

# Supporting Information

for

## **Activation of cryptic metabolite production through gene disruption: Dimethyl furan-2,4-dicarboxylate produced by *Streptomyces sahachiroi***

Dinesh Simkhada, Huitu Zhang, Shogo Mori, Howard Williams, Coran M. H. Watanabe\*

Address: Texas A&M University, Department of Chemistry, College Station, TX 77843,  
USA

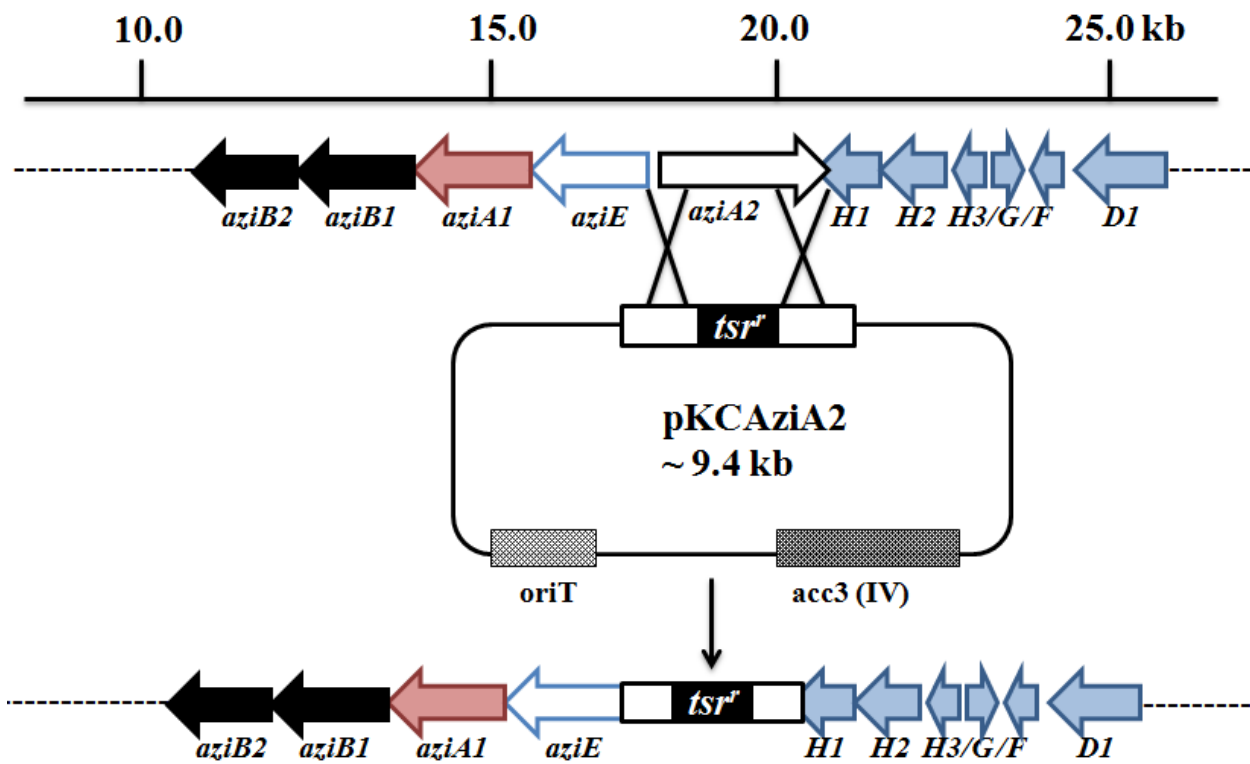
Email: Coran M. H. Watanabe – [watanabe@chem.tamu.edu](mailto:watanabe@chem.tamu.edu)

\* Corresponding author

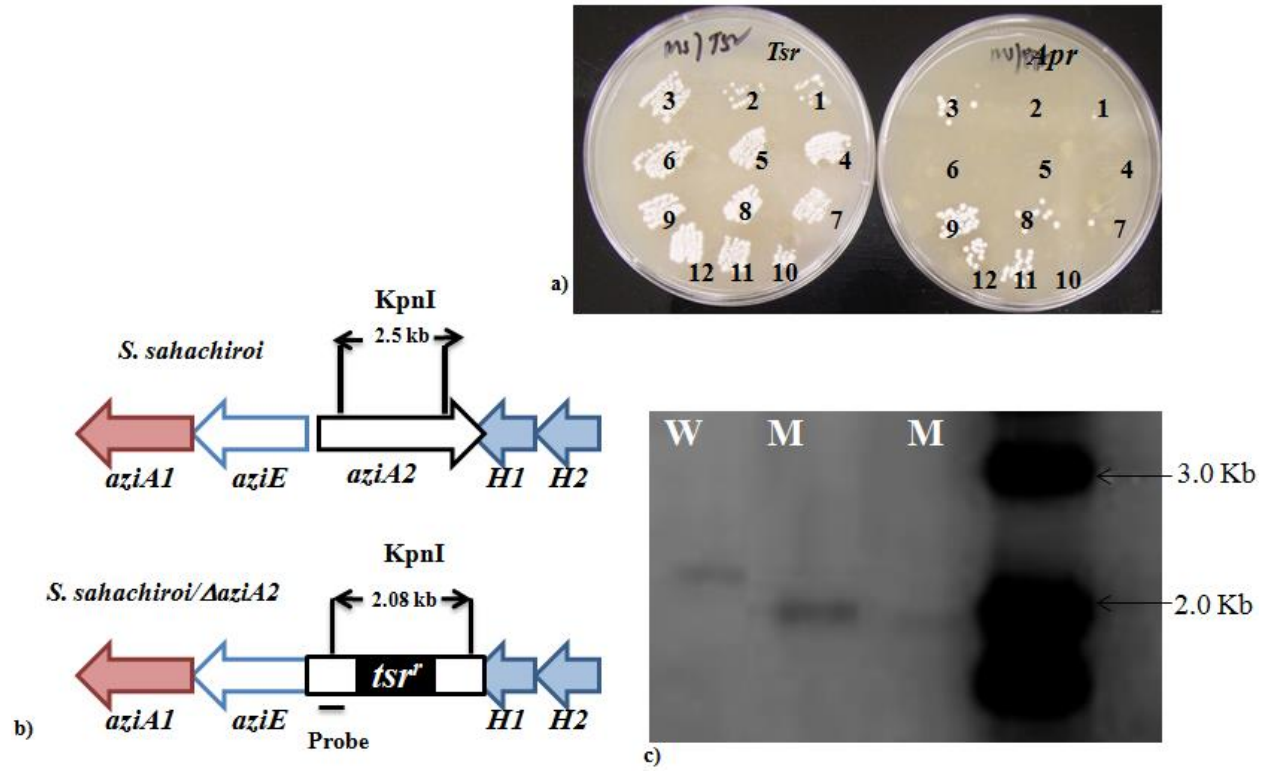
**Details on genetic knockout, Southern blot analysis,  
LC–MS analysis and NMR data**

