Supporting Information

for

Chemoenzymatic synthesis and biological evaluation of enantiomerically enriched 1-(β-hydroxypropyl)imidazolium- and triazolium-based ionic liquids

Paweł Borowiecki*, Małgorzata Milner-Krawczyk and Jan Plenkiewicz

Address: Warsaw University of Technology, Faculty of Chemistry, Noakowskiego St.

3, 00-664 Warsaw, Poland

Email: Paweł Borowiecki* - pawel_borowiecki@onet.eu

* Corresponding author

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General details

All commercially available reagents (Aldrich, Fluka and POCH) were used without further purification. Novozym SP 435 (lipase from Candida antarctica B immobilized on a macroporous acrylic resin, >10000 U/g), Amano PS (native lipase from Burkholderia cepacia ealier Pseudomonas cepacia, >23.000 U/g), Amano PS-C (immobilized on ceramic particles) and Amano PS-IM (immobilized on diatomite, >500 U/g), Amano AK (native lipase from *Pseudomonas fluorescens*, >20.000 U/g) were purchased from Novo Nordisk Co. and Amano Pharmaceutical Co., respectively and were used without any pretreatment. Melting points were obtained with an MPA100 Optimelt SRS apparatus. Thin-layer chromatography was carried on TLC aluminium plates with silica gel Kieselgel 60 F₂₅₄ (Merck) (0.2 mm thickness film) and the compounds were visualized in iodine vapor. Preparative plate chromatography was performed with PSC-Fertigplatten Kieselgel 60 F_{254} (20 x 20 cm with 2 mm thickness layer). The chromatographic analyses (GLC) were performed with an HP Series II 5890 instrument equipped with a flame ionization detector (FID) and fitted with HP-50+ (30 m) semipolar column. Helium (2 ml/min) was used as carrier gas; $T_{\text{injector}} = 280 \, ^{\circ}\text{C}$, $T_{\text{column}} = 100 \, ^{\circ}\text{C}$ (3 min) and 100–280 $^{\circ}\text{C}$ (10 $^{\circ}\text{C/min}$); retention times (t_R) are given in minutes under these conditions. Column chromatography was performed by using Silica gel 60 (Merck) of 40-63 µm. A mixture of chloroform/methanol was used as eluent (9:1 or 8:2 v/v depending on substance purified). The enantiomeric excesses of the resulting esters and alcohols were determined by HPLC analysis performed on Shimadzu CTO-10ASV chromatograph equipped with STD-20A UV detector and Chiralcel OD-H (Diacel) chiral column using mixtures of *n*-hexane/isopropyl alcohol as the mobile phase in appropriate ratios given in experimental section; flow (f) is given in ml/min; racemic alcohols and esters

were used as standards. Optical rotations were measured with a P20 polarimeter (Belligham & Stanley Ltd., line D spectrum of sodium) in a 2 dm long cuvette. UV–vis absorption spectra were measured with Cary 3 spectrometer (data were collected on diluted solutions in quartz spectroscopy cells). ¹H and ¹³C NMR spectra were measured with a Varian Mercury 400BB spectrometer operating at 400 MHz for ¹H and 100 MHz for ¹³C nuclei, chemical shifts (δ) are given in parts per million (ppm) related to tetramethylsilane (TMS) as internal standard; signal multiplicity assignment: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; coupling constant (*J*) are given in hertz (Hz). Mass spectra were recorded with a Micro-mass ESI Q-TOF spectrometer at the Mass Spectrometry Laboratory, Institute of Biochemistry and Biophysics (IBB), PAN. IR spectra of neat samples were recorded on a PerkinElmer System 2000 FT-IR Spectrometer equipped with a Pike Technologies GladiATRTM attenuated total reflectance (ATR) accessory with a monolithic diamond crystal stage and a pressure clamp. Elemental analyses were performed with a VARIO EL III (Elementar Analysensysteme GmbH) elemental analyzer.

Procedures for gram-scale preparation of racemic alcohols and acetates

1-(1*H*-Imidazol-1-yl)propan-2-ol (±)-3a: A mixture of imidazole (8 g; 117.52 mmol) and 1,2-propylene oxide (7.5 g; 129.28 mmol; 9 ml) was placed in a round-bottomed flask and stirred at 32 °C for 24 h. The resulting crude product was purified by column chromatography [CHCl₃–MeOH (9:1)] yielding a yellowish oil (13.2 g; 104.6 mmol; yield = 89%). $R_f = 0.41$ [CHCl₃–MeOH (9:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.17 (d, J = 8 Hz, 3H, C H_3 CH(OH)CH₂), 3.77–3.91 (m, 2H, HOCHC H_2 N¹), 3.98–

4.01 (m, 1H, HOC*H*CH₂N¹), 5.69 (s, 1H, O*H*), 6.83 (s, 1H, CH₂N¹CH=C*H*N³), 6.87 (s, 1H, CH₂N¹C*H*=CHN³), 7.32 (s, 1H, CH₂N¹C*H*=N³); ¹³C NMR (CDCl₃, 100 MHz) δ 20.5 (*C*H₃CH(OH)CH₂), 54.6 (HOCH*C*H₂N¹), 66.3 (HO*C*HCH₂N¹), 119.7 (CH₂N¹CH=CHN³), 128.1 (CH₂N¹CH=*C*HN³), 137.3 (CH₂N¹CH=N³); HRMS (ESI⁺, *m/z*): [M + H]⁺_{calcd} = 127.0865, [M + H]⁺_{found} = 127.1009; anal. calcd. (%) for C₆H₁₀N₂O: C, 57.12 H, 7.99 N, 22.21; found: C, 57.16 H, 7.98 N, 22.25; IR (neat, v cm⁻¹): 3111, 2971, 2869, 1734, 1509, 1437, 1373, 1285, 1233, 1107, 1076, 919, 820, 735, 661, 625, 472; UV–vis: λ_{max} = 201 nm; GC: t_R = 5.960; HPLC [hexane–iPrOH (9:1); f = 0.8]: t_R = 20.056, 34.482.

1-(1*H***-1,2,4-Triazol-1-yl)propan-2-ol (±)-3b**: A mixture of 1,2,4-triazole (7 g; 101.36 mmol) and 1,2-propylene oxide (6.47 g; 111.49 mmol; 7.8 ml) was placed in a round-bottomed flask and stirred at 32 °C for 24 h. The resulting crude product was purified by column chromatography [CHCl3–MeOH (4:1)] to afford a yellowish oil (10.9 g; 85.7 mmol; yield = 85%). $R_f = 0.42$ [CHCl3–MeOH (4:1)]; ¹H NMR (CDCl3, 400 MHz) δ 1.11 (d, J = 8 Hz, 3H, CH_3 CH(OH)CH2), 3.85–3.94 (m, 1H, HOC*H*CH2N¹), 3.98–4.10 (m, 2H, HOCHC*H*2N¹), 5.00 (s, 1H, O*H*), 7.66 (s, 1H, CH2N¹N²=C*H*N⁴), 7.98 (s, 1H, CH2N¹C*H*=N⁴); ¹³C NMR (CDCl3, 100 MHz) δ 20.17 (CH3CH(OH)CH2), 56.6 (HOCHCH2N¹), 65.3 (HOCHCH2N¹), 143.5 (CH2N¹CH=N⁴), 150.9 (CH2N¹N²=CHN⁴); HRMS (ESI⁺, m/z): [M + H]⁺calcd = 128.0818, [M + H]⁺found = 128.1116; anal. calcd. (%) for C₅H₉N₃O: C, 47.23 H, 7.13 N, 33.05; found: C, 47.27 H, 7.13 N, 33.07; IR (neat, v cm⁻¹): 3268, 3116, 2971, 1508, 1434, 1375, 1274, 1210, 1188, 1134, 1077, 1012, 938, 844, 752, 678, 638, 472; UV–vis: λ_{max} = 201 nm; GC: t_R = 4.379; HPLC [hexane–iPrOH (9:1); f = 0.8]: t_R = 14.716, 17.830.

1-(1H-Imidazol-1-yl)propan-2-yl acetate (±)-4a: To a solution of 1-(1H-imidazol-1yl)propan-2-ol (±)-3a (0.3 g; 2.38 mmol) in 2-methyl-2-butanol (2.5 ml), vinyl acetate (1.02 g; 11.89 mmol; 1.1 ml) and Novozym SP 435 (0.1 g) were added. The reaction mixture was stirred at room temperature with a magnetic stirrer for 48 h. Aliquots were regularly analyzed by GC until almost 100% substrate conversion was reached. The enzyme was removed by filtration, and the solution was evaporated to dryness. The crude product was purified by chromatography [CHCl3-MeOH (9:1)] to afford a yellowish oil (0.36 g; 2.14 mmol; yield = 90%). $R_f = 0.55$ [CHCl₃-MeOH (9:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.14 (d, J = 6.24 Hz, 3H, CH_3 CH(OH)CH₂), 1.94 (s, 3H, $CH_3C=O$), 3.90–4.03 (m, 2H, HOCHC H_2N^1), 5.01–5.05 (m, 1H, HOC H_2N^1), 6.85 (s, 1H, $CH_2N^1CH=CHN^3$), 6.97 (s, 1H, $CH_2N^1CH=CHN^3$), 7.43 (s, 1H, $CH_2N^1CH=N^3$); ¹³C NMR (CDCl₃, 100 MHz) δ 16.9 (CH₃CH(OH)CH₂), 20.8 (CH₃C=O), 50.73 (HOCHCH₂N¹), (HOCHCH₂N¹), $(CH_2N^1CH=CHN^3)$, 68.9 119.4 128.8 $(CH_2N^1CH=CHN^3)$, 137.4 $(CH_2N^1CH=N^3)$, 169.8 (C=O); HRMS $(ESI^+, m/z)$: [M + $H_{calcd}^{\dagger} = 169.0971$, $[M + H]_{found}^{\dagger} = 169.1240$; anal. calcd. (%) for $C_8H_{12}N_2O_2$: C, 57.13 H, 7.19 N, 16.66; found: C, 57.10 H, 7.20 N, 16.63; UV-vis: $\lambda_{max} = 201$ nm; GC: $t_R =$ 6.735; HPLC [hexane-iPrOH (9:1); f = 0.8]: $t_R = 32.938$, 39.637.

1-(1*H*-1,2,4-Triazol-1-yl)propan-2-yl acetate (±)-4b: To the solution of 1-(1*H*-1,2,4-triazol-1-yl)propan-2-ol (±)-3b (0.3 g; 2.36 mmol) in 2-methyl-2-butanol (2.5 ml), vinyl acetate (1.02 g; 11.79 mmol; 1.1 ml) and Novozym SP 435 (0.1 g) were added. The reaction mixture was stirred at room temperature with a magnetic stirrer for 48 h. Aliquots were regularly analyzed by GC until almost 100% substrate conversion was reached. The reaction was stopped by draining of the enzyme. After solvent evaporation the crude product was purified by chromatography [CHCl₃–MeOH (9:1)]

to yield product as a yellowish oil (0.32 g; 1.9 mmol; yield = 81%). $R_f = 0.64$ [CHCl₃–MeOH (9:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.14 (d, J = 6.46 Hz, 3H, C H_3 CH(OH)CH₂), 1.87 (s, 3H, C H_3 C=O), 4.11–4.26 (m, 2H, HOCHC H_2 N¹), 5.08–5.13 (m, 1H, HOCHCH₂N¹), 7.80 (s, 1H, CH₂N¹N²=CHN⁴), 7.99 (s, 1H, CH₂N¹CH=N⁴); ¹³C NMR (CDCl₃, 100 MHz) δ 17.0 (CH₃CH(OH)CH₂), 20.7 (CH₃C=O), 53.1 (HOCHCH₂N¹), 68.3 (HOCHCH₂N¹), 143.5 (CH₂N¹CH=N⁴), 151.6 (CH₂N¹N²=CHN⁴), 169.6 (C=O); HRMS (ESI⁺, m/z): [M + H]⁺_{calcd} = 170.0924, [M + H]⁺_{found} = 170.1215; anal. calcd. (%) for C₇H₁₁N₃O₂: C, 49.70 H, 6.55 N, 24.84; found: C, 49.67 H, 6.55 N, 24.86; UV-vis: λ_{max} = 201 nm; GC: t_R = 5.258; HPLC [hexane–iPrOH (9:1); t_R = 19.231, 24.214.

General procedure for the lipase-catalyzed kinetic resolution of racemic alcohols (±)-3a and (±)-3b

In a typical experiment, the reaction mixture was composed of 0.3 g of the appropriate racemic alcohol (±)-3a or (±)-3b, 5-fold molar excess of vinyl acetate, 3 ml of organic solvent, and 50–100 mg of lipase (depending of the enzyme used – see Table 1 and 2). The mixture was agitated (250 rpm) at room temperature. Conversion rate was measured by GC. The reaction was stopped by filtering off the enzyme. To the remaining lipase the same organic solvent (3 ml) as used in the reaction was added and suspension was stirred for a minimum of 10 min. Then, the enzyme was filtered off via a funnel stocked with cotton and washed with methanol (3 ml) in order to elute residuals of the products. The crude product was evaporated to dryness and purified by chromatography using a mixture of CHCl₃–MeOH (9:1) as eluent (details are given in Tables 1 and 2; physical, spectroscopic and analytical

data are identical as for the racemic standard compounds). The specific rotations were measured in CHCl₃ solution for the enantioenriched alcohols and esters, and are as follows:

$$(S)$$
-(+)-**5a**: $[\alpha]_D^{29}$ = +35.2 (*c* 1.0, CHCl₃) for ee = 98%

$$(R)$$
-(+)-6a: $[\alpha]_D^{29}$ = +9.2 (c 1.0, CHCl₃) for ee > 99%

(S)-(+)-**5b**:
$$[\alpha]_D^{29}$$
 = +44 (*c* 1.0, CHCl₃) for ee = 98%

$$(R)$$
-(+)-**6b**: $[\alpha]_D^{29}$ = +5.2 (c 1.0, CHCl₃) for ee = 90%

General procedure for the determination of the absolute configuration of (+)-5a (+)-5b

Esterification of (*S*)-(+)-1-(1*H*-imidazol-1-yl)propan-2-ol (+)-5a with (*R*)- or (*S*)-α-methoxy-α-phenylacetic acid. A catalytic amount of DMAP (5 mg) was added to a solution of (*S*)-(+)-1-(1*H*-imidazol-1-yl)propan-2-ol (+)-5a (50 mg; 0.4 mmol), (*R*)- or (*S*)-α-methoxy-α-phenylacetic acid (66 mg; 0.4 mmol), as appropriate, and DCC (98 mg; 0.48 mmol) in dry CH_2CI_2 (2.5 ml). After 24 h of stirring at rt, precipitated dicyclohexylurea was removed by filtration and then the urea cake was rinsed with toluene (3 × 5 ml). The combined toluene solution was evaporated in vacuum. Crude product as a yellow oil was purified by preparative chromatographic plate with a mixture of hexane—ethyl acetate (1:1). Appropriately, the separated fraction was removed from the plate together with silica gel, placed in a round-bottomed flask, and stirred with 10 ml of PhCH₃–AcOEt (8:2) for over 1 h. Finally, silica gel was filtered off, the fraction was rinsed with PhCH₃ (2 × 2.5 ml), and the solvents were removed under reduced pressure to afford the pure product.

(S)-((S)-1-(1*H*-Imidazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (9a): Yellow oil; yield 68%; $R_f = 0.09$ [hexane—ethyl acetate (1:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.19 (d, J = 6.32 Hz, 3H, C H_3 CH(OH)CH₂), 3.36 (s, 3H, OC H_3), 3.85–3.97 (m, 2H, MPAOCHC H_2 N¹), 4.72 (s, 1H, CHOCH₃), 5.14–5.24 (m, 1H, MPAOCHCH₂N¹), 6.45 (s, 1H, CH₂N¹CH=N³), 6.88 (s, 1H, CH₂N¹CH=CHN³), 7.07 (s, 1H, CH₂N¹CH=CHN³), 7.36–7.44 (m, 5H, Ph); ¹³C NMR (CDCl₃, 100 MHz) δ 16.9 (CH₃CH(OMPA)CH₂), 50.7 (MPAOCHCH₂N¹), 57.2 (OCH₃), 70.0 (MPAOCHCH₂N¹), 82.4 (CHOCH₃), 119.6 (CH₂N¹CH=CHN³), 127.3 (2C, Ph), 128.8 (CH₂N¹CH=CHN³), 129.0 (2C, Ph), 129.2 (Ph), 135.8 (Ph), 137.4 (CH₂N¹CH=N³), 169.8 (C=O).

((R)-((S)-1-(1H-Imidazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (10a): Yellow oil; yield 74%; $R_f = 0.09$ [hexane-ethyl acetate (1:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.10 (d, J = 6.32 Hz, 3H, $CH_3CH(OMPA)CH_2$), 3.35 (s, 3H, OCH_3), 3.95– 4.09 (m, 2H, MPAOCHC H_2N^1), 4.71 (s, 1H, CHOCH₃), 5.10–5.22 (m, 1 H, MPAOCHCH₂N¹), 6.82 (s, 1H, CH₂N¹CH=CHN³), 7.01 (s, 1H, CH₂N¹CH=CHN³), 7.32-7.38 (m, 5H, Ph), 7.39 (s, 1H, $CH_2N^1CH=N^3$); ^{13}C NMR (CDCl₃, 100 MHz) δ 16.8 $(CH_3CH(OMPA)CH_2),$ 50.8 (MPAOCH CH_2N^1), 57.4 (OCH₃),70.1 (MPAOCHCH₂N¹), 82.5 (CHOCH₃), 119.5 (CH₂N¹CH=CHN³), 126.9 (2C, Ph), 128.7 (2C, Ph), 128.9 (Ph), 129.1 $(CH_2N^1CH=CHN^3)$, 135.7 (Ph), 137.6 $(CH_2N^1CH=N^3)$, 169.9 (C=O).

