J = 5.4 Hz, 1 H), 6.24 (br s, 1 H), 6.46 (d, J = 5.9 Hz, 2H), 8.27 (d, J = 5.9 Hz, 2H). ¹³C NMR (50 MHz, NMR (CDCl₃) δ 14.1, 22.6, 24.0, 26.6, 29.7, 31.4, 31.5, 39.5, 48.9, 63.5, 108.2, 150.0, 152.1, 172.0. IR (KBr) 3299, 2928, 1650, 1602, 1551, 1518 cm⁻¹. MS (FAB) m/z (rel intensity) 275 (M⁺, 42), 147 (100), 105 (40), 78 (38). HRMS (FAB) Calcd for $C_{16}H_{25}ON_3$ 275.1998, found 275.1999.

Typical procedure for asymmetric desymmetrization

To a solution of substrate (0.1 mmol), catalyst (5 mol %) and 2,4,6-collidine (0.14 mmol, $18.5~\mu L$) in solvent (0.5 mL), isobutyric anhydride (0.13 mmol, $21.5~\mu L$) was added. After the mixture was stirred, it was diluted with EtOAc and washed with 1 M aq. HCl, sat. aq. NaHCO₃ and brine. The organic layer was dried over Na₂SO₄, filtered and evaporated in vacuo. Yields of monoacylate, diacylate and recovered substrate were determined by 1H NMR integration with dibenzyl ether as an internal standard.

Enantiomeric purities of 1,2- and 1,3-cyclohexanediol monoisobutyrate were determined by GLC analysis with beta-DEX 225 at 150 °C (Tables 1–5 in the manuscript).

Enantiomeric purities of *cis*-2,3-butanediol isobutyrate (**23a**) and hydrobenzoin isobutyrate (**23b**) were determined by GLC analysis with beta-DEX 225 at 90 °C and HPLC analysis with chiral stationary phase [column: CHIRALCEL OJ (4.6 mm x 25 cm), eluent: hexane—iPrOH (95:5), flow rate: 0.5 mL/min], respectively (Table 6 in the main manuscript).

1,3-Cyclohexanediol isobutyrate (20)

¹H-NMR (400 MHz, CDCl₃) δ: 4.65–4.54 (m, 1H), 3.61–3.50 (m, 1H), 3.28 (br s, 1H), 2.40 (sept, J = 7.1 Hz, 1H), 2.15–2.07 (m, 1H), 1.85–1.74 (m, 2H), 1.74–1.64 (m, 1H), 1.31–0.99 (m, 4H), 1.04 (d, J = 6.9 Hz, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 176.5, 70.6, 68.1, 40.4, 34.0, 33.9, 30.4, 19.9, 18.7. IR (neat) 3400, 2880, 2875, 2835, 1730, 1718 cm⁻¹; MS (EI) m/z (rel intensity) 186 (M⁺, 0.15), 168 (5), 125 (10), 98 (80), 81 (100); HRMS (EI) calcd for C₁₀H₁₈O₃ (M⁺), 186.1256, found 186.1248.

Acylative kinetic resolution of racemic-6 with catalyst 12b

To a solution of racemic-6 (18.6 mg, 0.1 mmol), catalyst 12b (3.3 mg, 5 mol%) and 2,4,6-collidine (13.2 μ L, 0.1 mmol) in CDCl₃ (0.7 mL), isobutyric anhydride (11.6 μ L, 0.07 mmol) was added at 20 °C. After the mixture was stirred at 20 °C for 23 h, conversion of 6 was determined by ¹H NMR integration with dibenzyl ether as an internal standard. Then, the mixture was diluted with EtOAc and washed with 1 M aq. HCl, sat. aq. NaHCO₃, and brine. The organic layer was dried over Na₂SO₄, filtered and evaporated in vacuo. Enantiomeric purity of (1*R*,2*S*)-6 (67% ee) was determined by GLC analysis with beta-DEX 225 at 150 °C.