## Contents

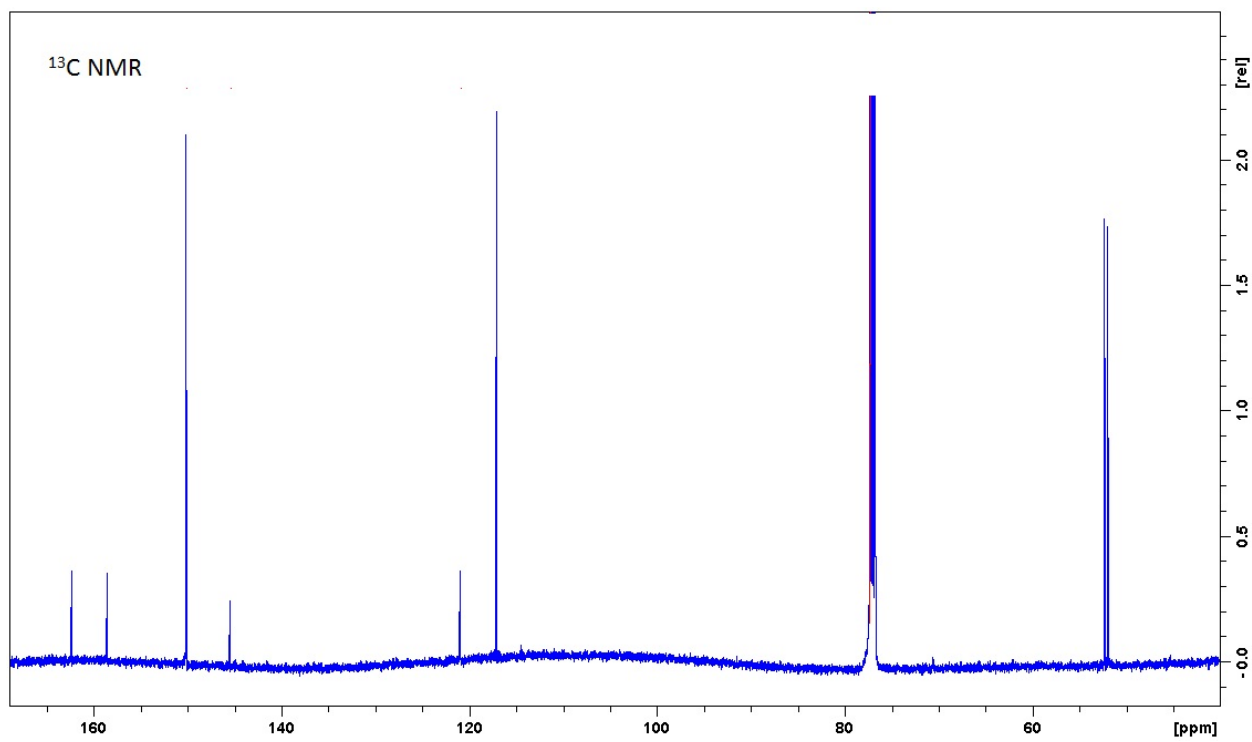
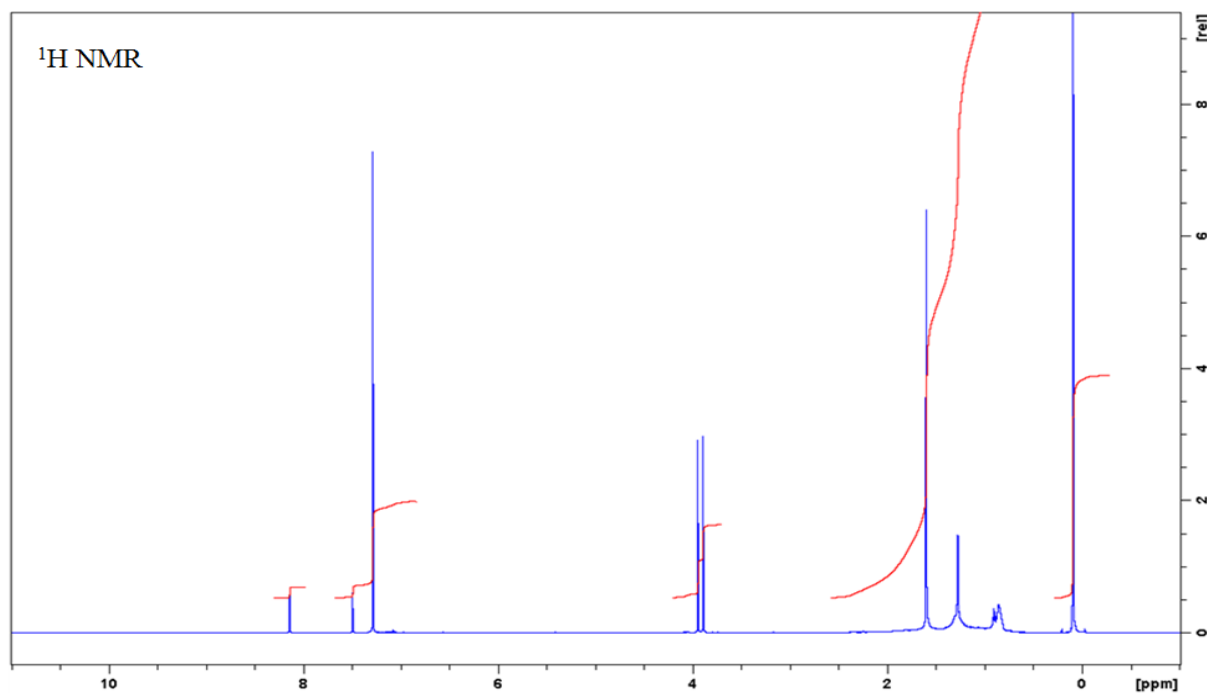
<b>Figure S1:</b> Strategy for genetic knockout of <i>aziA2</i> .....	S3
<b>Figure S2:</b> Confirmation of disruptants: Southern blot.....	S4
<b>Figure S3:</b> NMR profile analysis of dimethyl furan-2,4-dicarboxylate (in order: <sup>1</sup> H NMR, <sup>13</sup> C NMR, Dept 90, Dept 135, HMBC and HMQC).....	S5–S10
<b>Figure S4:</b> LC–APCIMS profile of dimethyl furan-2,4-dicarboxylate.....	S11
<b>Figure S5:</b> LC–APCIMS/MS profile of dimethyl furan-2,4-dicarboxylate.....	S11
<b>Figure S6:</b> GC–MS profile of dimethyl furan-2,4-dicarboxylate.....	S12
<b>Supplemental experimental procedures</b> .....	S13
<b>Table S1:</b> List of strains and plasmids used in this study.....	S13
<b>Table S2:</b> List of primers used in this study.....	S14
<b>References</b> .....	S15