Esterification of (*S*)-(+)-1-(1*H*-1,2,4-triazol-1-yl)propan-2-ol (+)-5b with (*R*)- or (*S*)- α -methoxy- α -phenylacetic acid. A catalytic amount of DMAP (5 mg) was added to a solution of (*S*)-(+)-1-(1*H*-1,2,4-triazol-1-yl)propan-2-ol (+)-5b (50 mg; 0.39 mmol), (*R*)- or (*S*)- α -methoxy- α -phenylacetic acid (66 mg; 0.39 mmol), as appropriate, and

DCC (97.4 mg; 0.47 mmol) in dry methylene chloride (2.5 ml). After 24 h of stirring at room temp., precipitated dicyclohexylurea was removed by filtration and then the urea cake was rinsed with toluene (3 \times 5 ml). The combined organic solutions were washed with cold 1 M HCl (2 \times 1 ml), saturated NaHCO₃ (2 \times 1 ml), and saturated NaCl (1 \times 1 ml). Then the organic layer was dried over MgSO₄ and filtered, and the solvent was evaporated under reduced pressure. The crude product was purified by preparative plate chromatography using hexane—ethyl acetate (1:1) mixture.

(S)-((S)-1-(1*H*-1,2,4-Triazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (9b): Yellow oil; yield 63%; $R_f = 0.09$ [hexane–ethyl acetate (1:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.25 (d, J = 6.55 Hz, 3H, $CH_3CH(OMPA)CH_2$), 3.34 (s, 3H, OCH_3), 4.16 (d, J = 4.52 Hz, 2H, MPAOCHC H_2N^1), 4.69 (s, 1H, $CHOCH_3$), 5.23–5.36 (m, 1H, MPAOC HCH_2N^1), 7.28 (s, 1H, $CH_2N^1CH=N^4$), 7.32–7.47 (m, 5H, Ph), 7.79 (s, 1H, $CH_2N^1N^2=CHN^4$); ¹³C NMR (CDCl₃, 100 MHz) δ 17.1 ($CH_3CH(OMPA)CH_2$), 53.1 (MPAOCH CH_2N^1), 57.2 (OCH_3), 69.3 (MPAOCH CH_2N^1), 82.3 ($CHOCH_3$), 126.9 ($CH_2N^1CH=N^4$), 127.2 (Ph), 128.9 (2C, Ph), 129.2 (2C, Ph), 135.8 (Ph), 151.6 ($CH_2N^1N^2=CHN^4$), 169.6 (C=O).

(*R*)-((*S*)-1-(1*H*-1,2,4-Triazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (10b): Yellow oil; yield 59%; $R_f = 0.09$ [hexane—ethyl acetate (1:1)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.17 (d, J = 6.32 Hz, 3H, C H_3 CH(OMPA)CH₂), 3.34 (s, 3H, OC H_3), 4.18–4.34 (m, 2H, MPAOCHC H_2 N¹), 4.69 (s, 1H, C H_3 OCH₃), 5.26–5.30 (m, 1H, MPAOC H_3 CH₂N¹), 7.28–7.41 (m, 5H, Ph), 7.89 (d, J = 5.87 Hz, 2H, CH₂N¹N²=C H_3 C and CH₂N¹C $H=N^4$); ¹³C NMR (CDCl₃, 100 MHz) δ 16.9 (C_3 CH(OMPA)CH₂), 53.2 (MPAOCH C_3 CH₂N¹), 57.3 (O C_3 H₃), 69.4 (MPAOCHCH₂N¹), 82.4 (C_3 HOCH₃), 126.8 (Ph),

128.7 (2C, Ph), 128.8 (2C, Ph), 135.7 (Ph), 143.8 ($CH_2N^1CH=N^4$), 151.9 ($CH_2N^1N^2=CHN^4$), 169.7 (C=O).

General methods for the preparation of chiral ionic liquids

Synthesis and characterization of optically active imidazolium salts [(+)-7a-h]. Optically active (S)-(+)-1-(1H-imidazol-1-yl)propan-2-ol (+)-5a (0.5 g; 3.96 mmol) was dissolved in dry CH₃CN (5 ml), and subsequently a 3-fold molar excess of the appropriate freshly distilled alkyl bromide or iodide was added. The reaction mixture was stirred under the conditions given in Table 3. The reaction progress was monitored by TLC with a chloroform–methanol (8:2) mixture as the eluent. The reaction mixture was cooled to room temperature, and the separated salt was washed with Et₂O (3 × 5 ml) and n-hexane (5 ml) to afford a dark oil, which was sequentially purified from tars with a column filled with activated carbon. The resulting product was a colored oil.

(*S*)-(+)-1-(2-Hydroxypropyl)-3-propyl-1*H*-imidazol-3-ium bromide (+)-7a: White oil; yield 92%; $R_f = 0.26$ [CHCl₃–MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.97 (t, J = 7.34 Hz, 3H, CH₂CH₃), 1.25 (d, J = 6.32 Hz, 3H, CH₃CH(OH)CH₂), 1.89–2.01 (m, 2H, CH₂CH₃), 4.13–4.26 (m, 2H, +N³CH₂CH₂CH₃), 4.28–4.36 (m, 2H, HOCHCH₂N¹), 4.50 (m, 1H, HOCHCH₂N¹), 7.40 (t, J = 1.58 Hz, 1H, CH₂N¹CH=CHN³), 7.64 (t, J = 1.47 Hz, 1H, CH₂N¹CH=CHN³), 9.74 (s, 1H, CH₂N¹CH=N³); ¹³C NMR (CDCl₃, 100 MHz) δ 10.7 (CH₂CH₃), 20.3 (CH₂CH₃), 23.4 (CH₃CH(OH)CH₂), 51.5 (+N³CH₂CH₂CH₃), 56.1 (HOCHCH₂N¹), 65.5 (HOCHCH₂N¹), 121.2 (CH₂N¹CH=CHN³), 123.6 (CH₂N¹CH=CHN³), 136.7 (CH₂N¹CH=N³); HRMS (ESI⁺,

m/z): [M⁺]_{calcd} = 169.1329, [M⁺]_{found} = 169.4186; anal. calcd. (%) for C₉H₁₇BrN₂O: C, 43.39 H, 6.88 N, 11.24; found: C, 43.34 H, 6.89 N, 11.21; IR (neat, v cm⁻¹): 3233, 3070, 2968, 1563, 1451, 1375, 1274, 1163, 1135, 1072, 937, 846, 782, 753, 627, 472; [α]_D²⁹ = +23.6 (c 1.0, CHCl₃) for ee = 98%.

(S)-(+)-1-(2-Hydroxypropyl)-3-(prop-2-en-1-yl)-1*H*-imidazol-3-ium bromide (+)-7b: Dark brown oil; yield 93%; $R_f = 0.31$ [CHCl₃–MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.24 (d, J = 6.32 Hz, 3H, $CH_3CH(OH)CH_2$), 4.13–4.22 (m, 1H, $HOCHCH_2N^1$), 4.23– 4.31 (m, 1H, HOCHC H_2N^1), 4.47 (dd, J = 13.55, 2.71 Hz, 1H, HOCHC H_2N^1), 4.92 (d, $J = 6.32 \text{ Hz}, 2H, {}^{+}N^{3}CH_{2}CH_{a} = CH_{a}H_{b}), 5.39 - 5.50 \text{ (m, 2H, } {}^{+}N^{3}CH_{2}CH_{a} = CH_{a}H_{b}), 5.95 -$ 6.09 (m, 1H, ${}^{+}N^{3}CH_{2}CH_{a}=CH_{a}H_{b}$), 7.37 (t, J=1.81 Hz, 1H, $CH_{2}N^{1}CH=CHN^{3}$), 7.66 (t, $J = 1.81 \text{ Hz}, 1\text{H}, \text{CH}_2\text{N}^1\text{CH} = \text{C}H\text{N}^3), 9.65 - 9.70 \text{ (m, 1H, CH}_2\text{N}^1\text{C}H = \text{N}^3); ^{13}\text{C NMR}$ (CDCl₃, 100 MHz) δ 20.3 (CH₃CH(OH)CH₂), 52.1 (${}^{\dagger}N^{3}$ CH₂CH=CH₂), 56.3 (HOCHCH₂N¹), (HOCHCH₂N¹), 121.2 (CH₂N¹CH=CHN³),65.4 122.6 $(^{\dagger}N^{3}CH_{2}CH=CH_{2}), 123.7 (CH_{2}N^{1}CH=CHN^{3}), 129.7 (^{\dagger}N^{3}CH_{2}CH=CH_{2}),$ 136.6 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 167.1173$, $[M^+]_{found} = 167.0548$; anal. calcd. (%) for C₉H₁₅BrN₂O: C, 43.74 H, 6.12 N, 11.34; found: C, 43.69 H, 6.11 N, 11.37; IR (neat, v cm⁻¹): 3297, 3072, 2970, 1560, 1444, 1423, 1161, 1135, 1078, 994, 938, 840, 759, 623, 579, 471; $[\alpha]_D^{29} = +25.3$ (c 1.0, CHCl₃) for ee = 98%.

(*S*)-(+)-3-Butyl-1-(2-hydroxypropyl)-1*H*-imidazol-3-ium bromide (+)-7c: Yellowish oil; yield 94%; $R_f = 0.37$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.76 (t, J = 7.34 Hz, 3H, CH₂CH₃), 1.07 (d, J = 6.32 Hz, 3H, CH₃CH(OH)CH₂), 1.13–1.25 (m, 2H, CH₂CH₃), 1.67–1.79 (m, 2H, CH₂CH₂CH₂CH₃), 3.96–4.05 (m, 1H, HOC*H*CH₂N¹), 4.07–4.21 (m, 2H, $^{+}$ N³CH₂CH₂CH₂CH₃), 4.29 (s, 1H, HOCHCH₂N¹), 4.37 (dd, J = 0.00

13.55, 2.94 Hz, 1H, HOCHC H_2N^1), 7.37 (t, J = 1.81 Hz, 1H, $CH_2N^1CH = CHN^3$), 7.60 $(t, J = 1.69 \text{ Hz}, 1H, CH_2N^1CH = CHN^3), 9.50 \text{ (s, } 1H, CH_2N^1CH = N^3); ^{13}C \text{ NMR (CDCl}_3,$ 100 MHz) δ 13.0 (CH₂CH₃), 19.0 (CH₃CH(OH)CH₂), 19.8 (CH₂CH₃), 31.5 (⁺N³CH₂CH₂CH₂CH₃), (CH₂CH₂CH₂CH₃),49.3 55.5 (HOCHCH₂N¹), 65.1 121.1 (CH₂N¹CH=CHN³), 123.4 (CH₂N¹CH=CHN³), (HOCHCH₂N¹), 135.9 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 183.1486$, $[M^+]_{found} = 183.5657$; anal. calcd. (%) for C₁₀H₁₉BrN₂O: C, 45.64 H, 7.28 N, 10.64; found: C, 45.60 H, 7.30 N, 10.61; IR (neat, v cm⁻¹): 3294, 3068, 2962, 1562, 1460, 1372, 1164, 1137, 1078, 948, 841, 753, 623, 470; $[\alpha]_D^{29} = +17.2$ (c 1.0, CHCl₃) for ee = 98%.

(S)-(+)-1-(2-Hydroxypropyl)-3-pentyl-1H-imidazol-3-ium bromide (+)-7d: Red oil; yield 82%; $R_f = 0.43$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.72 (t, J =6.99 Hz, 3H, CH_2CH_3), 1.10 (d, J = 6.32 Hz, 3H, $CH_3CH(OH)CH_2$), 1.13–1.21 (m, 4H, ⁺N³CH₂CH₂CH₂CH₂CH₃), 1.74–1.78 (m, 2H, ⁺N³CH₂CH₂CH₂CH₂CH₃), 3.99–4.23 (m, 4H, ${}^{+}N^{3}CH_{2}CH_{2}CH_{2}CH_{3}$, $HOCHCH_{2}N^{1}$ and $HOCHCH_{2}N^{1}$), 4.39 (dd, J = 13.66, 2.82 Hz, 1H, HOCHC H_2N^1), 4.77 (br. s., 1H, OH), 7.36 (d, J = 1.58 Hz, 1H, $CH_2N^1CH=CHN^3$), 7.63 (d, J=1.35 Hz, 1H, $CH_2N^1CH=CHN^3$), 9.52 (s, 1H, $CH_2N^1CH=N^3$); ^{13}C NMR (CDCl₃, 100 MHz) δ 13.43 (CH₂CH₃), 19.9 (CH₃CH(OH)CH₂). 21.6 (CH₂CH₃),27.8 ([†]N³CH₂CH₂CH₂CH₂CH₃), 29.4 ([†]N³CH₂CH₂CH₂CH₂CH₃), 49.6 ([†]N³CH₂CH₂CH₂CH₂CH₃), 55.6 (HOCHCH₂N¹), 65.2 $(CH_2N^1CH=CHN^3)$, 123.5 $(CH_2N^1CH=CHN^3)$, $(HOCHCH_2N^1), 121.0$ 135.9 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 197.1643$, $[M^+]_{found} = 197.0844$; anal. calcd. (%) for C₁₁H₂₁BrN₂O: C, 47.66 H, 7.64 N, 10.11; found: C, 47.61 H, 7.66 N, 10.15; IR (neat, v cm⁻¹): 3298, 3071, 2959, 2931, 1562, 1457, 1373, 1163, 1137, 1078, 937, 841, 747, 623, 473; $[\alpha]_D^{29} = +15.4$ (c 1.0, CHCl₃) for ee = 98%.

(S)-(+)-3-Heptyl-1-(2-hydroxypropyl)-1*H*-imidazol-3-ium iodide (+)-7e: Cherry-red oil; yield 92%; $R_f = 0.52$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.64 (t, J = 6.44 Hz, 3H, CH_2CH_3), 0.94–1.22 (m, 11H, ${}^{+}N^{3}CH_2CH_2CH_2CH_2CH_2CH_2CH_3$ and $CH_3CH(OH)CH_2$), 1.70–1.74 (m, 2H, ${}^+N^3CH_2CH_2CH_2CH_2CH_2CH_2CH_3$), 3.88 (br. s., OH), 3.98-4.18 (m, 4H, $HOCHCH_2N^1$ one of $HOCHCH_2N^1$ $^{+}N^{3}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}$), 4.36 (d, J = 11.29 Hz, 1H, one of $HOCHCH_{2}N^{1}$), 7.32 (s, 1H, $CH_2N^1CH=CHN^3$), 7.55 (s, 1H, $CH_2N^1CH=CHN^3$), 9.26 (s, 1H, $CH_2N^1CH=N^3$); ¹³C NMR (CDCI₃, 100 MHz) δ 13.6 (CH₂CH₃), 19.9 (CH₃CH(OH)CH₂), 22.0 $(CH_2CH_3),$ ([†]N³CH₂CH₂CH₂CH₂CH₂CH₂CH₃), 25.7 28.1 (⁺N³CH₂CH₂CH₂CH₂CH₂CH₂CH₃), ([†]N³CH₂CH₂CH₂CH₂CH₂CH₂CH₃), 29.6 31.0 $(^{\dagger}N^{3}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}), 49.8 (^{\dagger}N^{3}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}),$ 55.7 (HOCHCH₂N¹),(HOCHCH₂N¹),65.1 121.2 $(CH_2N^1CH=CHN^3)$, 123.4 $(CH_2N^1CH=CHN^3)$, 135.4 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 225.1955$, $[M^+]_{found}$ = 225.0930; anal. calcd. (%) for $C_{13}H_{25}IN_2O$: C, 44.33 H, 7.15 N, 7.95; found: C, 44.37 H, 7.14 N, 7.99; IR (neat, v cm⁻¹): 3337, 3080, 2926, 2856, 1561, 1457, 1374, 1162, 1136, 1092, 1076, 936, 838, 745, 635, 473; $[\alpha]_D^{29} = +16.3$ (c 1.0, CHCl₃) for ee = 98%.

 7.58 (t, J = 1.69 Hz, 1H, $CH_2N^1CH=CHN^3$), 9.29 (s, 1H, $CH_2N^1CH=N^3$); ¹³C NMR (CDCl₃, 100 MHz) δ 13.35 (CH_2CH_3), 19.7 ($CH_3CH(OH)CH_2$), 21.7 (CH_2), 25.4 (CH_2), 27.4 (CH_2), 27.9 (CH_2), 29.3 (CH_2), 29.7 (CH_2), 30.8 (CH_2), 32.8 (CH_2), 49.5 ($^{\dagger}N^3CH_2$), 55.4 (HOCH CH_2N^1), 64.9 (HO $CHCH_2N^1$), 121.0 ($CH_2N^1CH=CHN^3$), 123.2 ($CH_2N^1CH=CHN^3$), 135.2 ($CH_2N^1CH=N^3$); HRMS (ESI⁺, m/z): [M^+]_{calcd} = 267.2425, [M^+]_{found} = 267.1200; anal. calcd. (%) for $C_{16}H_{31}IN_2O$: C, 48.73 H, 7.92 N, 7.10; found: C, 48.76 H, 7.91 N, 7.12; IR (neat, v cm⁻¹): 3333, 3081, 2922, 2853, 1561, 1458, 1376, 1163, 1137, 1076, 936, 745, 635, 472; [α]_D²⁹ = +15.8 (α 1.0, α) CHCl₃) for ee = 98%.