Determination of the absolute configuration of 2,3-butanediol isobutyrate (23a)

Asymmetric desymmetrization of 2,3-butanediol (**22a**) with benzoic anhydride in the presence of cat. **11**, according to the general procedure, afforded the monobenzoate (**S1**) in 50% ee in 39% yield (HPLC conditions: CHIRALPAK AD, n-hexane/2-propanol = 97:3, flow rate: 0.2 mL/min, t_R = 33 min, 37 min, Scheme I). The absolute configuration of **S1** was

determined to be (2R,3S) by comparison with the optical rotation of the literature data [2]. Monobenzoate (**S1**) was further converted to (2R,3S)-**S2** [[α]_D²¹ –13.8 (c 1.0, CHCl₃)]. On the other hand, 2,3-butanediol isobutyrate (**23a**) obtained in Table 5 was converted to the corresponding benzoate (**S2**), which was assigned to be (2S,3S) by comparison of the optical rotation of **S2** from Scheme I. Thus, the absolute configuration of **23a** was determined to be (2S,3S).

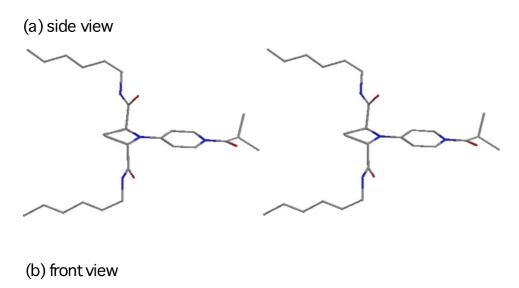
$$\begin{array}{c} \text{Cat 11 (5 \text{ mol\%})} \\ \text{OCOPh} \\ \text{Me} \\ \textbf{22a} \\ \textbf{22a} \\ \textbf{23a} \\ \textbf{DMAP (10 \text{ mol\%})} \\ \text{Collidine (1.4 equiv)} \\ \text{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \text{39\%} \\ \textbf{S1 50\% ee} \\ [\alpha]_D^{21} - 6.4 \\ (c 1.2, \text{CHCl}_3) \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{39\%} \\ \textbf{S2 51\% ee} \\ [\alpha]_D^{21} - 6.4 \\ (c 1.2, \text{CHCl}_3) \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{30\%} \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{30\%} \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{30\%} \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{30\%} \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{Me} \\ \textbf{CHCl}_3, 20 \, ^{\circ}\text{C}, 24 \, h} \\ \textbf{30\%} \\ \textbf{Me} \\ \textbf{21} \\ \textbf{Me} \\ \textbf{21} \\ \textbf{Me} \\ \textbf{224} \\ \textbf{Me} \\ \textbf{23a} \\ \textbf{21} \\ \textbf{23a} \\ \textbf{21} \\ \textbf{23a} \\ \textbf{21} \\ \textbf{22} \\ \textbf{23a} \\ \textbf{23a} \\ \textbf{24} \\ \textbf{24} \\ \textbf{25} \\ \textbf{26} \\ \textbf{21} \\ \textbf{278\% ee} \\ \textbf{26} \\ \textbf{21} \\ \textbf{278\% ee} \\ \textbf{278\% ee} \\ \textbf{278\% ee} \\ \textbf{278\% ee} \\ \textbf{21} \\ \textbf{21} \\ \textbf{21} \\ \textbf{22} \\ \textbf{23} \\ \textbf{24} \\ \textbf{24} \\ \textbf{25} \\ \textbf{278\% ee} \\ \textbf{26} \\ \textbf{21} \\ \textbf{278\% ee} \\ \textbf{21.1, CHCl}_3) \\ \textbf{21.1, CHCl}_3) \\ \textbf{21.1, CHCl}_3) \\ \textbf{22.1} \\ \textbf{23a} \\ \textbf{23a} \\ \textbf{24a} \\ \textbf{25a} \\$$