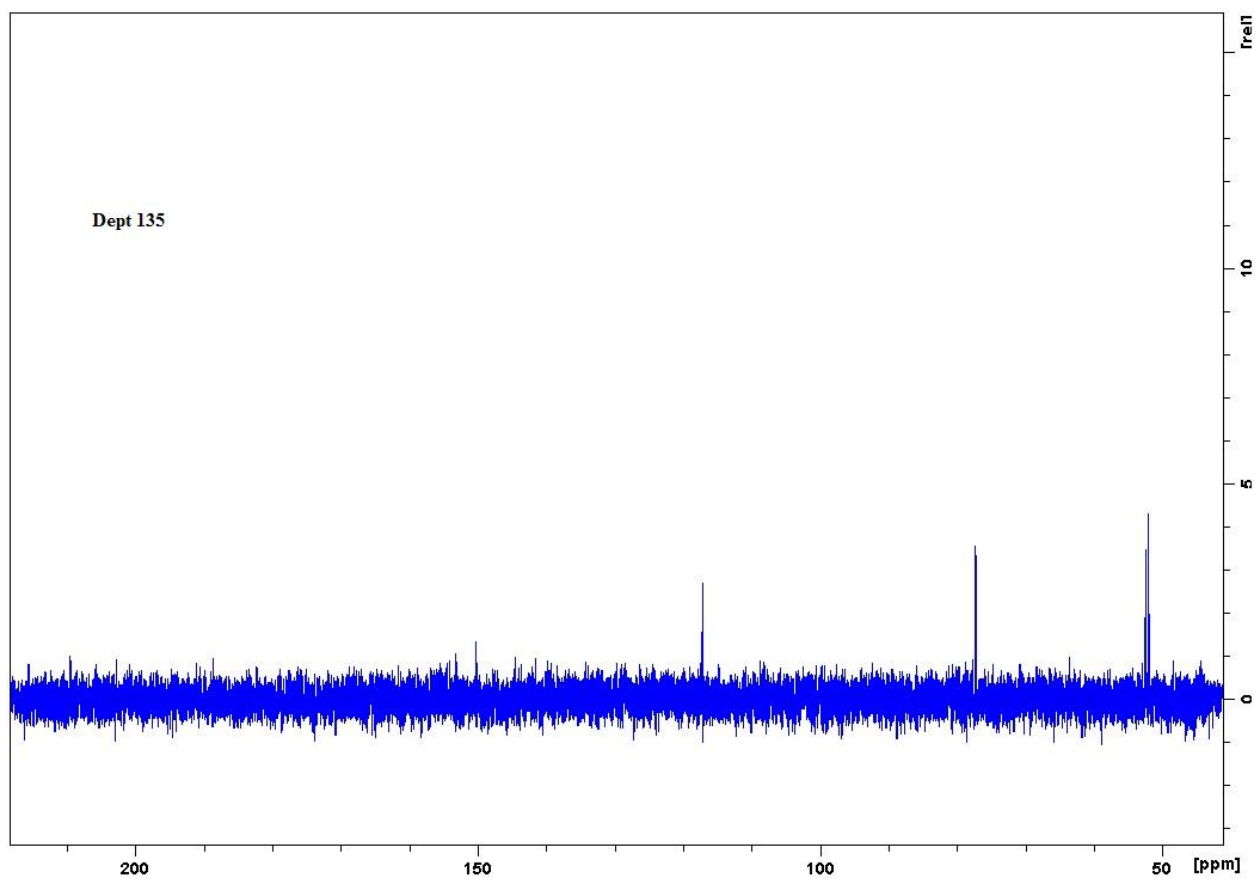
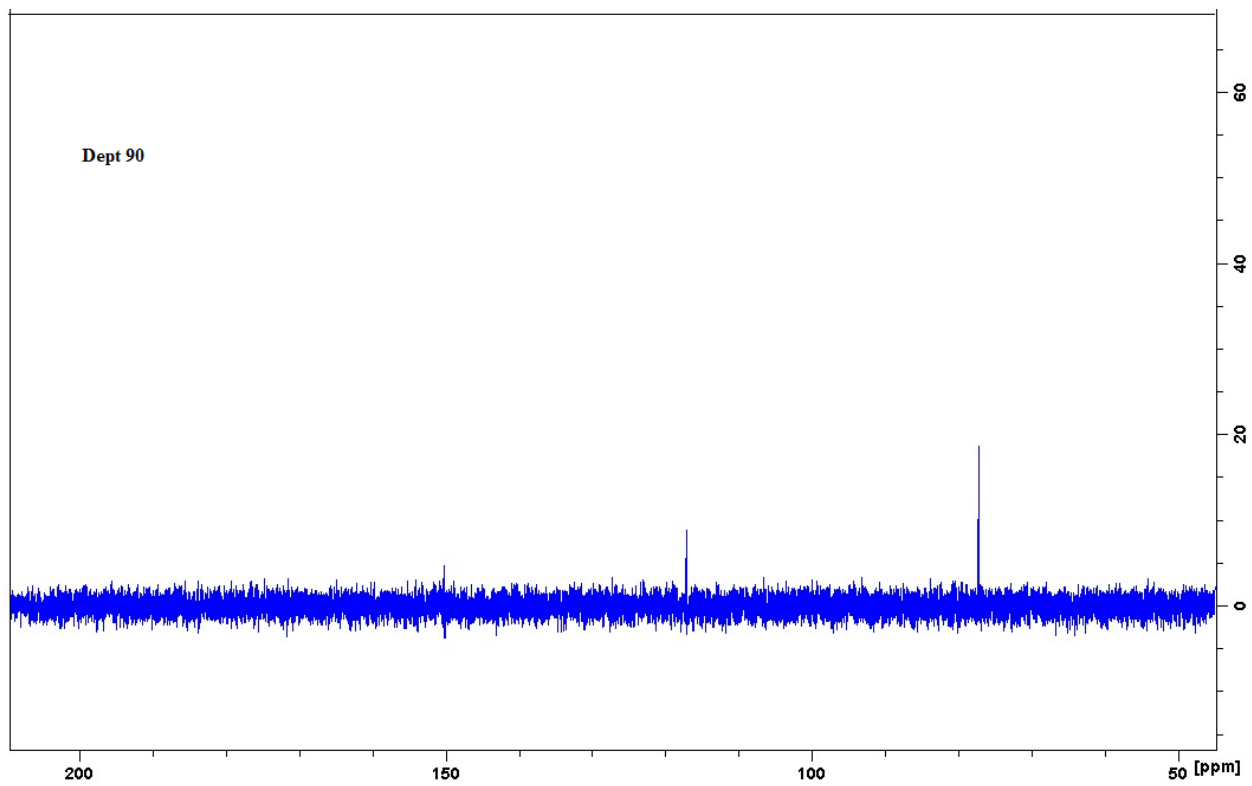


