

Supporting Information
for
Synthesis and stability study of a new major metabolite of
 γ -hydroxybutyric acid

Ida Nymann Petersen¹, Jesper Langgaard Kristensen¹, Christian Tortzen², Torben Breindahl³ and Daniel Sejer Pedersen^{*1}

Address: ¹Department of Drug Design and Pharmacology, University of Copenhagen, Universitetsparken 2, DK-2100 Copenhagen, Denmark, ²Department of Chemistry, University of Copenhagen, Universitetsparken 5, DK-2100 Copenhagen, Denmark and ³Department of Clinical Biochemistry, Vendsyssel Hospital, Bispensgade 37, DK-9800 Hjørring, Denmark

Email: Daniel Sejer Pedersen – daniel.pedersen@sund.ku.dk

* Corresponding author

1D and 2D NMR spectra for 2 and d_4 -2 and all details for the NMR stability study of GHB glucuronide 2

Table of Contents

1. NMR characterisation

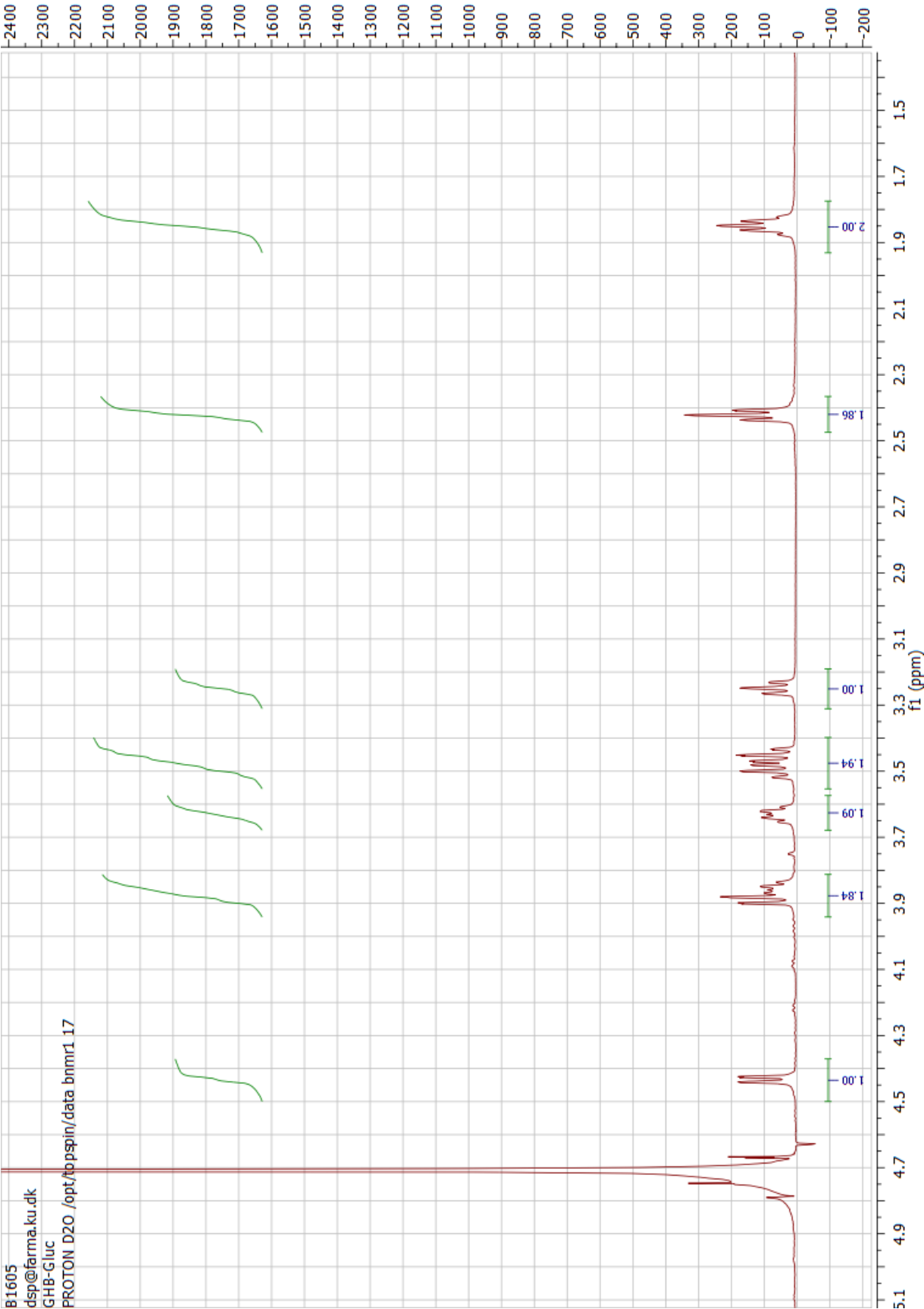
1.1 1D and 2D NMR spectra of glucuronide 2	S3
1.2 1D and 2D NMR spectra of glucuronide <i>d</i> ₄ - 2	S8

2. NMR stability study for GHB-GLUC (2)

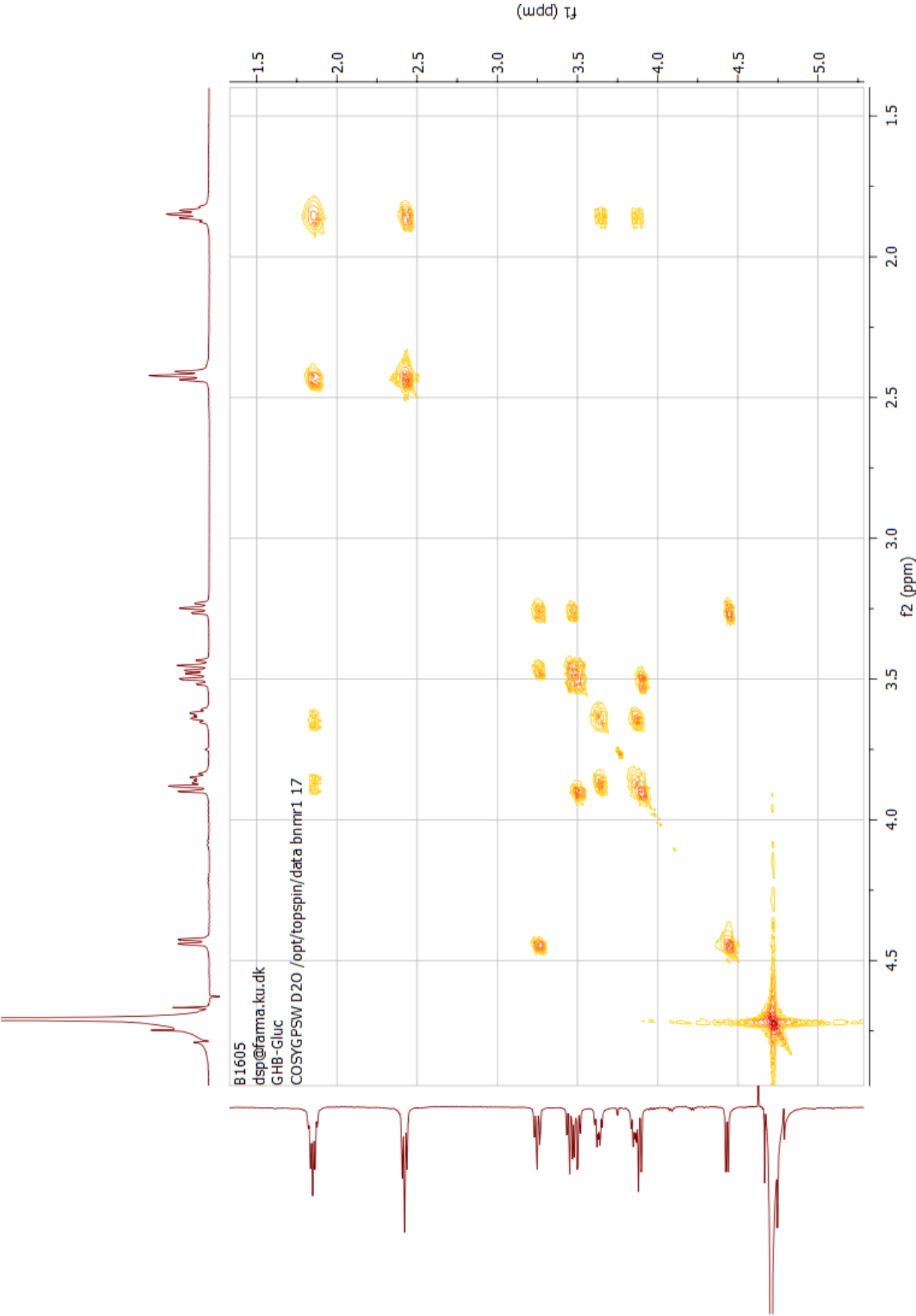
2.1 General NMR experimental	S13
2.2 NMR stability study ¹ H NMR spectra	S15
2.3 NMR reference spectra	S18