(S)-(+)-3-Dodecyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7g: Cherryred oil; yield 86%; $R_f = 0.51$ [CHCl₃: MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.83 (t, 6.59 Hz, 3H, CH_2CH_3), 1.15-1.25 (m, 18H, $CH_3CH(OH)CH_2$), 1.81–2.02 (m, 2H, ${}^+N^3CH_2CH_2$), 3.78 (d, J = 4.97 Hz, 1H, CH), 4.12–4.35 (m, 3H, CH_2), 4.46 (d, J = 11.07 Hz, 1H, CH), 7.35 (t, J = 1.81 Hz, 1H, $CH_2N^1CH=CHN^3$), 7.59 (t, J=1.69 Hz, 1H, $CH_2N^1CH=CHN^3$), 9.21 (s, 1H, $CH_2N^1CH=N^3$); ¹³C NMR (CDCl₃, 100 Hz) δ 14.0 (CH₂CH₃), 20.4 (CH₃CH(OH)CH₂), 22.5 (CH₂CH₃), 26.2 (CH₂), 28.3 (CH₂), 28.8 (CH₂), 29.1 (CH₂), 29.4 (2C, CH₂), 29.9 (CH_2) , 30.3 (CH_2) , 31.7 (CH_2) , 50.5 $({}^{\dagger}N^3CH_2)$, 56.4 $(HOCHCH_2N^1)$, 65.5 $(CH_2N^1CH=CHN^3)$, 123.8 $(CH_2N^1CH=CHN^3)$, $(HOCHCH_2N^1), 121.6$ 135.8 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 295.2738$, $[M^+]_{found} = 295.8576$; anal. calcd. (%) for C₁₈H₃₅IN₂O: C, 51.18 H, 8.35 N, 6.63; found: C, 51.22 H, 8.32 N, 6.67; IR (neat, v cm⁻¹): 3332, 2921, 2852, 1560, 1466, 1376, 1162, 1091, 1075, 936, 832, 745, 622, 472; $[\alpha]_D^{29} = +14.2$ (c 1.0, CHCl₃) for ee = 98%.

(S)-(+)-3-Hexadecyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7h: Colorless oil; yield 81%; $R_f = 0.58$ [CHCl₃: MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.82 (t, J = 6.78 Hz, 3H, CH₂CH₃), 1.17–1.31 (m, 29H, ${}^{+}N^{3}$ CH₂CH₂ (CH₂)₁₃CH₃ and $CH_3CH(OH)CH_2$), 1.80–1.94 (m, 2H, ${}^+N^3CH_2CH_2$), 4.15–4.29 (m, 4H, HOCHC H_2N^1 and ${}^{+}N^{3}CH_{2}$), 4.43 (s, 1H, HOCHCH₂N¹), 7.33 (t, J = 1.69 Hz, 1H, CH₂N¹CH=CHN³), 7.59 (t, J = 1.58 Hz, 1H, $CH_2N^1CH=CHN^3$), 9.39 (s, 1H, $CH_2N^1CH=N^3$); ¹³C NMR (CDCl₃, 100 Hz) δ 14.0 (CH₂CH₃), 20.1 (CH₃CH(OH)CH₂), 22.5 (CH₂CH₃), 26.1 (CH₂), 28.8 (CH₂), 29.2 (CH₂), 29.2 (CH₂), 29.4 (CH₂), 29.5 (CH₂), 29.5 (CH₂), 29.5 (CH_2) , 29.5 (3C, CH_2), 29.9 (CH_2) , 31.7 (CH_2) , 50.2 $({}^{\dagger}N^3CH_2)$, 56.2 $(HOCHCH_2N^1)$, 65.4 (HOCHCH₂N¹), 121.3 (CH₂N¹CH=CHN³), 123.6 (CH₂N¹CH=CHN³), 136.0 $(CH_2N^1CH=N^3)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 351.3364$, $[M^+]_{found} = 351.2940$; anal. calcd. (%) for C₂₂H₄₃IN₂O: C, 55.22 H, 9.06 N, 5.85; found: C, 55.17 H, 9.08 N, 5.81; IR (neat, v cm⁻¹): 3328, 2914, 2850, 1561, 1473, 1374, 1164, 1136, 1021, 936, 839, 749, 715, 626, 472; $[\alpha]_D^{29} = +11.4$ (c 1.0, CHCl₃) for ee = 98%.

Synthesis and characterization of optically active triazolium salts [(+)-8a-f]. Optically active (S)-(+)-1-(1H-1,2,4-triazol-1-yl)propan-2-ol (+)-5b (0.5 g; 3.93 mmol) was dissolved in dry CH₃CN (5 ml), and a 3-fold molar excess of the appropriate freshly distilled alkyl bromide or iodide was added. The reaction mixture was stirred under the conditions given in Table 4. The reaction progress was monitored by TLC using a solution of chloroform–methanol (8:2) as the eluent. After stirring had been stopped and the mixture cooled to 0–5 °C, the separated salt was filtered off, and washed with Et₂O (3 × 5 ml) and toluene (5 ml). Next, the solvents were removed under reduced pressure to yield the product as a viscous liquid (gum) or sticky oil.

(*S*)-(+)-1-(2-Hydroxypropyl)-4-propyl-1*H*-1,2,4-triazol-4-ium bromide (+)-8a: White gum; yield 98%; $R_f = 0.31$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.94 (t, J = 7.38 Hz, 3H, CH₂CH₃), 1.23 (d, J = 6.43 Hz, 3H, CH₃CH(OH)CH₂), 1.97 (m, 2H, CH₂CH₃), 4.06–4.25 (m, 1H, HOC*H*CH₂N¹), 4.27–4.53 (m, 4H, HOCHCH₂N¹ and ${}^{\dagger}N^4$ CH₂), 8.99 (s, 1H, CH₂N¹N²=C*H*N⁴), 10.62 (s, 1H, CH₂N¹C*H*=N⁴); ¹³C NMR (CDCl₃, 100 MHz) δ 10.6 (CH₂CH₃), 20.0 (CH₂CH₃), 23.0 (CH₃CH(OH)CH₂), 49.9 (${}^{\dagger}N^4$ CH₂), 59.3 (HOCHCH₂N¹), 64.6 (HOCHCH₂N¹), 142.8 (CH₂N¹N²=CHN⁴), 143.7 (CH₂N¹CH=N⁴); HRMS (ESI[†], m/z): [M[†]]_{calcd} = 170.1282, [M[†]]_{found} = 170.1560; anal. calcd. (%) for C₈H₁₆BrN₃O: C, 38.41 H, 6.45 N, 16.80; found: C, 38.36 H, 6.46 N, 16.85; IR (neat, v cm⁻¹): 3308, 3038, 2971, 1577, 1446, 1374, 1272, 1159, 1137, 1067, 999, 936, 909, 843, 808, 626, 583, 478; [α]_D²⁹ = +18.8 (*c* 1.0, CHCl₃) for ee = 98%.

(S)-(+)-1-(2-Hydroxypropyl)-4-(prop-2-en-1-yl)-1*H*-1,2,4-triazol-4-ium bromide (+)-8b: Brown gum; yield 94%; $R_f = 0.18$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 1.23 (d, J = 6.43 Hz, 3H, $CH_3CH(OH)CH_2$), 4.12–4.23 (m, 1H, HOC*H*CH₂N¹), 4.33–4.48 (m, 2H, HOCHCH₂N¹), 4.82 (br. s., 1H, O*H*), 5.10 (d, J = 6.67 Hz, 2H, [†]N⁴C*H*₂), 5.44 (dd, J = 10.12, 0.83 Hz, 1H, [†]N⁴CH₂CH_a=C*H*_aH_b), 5.54 (dd, J = 16.90, 0.71 Hz, 1H, [†]N⁴CH₂CH_a=CH_aH_b), 6.10 (ddt, J = 16.96, 10.18, 6.55, 6.55 Hz, 1H, [†]N⁴CH₂C*H*_a=CH_aH_b), 8.89 (s, 1H, CH₂N¹N²=C*H*N⁴), 10.58 (s, 1H, CH₂N¹C*H*=N⁴); ¹³C NMR (CDCl₃, 100 MHz) δ 20.0 (*C*H₃CH(OH)CH₂), 50.6 ([†]N⁴CH₂), 59.4 (HOCHCH₂N¹), 64.6 (HOCHCH₂N¹), 123.6 ([†]N⁴CH₂CH=CH₂), 128.8 ([†]N⁴CH₂CH=CH₂), 142.8 (CH₂N¹N²=CHN⁴), 143.4 (CH₂N¹CH=N⁴); HRMS (ESI[†], *m/z*): [M[†]]_{calcd} = 168.1126, [M[†]]_{found} = 168.1497; anal. calcd. (%) for C₈H₁₄BrN₃O: C,

38.73 H, 5.69 N, 16.94; found: C, 38.75 H, 5.68 N, 16.92; IR (neat, v cm⁻¹): 3308, 3039, 2979, 1575, 1443, 1385, 1274, 1243, 1151, 1135, 1078, 988, 944, 842, 780, 626, 578, 479; $[\alpha]_D^{29} = +15.6$ (*c* 1.0, CHCl₃) for ee = 98%.

(S)-(+)-4-Butyl-1-(2-hydroxypropyl)-1*H*-1,2,4-triazol-4-ium bromide (+)-8c: White gum; yield 90%; $R_f = 0.35$ [CHCl₃–MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.88 (t, J = 7.26 Hz, 3H, CH₂CH₃), 1.21 (d, J = 6.43 Hz, 3H, CH₃CH(OH)CH₂), 1.28–1.39 (m, 2H, CH₂CH₃), 1.84–1.95 (m, 2H, ⁺N⁴CH₂CH₂CH₂CH₃), 4.13–4.22 (m, 1H, HOC*H*CH₂N¹), 4.33–4.49 (m, 4H, HOCHCH₂N¹ and ⁺N⁴CH₂), 4.67 (br. s., 1H, O*H*), 8.99 (s, 1H, CH₂N¹N²=C*H*N⁴), 10.63 (s, 1H, CH₂N¹C*H*=N⁴); ¹³C NMR (CDCl₃, 100 MHz) δ 13.2 (CH₂CH₃), 19.3 (CH₃CH(OH)CH₂), 20.0 (CH₂CH₃), 31.3 (⁺N⁴CH₂CH₂CH₂CH₃), 48.3 (⁺N⁴CH₂), 59.2 (HOCHCH₂N¹), 64.5 (HOCHCH₂N¹), 142.8 (CH₂N¹N²=CHN⁴), 143.6 (CH₂N¹CH=N⁴); HRMS (ESI⁺, *m/z*): [M⁺]_{calcd} = 184.1439, [M⁺]_{found} = 184.1733; anal. calcd. (%) for C₉H₁₈BrN₃O: C, 40.92 H, 6.87 N, 15.91; found: C, 40.89 H, 6.87 N, 15.93; IR (neat, v cm⁻¹): 3320, 2963, 2939, 1778, 1464, 1369, 1308, 1277, 1203, 1165, 1140, 1100, 1068, 1000, 935, 843, 780, 629, 570, 478; [α]_D²⁹ = +17.7 (c 1.0, CHCl₃) for ee = 98%.

(*S*)-(+)-1-(2-Hydroxypropyl)-4-pentyl-1*H*-1,2,4-triazol-4-ium bromide (+)-8d: Brownish oil; yield 78%; $R_f = 0.40$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.78 (t, J = 6.83 Hz, 3H, CH₂CH₃), 1.17 (d, J = 6.55 Hz, 3H, CH₃CH(OH)CH₂), 1.21–1.26 (m, 4H, ⁺N⁴CH₂CH₂CH₂CH₂CH₃), 1.81–1.94 (m, 2H, ⁺N⁴CH₂CH₂CH₂CH₂CH₃), 4.13 (m, 1H, HOC*H*CH₂N¹), 4.31–4.39 (m, 4H, HOCHC*H*₂N¹ and ⁺N⁴C*H*₂), 5.03 (br. s., 1H, O*H*), 9.00 (s, 1H, CH₂N¹N²=C*H*N⁴), 10.56 (s, 1H, CH₂N¹C*H*=N⁴); ¹³C NMR (CDCl₃, 100 MHz) δ 13.5 (CH₂CH₃), 19.8 (*C*H₃CH(OH)CH₂), 21.6 (*C*H₂CH₃), 27.9

(${}^{\dagger}N^{4}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}$), 29.0 (${}^{\dagger}N^{4}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}$), 48.4 (${}^{\dagger}N^{4}CH_{2}$), 59.1 (${}^{\dagger}N^{4}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}CH_{3}$), 48.4 (${}^{\dagger}N^{4}CH_{2}$), 59.1 (${}^{\dagger}N^{4}CH_{2}N^{1}$), 64.4 (${}^{\dagger}N^{4}CH_{2}N^{1}$), 142.6 (${}^{\dagger}CH_{2}N^{1}N^{2}=CHN^{4}$), 143.6 (${}^{\dagger}CH_{2}N^{1}CH_{2}N^{4}$); HRMS (${}^{\dagger}ESI^{\dagger}$, m/z): [${}^{\dagger}M^{\dagger}$]_{calcd} = 198.1595, [${}^{\dagger}M^{\dagger}$]_{found} = 198.1956; anal. calcd. (%) for ${}^{\dagger}C_{10}H_{20}BrN_{3}O$: C, 43.17 H, 7.25 N, 15.10; found: C, 43.19 H, 7.24 N, 15.14; IR (neat, v cm⁻¹): 3328, 2958, 2931, 1576, 1458, 1376, 1160, 1136, 1075, 996, 937, 907, 846, 761, 624, 475; [${}^{\dagger}Q$]_D²⁹ = +15.4 (${}^{\dagger}C$ 1.0, ${}^{\dagger}CH_{2}$) for ee = 98%.

(S)-(+)-4-Heptyl-1-(2-hydroxypropyl)-1*H*-1,2,4-triazol-4-ium iodide (+)-8e: Amber gum; yield 87%; $R_f = 0.51$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz) δ 0.79 (t, J = 6.66 Hz, 3H, CH_2CH_3), 1.00–1.48 (m, 11H, $^{\dagger}N^4CH_2CH_2CH_2CH_2CH_2CH_2CH_3$ and $CH_3CH(OH)CH_2$), 1.77–2.06 (m, 2H, ${}^+N^4CH_2CH_2CH_2CH_2CH_2CH_2CH_3$), 4.14 (br. s., 1H, OH), 4.27 (m, 1H, HOCHCH₂N¹), 4.33–4.54 (m, 4H, HOCHCH₂N¹ and ${}^{\dagger}N^{4}CH_{2}$), 8.84 (s, 1H, $CH_2N^1N^2=CHN^4$), 10.45 (s, 1H, $CH_2N^1CH=N^4$); ¹³C NMR (CDCl₃. 100 MHz) δ 13.8 (CH₂CH₃), 20.0 (CH₃CH(OH)CH₂), 22.2 (CH₂CH₃), 25.9 $(^{+}N^{4}CH_{2}CH_{2}),$ ([†]N⁴CH₂CH₂CH₂CH₂CH₂CH₃), 28.3 29.4 ([†]N⁴CH₂CH₂CH₂CH₂CH₂CH₃), 31.2 (${}^{\dagger}N^{4}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{2}CH_{3}$), 48.8 $(^{+}N^{4}CH_{2})$, 59.3 (HOCHCH₂N¹), 64.5 (HOCHCH₂N¹), 142.4 (CH₂N¹N²=CHN⁴), 143.4 $(CH_2N^1CH=N^4)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 226.1908$, $[M^+]_{found} = 226.2362$; anal. calcd. (%) for C₁₂H₂₄IN₃O: C, 40.80 H, 6.85 N, 11.90; found: C, 40.83 H, 6.84 N, 11.92; IR (neat, v cm⁻¹): 3315, 3054, 2956, 2930, 2857, 1574, 1455, 1412, 1372, 1308, 1162, 1135, 1094, 1073, 1001, 935, 891, 846, 767, 721, 619, 526, 477; $[\alpha]_D^{29} =$ +10.2 (c 1.0, CHCl₃) for ee = 98%.

(S)-(+)-4-Decyl-1-(2-hydroxypropyl)-1H-1,2,4-triazol-4-ium iodide (+)-8f: Amber gum; yield 92%; $R_f = 0.55$ [CHCl₃-MeOH (8:2)]; ¹H NMR (CDCl₃, 400 MHz)

3H, δ 0.78 J 6.78 Hz, CH_2CH_3), 1.10-1.30 17H, (t, (m, † N 4 CH $_{2}$ CH $_{2}$ CH $_{3}$ CH $_{4}$ CH $_{5}$ CH $_{6}$ CH $_{7}$ 2H, ${}^{+}N^{4}CH_{2}CH_{2}$), 4.13–4.14 (m, 1H, HOCHCH₂N¹), 4.25 (br.s., 1H, OH), 4.32–4.51 (m, 4H, HOCHC H_2 N¹ and ⁺N⁴C H_2), 8.83 (s, 1H, CH₂N¹N²=CHN⁴), 10.42 (s, 1H, $CH_2N^1CH=N^4$); ¹³C NMR (CDCI₃, 100 MHz) δ 13.8 (CH₂CH₃), 19.9 (CH₃CH(OH)CH₂), 22.3 (CH₂), 26.0 (CH₂), 28.6 (CH₂), 28.9 (CH₂), 29.1 (CH₂), 29.1 (CH₂), 29.4 (CH₂), 31.5 (CH₂), 48.8 ($^{+}N^{4}$ CH₂), 59.3 (HOCHCH₂N¹), 64.4 (HOCHCH₂N¹), 142.3 $(CH_2N^1N^2=CHN^4)$, 143.3 $(CH_2N^1CH=N^4)$; HRMS $(ESI^+, m/z)$: $[M^+]_{calcd} = 268.2378$, $[M^+]_{found}$ = 268.2930; anal. calcd. (%) for $C_{15}H_{30}IN_3O$: C, 45.57 H, 7.65 N, 10.63; found: C, 45.55 H, 7.65 N, 10.66; IR (neat, v cm⁻¹): 3310, 3056, 2922, 2852, 1574, 1454, 1412, 1364, 1337, 1308, 1290, 1162, 1136, 1095, 1073, 1001, 935, 893, 846, 767, 721, 641, 620, 526, 463; $[\alpha]_D^{29} = +8.5$ (c 1.0, CHCl₃) for ee = 98%.