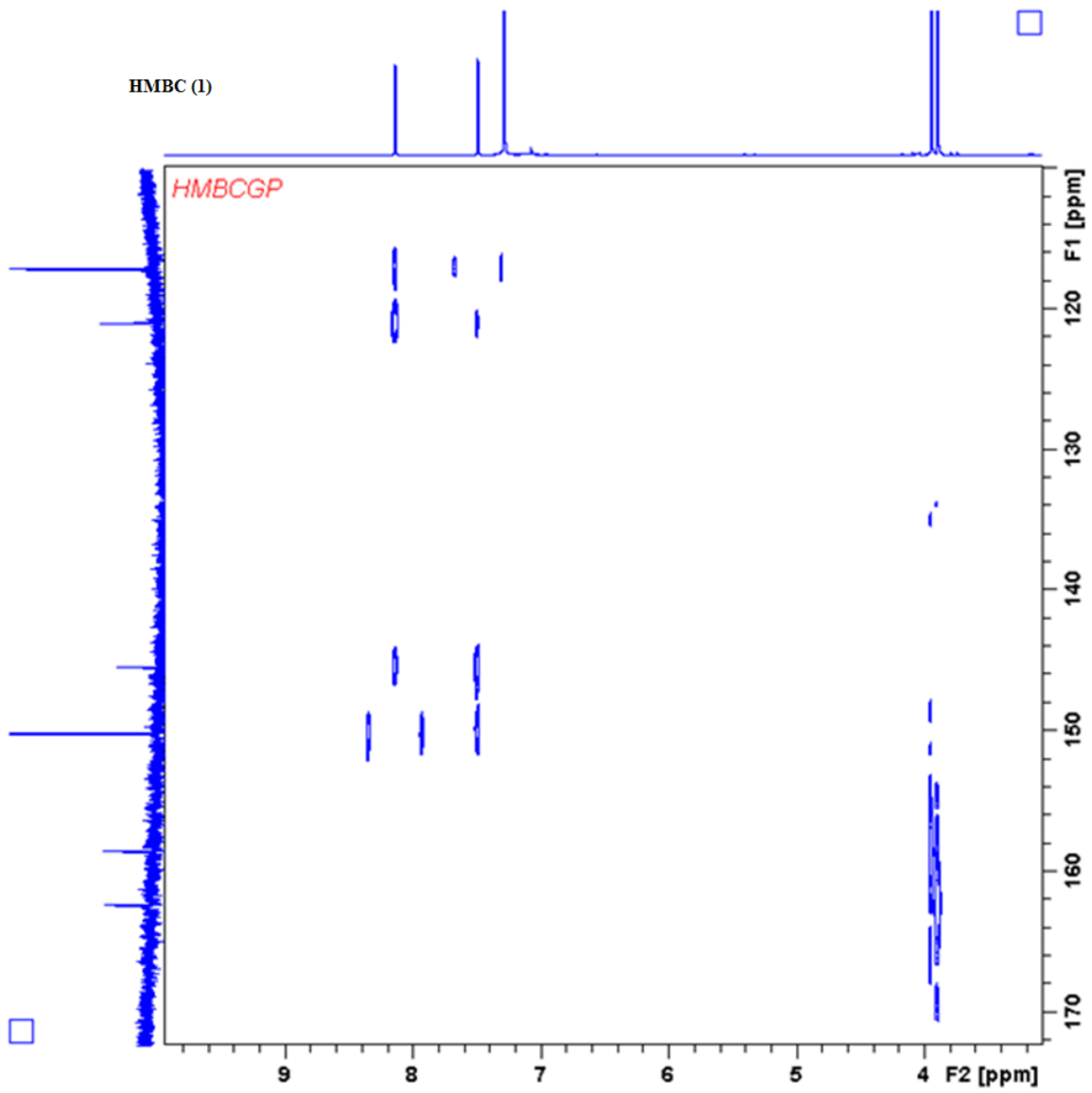
**Figure S1:** Strategy for genetic knockout of *aziA2*.