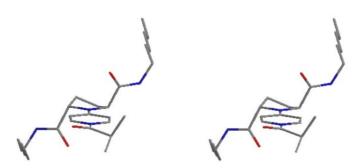
cis-2,3-Butanediol diisobutyrate (24a)

¹H NMR (200 MHz, CDCl₃) δ 1.00-1.17 (m, 12H), 1.18 (d, J = 5.4 Hz, 6H), 2.40–2.62 (m, 2H), 4.90–5.10 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 176.1, 70.8, 34.0, 18.9, 18.7, 14.9. IR (film) 2890, 2870, 2840, 1730 cm⁻¹. MS m/z (rel intensity) 230 (M, 0.03), 215 (2), 142 (70), 115 (72), 89 (68), 71 (100), 55 (58). HRMS calcd for C₁₂H₂₂O₄ 230.1518, found 230.1521. HRMS calcd for C₁₁H₁₉O₄ (M–CH₃) 215.1284, found 215.1287.

Conformation search of catalyst 11.

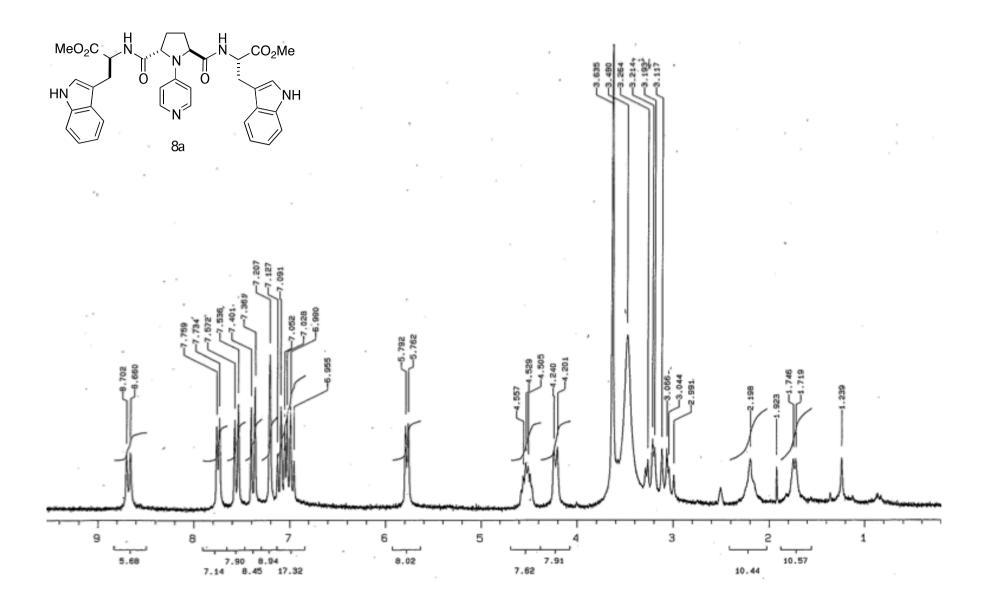
The most stable conformer of **11** was generated by a molecular modeling search with AMBER* force field with the GB/SA solvation model for chloroform using MacroModel V 9.0 (50,000 steps MCMM). Stereoview of the most stable conformation was shown below.

