

Supporting Information

for

**The structural and chemical basis of temporary adhesion
in the sea star *Asterina gibbosa***

Birgit Lengerer^{*1}, Marie Bonneel¹, Mathilde Lefevre², Elise Hennebert², Philippe Leclère³, Emmanuel Gosselin⁴, Peter Ladurner⁵ and Patrick Flammang^{*1}

Address: ¹Biology of Marine Organisms and Biomimetics Unit, Research Institute for Biosciences, University of Mons, 23 Place du Parc, 7000 Mons, Belgium, ²Cell Biology Unit, Research Institute for Biosciences, University of Mons, 23 Place du Parc, 7000 Mons, Belgium, ³Laboratory for Chemistry of Novel Materials, Center for Innovation and Research in Materials and Polymers (CIRMAP), University of Mons, 20 Place du Parc, 7000 Mons, Belgium, ⁴Laboratory of Physics of Surfaces and Interfaces (LPSI), University of Mons, 23 Place du Parc, 7000 Mons, Belgium and ⁵Institute of Zoology and Center of Molecular Bioscience Innsbruck, University of Innsbruck, Technikerstr. 25, A-6020 Innsbruck, Austria

E-mail: Birgit Lengerer^{*} - birgit.lengerer@umons.ac.be, Marie Bonneel - marie.bonneel@umons.ac.be, Mathilde Lefevre - mathilde.lefevre@umons.ac.be, Elise Hennebert - elise.hennebert@umons.ac.be, Philippe Leclère - philippe.leclere@umons.ac.be, Emmanuel Gosselin - emmanuel.gosselin@umons.ac.be, Peter Ladurner - peter.ladurner@uibk.ac.at, Patrick Flammang^{*} - patrick.flammang@umons.ac.be

* Corresponding author

Antibody labelling of tube foot sections

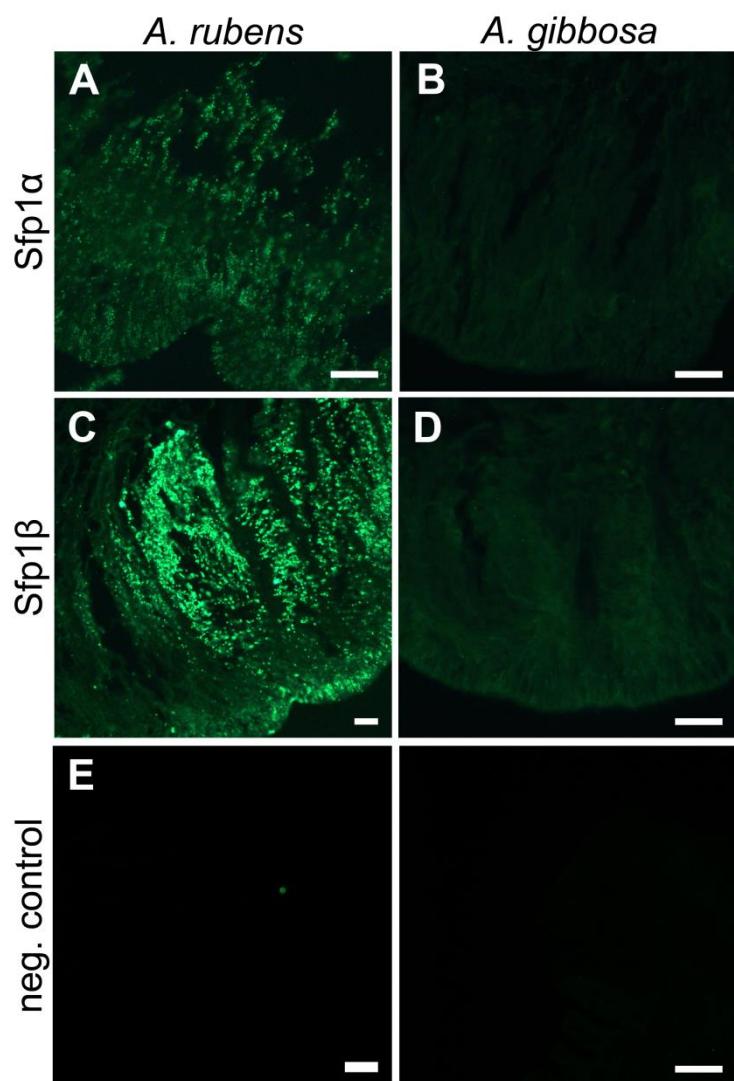


Figure S1: Antibody labelling of tube foot sections from *Asterias rubens* and *Asterina gibbosa*. Tube foot sections of *Asterias rubens* are on the left (A,C,E) and *Asterina gibbosa* to the right (B,D,F). Antibody directed against Sfp1 α (A,B), against Sfp1 β (C,D) and negative controls skipping the primary antibody (E,F). Scale bars: 20 μ m.