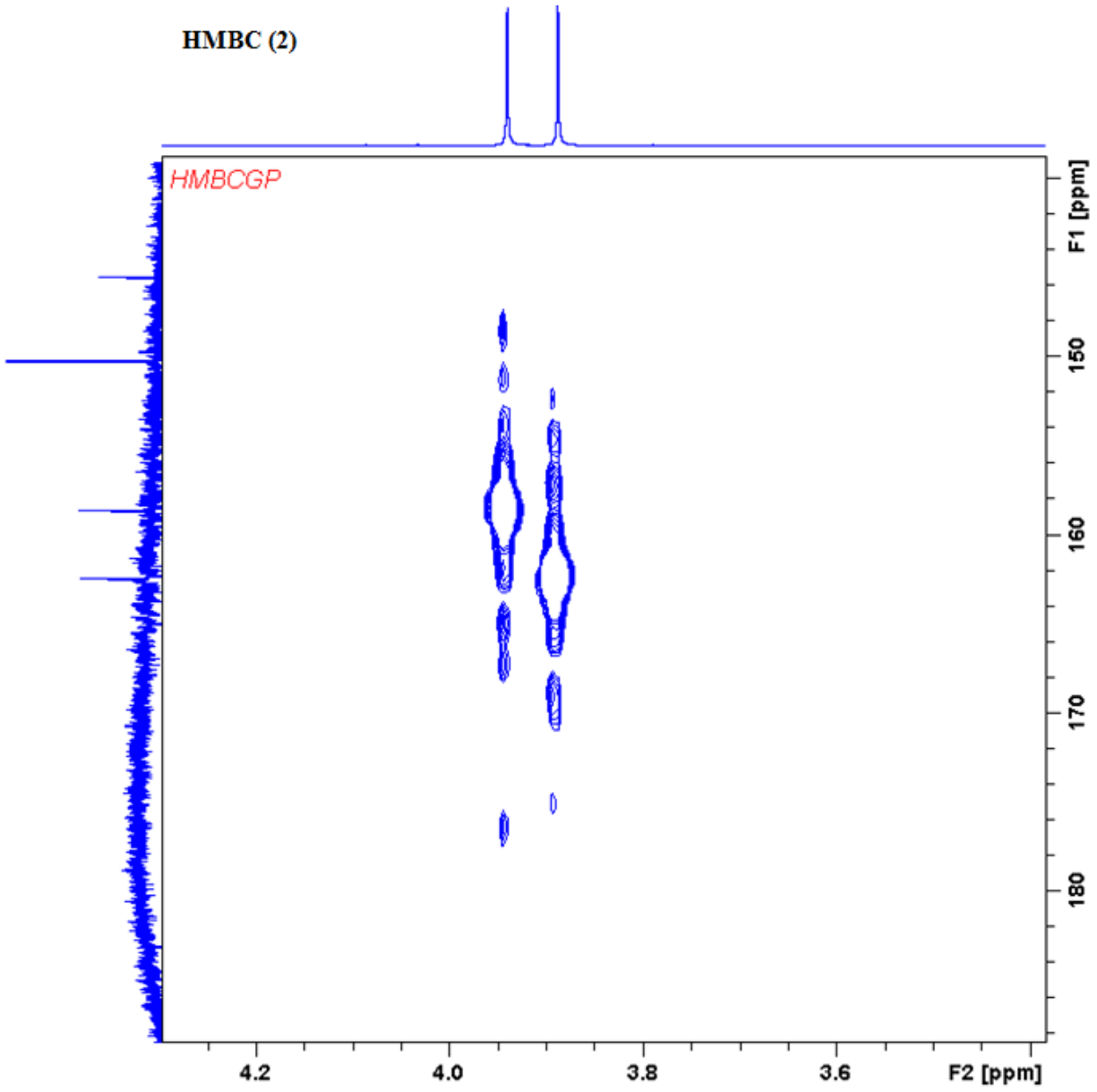


**Figure S2:** Confirmation of *aziA2* knockout: a) Screening for positive colonies, b) Strategy for Southern hybridization, c) Southern blot; W: wild type, M: mutants.