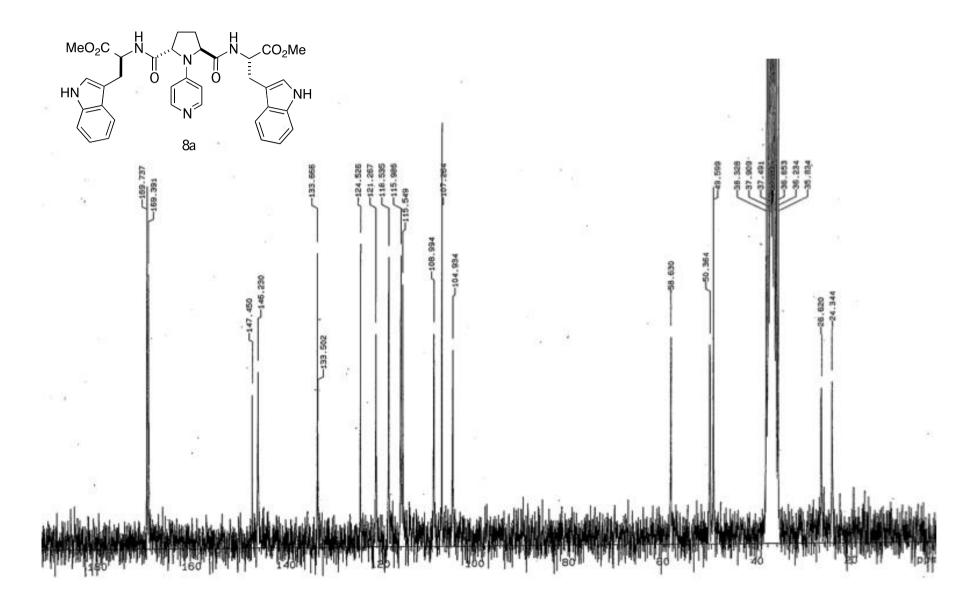


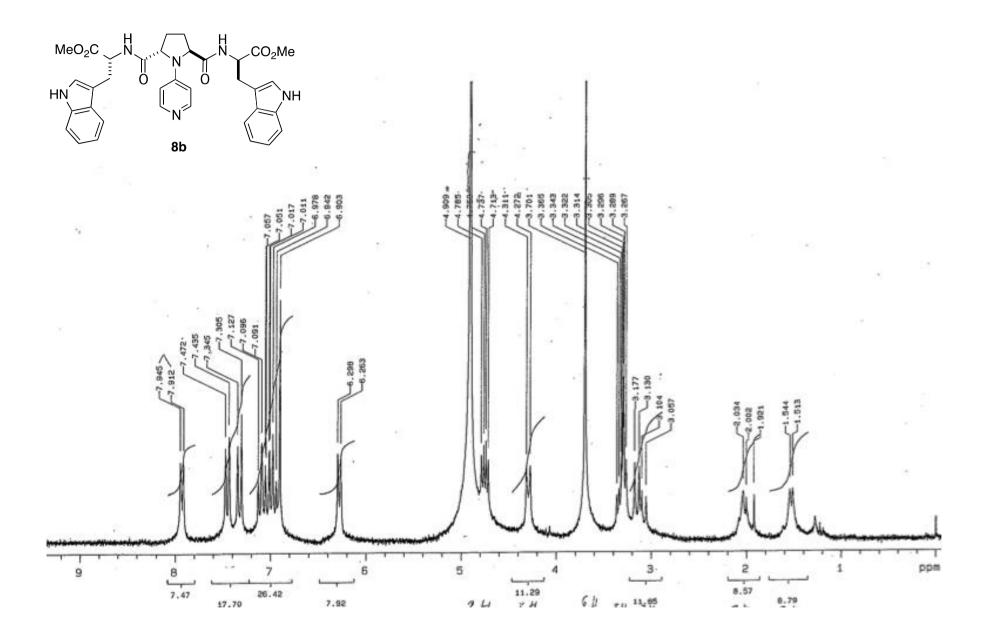


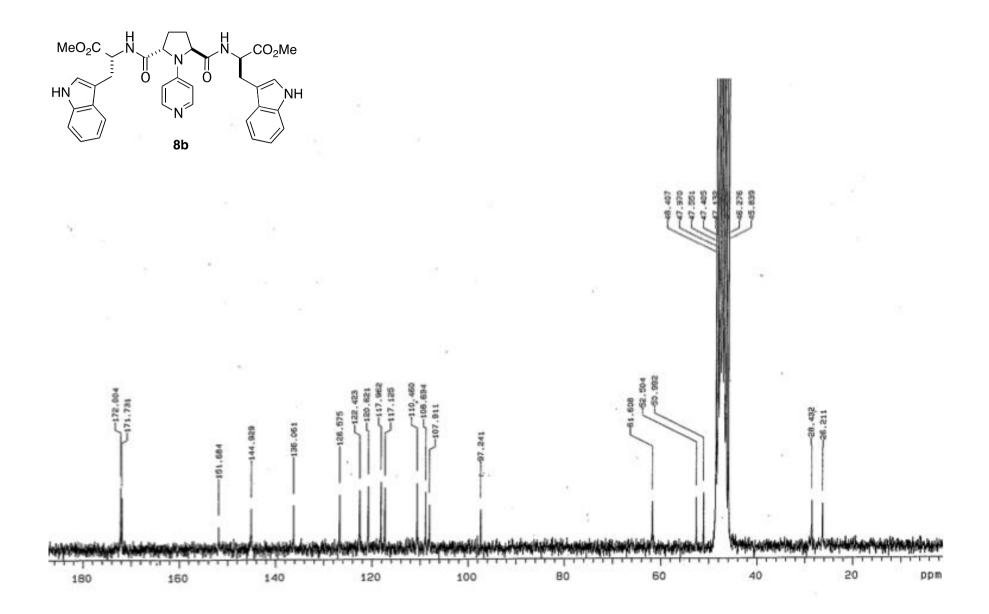
References

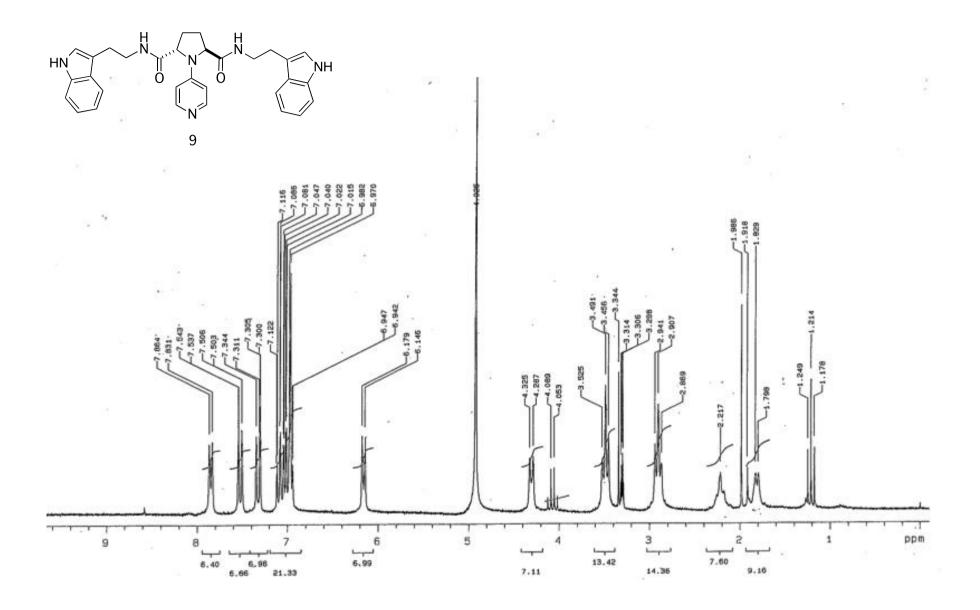
- 1. Kawabata, T.; Muramatsu, W.; Nishio, T.; Shibata, T.; Schedel, H. J. Am. Chem. Soc., 2007, 129, 12890.
- 2. Mazet, C.; Köhler, V.; Pfaltz, A. Angew. Chem. Int. Ed. 2005, 44, 4888.