Table S1: Overview of lectin binding specificity according to manufacturer Vector laboratories.

Lectin	Acronym	Preferred sugar specificity	Comments
Concanavaline A	Con A	α Man, α Glc	Recognizes α -linked mannose present as part of a core oligosaccharide in many serum and membrane glycoproteins
Jacalin	Jacalin	Gal β 3GalNAc	Binds also in the presence of conjugated sialic acid.
Wheat germ agglutinin	WGA	GlcNAc	Preferable binds to dimer and trimers of GlcNAc. Can also bind terminal GlcNAc and chitobiose.
Datura Stramonium lectin	DSL	(GlcNAc)2-4	Preferable binds to chitobiose or chitotriose over single GlcNAc residues.
Peanut agglutinin	PNA	Gal β 3GalNAc	Does not bind in the presence of conjugated sialic acid.
Soybean agglutinin	SBA	α \sim β GalNAc	Binds to oligosaccharide structures with terminal α - or β -linked N-acetylgalactosamine, and to a lesser extent, galactose residues.
Griffonia (Bandeiraea) simplicifolia lectin I	GSL I	α Gal, α GalNAc	GSL I is a mixture of the five isolectins recognizing α Gal and α GalNAc.
Vicia villosa agglutinin	VVA	GalNAc	Recognizes preferentially α - or β -linked terminal GalNAc, especially a single GalNAc residue linked to serine or threonine in a polypeptide.
Succinylated wheat germ agglutinin	sWGA	GlcNAc	This derivative does not bind to sialic acid residues, unlike the native form.
Lens culinaris agglutinin	LCA	α Man, α Glc	Recognizes sequences containing α Man residues but recognizes additional sugars as part of the receptor structure, giving it a narrower specificity than Con A.
Pisum sativum agglutinin	PSA	α Man, α Glc	Is nearly identical in structure and carbohydrate specificity to LCA.
Ricinus communis agglutinin	RCA I	Gal, GalNAc	Binds to Gal or GalNAc residues of membrane glycoconjugates.
Griffonia (Bandeiraea) simplicifolia lectin II	GSL II	α or β GlcNAc	Is unique in its ability to recognize exclusively α - or β -linked GlcNAc residues on the nonreducing terminal of oligosaccharides.
Lycopersicon esculentum (tomato) lectin	LEL	(GlcNAc)2-4	
Elderberry bark Lectin	EBL	Neu5Aca6Gal/GaINAc	Binds preferentially to sialic acid attached to terminal Gal in α -2,6 and to a lesser degree, α -2,3 linkage. This lectin does not appear to bind sialic acid linked to GalNAc.
Ulex europeus agglutinin 1	UEA 1	L-Fuc	Binds to many glycoproteins and glycolipids containing α -linked fucose residues.
Maackia amurensis lectin II	MAL II	Neu5Aca3Gal β 4GalNAc	Binds only particular carbohydrate structures that contain sialic acid. Unlike SNA which seems to prefer structures with (α -2,6) linked sialic acid, MAL II appears to bind sialic acid in an (α -2,3) linkage.
Dolichos biflorus agglutinin	DBA	α GalNAc	
Sambucus nigra	SNA	Neu5Aca6Gal/Ga	Binds preferentially to sialic acid attached to terminal

agglutinin		INAc	Gal in α -2,6 and to a lesser degree, α -2,3 linkage. This lectin does not appear to bind sialic acid linked to GalNAc.
<i>Phaseolus vulgaris</i> erythro agglutinin	PHA-E	Gal β 4GlcNAc β 2 Man α 6(GlcNAc β 4)(GlcNAc β 4Man α 3)Man β 4	
<i>Phaseolus vulgaris</i> leuco agglutinin	PHA-L	Gal β 4GlcNAc β 6(GlcNAc β 2Man α 3)Man α 3	
<i>Sophora Japonica</i> agglutinin	SJA	β GalNAc	
<i>Erythrina cristagalli</i> lectin	ECL	Gal β 4GlcNAc	Sialic acid substitution appears to prevent the lectin from binding.
<i>Solanum tuberosum</i> (potato) lectin	STL	(GlcNAc)2-4	Binds oligomers of GlcNAc and some bacterial cell wall oligosaccharides containing GlcNAc and <i>N</i> -acetylmuramic acid.