1.1 1D and 2D NMR spectra of glucuronide 2

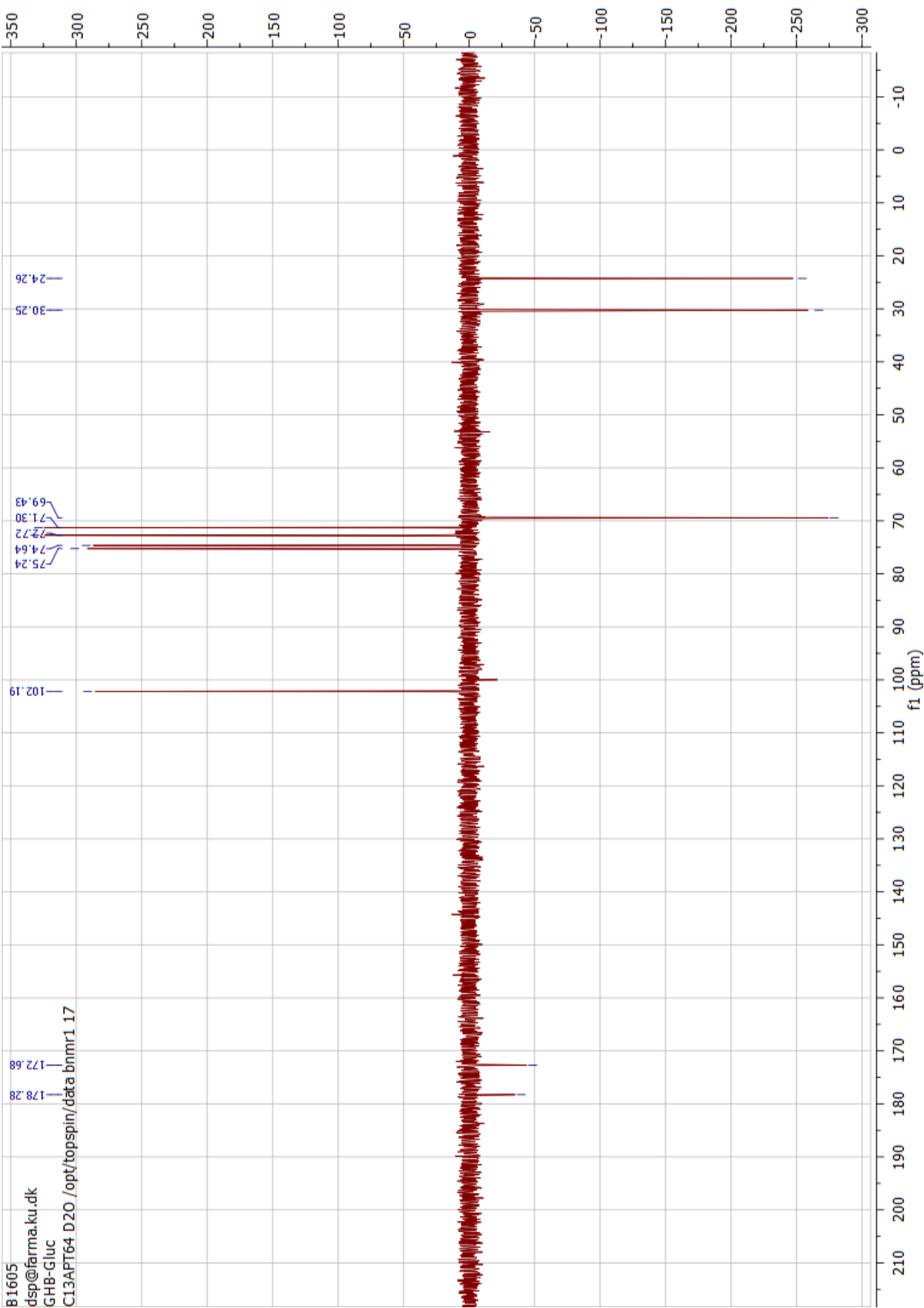
¹H NMR spectrum of GHB glucuronide 2



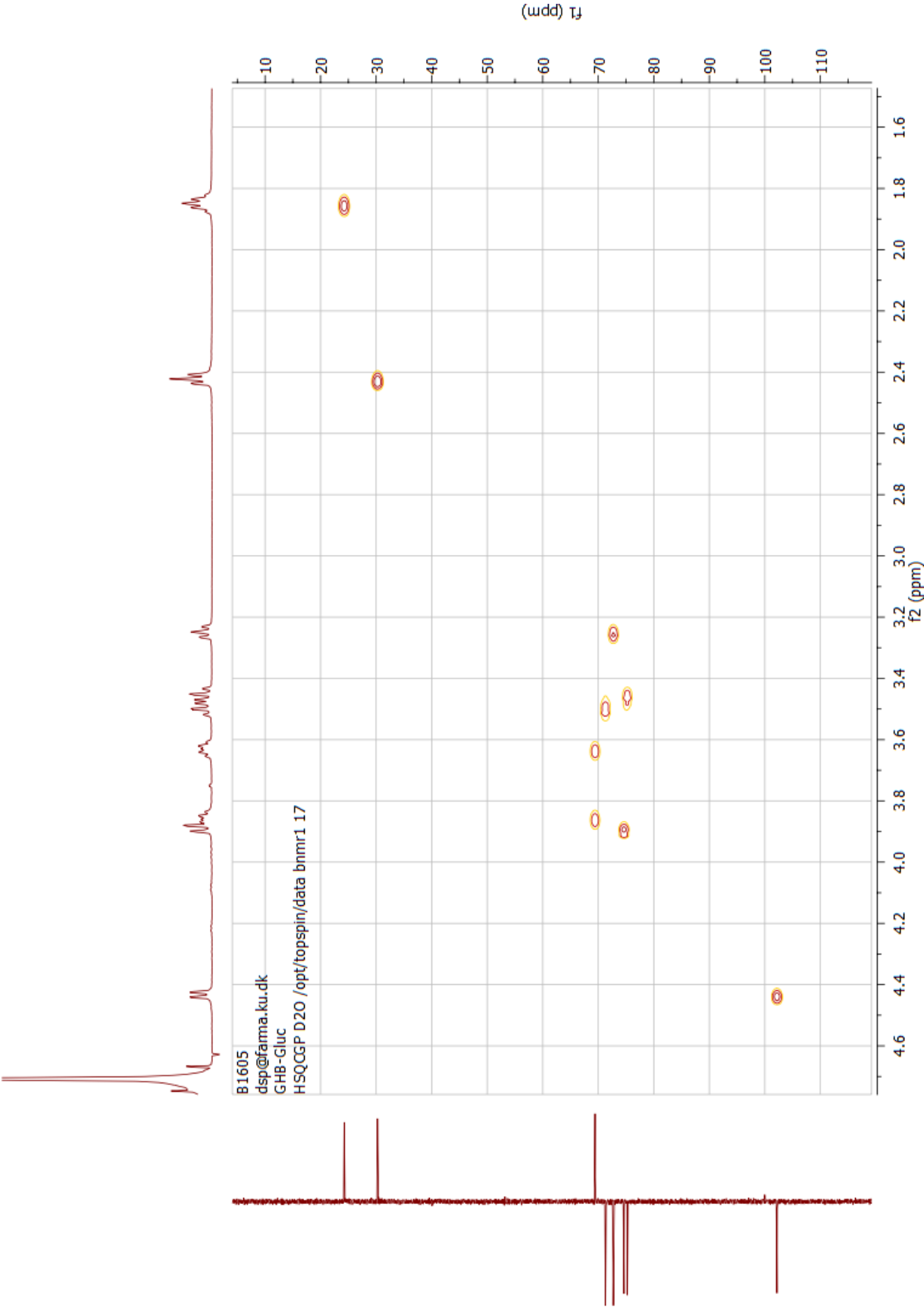
COSY NMR spectrum of GHB glucuronide 2



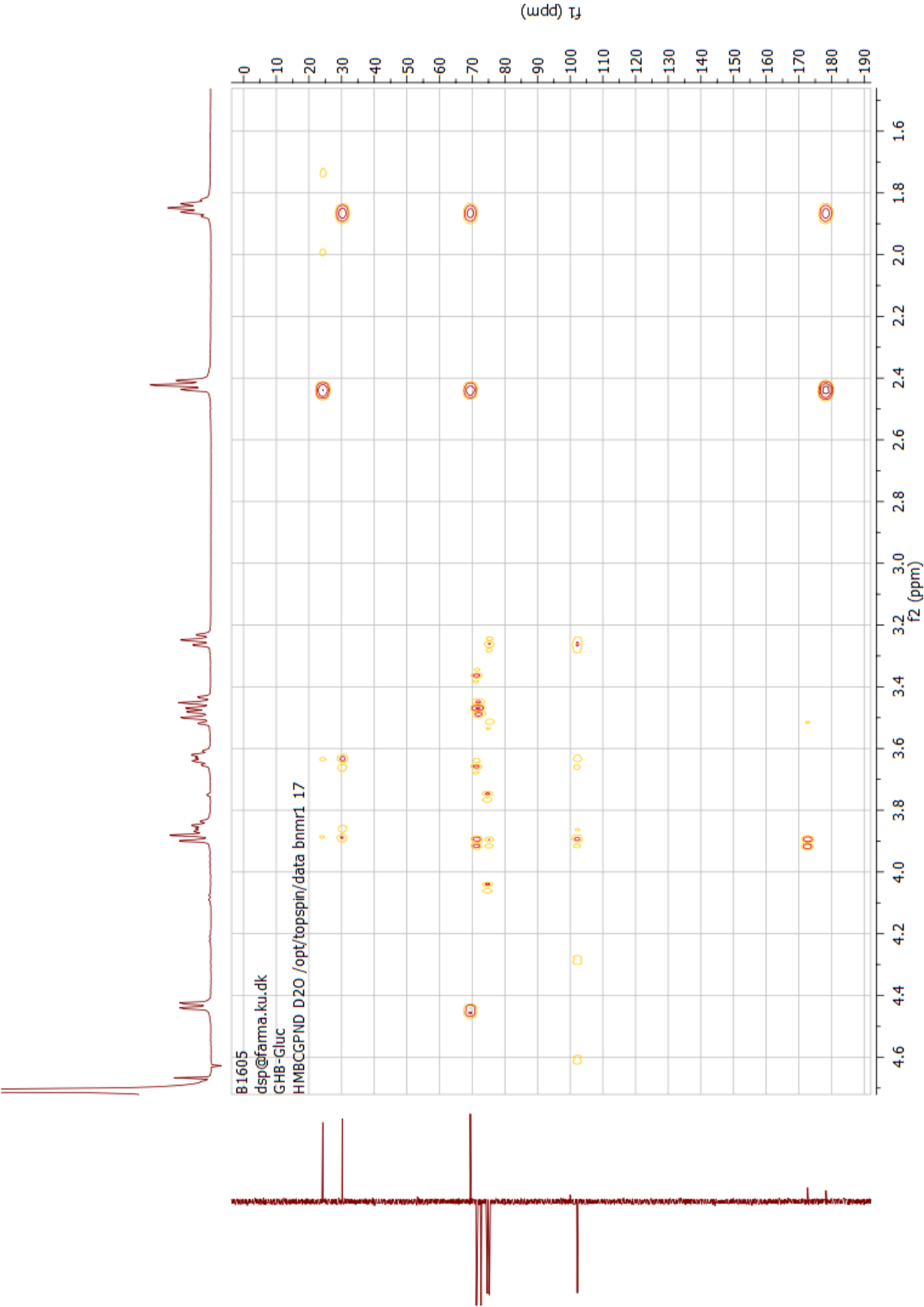
¹³C APT NMR spectrum of GHB glucuronide 2