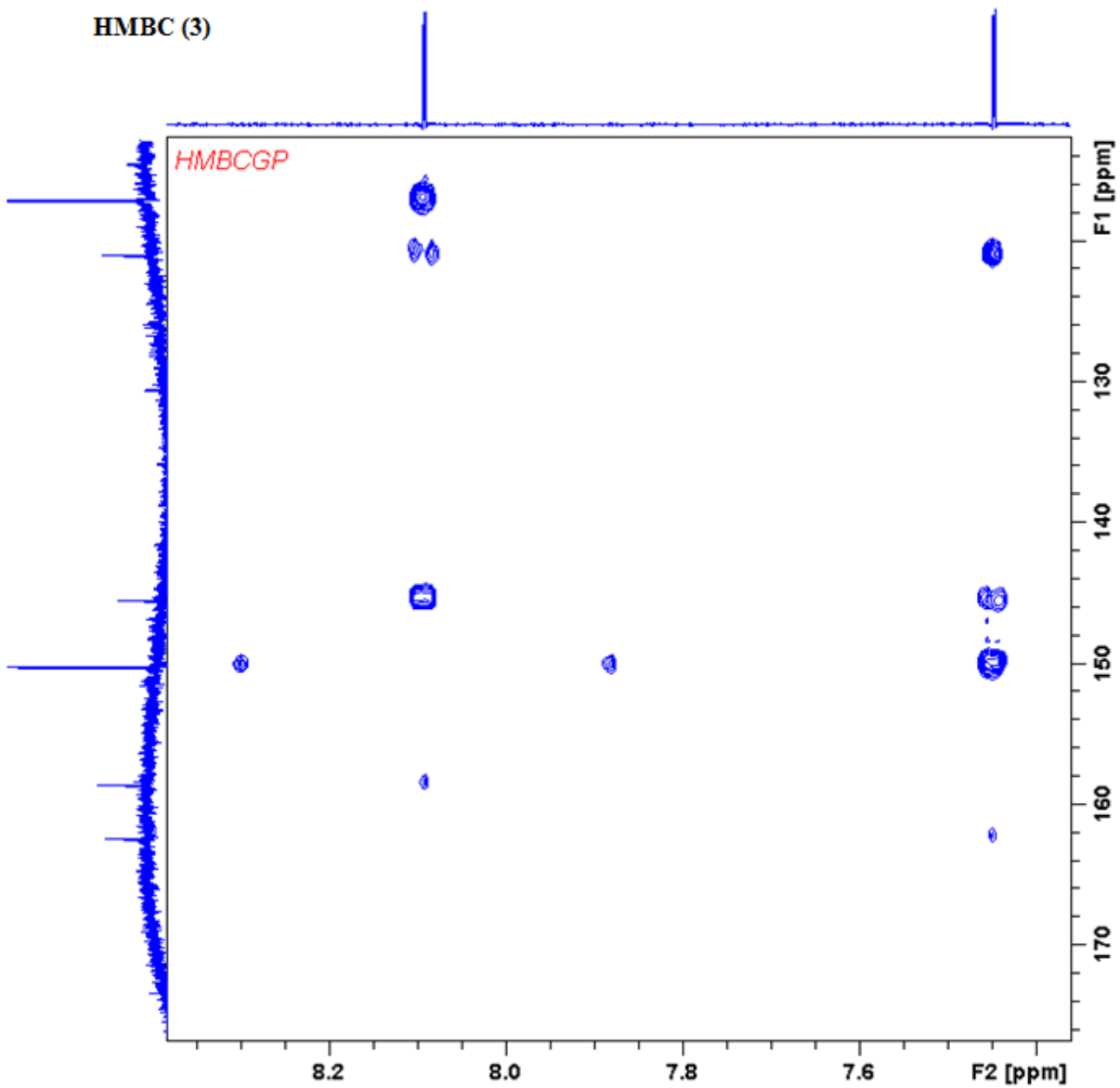


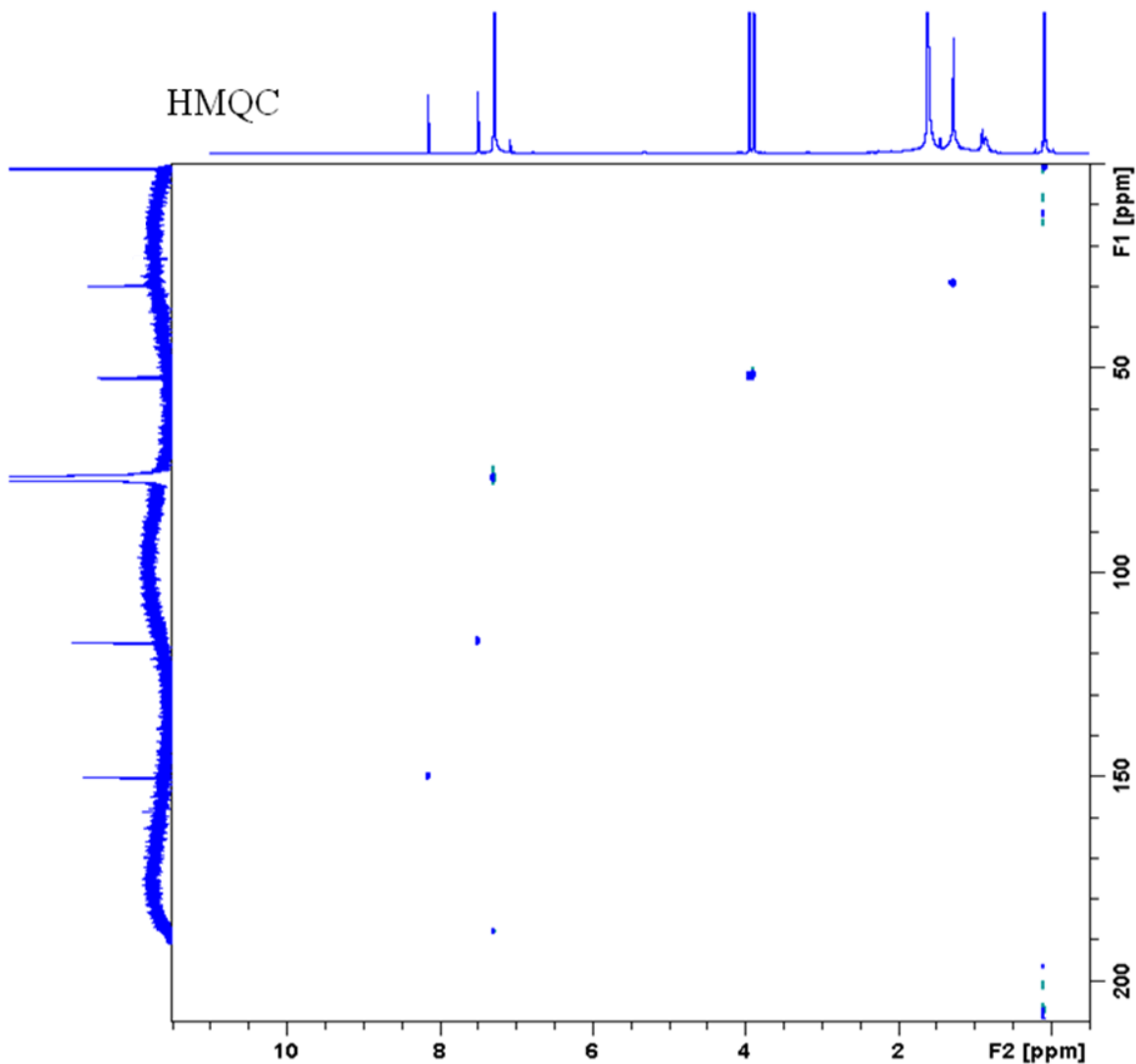




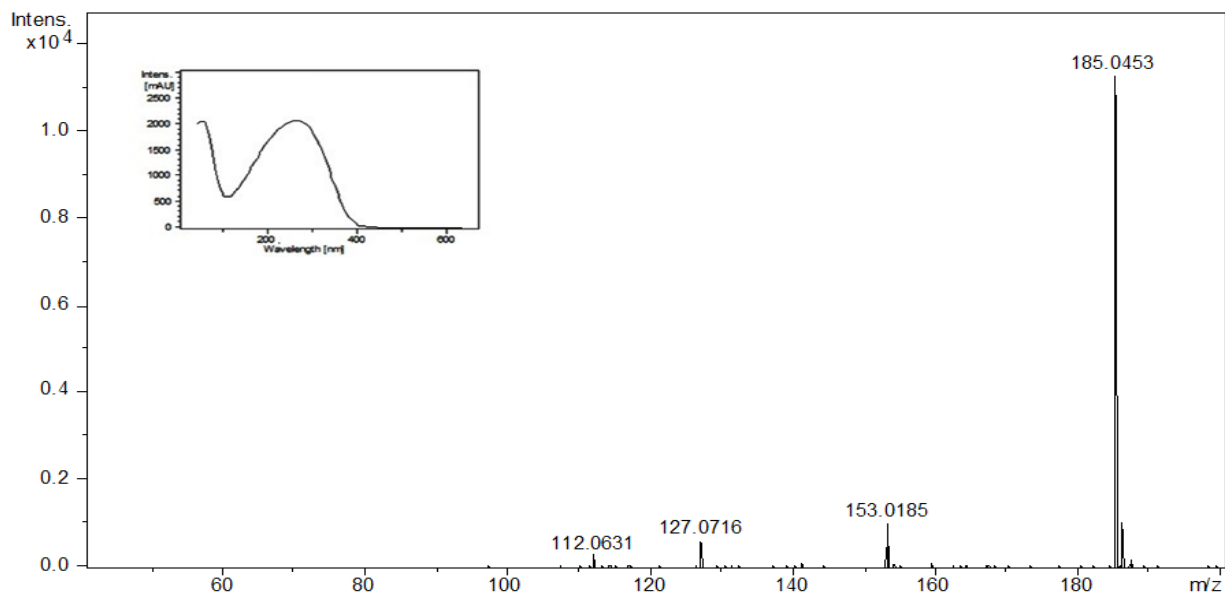


HMBC (3)

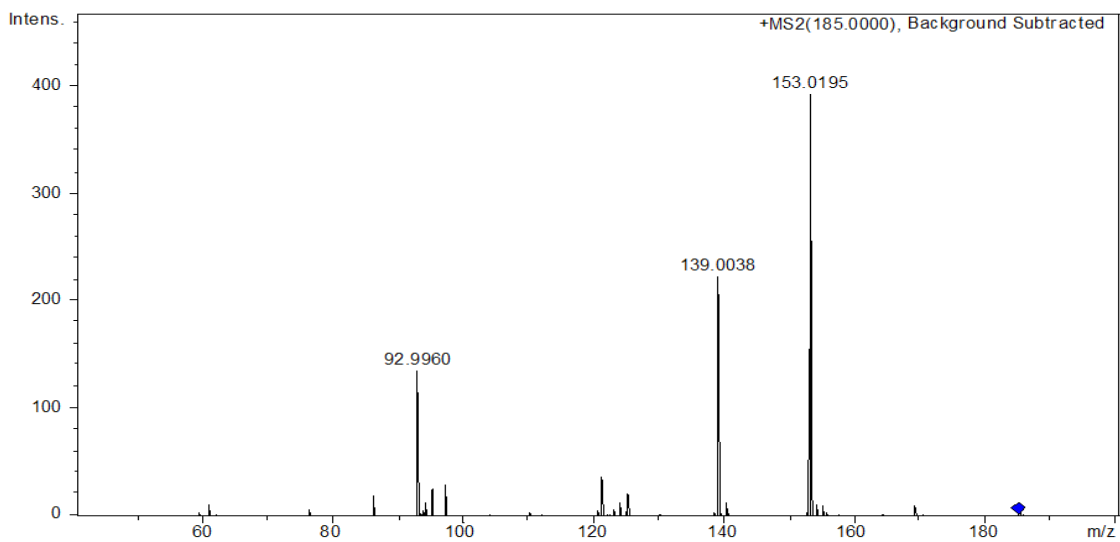




**Figure S3:** NMR Profile for dimethyl furan-2,4-dicarboxylate (in order:  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, Dept 90, Dept 135, HMBC (1,2 and 3) and HMQC).