Sugar Abbreviations:

Fuc	L-Fucose
Gal	D-Galactose
GalNAc	<i>N</i> -Acetylgalactosamine
Glc	D-Glucose
GlcNAc	<i>N</i> -Acetylglucosamine
Man	Mannose
Neu5Ac	<i>N</i> -Acetylneurameric acid (sialic acid)

Lectin labelling of tube foot sections from *Asterina gibbosa*.

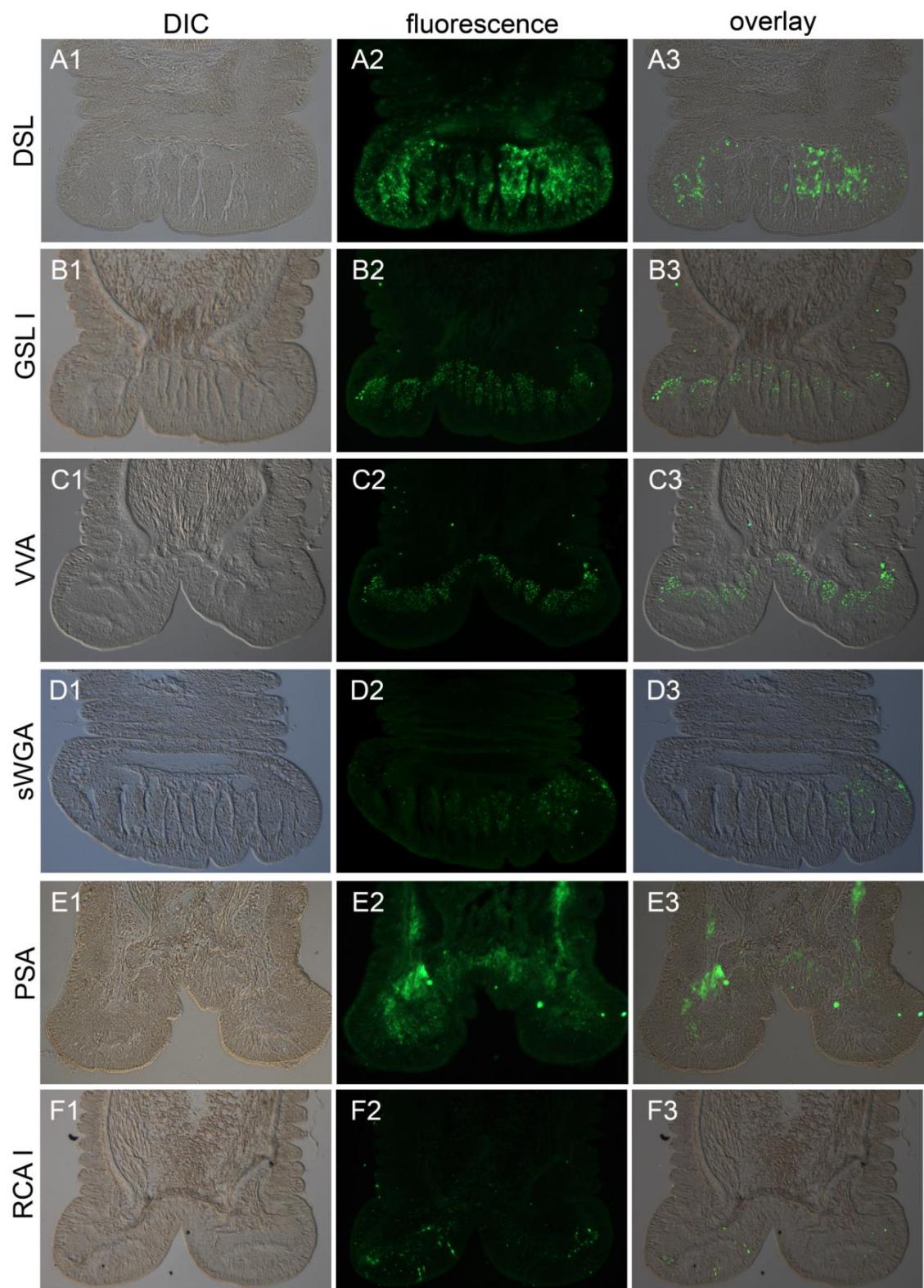


Figure S2: Lectin labelling of tube foot sections from *Asterina gibbosa*; with (A1-3) DSL, (B1-3) GSL I, (C1-3) VVA, (D1-3) sWGA, (E1-3) PSA, and (F1-3) RCA.