HSQC NMR spectrum of GHB glucuronide 2

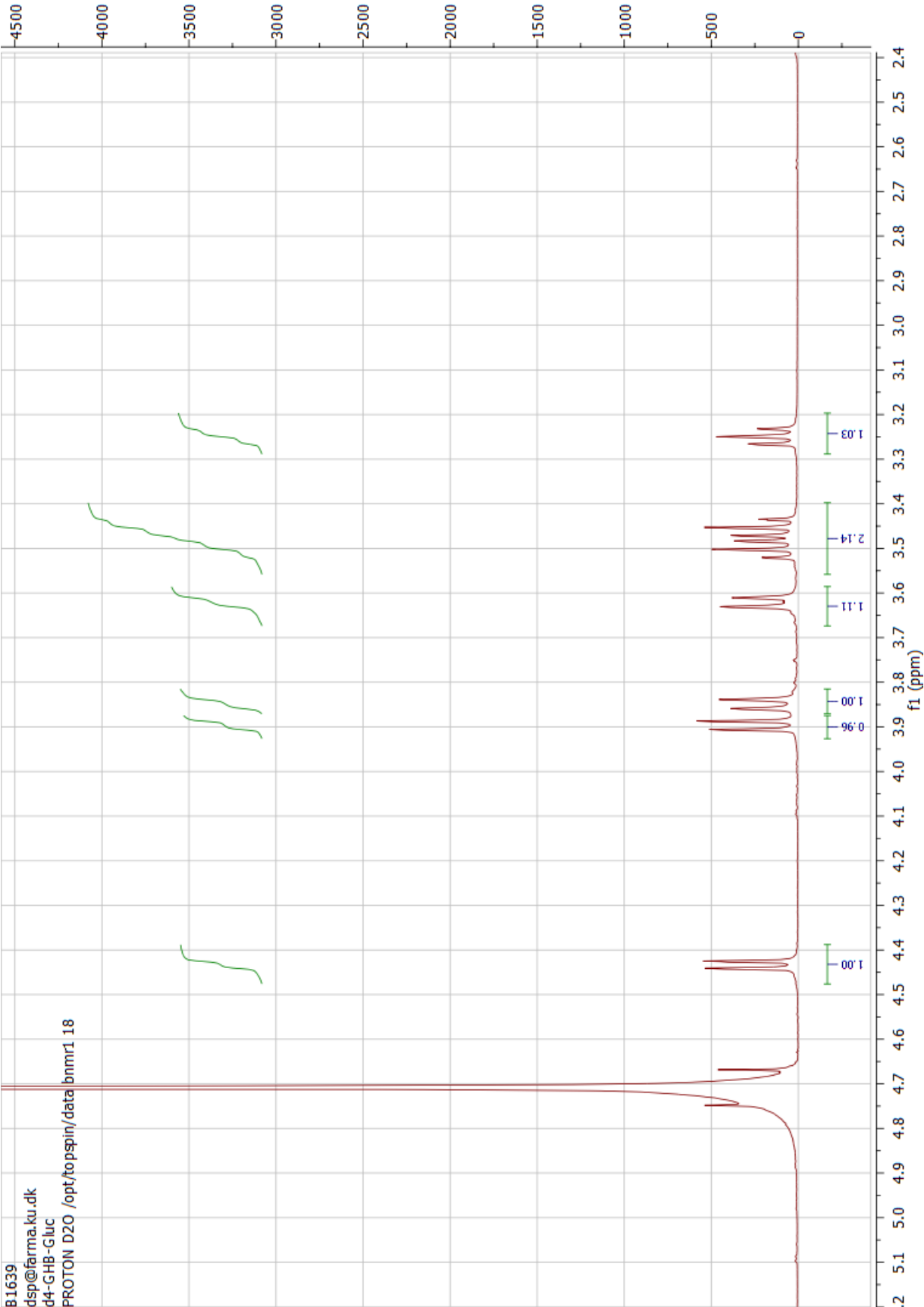


HMBC NMR spectrum of GHB glucuronide 2



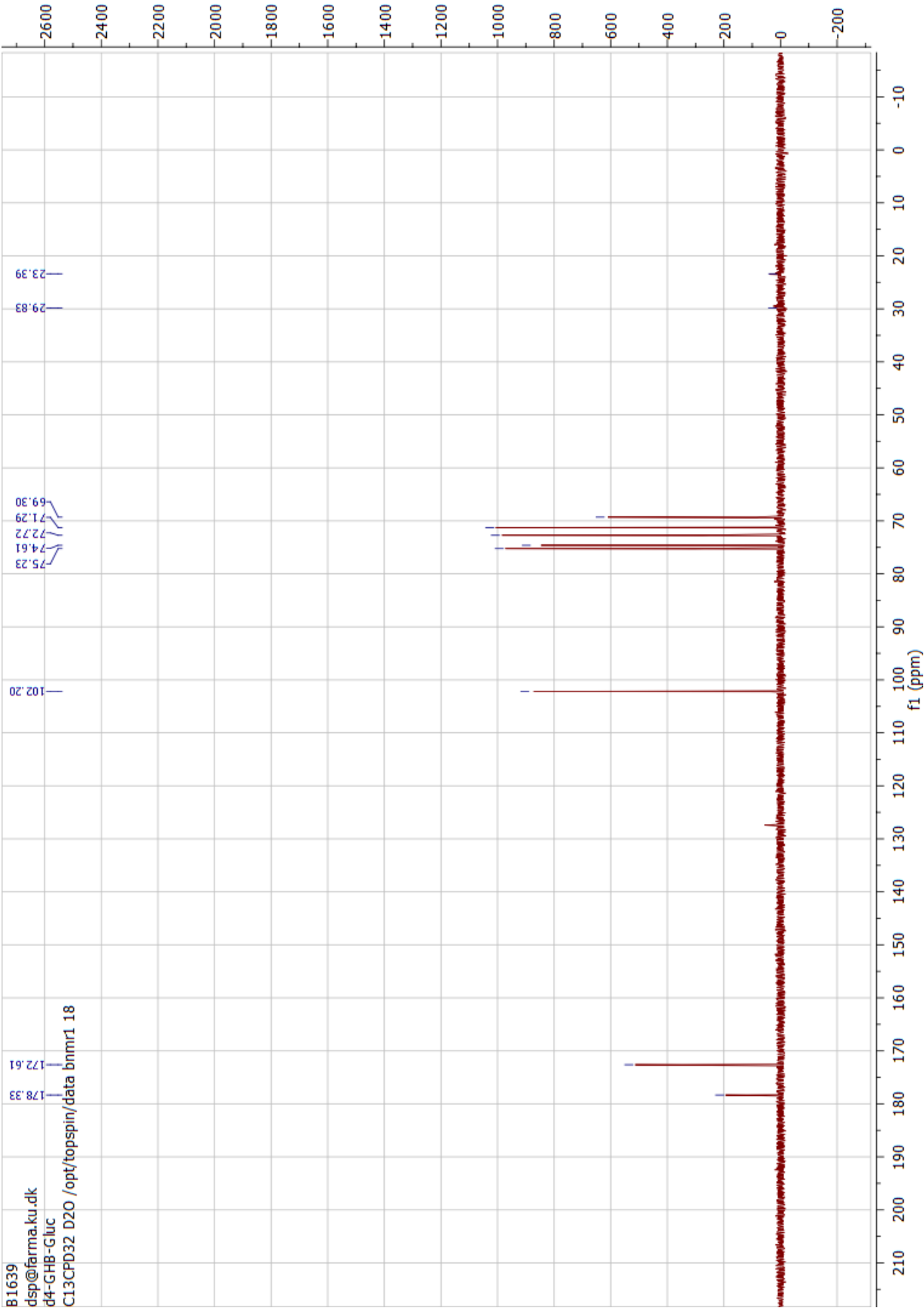
1.2 1D and 2D NMR spectra of glucuronide *d*₄-2