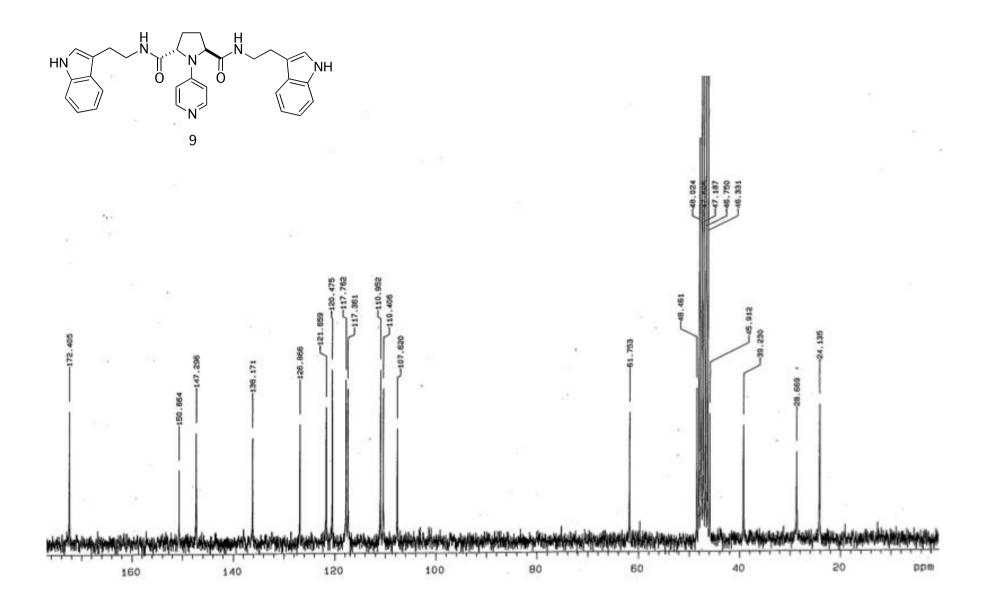


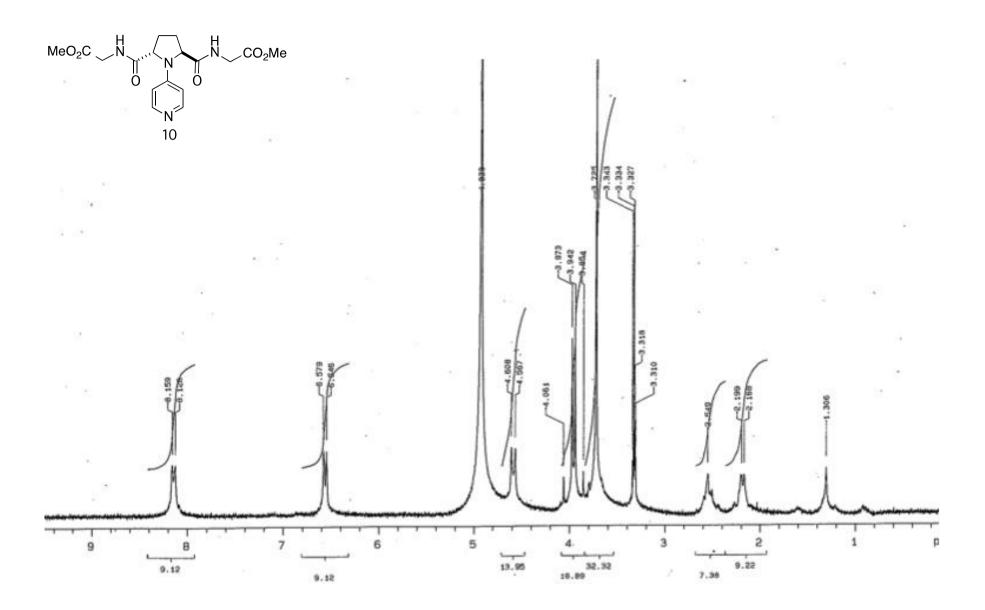