Lectin labelling of tube foot sections from *Asterina gibbosa*.

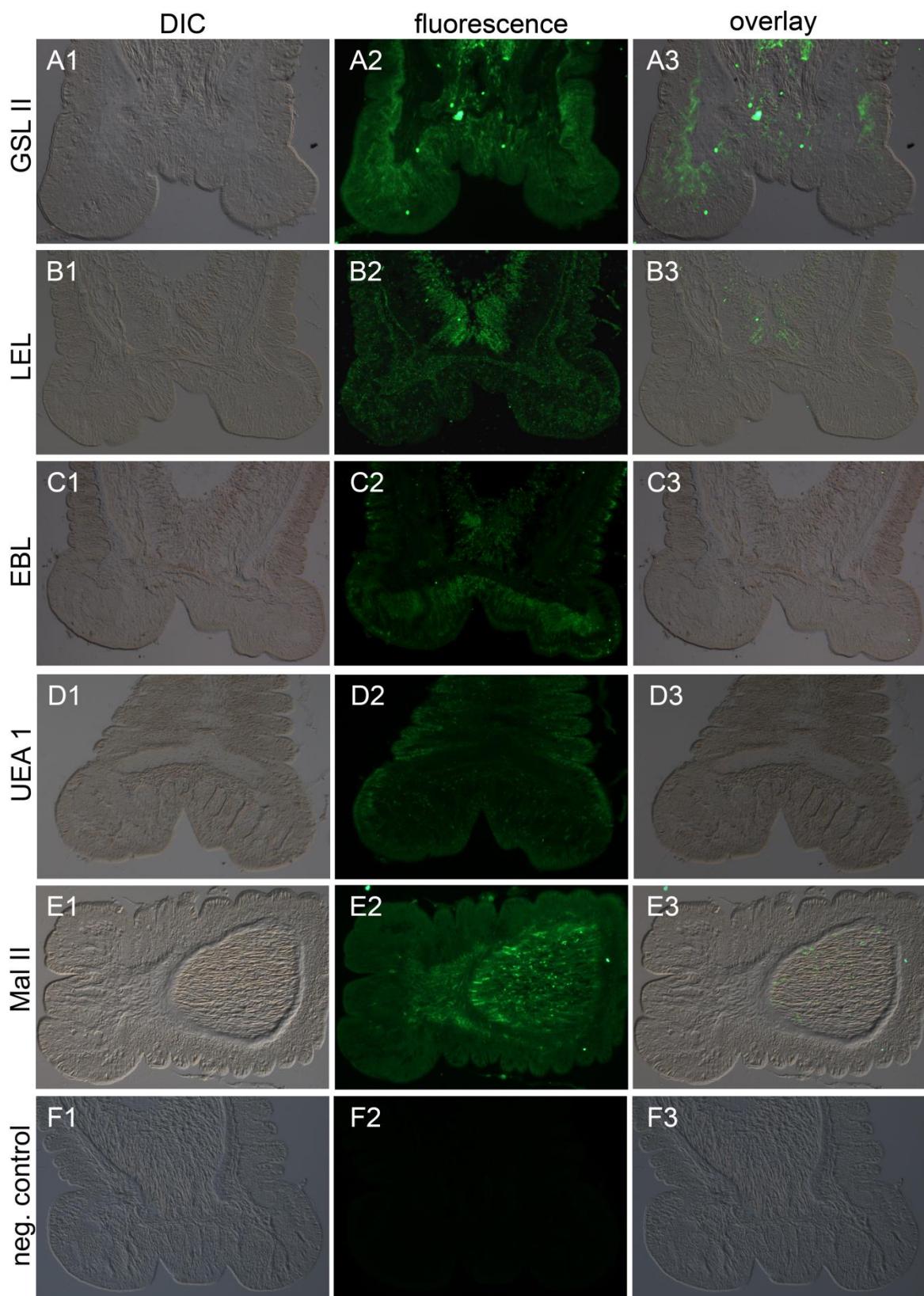


Figure S3: Lectin labelling of tube foot sections from *Asterina gibbosa*, with (A1-3) GSL II, (B1-3)

LEL, (C1-3) EBL, (D1-3) UEA 1, (E1-3) Mal II and (F1-F3) negative control.