¹H NMR spectra of GHB glucuronide *d*₄-2

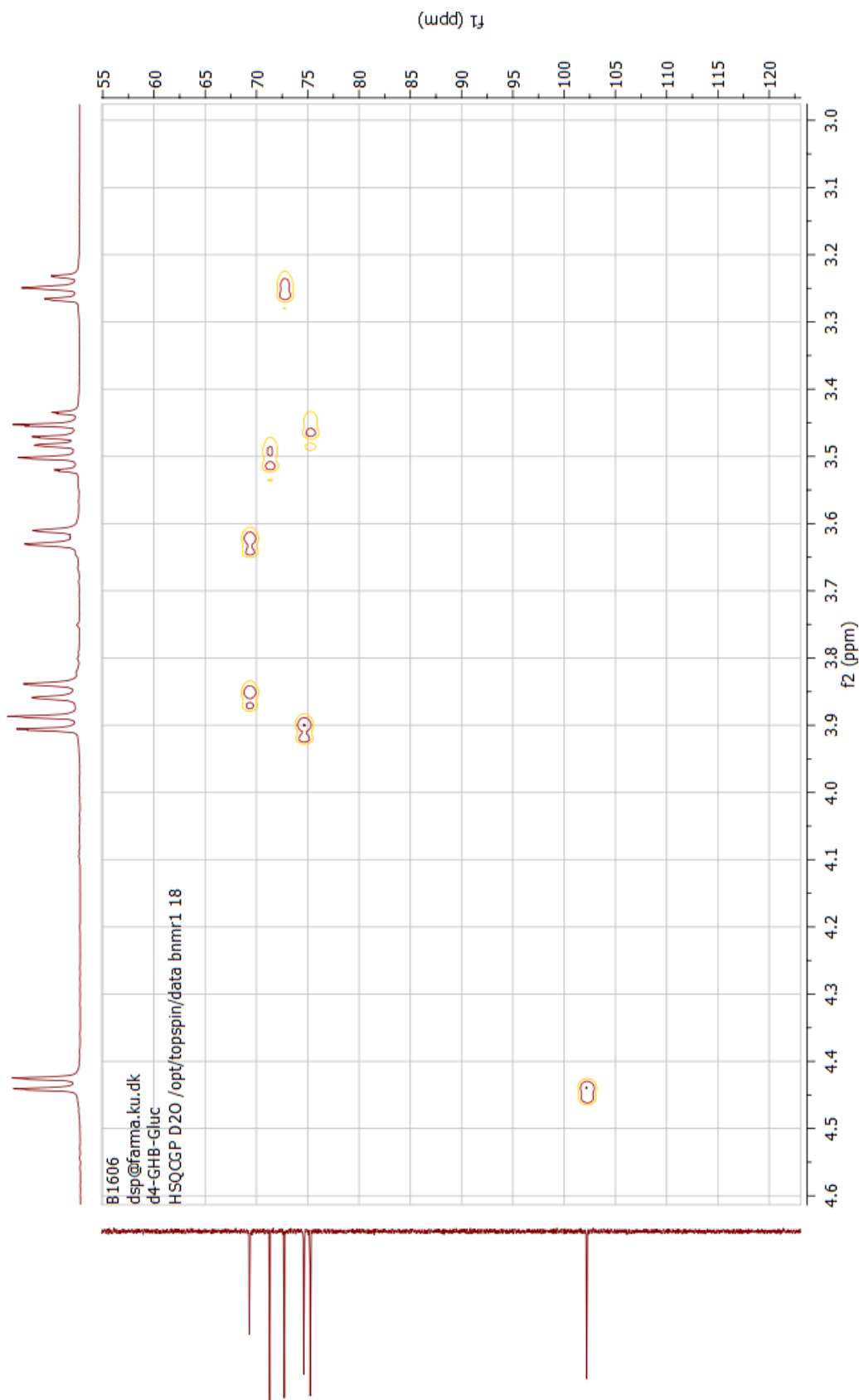


B1606
dsp@farma.ku.dk
d4-GHB-Gluc
COSYGPW D20 /opt/topspin/data/bnmr1 18

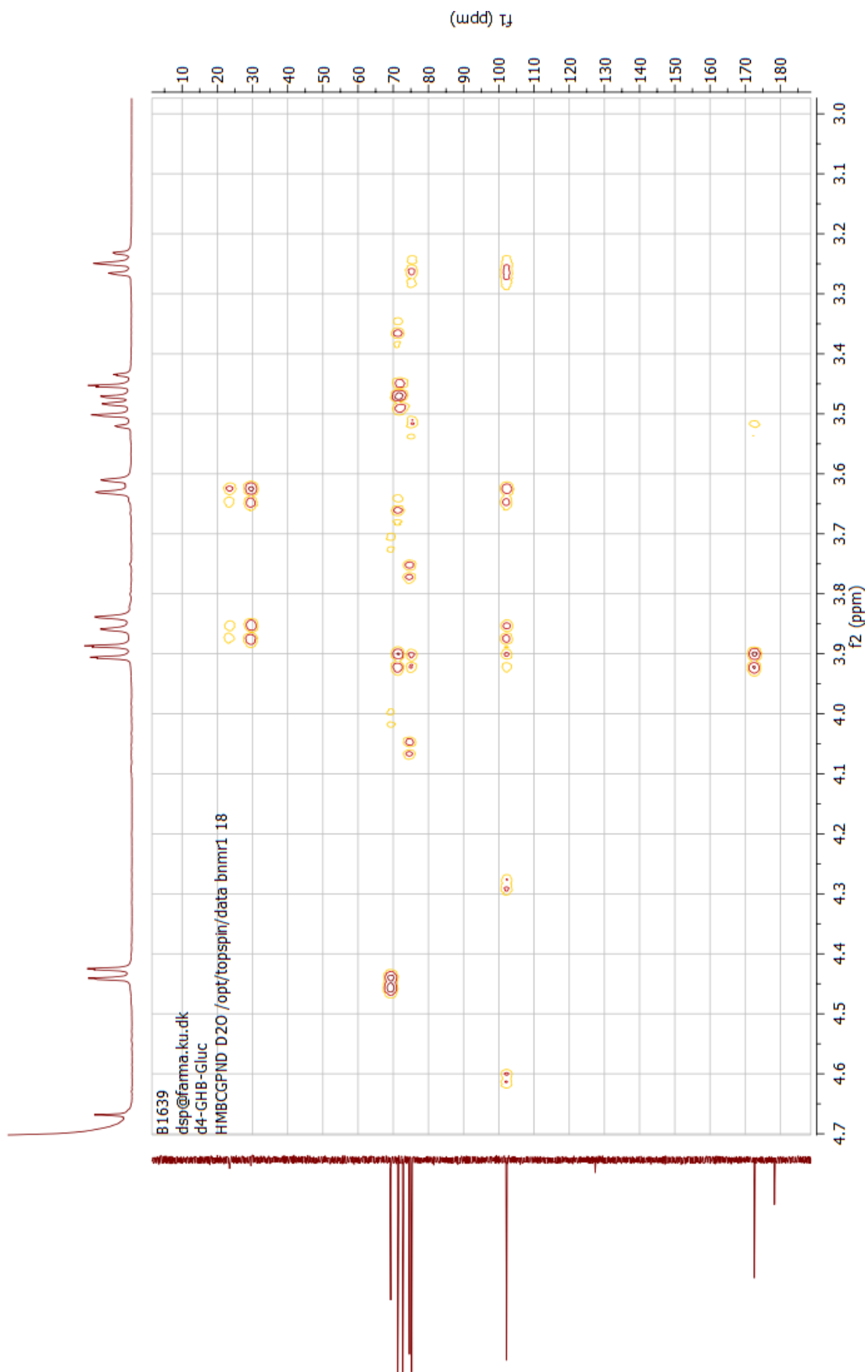
¹³C NMR of GHB glucuronide *d*₄-2



HSQC NMR of GHB glucuronide *d*₄-2



HMBC NMR of GHB glucuronide d_4 -2



2. NMR stability study for GHB-GLUC (2)

2.1 General NMR experimental

All NMR spectra were recorded on a Bruker 500 MHz Avance III NMR spectrometer. A Watergate-type water-suppression pulse sequence using excitation sculpting with gradients was employed (zgesgp) [1].

Stock solutions:

Buffer A: 10 mM sodium phosphate monobasic (monohydrate) in 90% H₂O/10 D₂O. pH = 4.8.

Buffer B: 10 mM sodium phosphate dibasic (heptahydrate) in 90% H₂O/10 D₂O. pH = 9.0.

Buffers were prepared by dissolving the appropriate salt (**A**: 1.38 g and **B**: 2.68 g) in 10 mL D₂O and adjusting the volume to 100 mL with mQ water.

pH was determined using a Radiometer Copenhagen PHM92 pH meter after calibration using IUPAC calibration solutions pH 4.005 and 10.012 from Radiometer Analytical.

Internal standard: 0.1 M *t*-BuOH in H₂O. 0.74 g *t*-BuOH was dissolved in H₂O to give a final volume of 100 mL.

NMR buffer A with internal standard: Prepared by taking 800 µL internal standard solution and adding Buffer A to give a final volume of 20 mL (final *t*-BuOH concentration 4 µM).

NMR buffer B with internal standard: Prepared by taking 800 µL internal standard solution and adding Buffer B to give a final volume of 20 mL (final *t*-BuOH concentration 4 µM).

Determination of integration accuracy:

Watergate-type water suppression during NMR experiments can affect integration accuracy. Hence the NMR method was validated by measuring known concentrations of GHB and comparing them to an internal standard (*t*-BuOH). Three samples with increasing

concentration of *t*-BuOH relative to GHB were analyzed by ^1H NMR. Integration in ^1H NMR spectra was accurate for all three GHB methylene groups over the examined concentration range (deviation $< \pm 1\%$).

NMR samples for determining integration accuracy:

GHB (3.0 mg) was dissolved in NMR buffer A with internal standard (1.0 mL) to give NMR sample C = 1.

NMR sample C = 1 (0.5 mL) was diluted with NMR buffer with internal standard (0.5 mL) to give NMR sample C = 2.

NMR sample C = 2 (0.5 mL) was diluted with NMR buffer with internal standard (0.5 mL) to give NMR sample C = 4.

NMR Samples

GHB-GLUC (1 mg, 3.6 μmol) was dissolved in NMR buffer A with internal standard (0.6 mL).

GHB-GLUC (1 mg, 3.6 μmol) was dissolved in NMR buffer B with internal standard (0.6 mL).

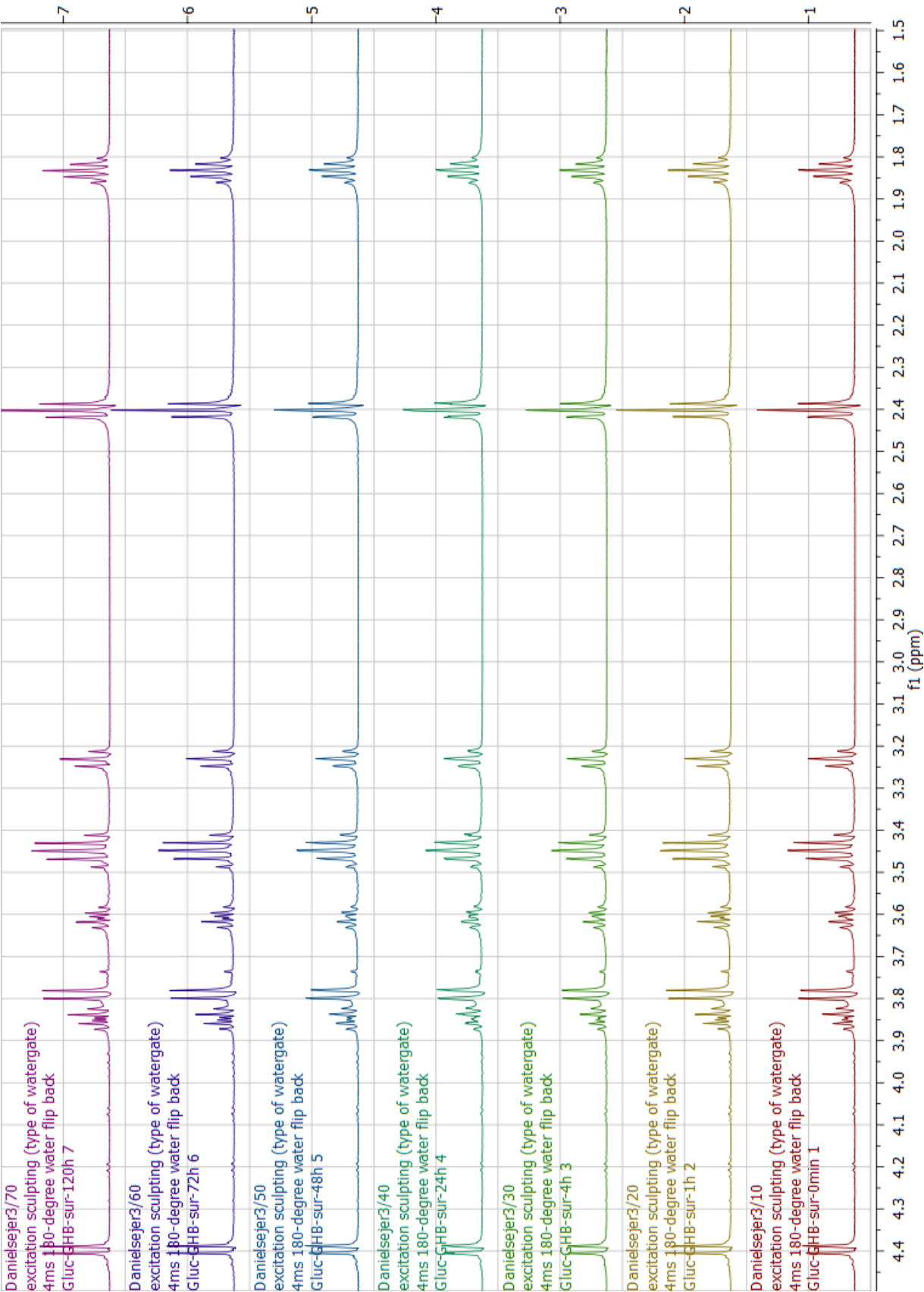
The stability of GHB-GLUC in both buffer systems was evaluated over time at 18, 60 and 90 °C.