Microorganisms used

The strains were of the American Type Culture Collection (ATCC): Escherichia coli ATCC 8739, Salmonella typhimurium ATCC 14028, Pseudomonas aeruginosa ATCC 9027, Bacillus subtilis ATCC 6633, Staphylococcus aureus ATCC 6538, Candida albicans ATCC 10231, Aspergillus brasiliensis ATCC 16404. The strains received from the IHAR-PIB collection of Plant Breeding and Acclimatization Institute, Młochów: Colletotrichum coccodis MC 1, Fusarium culmorum MF 18, Fusarium oxysporum MF 5, Fusarium sambucinum MF 1, Phytophthora infestans MP 324, Phytophthora infestans MP 1320.

Antimicrobal activity assays

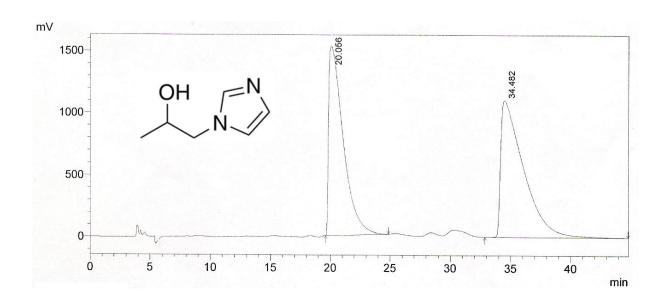
Qualitative evaluation of the antimicrobial activity was determined by the agar diffusion method according to Rebros et al. [1] with some modifications. Bacterial lawns were grown in liquid TSB (Merck) medium for 24 h. Inocula of approximately $2.5-3.5 \times 10^8$ CFU/ml (200 μ l) were spread on solid TSA (Merck) medium. Subsequently, sterile 8 mm paper discs were placed on the surface of the agar and 20 μ l of the tested ILs at the 0.025 M concentration was added. Plates were incubated for 24 h at 30 °C and the diameter of growth-inhibition zones were measured to the nearest millimeter. Each test was performed in duplicate. The ILs used in the experiment were diluted with 0.1–1% EtOH. It was verified in a separate experiment that EtOH at a concentration of 0.1–1% does not inhibit bacterial growth by itself (data not shown).

Quantitative evaluation of antimicrobal activity was determined by tube dilution method [2]. Bacterial and yeast strains were cultured for 24 h on TSB (Merck) medium. The ILs were diluted in TSB medium and subsequently the same amount of inoculum was added to each tube. Positive control was prepared without ILs. Growth (or lack thereof) of the microorganisms was determined visually after incubation for 24 h at 30 °C. The lowest concentration at which there was no visible growth (turbidity) was taken as the minimal inhibitory concentration (MIC).

Antifungal activity was evaluated by measuring inhibition of radial growth on an agar medium in a Petri dish (Liu et al. [3]). The ILs were dissolved in EtOH (96%). Subsequently, the appropriate diluted ILs were added to a PDA (Merck) medium or, in the case of *P. infestans* strains, to a Rye agar [4] medium to give a final

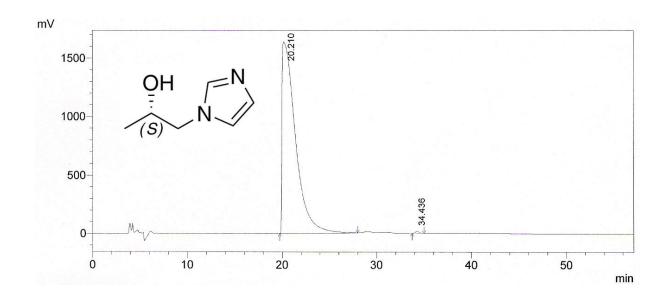
concentration of 1 mM of ILs and below 0.1% of ethanol. The 8 mm disk were cut from an actively growing mycelium of the fungi and placed in the center of the agar plate containing the proper dilution of ILs. The positive control was prepared without ILs. The radial growth of fungal colonies was measured after several days of incubation at 25 °C (in the case of *P. infestans* strains 18 °C). The inhibitory activity of ILs was expressed as a percentage of the positive control growth. Each experiment was carried out twice and the data presented here are the average of two experiments.

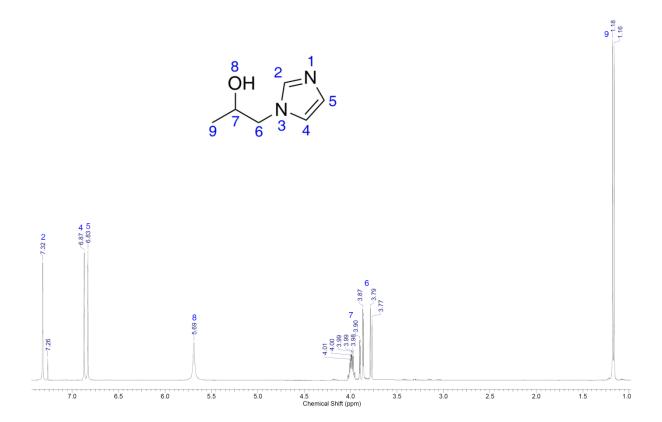
1-(1*H*-Imidazol-1-yl)propan-2-ol (±)-3a:

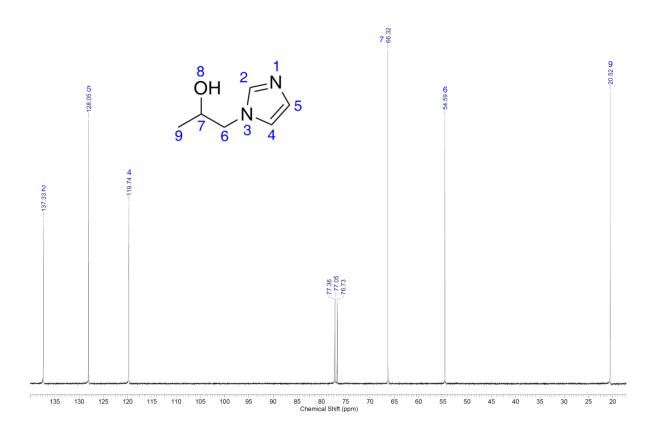


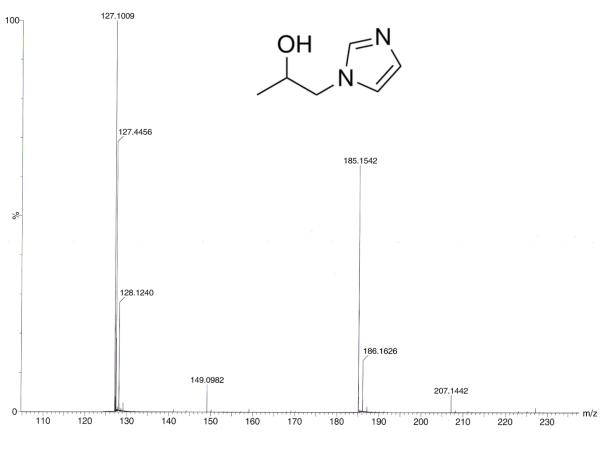
(S)-(+)-1-(1H-Imidazol-1-yl)propan-2-ol [(S)-(+)-5a]

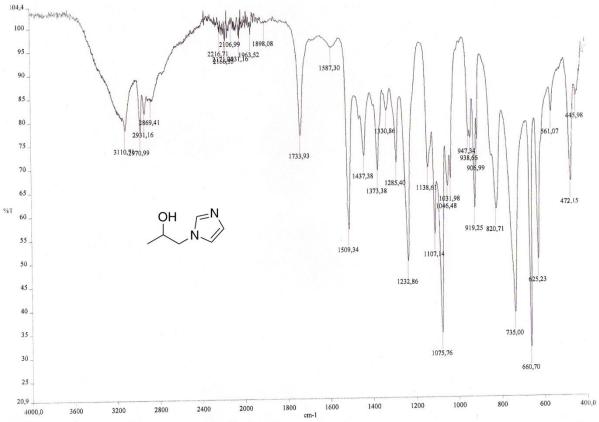
$$[\alpha]_D^{29}$$
 = +35.2 (*c* 1.0, CHCl₃) for ee = 98%



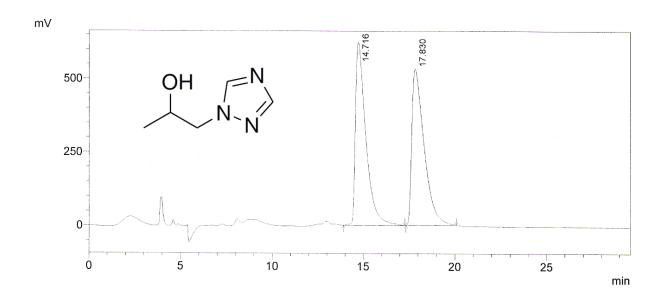






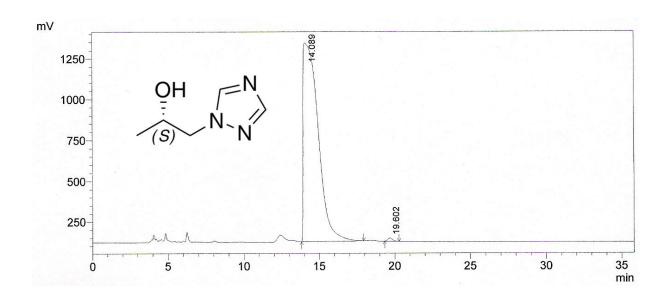


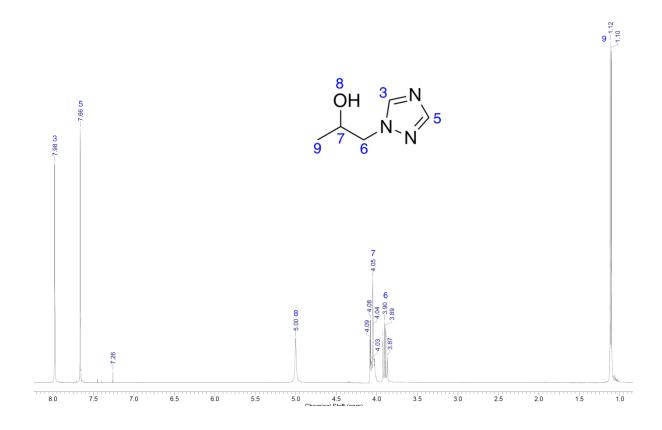
1-(1*H*-1,2,4-Triazol-1-yl)propan-2-ol (±)-3b:

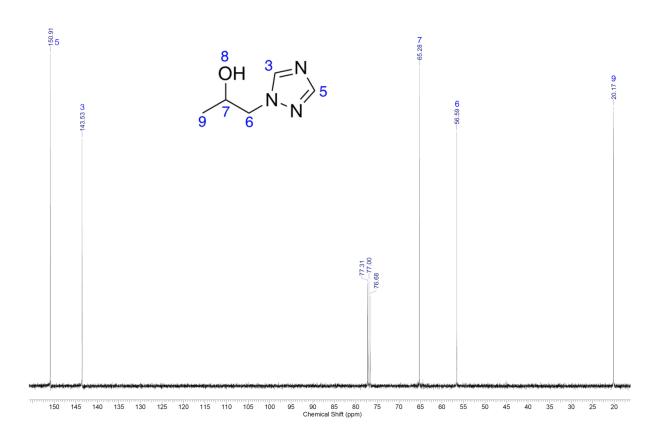


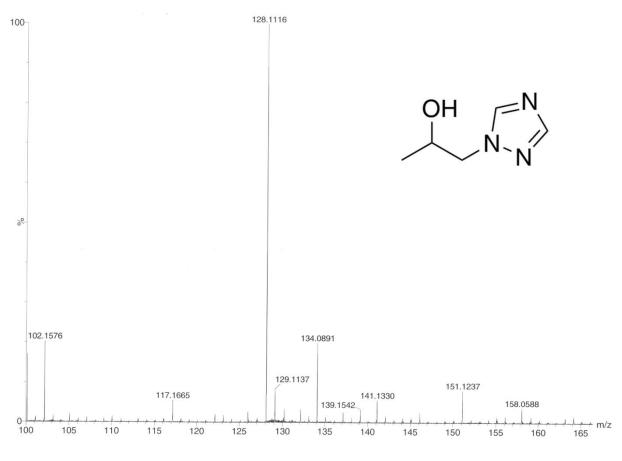
(S)-(+)-1-(1H-1,2,4-Triazol-1-yl)propan-2-ol [(S)-(+)-5b]

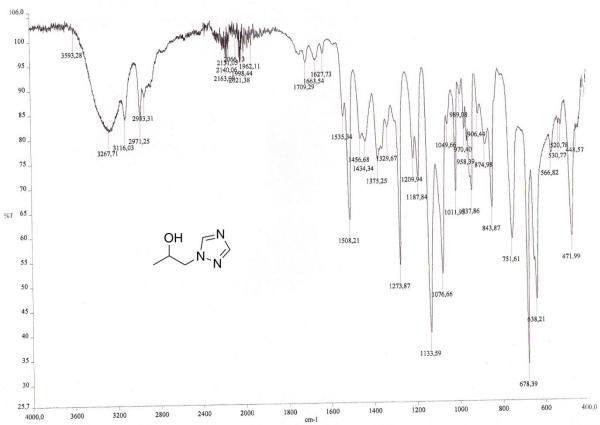
$$[\alpha]_D^{29}$$
 = +44 (c 1.0, CHCl₃) for ee = 98%



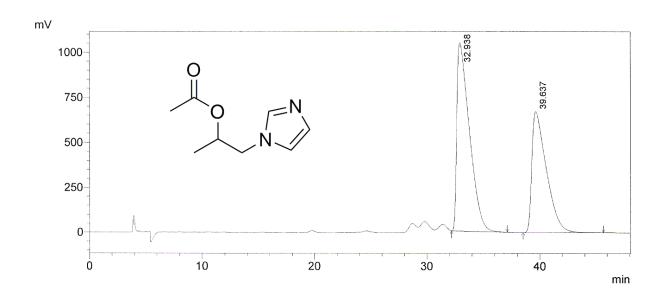






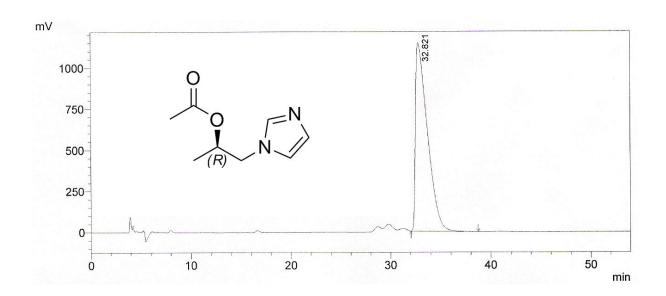


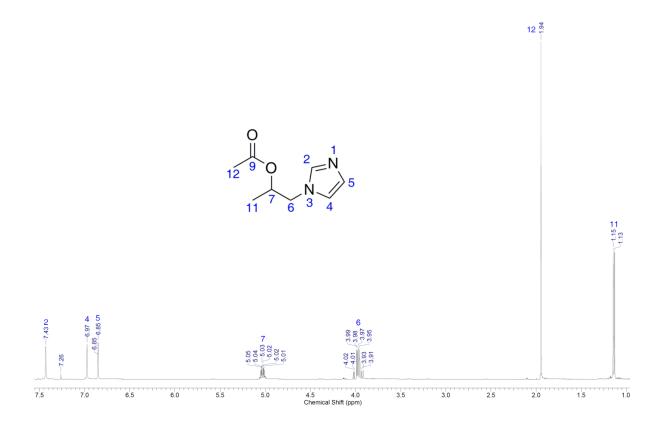
1-(1*H*-Imidazol-1-yl)propan-2-yl acetate (±)-4a:

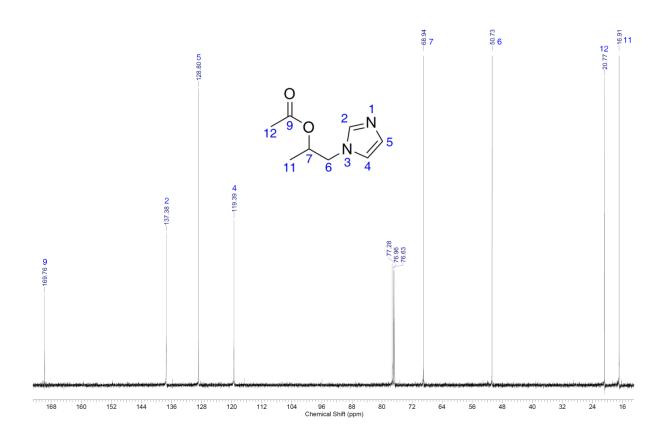


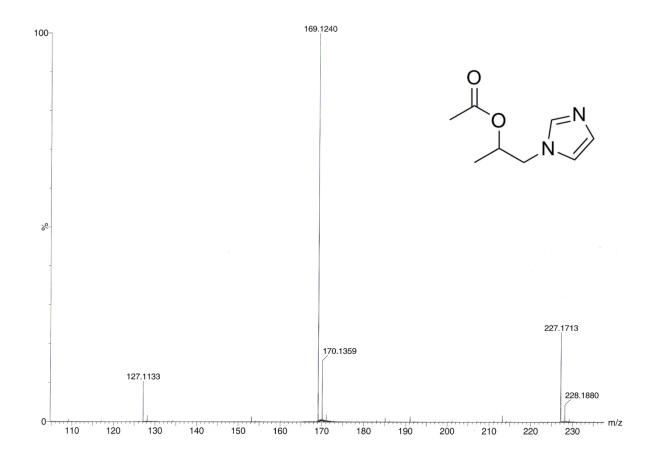
(R)-(+)-1-(1H-Imidazol-1-yl)propan-2-yl acetate [(R)-(+)-6a]