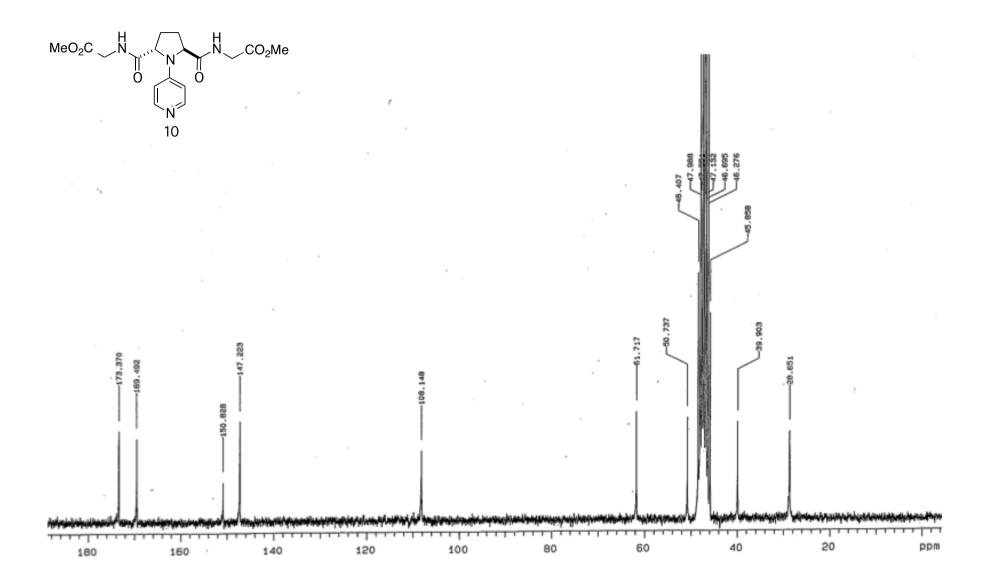


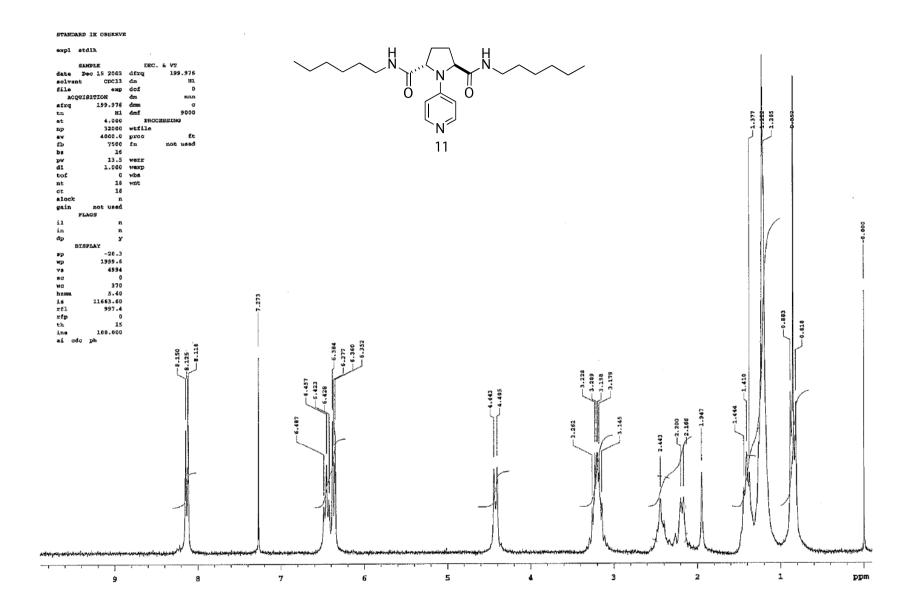














expl std13c