Reference spectra (^1H and ^{13}C):

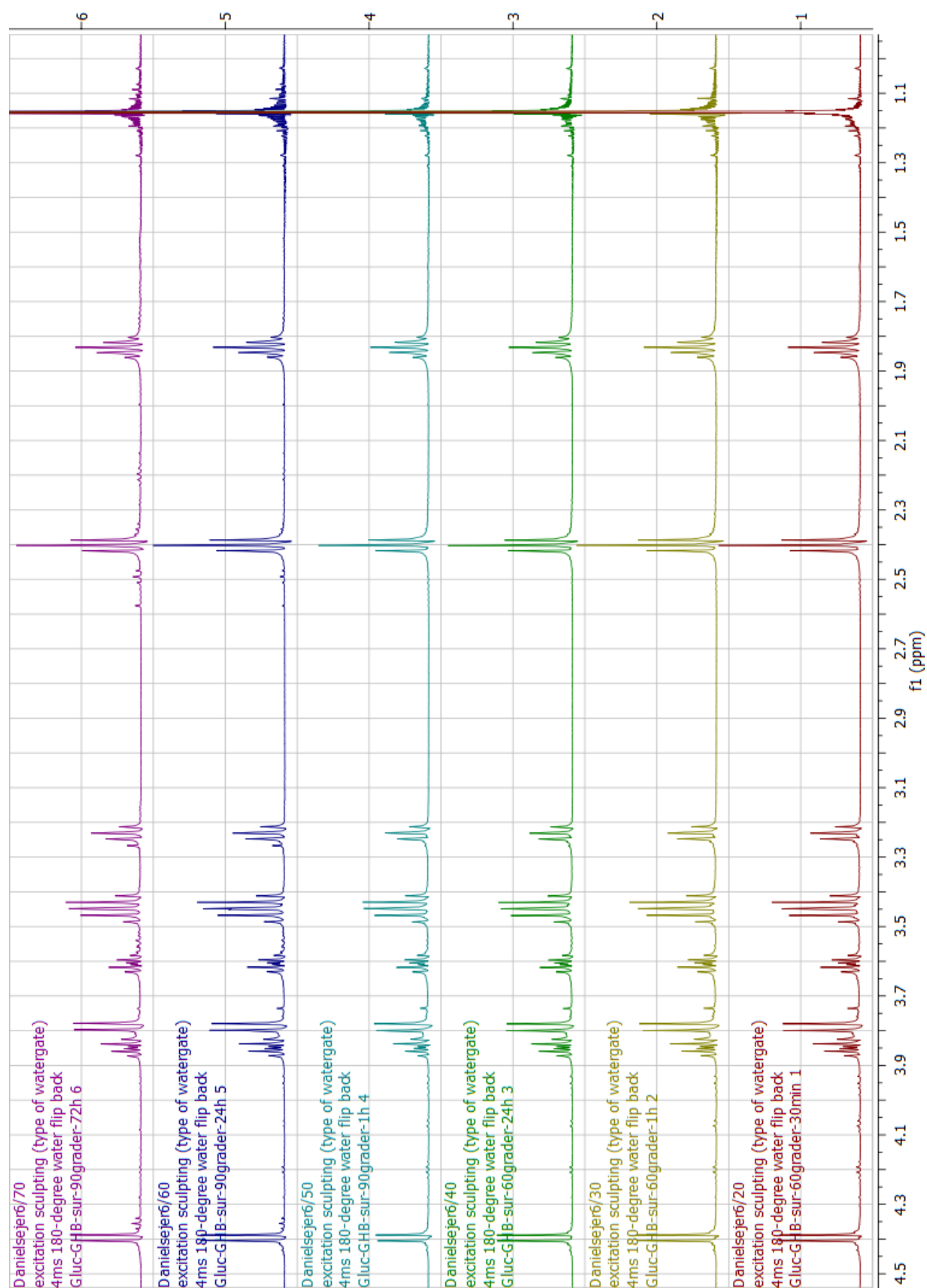
- a) *t*-BuOH (12 mg) in 10% D_2O in H_2O (1.0 mL).
- b) γ -Butyrolactone (2 mg) in NMR buffer A (1.0 mL).
- c) γ -Hydroxybutyric acid in NMR buffer A. Integration experiment at $[C] = 1, 2$ and 4 (vide supra).
- d) β -D-Glucuronic acid (2 mg) in NMR buffer A (1.0 mL).

2.2 NMR stability study ¹H NMR spectra

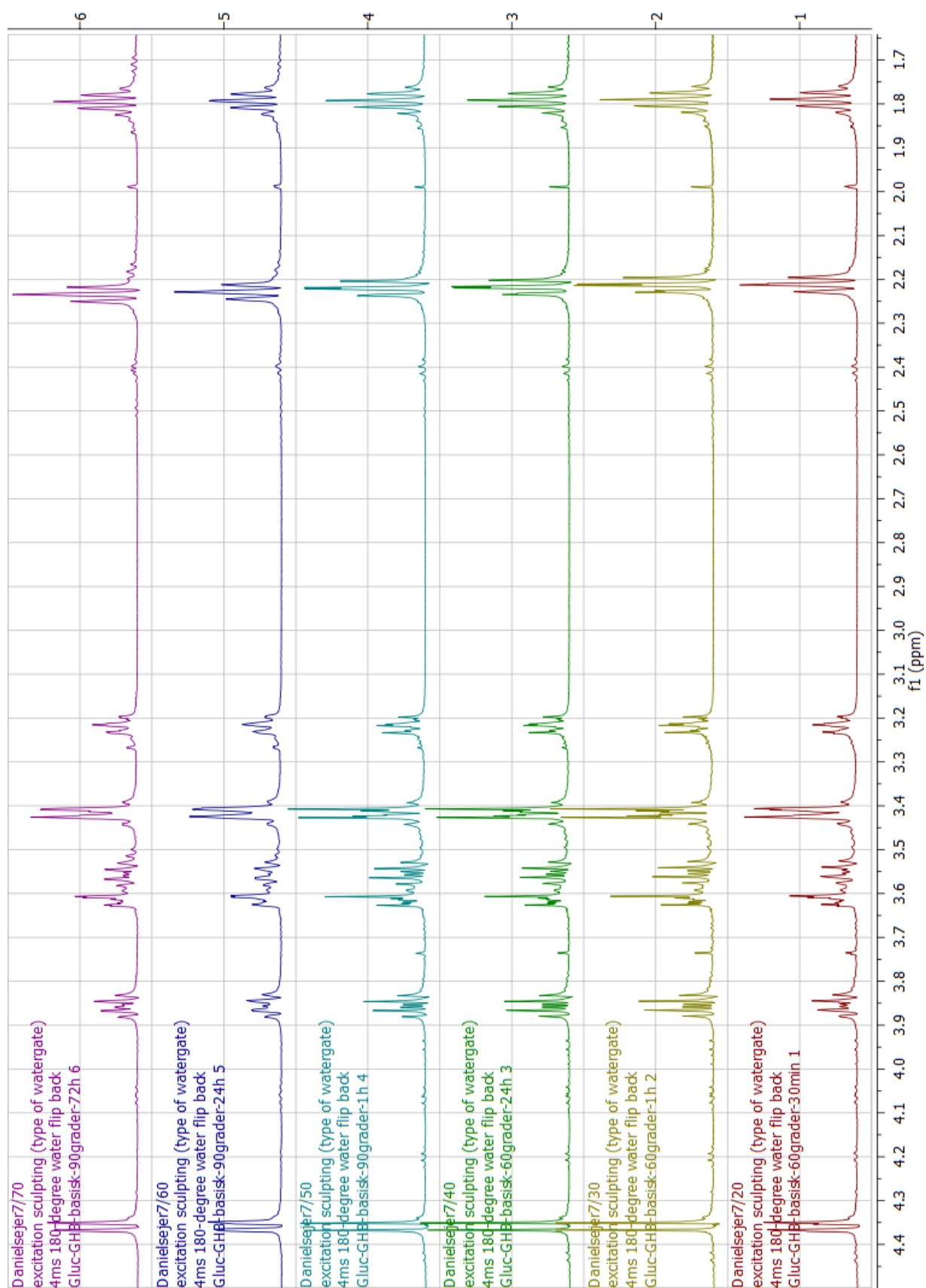
¹H NMR of 2 in NMR buffer A with internal standard at 18 °C (0–120 h)



^1H NMR of 2 in NMR buffer A with internal standard at 60 °C (0.5–24 h) and 90 °C (1–72 h)