$[\alpha]_D^{29}$ = +9.2 (*c* 1.0, CHCI₃) for ee > 99%

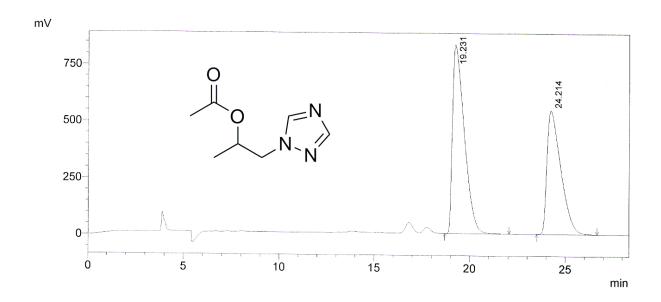






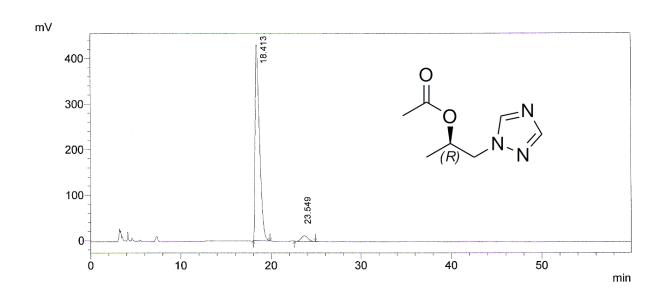


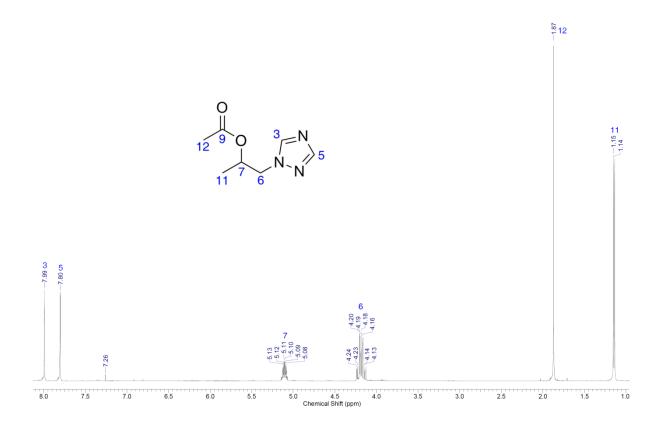
1-(1*H*-1,2,4-Triazol-1-yl)propan-2-yl acetate (±)-4b:

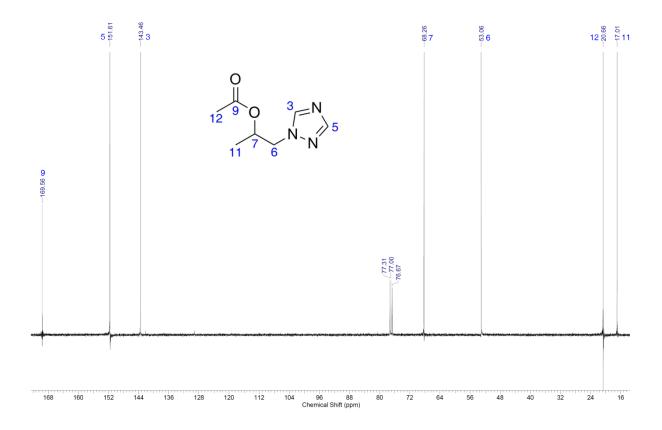


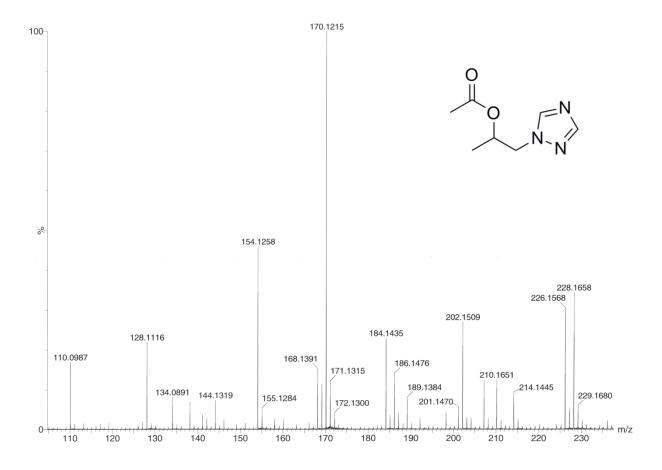
(R)-(+)-1-(1H-1,2,4-Triazol-1-yl)propan-2-yl acetate [(R)-(+)-6b]

$$[\alpha]_D^{29}$$
 = +5.2 (*c* 1.0, CHCI₃) for ee = 90%

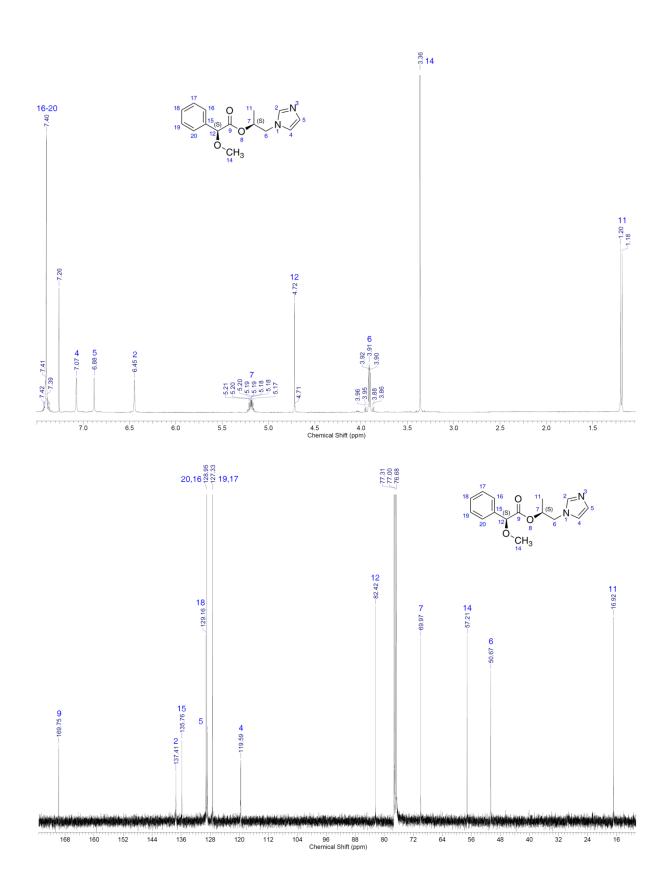




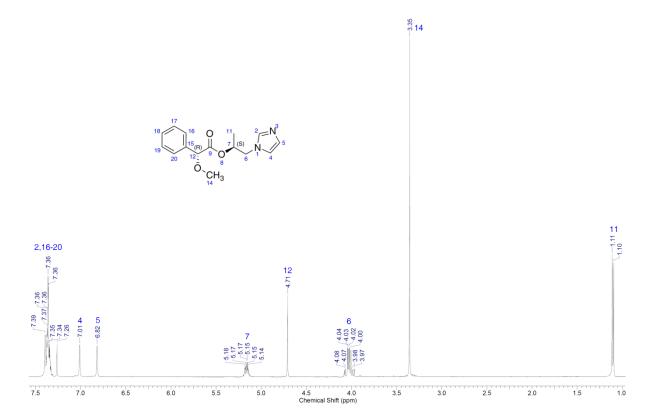


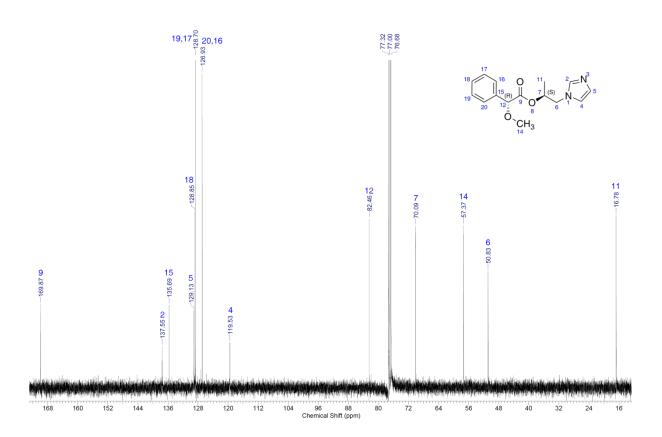


(S)-((S)-1-(1H-Imidazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (9a):

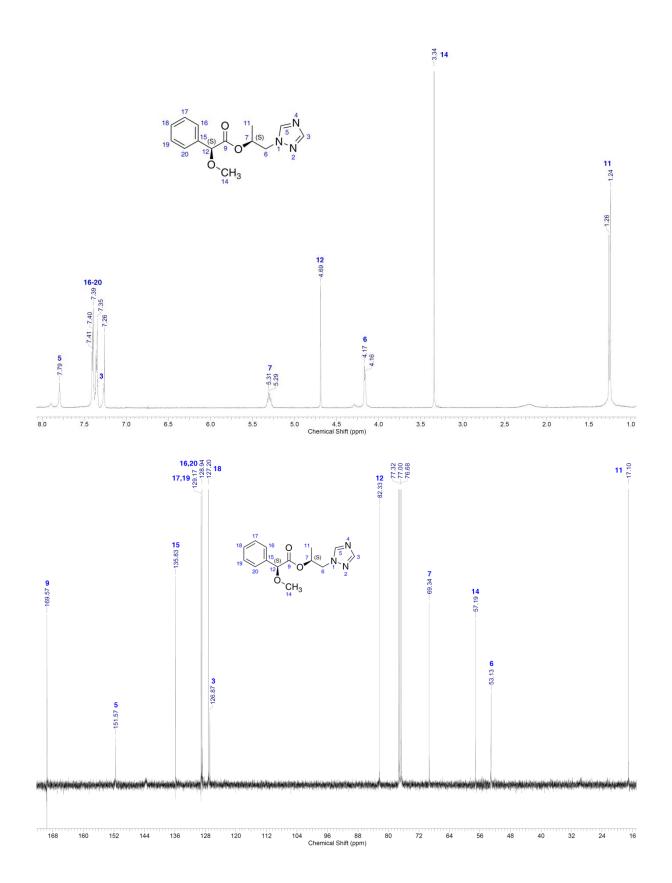


((R)-((S)-1-(1H-Imidazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (10a):

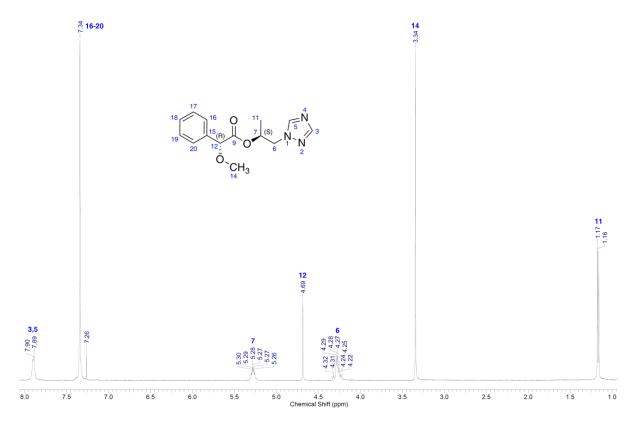


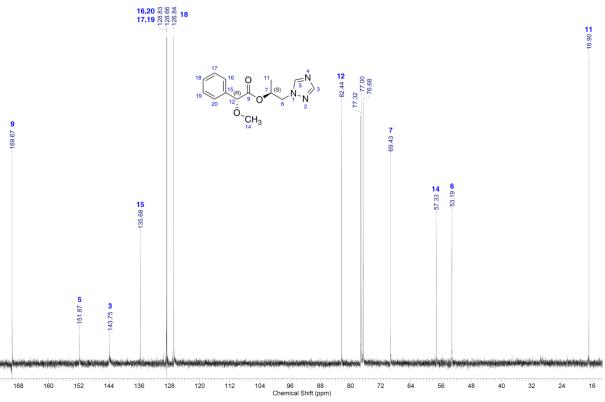


(S)-((S)-1-(1H-1,2,4-Triazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (9b):



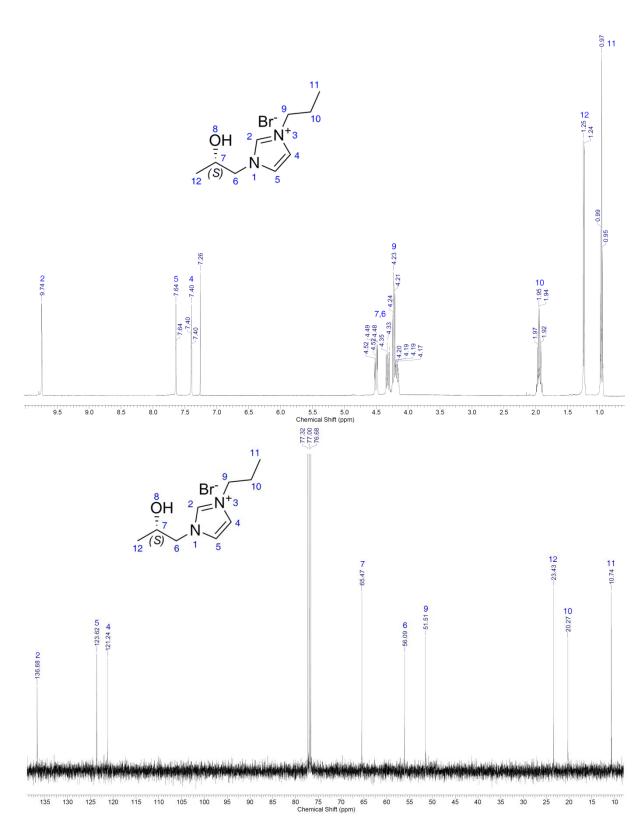
(R)-((S)-1-(1H-1,2,4-Triazol-1-yl)propan-2-yl) 2-methoxy-2-phenylacetate (10b):

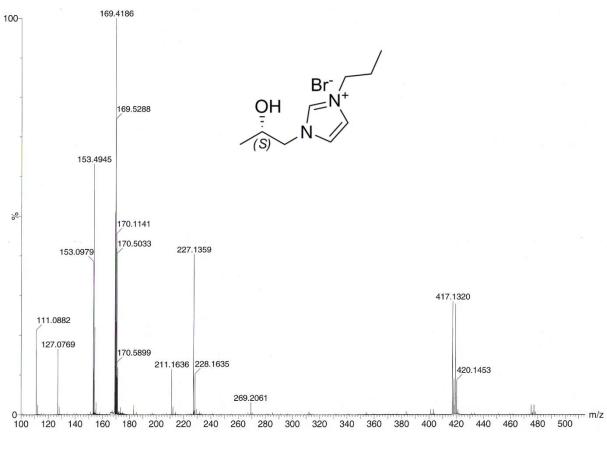


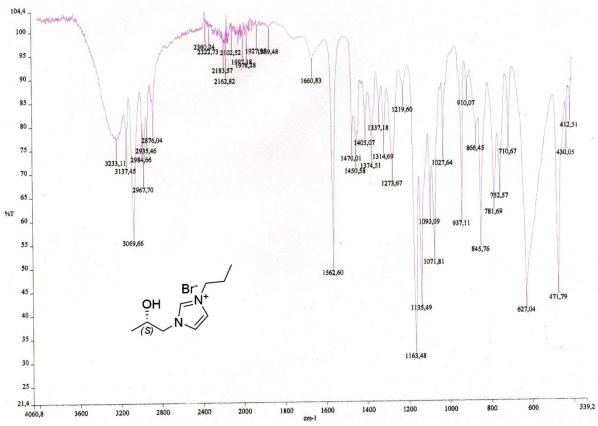


Spectra of the imidazolium salts [(+)-7a-h]

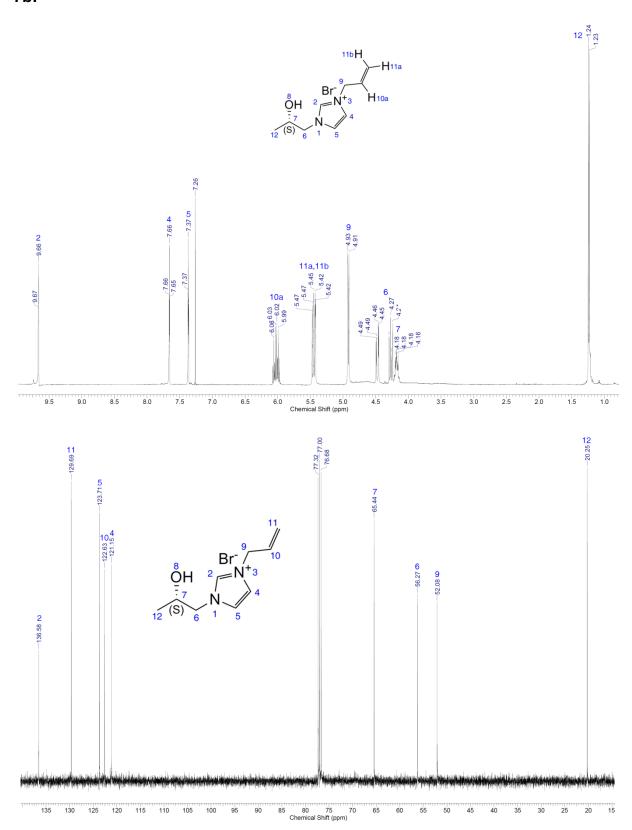
(S)-(+)-1-(2-Hydroxypropyl)-3-propyl-1*H*-imidazol-3-ium bromide (+)-7a:

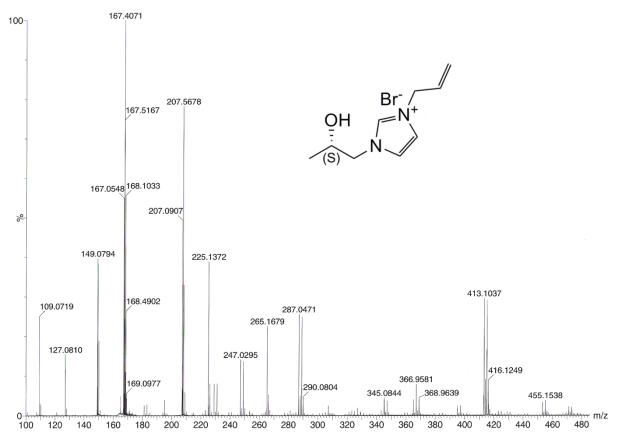


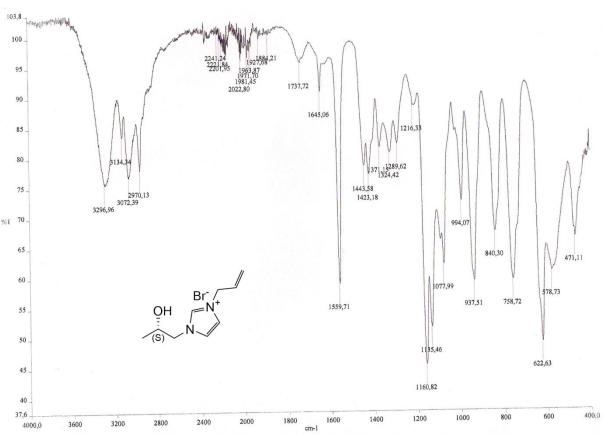




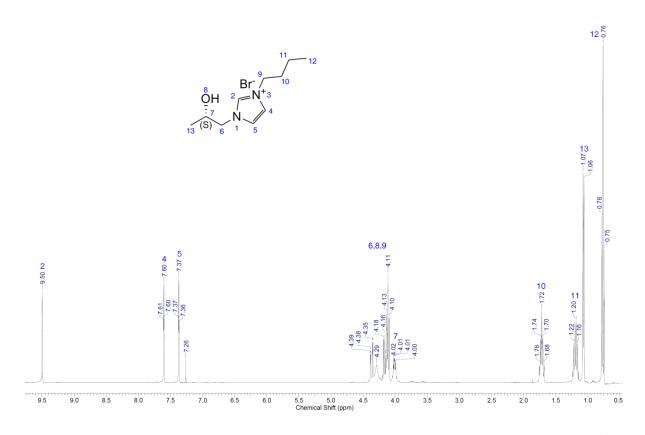
(*S*)-(+)-1-(2-Hydroxypropyl)-3-(prop-2-en-1-yl)-1*H*-imidazol-3-ium bromide (+)-7b:

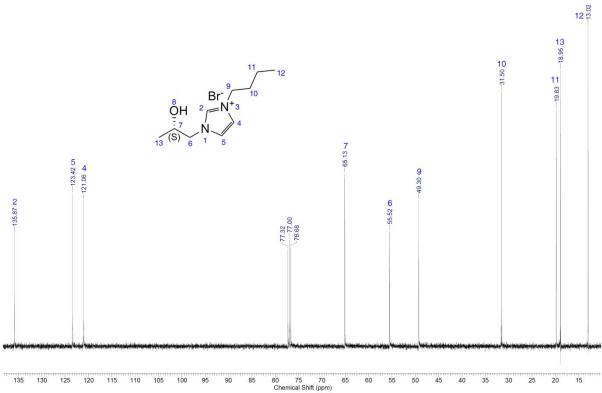


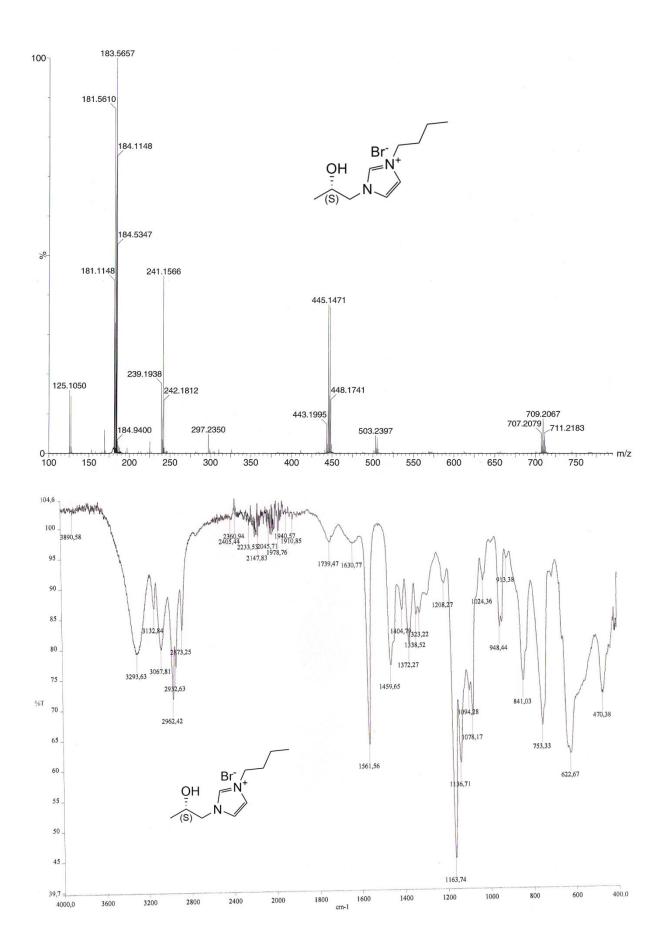




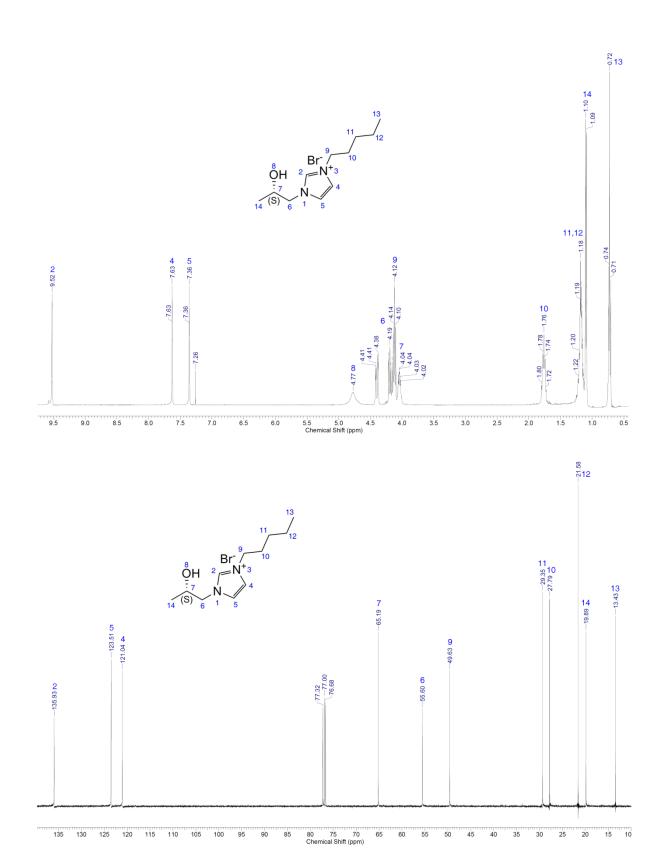
(S)-(+)-3-Butyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium bromide (+)-7c:

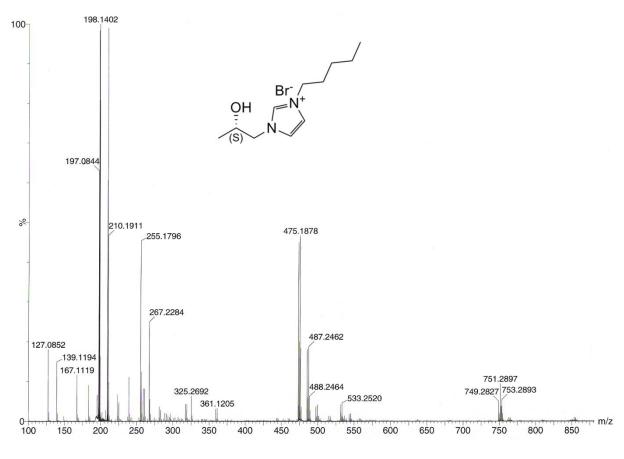


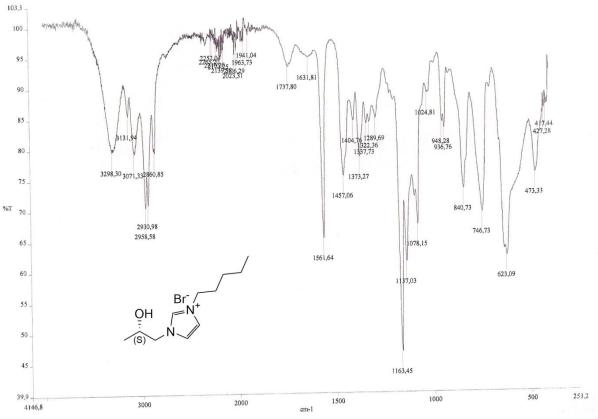




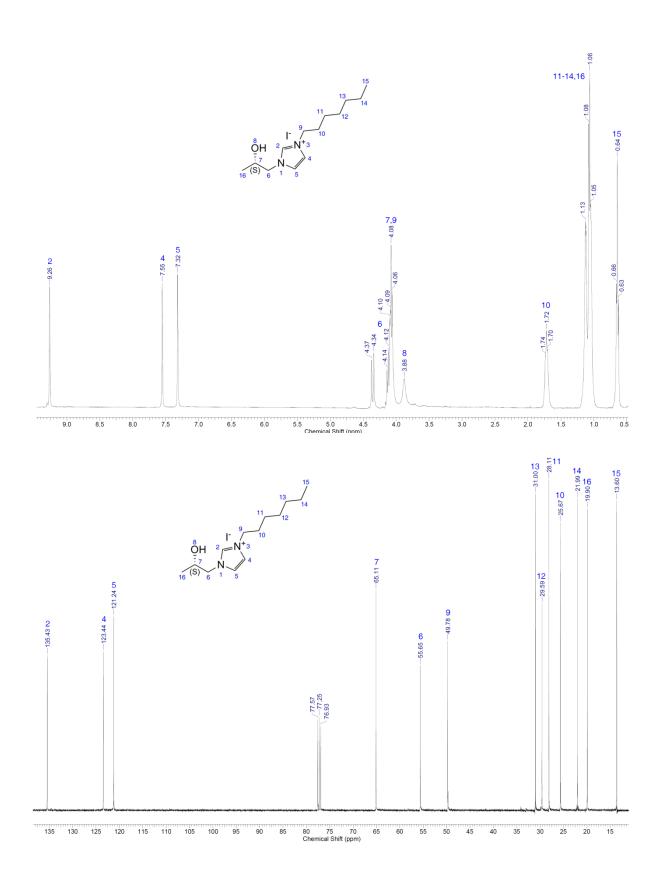
(S)-(+)-1-(2-Hydroxypropyl)-3-pentyl-1*H*-imidazol-3-ium bromide (+)-7d:

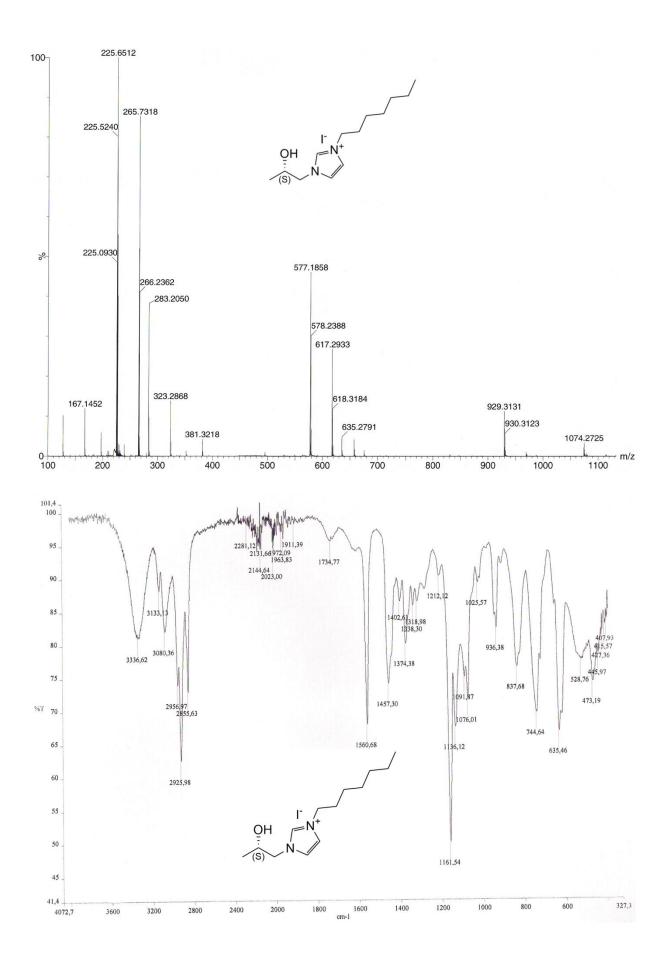




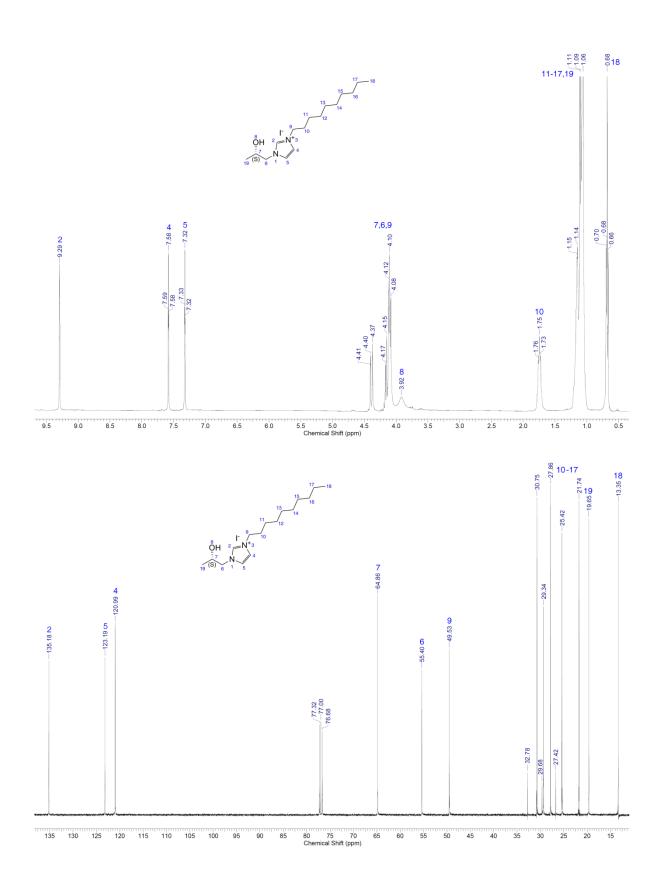


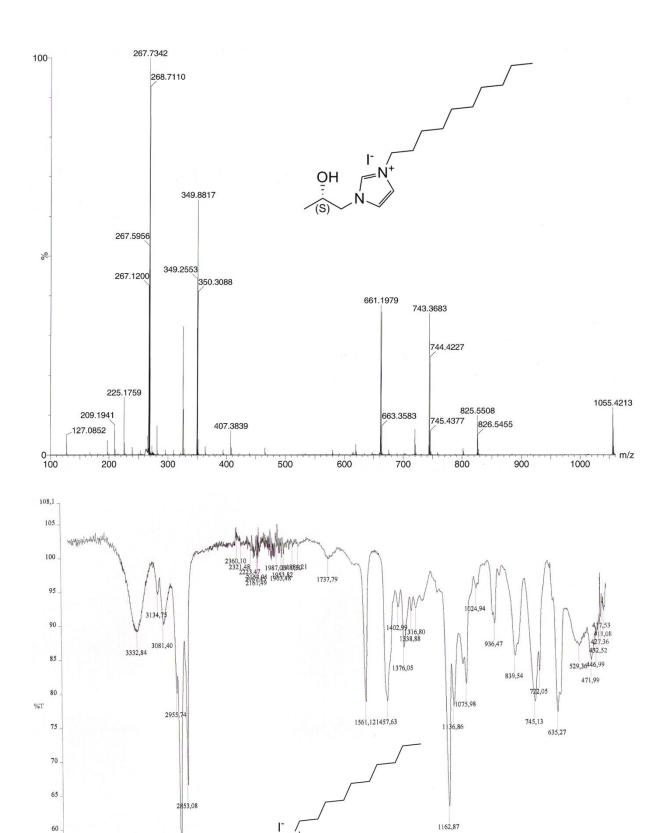
(S)-(+)-3-Heptyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7e:





(S)-(+)-3-Decyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7f:





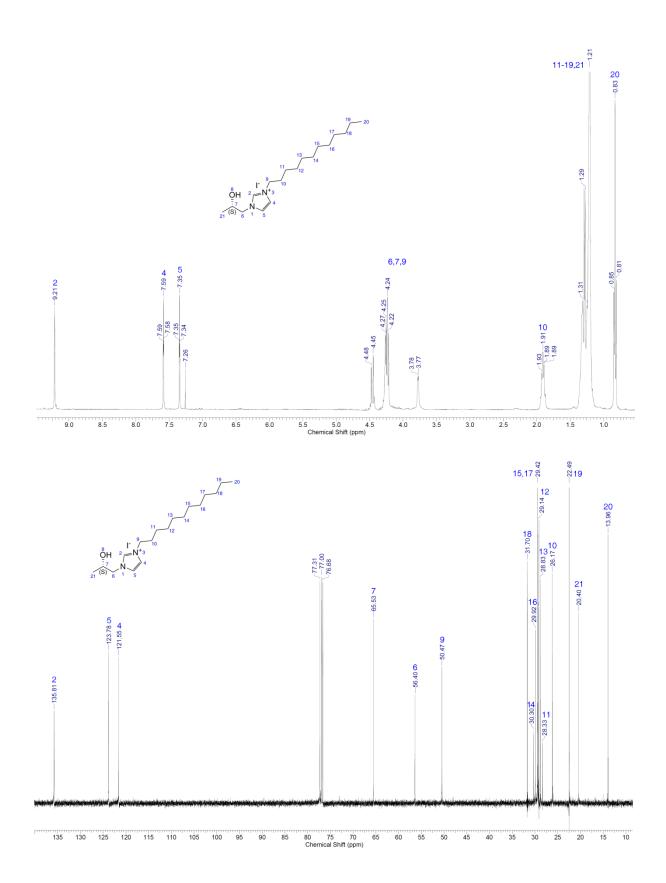
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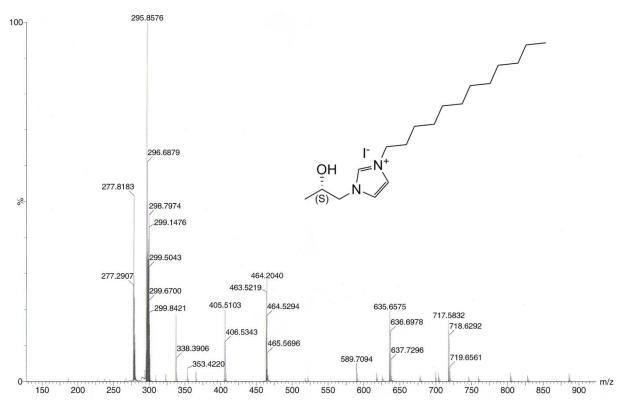
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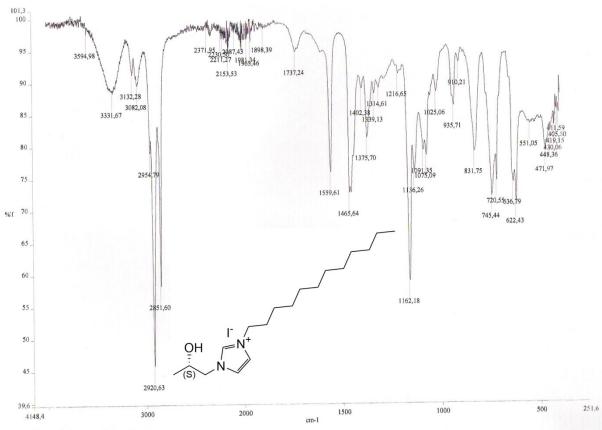
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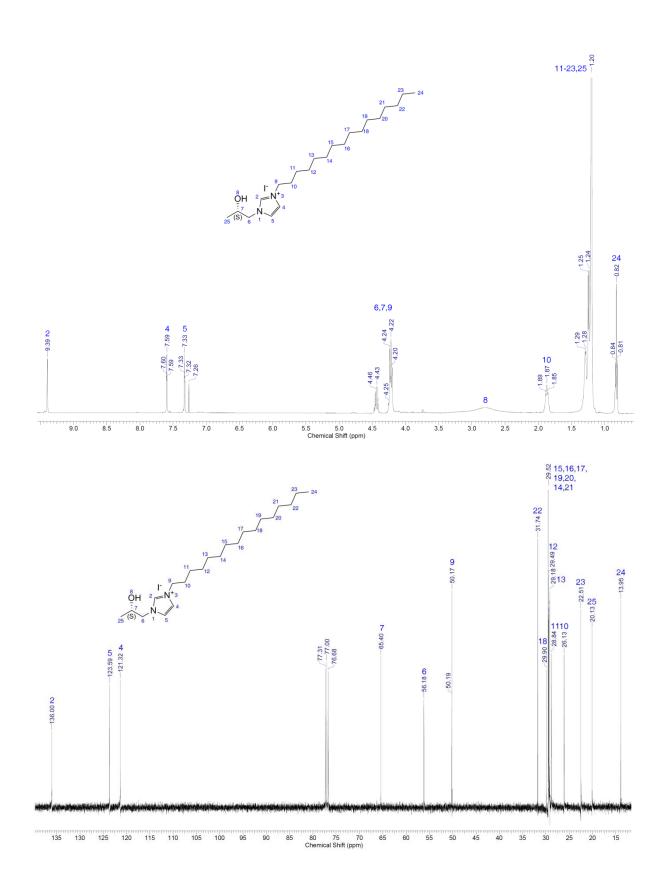
(S)-(+)-3-Dodecyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7g:

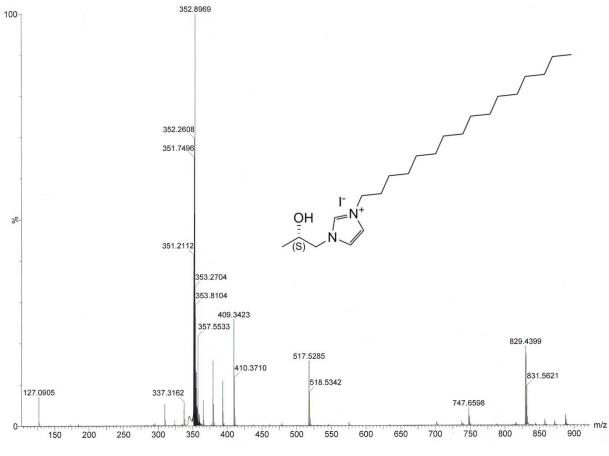


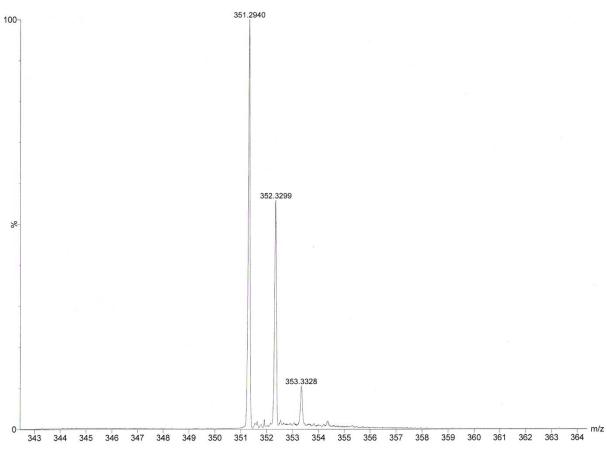


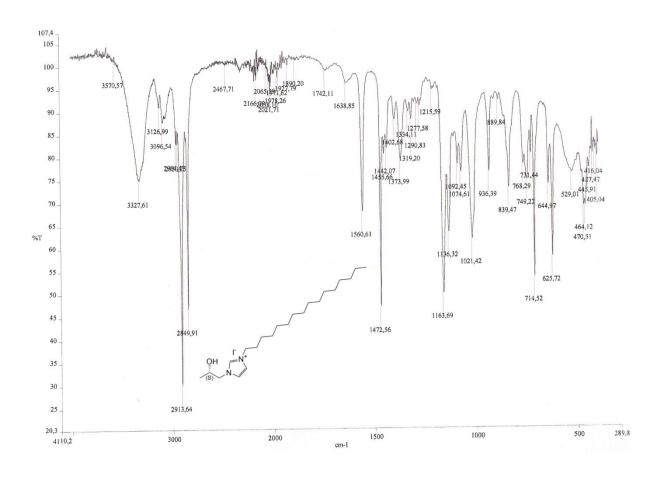


(S)-(+)-3-Hexadecyl-1-(2-hydroxypropyl)-1H-imidazol-3-ium iodide (+)-7h:



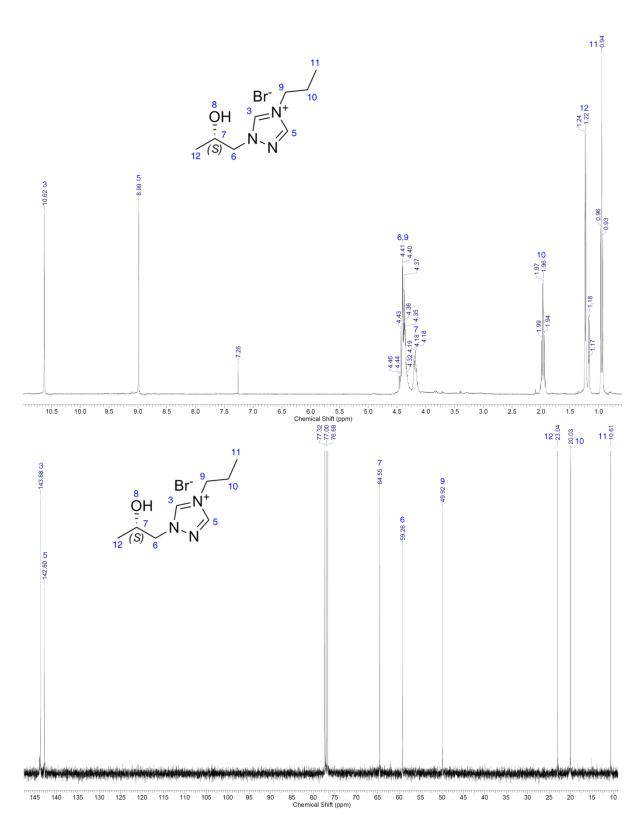


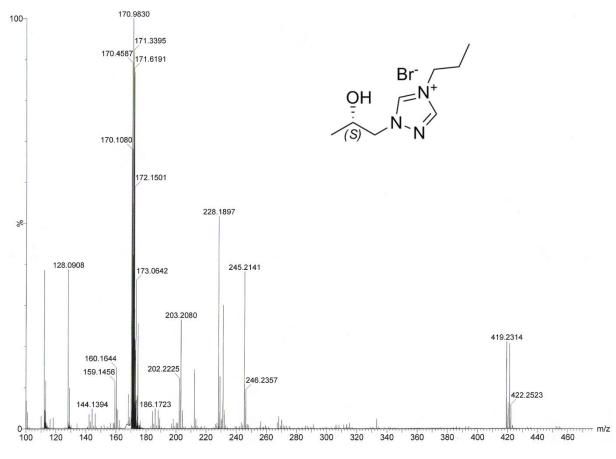


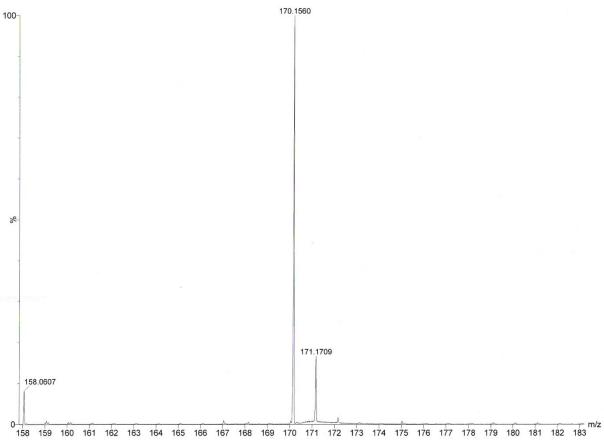


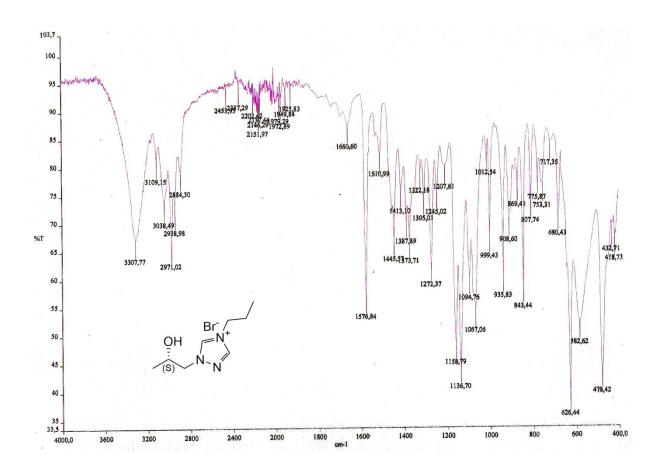
Spectra of the triazolium salts [(+)-8a-f]

(S)-(+)-1-(2-Hydroxypropyl)-4-propyl-1*H*-1,2,4-triazol-4-ium bromide (+)-8a:

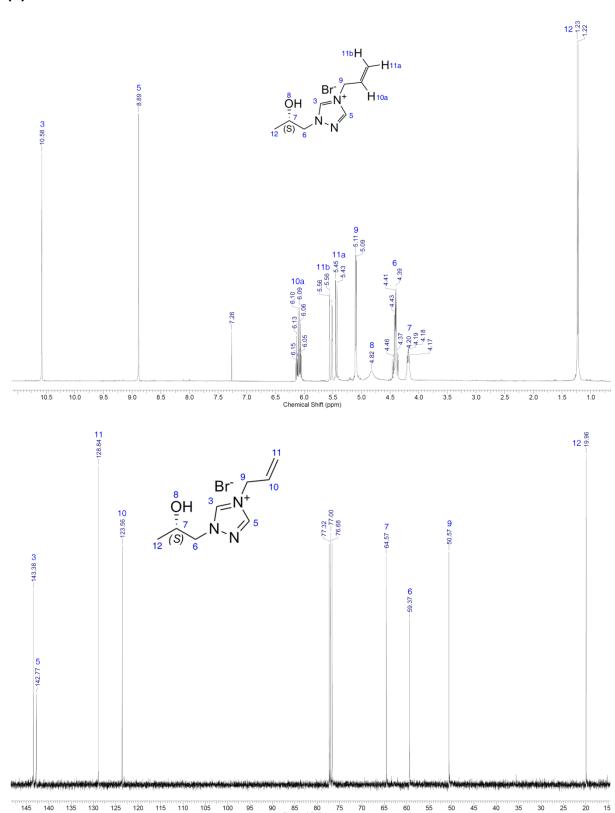


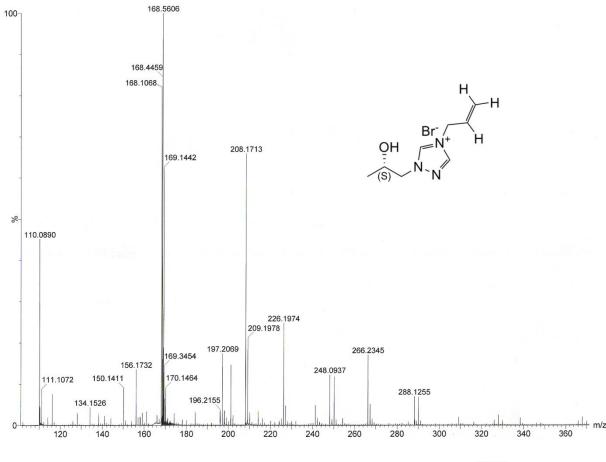


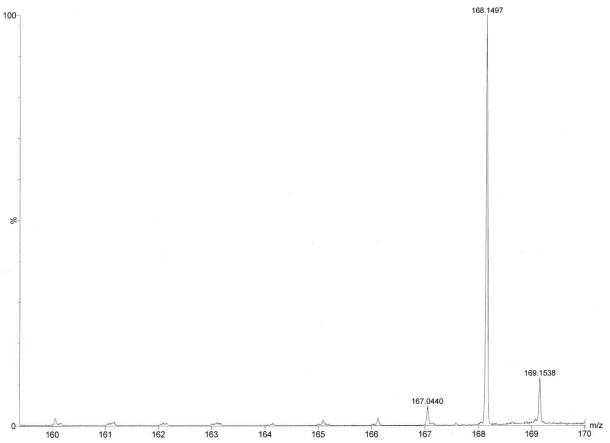


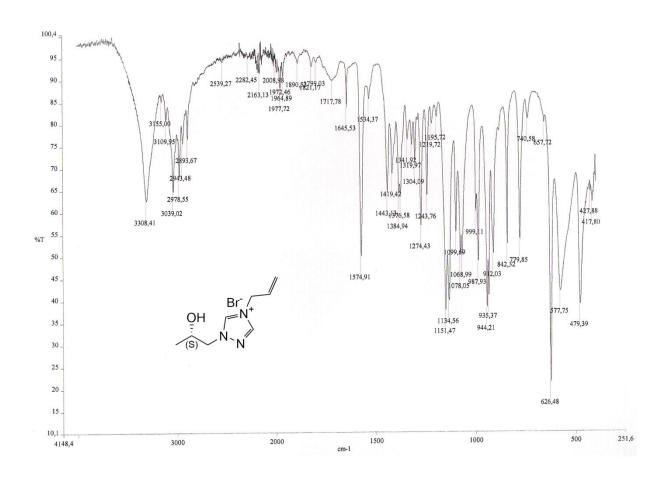


(*S*)-(+)-1-(2-Hydroxypropyl)-4-(prop-2-en-1-yl)-1*H*-1,2,4-triazol-4-ium bromide (+)-8b:

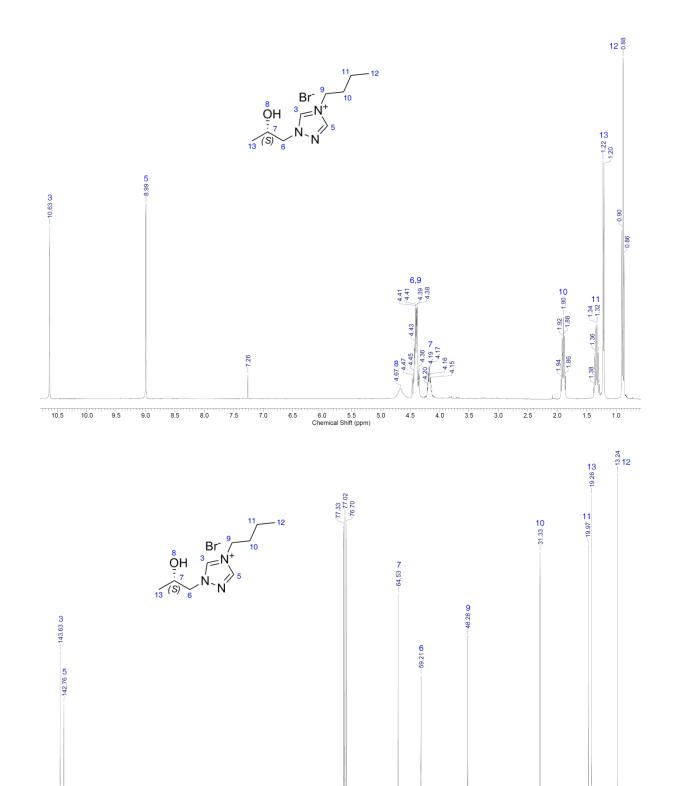




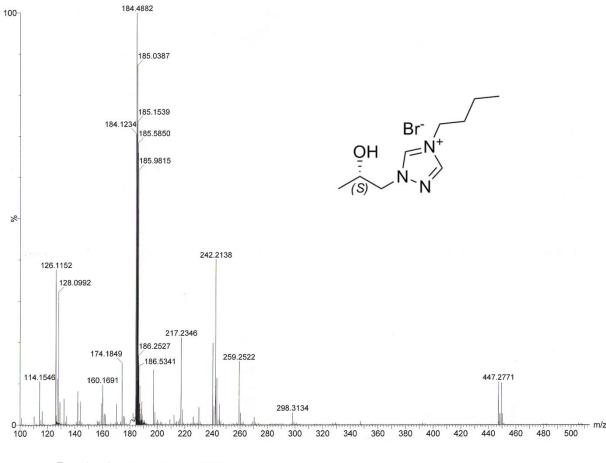


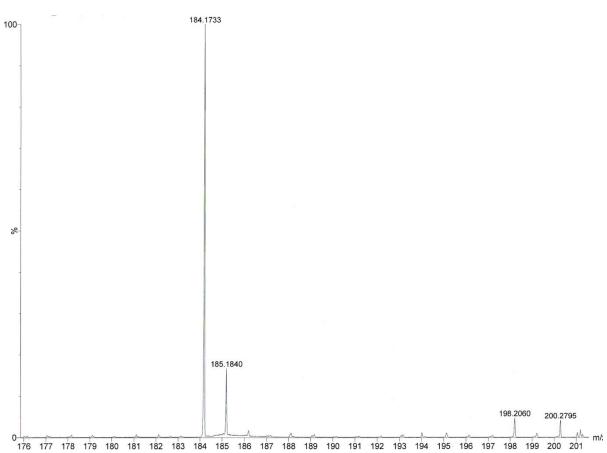


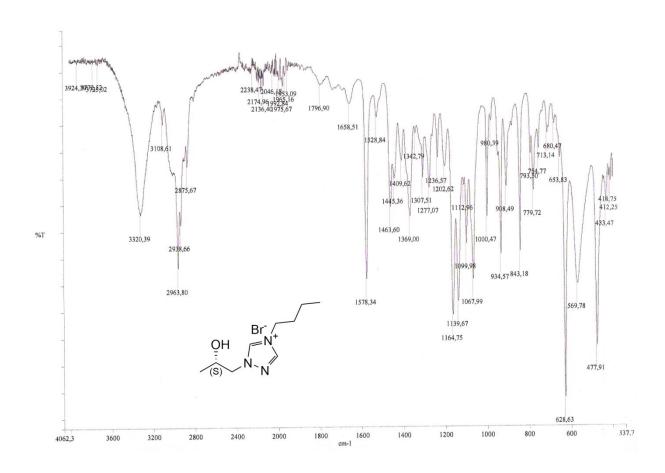
(S)-(+)-4-Butyl-1-(2-hydroxypropyl)-1H-1,2,4-triazol-4-ium bromide (+)-8c:



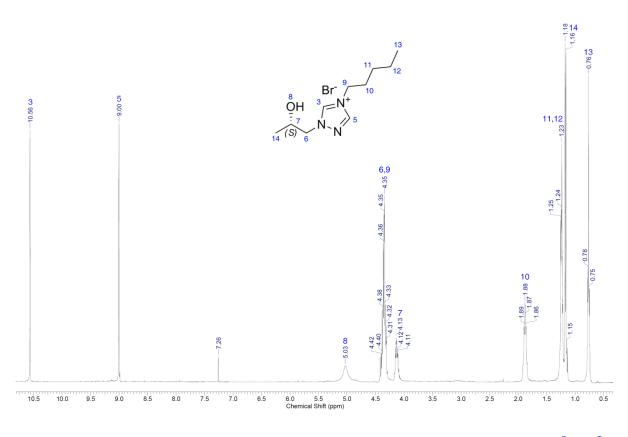
145 140 135 130 125 120 115 110

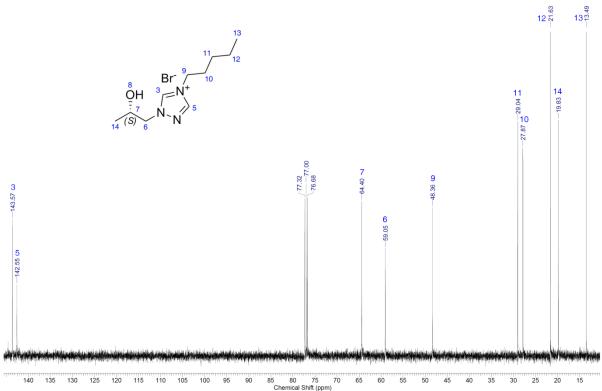


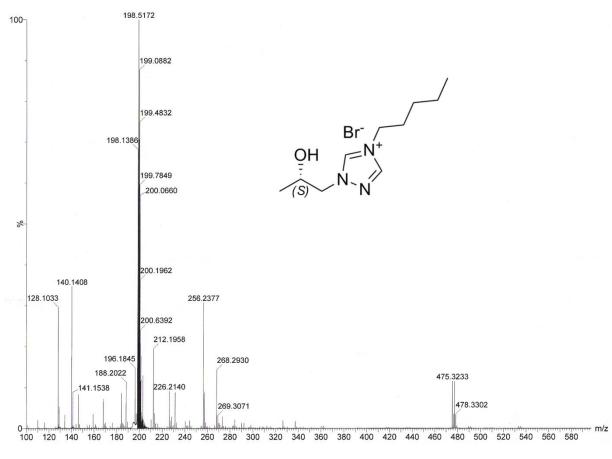


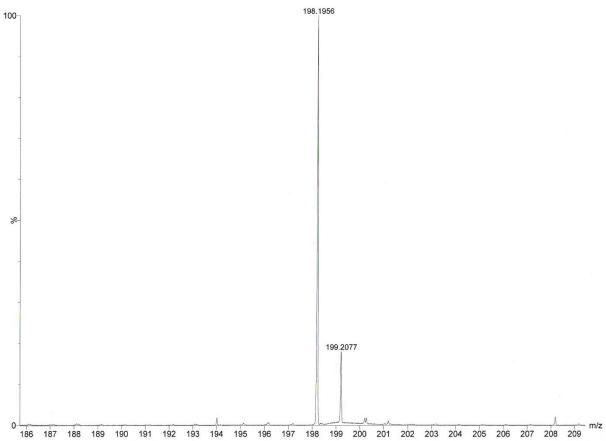


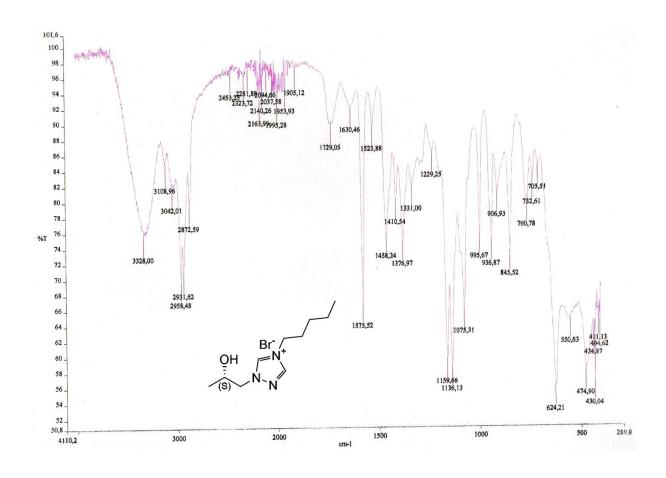
(S)-(+)-1-(2-Hydroxypropyl)-4-pentyl-1*H*-1,2,4-triazol-4-ium bromide (+)-8d:



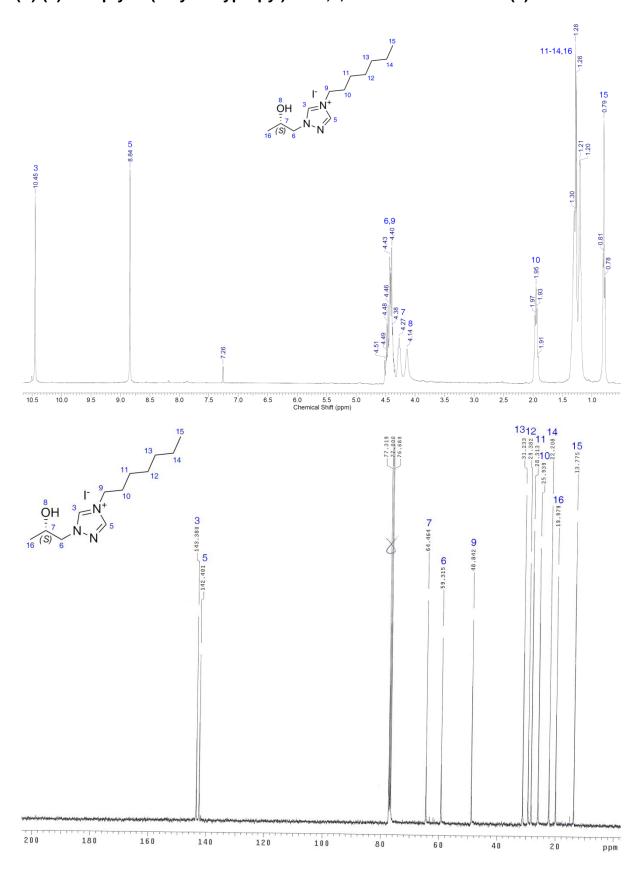


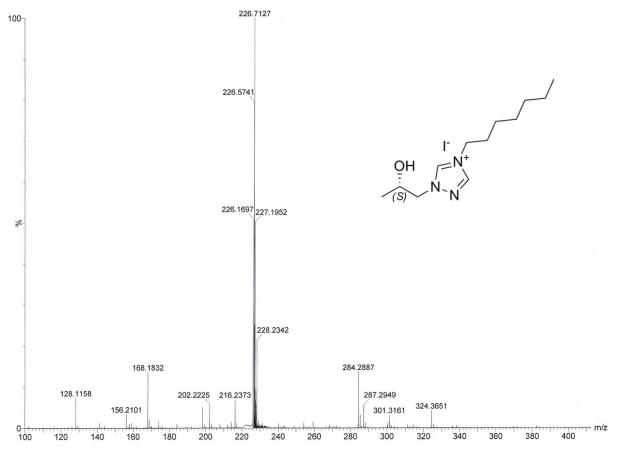


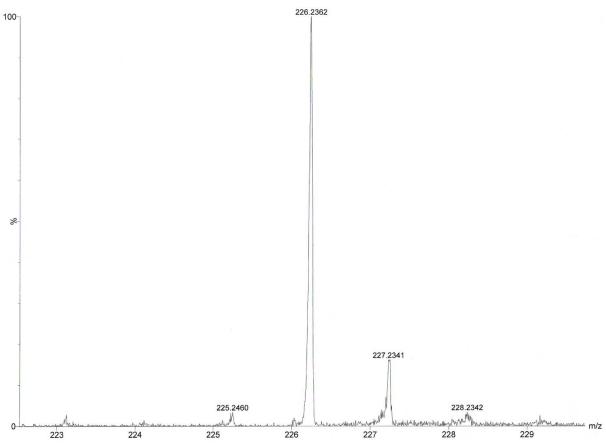


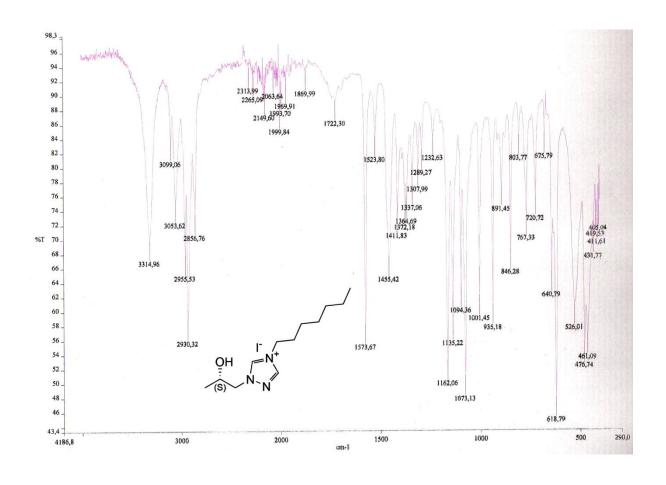


(S)-(+)-4-Heptyl-1-(2-hydroxypropyl)-1*H*-1,2,4-triazol-4-ium iodide (+)-8e:

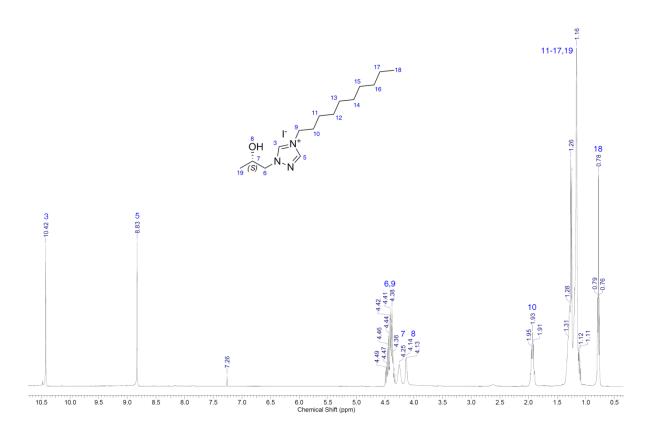


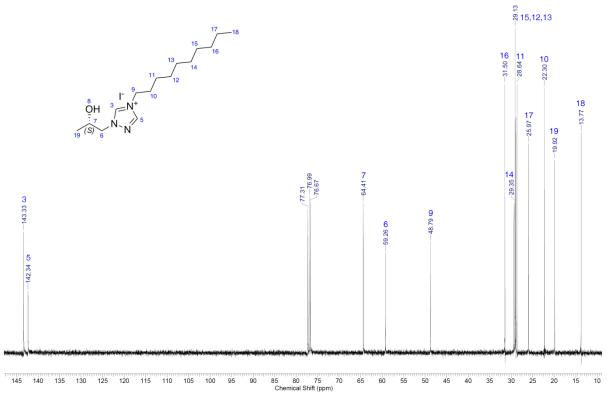


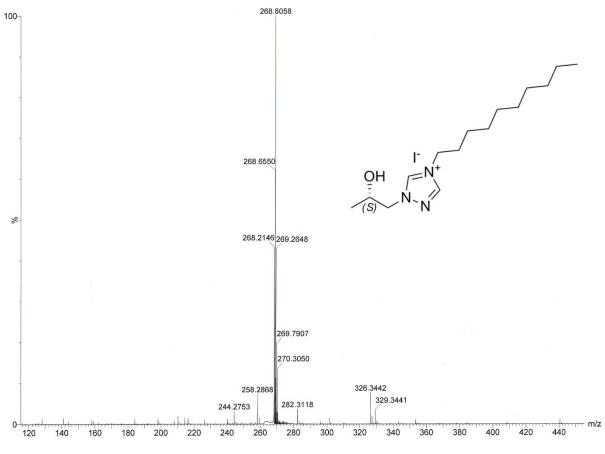


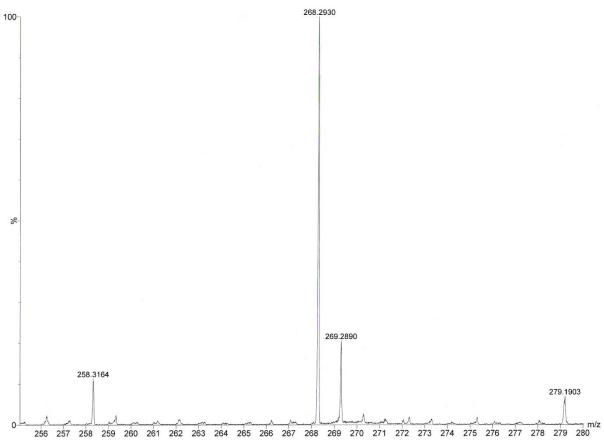


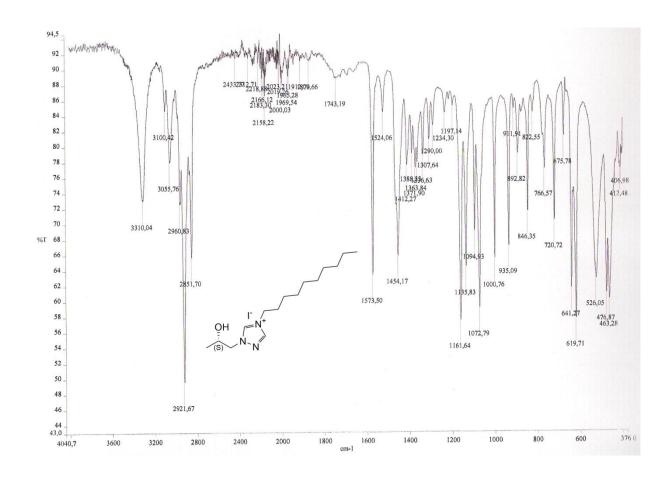
(S)-(+)-4-Decyl-1-(2-hydroxypropyl)-1H-1,2,4-triazol-4-ium iodide (+)-8f:











References

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