**Figure S4:** LC-APCIMS profile of dimethyl furan-2,4-dicarboxylate.



**Figure S5:** LC-APCIMS/MS profile of dimethyl furan-2,4-dicarboxylate.

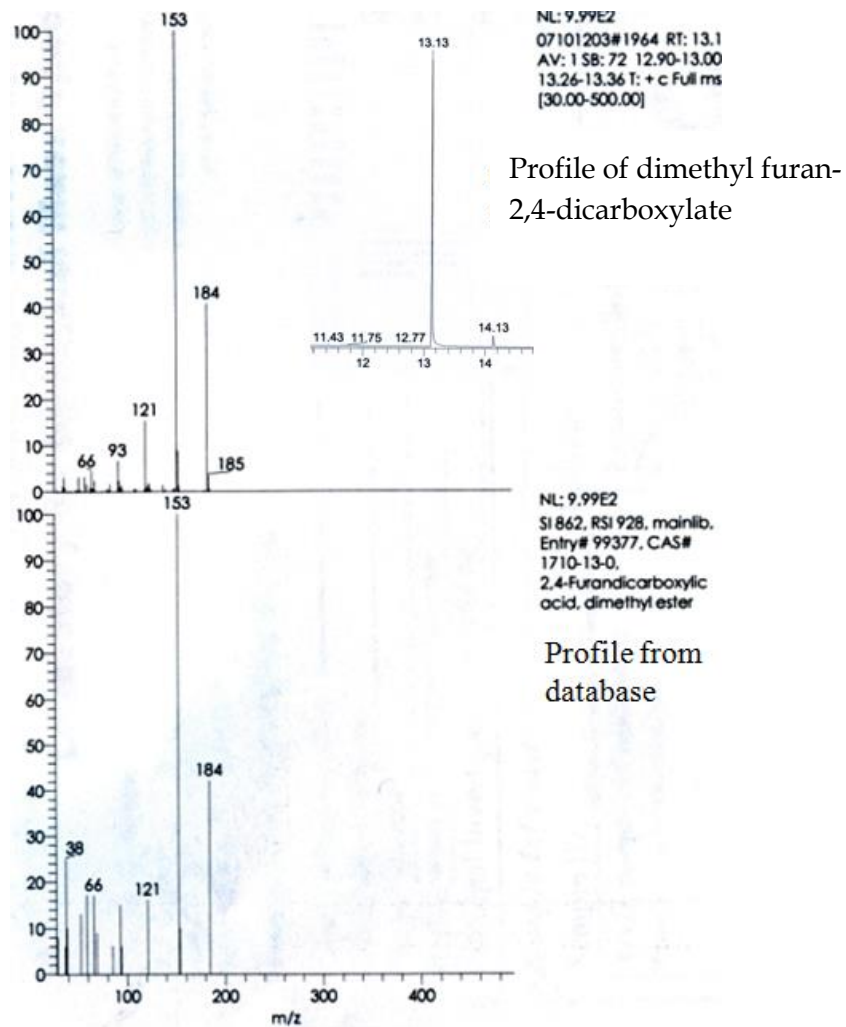


Figure S6: GC-MS profile of dimethyl furan-2,4-dicarboxylate.

## Supplemental experimental procedures

### Bacterial strains, plasmids and culture conditions

*Escherichia coli* (*E. coli*) strains (XL1 Blue and DH10B) were cultured in Luria Bertani (LB) medium at 37 °C. *E. coli* XL1 Blue was routinely used as a host for the recombinant plasmids. *E. coli* S17-1 was used as a donor strain for conjugative transfer of mobilizable plasmids into *Streptomyces sahachiroi*. The parental strain *S. sahachiroi* was cultured at 28 °C in R2YE medium [1] for 2–3 days for the preparation of genomic DNA. pGEM-T Easy (Promega, USA) was used as a cloning vector and pKC1139 was used as disruption vector. All of the bacterial strains and a list of vectors and recombinant plasmids including their relevant sources are provided in Table S1.

**Table S1: List of strains and plasmids used in this study.**

Strains/Plasmids	Relevant characteristics	References
<b>Strains</b>		
<i>E. coli</i> XL1 Blue	<i>recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac</i> [F' <i>proAB lacI<sup>q</sup> Z DM15 Tn10</i> (Tet <sup>r</sup> )]	Stratagene La Jolla, CA, USA
<i>E. coli</i> S17-1	<i>recA1 pro thi</i> ; has the <i>tra</i> genes from plasmid RP4 integrated in the chromosome	Simon <i>et. al.</i> , 1983 [2]
<i>E. coli</i> S17-1/pKC-AziA2	<i>recA1 pro thi</i> ; has the <i>tra</i> genes from plasmid RP4 integrated in the chromosome with <i>aziA2</i> disruption plasmid pKC-AziA2	This study
<i>Streptomyces sahachiroi</i>	Azinomycin B producer, wild type strain	ATCC
<i>S. sahachiroi</i> /Δ <i>aziA2</i>	<i>Streptomyces sahachiroi</i> with deletion of <i>aziA2</i>	This study
<b>Plasmids</b>		
pGEM <sup>®</sup> T-easy vector	<i>E. coli</i> general cloning vector, amp <sup>r</sup>	Promega, USA
pKC1139	<i>Streptomyces-E. coli</i> bifunctional vector, Apr <sup>r</sup>	Bierman <i>et. al.</i> , 1992 [3]
pKC-AziA2	Disruption recombinant plasmid for <i>aziA2</i>	This study

**Table S2: List of primers used in this study**

<b>Primers</b>	<b>Sequence (5'-3')*</b>
<b>Gene disruption primers</b>	
aziA2UF	<u>AAGCTT</u> GACTACGAAGACTACATGCCCC
aziA2UR	<u>TCTAGAGT</u> CGGCGAGGTCCGGTGCGCCC
aziA2DF	<u>TCT AGAGT</u> CCCAGACCCTGCTGATGCAGG
aziA2DR	<u>GAATTC</u> AGGGACCGCGGCTCCCGCTCCGT
TsrF	<u>TCTAGAGAT</u> CAAGGCGAATACTTCATATG
TsrR	<u>TCTAGA</u> ACGAATCGAGGTTCGAGGAACCGAG
<b>Primers for detection of genotype</b>	
aziA2 probe-F	TGATGCAGGAGCTGAGCACCGCCT
aziA2 Probe R	GACGATCTCCGCGAAGGGCAGCT

\* Restriction sites are underlined

## References

1. Kieser, T.; Bibb, M.; Butter, M.; Chater, K. F.; Hopwood, D. A., *Practical Streptomyces Genetics*. The John Innes Foundation: Norwich, UK, 2000.
2. Simon, R., Priefer U. and Pühler A., A Broad Host Range Mobilization System for In vivo Genetic Engineering: Transposon Mutagenesis in Gram Negative Bacteria. *Biotechnol.* **1983**, *1*, 784–791.
3. Bierman, M.; Logan, R.; O'Brien, K.; Seno, E. T.; Nagaraja Rao, R.; Schonher, B. E., Plasmid Cloning Vectors for the Conjugal Transfer of DNA from *Escherichia coli* to *Streptomyces spp.* *Gene* **1992**, *116*, 43–49.