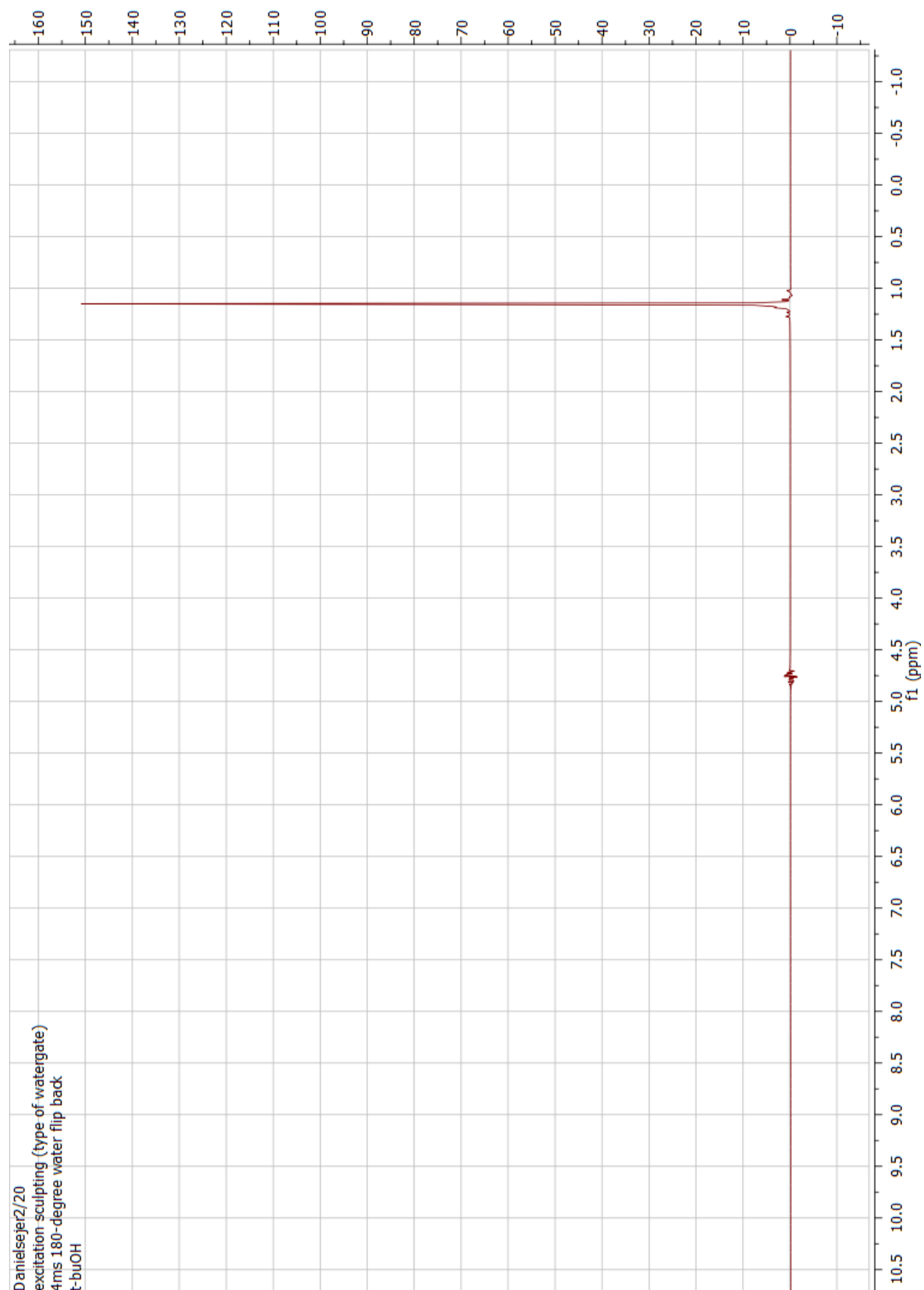


^1H NMR of 2 in NMR buffer B with internal standard at 60 °C (0.5–24 h) and 90 °C (1–72 h)

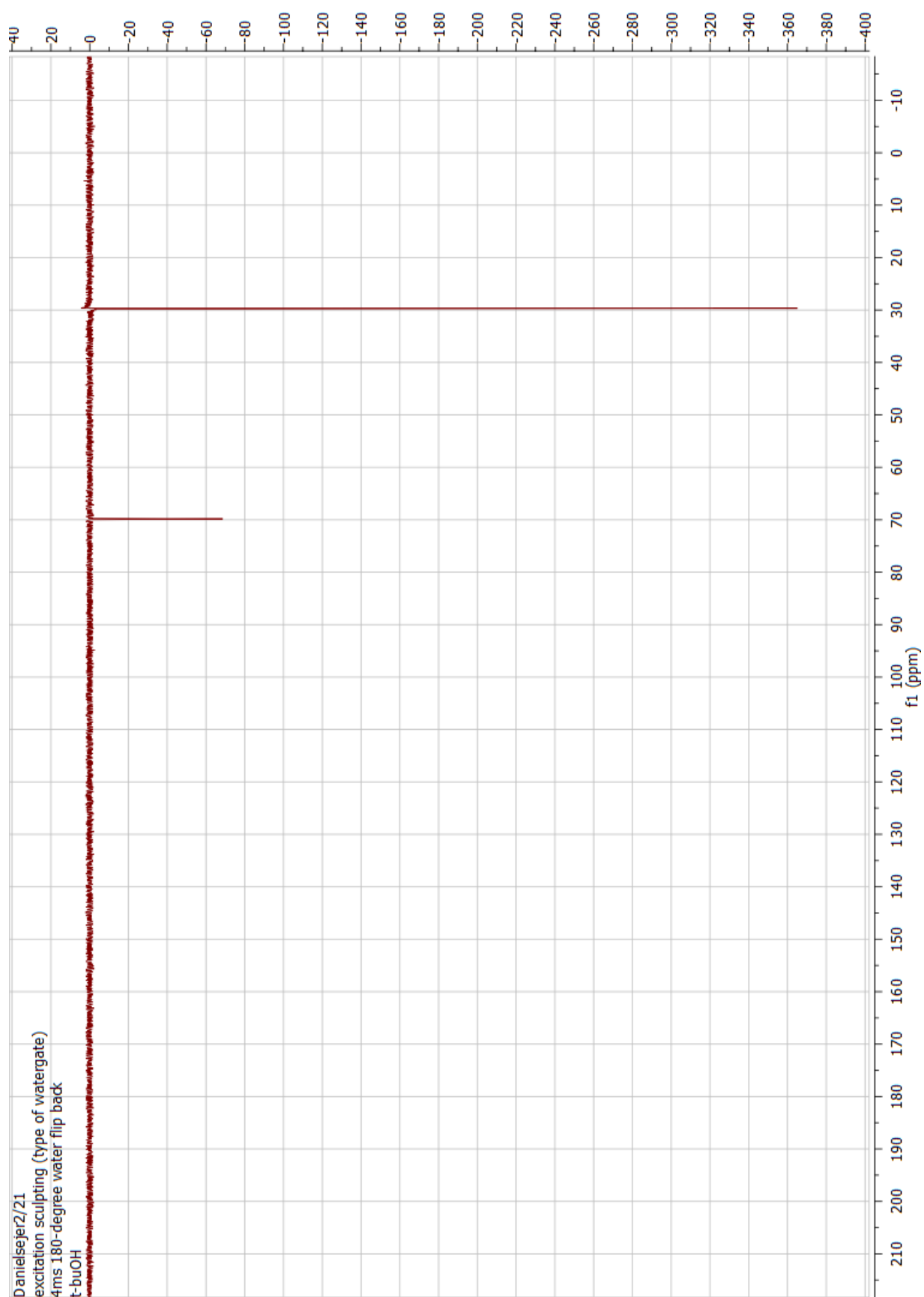


2.3 NMR reference spectra

^1H NMR of *t*-BuOH in 10% D_2O in H_2O



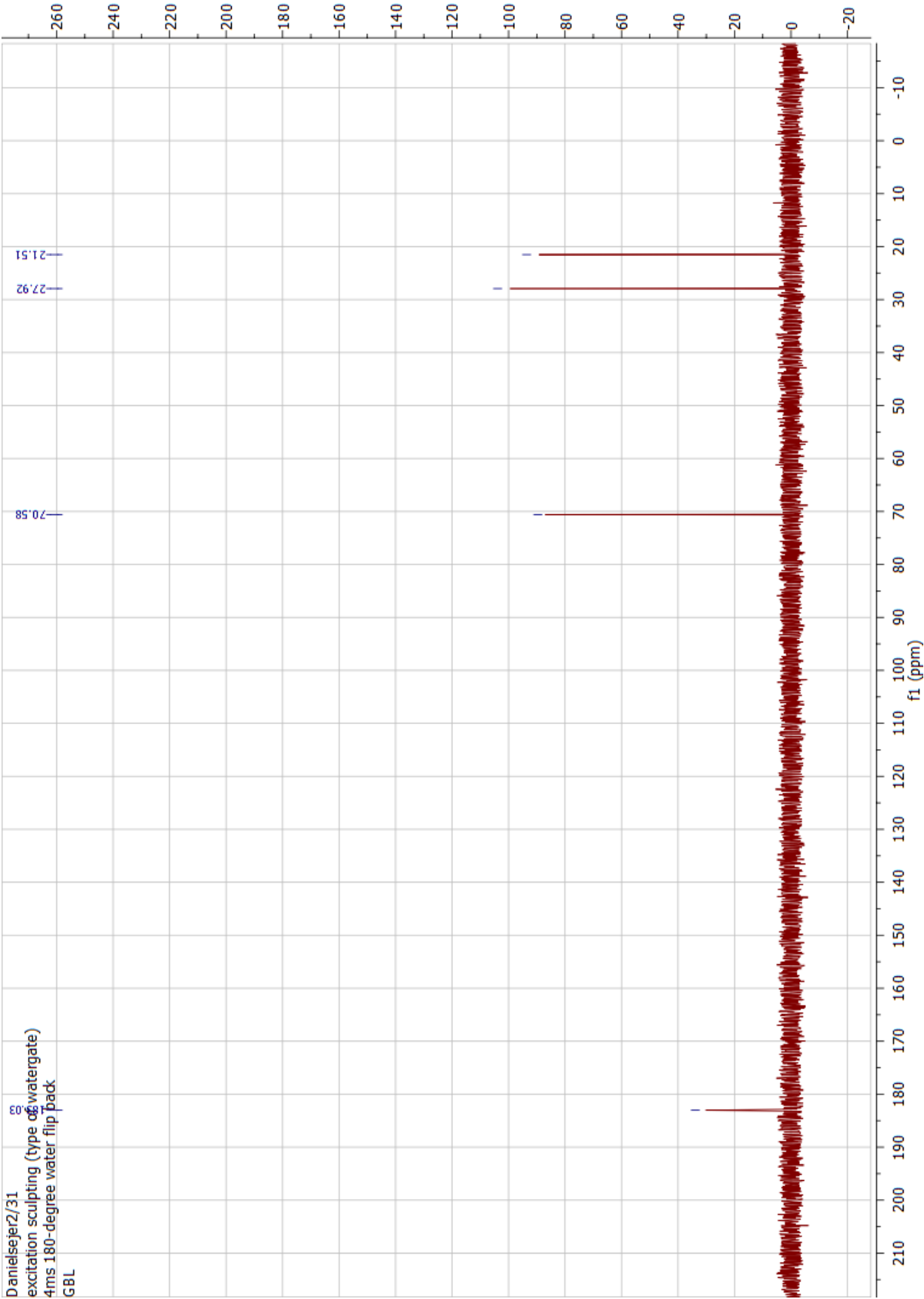
^{13}C APT NMR of *t*-BuOH in 10% D_2O in H_2O



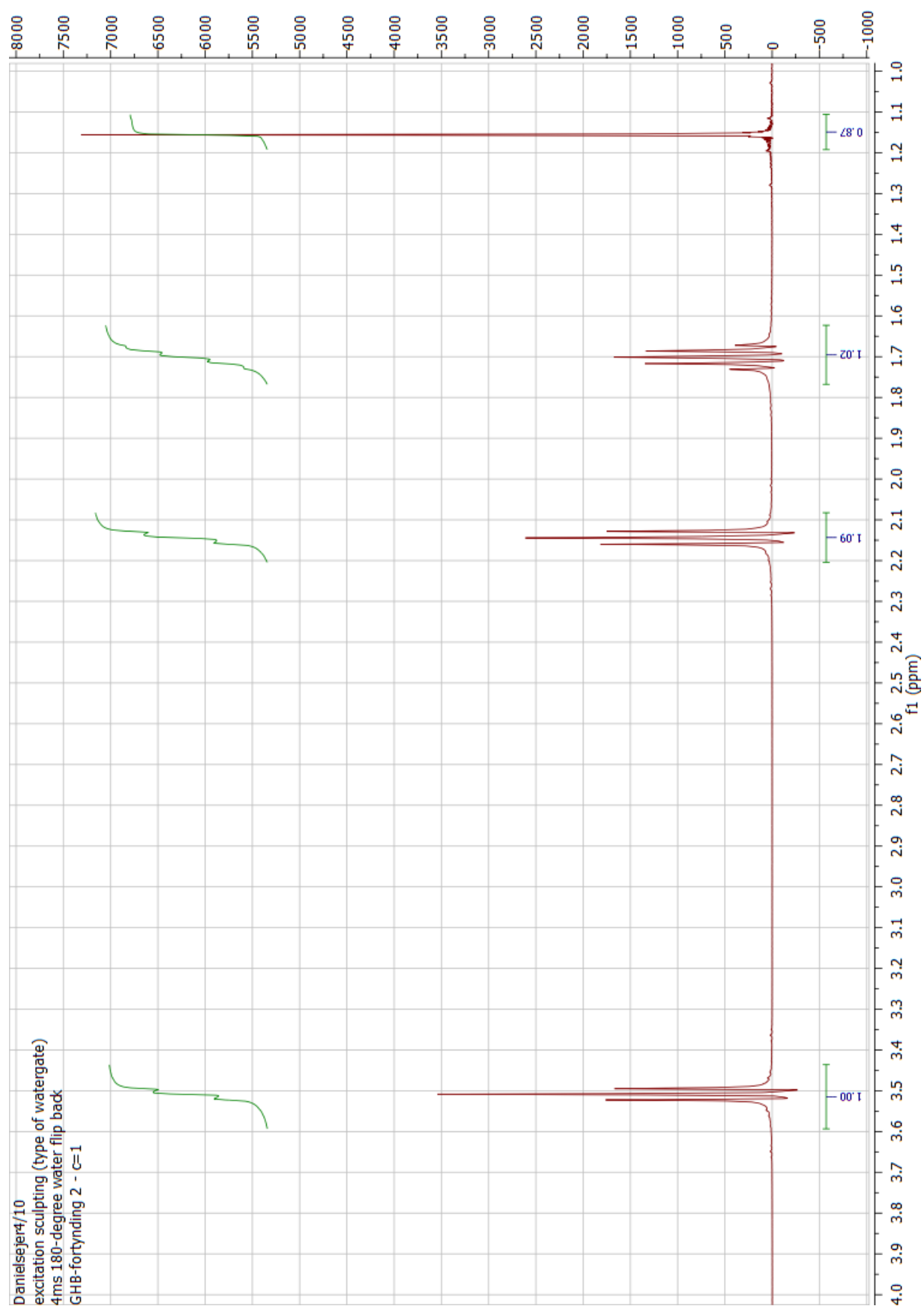
¹H NMR of γ-butyrolactone (GBL) in NMR buffer A



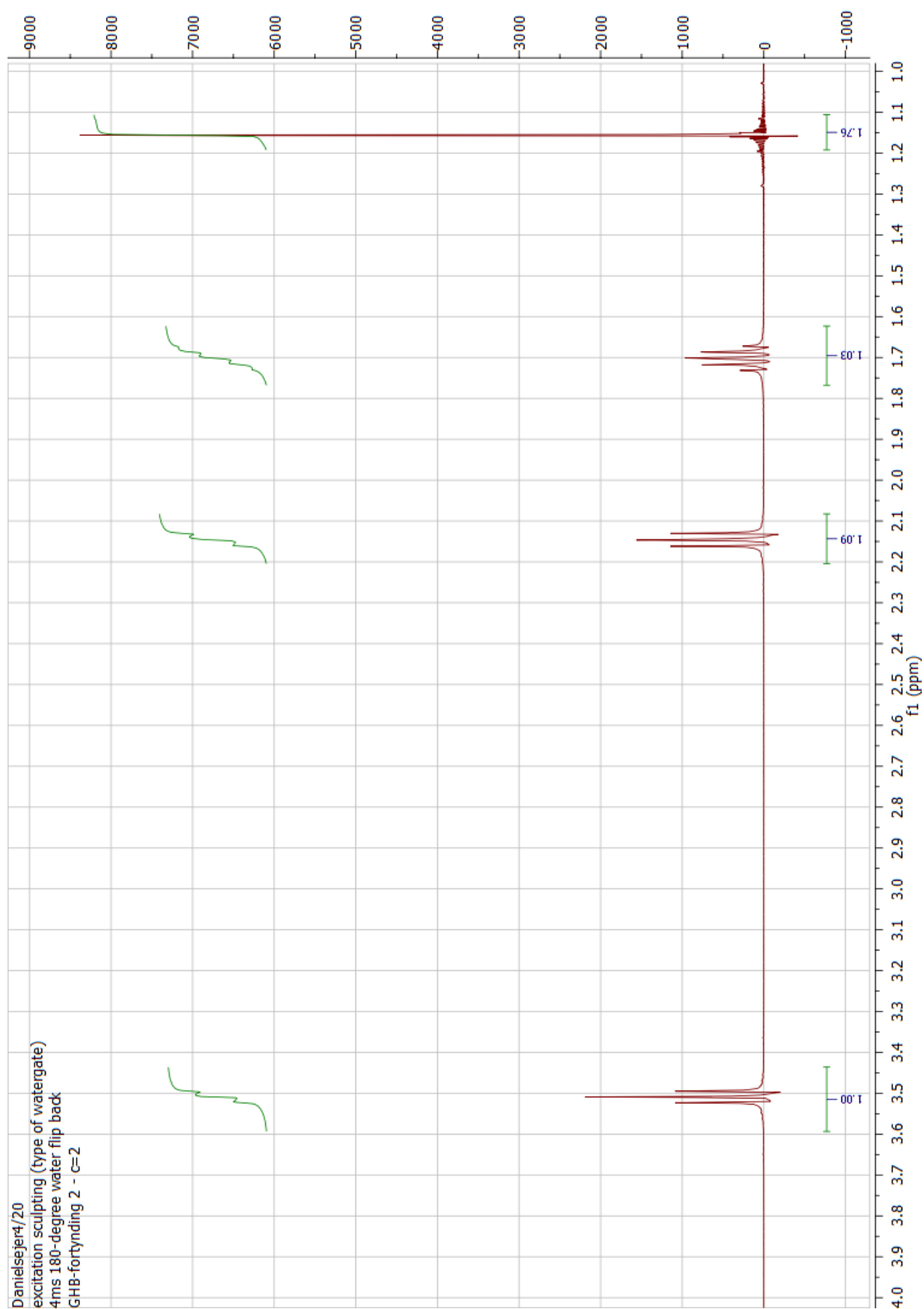
¹³C NMR of γ-butyrolactone (GBL) in NMR buffer A



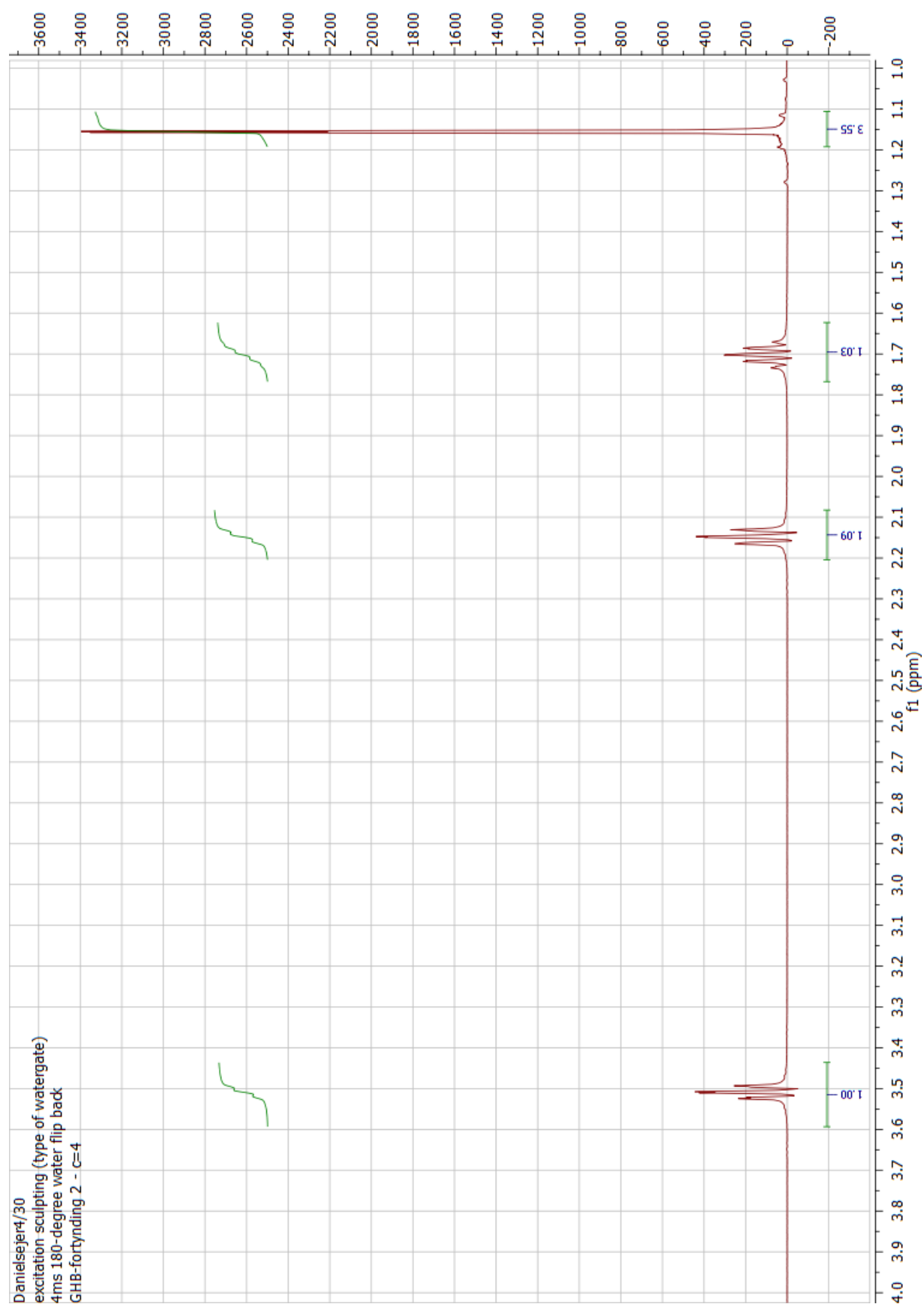
^1H NMR of γ -hydroxybutyric acid (GHB) in NMR buffer A, $[\text{C}] = 1$



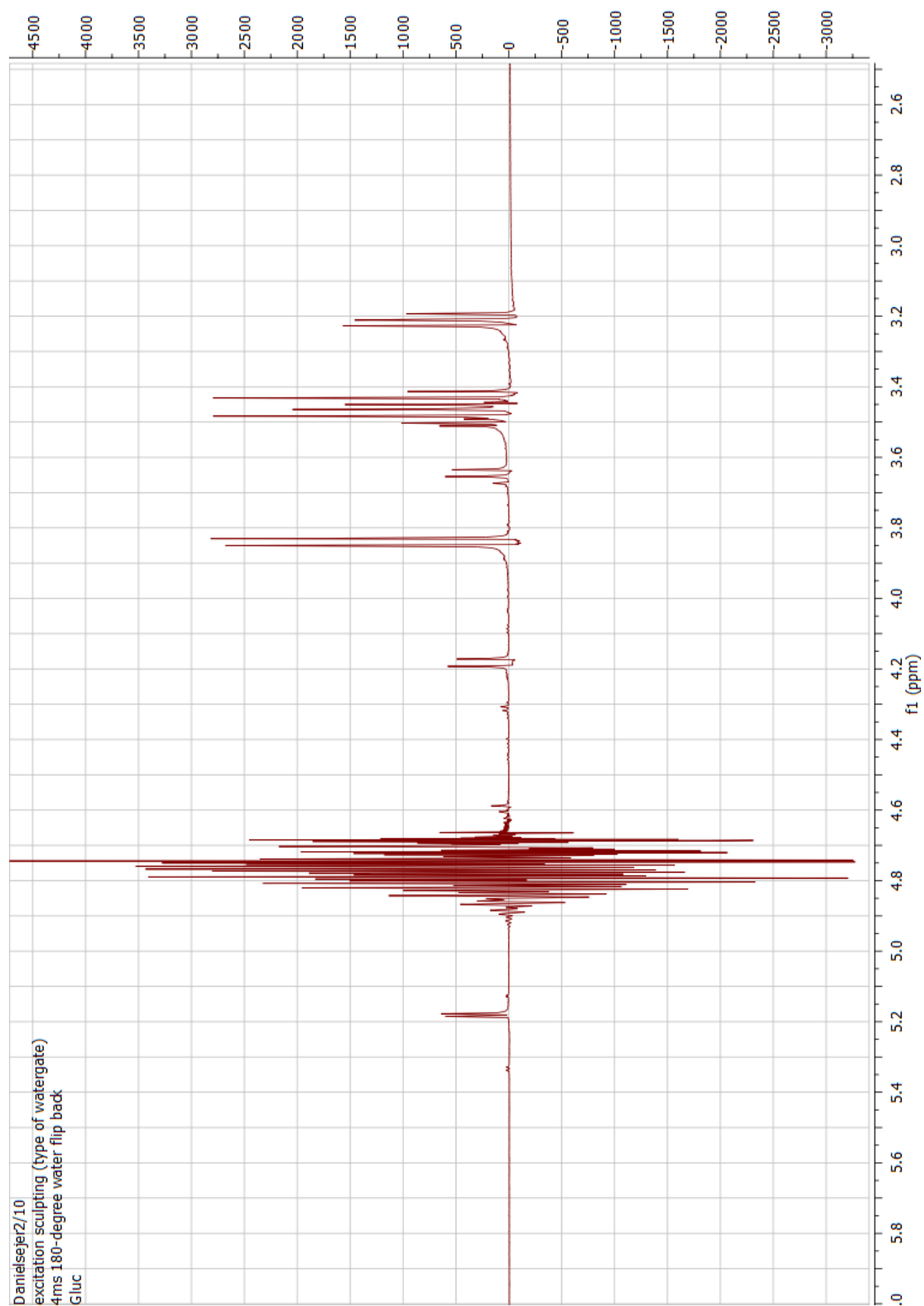
^1H NMR of γ -hydroxybutyric acid (GHB) in NMR buffer A, $[\text{C}] = 2$



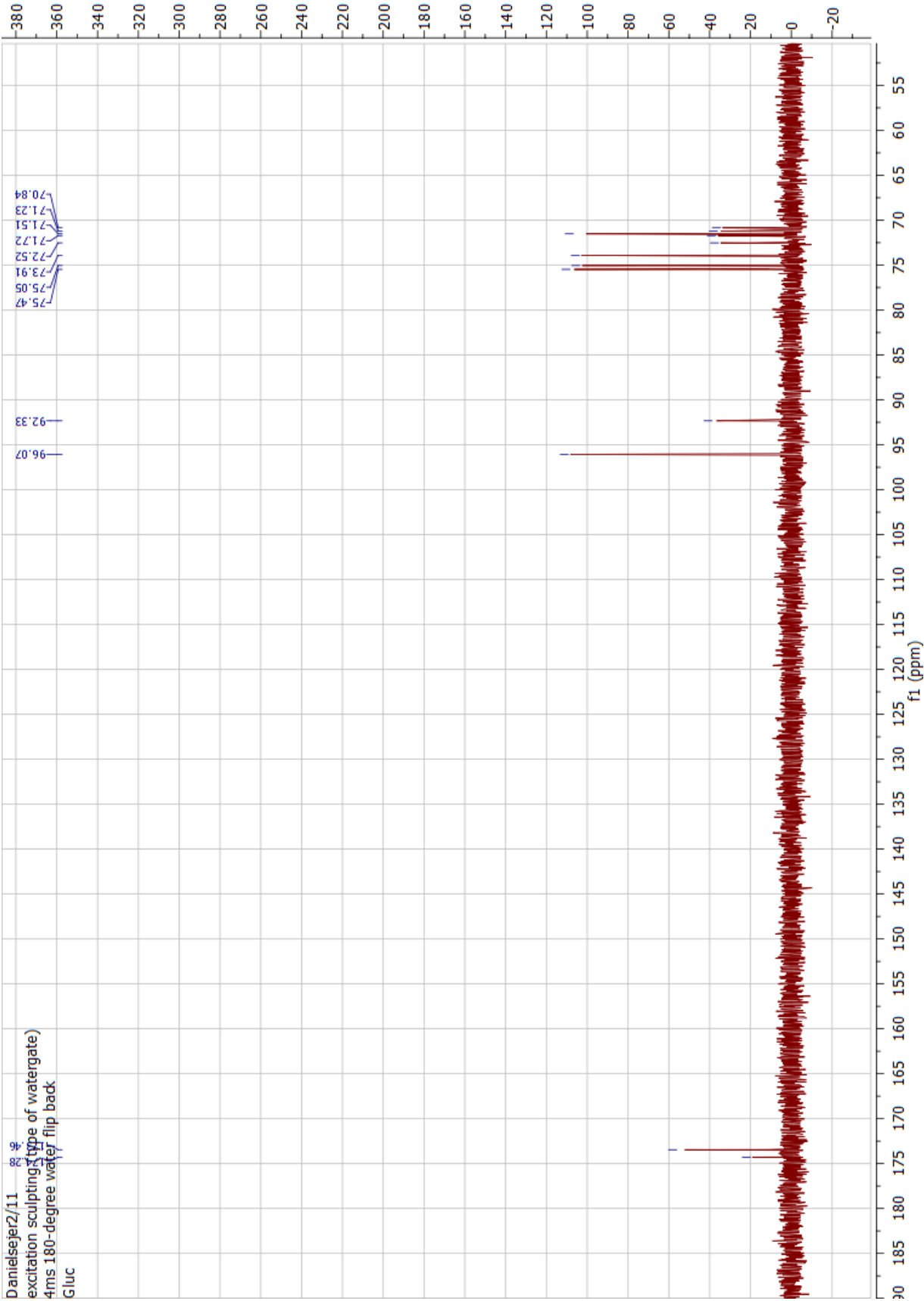
^1H NMR of γ -hydroxybutyric acid (GHB) in NMR buffer A, $[\text{C}] = 4$



¹H NMR of β-D-glucuronic acid in NMR buffer A



¹³C NMR of β-D-glucuronic acid in NMR buffer A



Reference

- [1] Hwang, T. L.; Shaka, A. J. *J. Mag. Reson. Series A* **1995**, 112, 275–279